A LYSIMETER STUDY OF DOMESTIC WASTE WATER RENOVATION BY FOREST SOIL FILTRATION

BY

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A THESIS SUBMITTED IN PARTIAL FULFILMENT OF

THE REQUIREMENTS FOR THE DEGREE OF

MASTER OF SCIENCE

in the Department
of
SOIL SCIENCE

We accept this thesis as conforming to the required standard

THE UNIVERSITY OF BRITISH COLUMBIA
September, 1973

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ABSTRACT

Laboratory lysimeters were used to investigate the behaviour, over time, of a humid west coast forest soil under intermittent primary municipal waste water irrigation. soil packed to a depth of 69 cm and to a uniform density of about 0.9 gm per cm³ was covered with a forest floor 9 cm thick. Sintered glass bead tensiometers were used to gauge the water potential distributions in the soil lysimeters. Irrigation and drainage systems were designed to maintain constant rates of waste water application and facilitate measurement of drainage rates. Two groups of soil lysimeters each with triplicate samples, were loaded with waste water at the rates of 0.23 cm per day (37 cm³ per day) and 0.47 cm per day (75 cm³ per day) for a period of 9 months. The soil lysimeters were incubated at a temperature of about 15.5 degrees Centigrade. amounts of nitrogen added to both groups of soil lysimeters were 223.7 gm and 436.9 gm or equivalent to 1.4 % and 2.7 % of the total nitrogen of the original soil, respectively. Renovations of wastewater in terms of nitrogen were 75 % and 43 % with respect to the two groups of soil lysimeters. Renovations in terms of phosphorus were more than 99 % in both groups of soil lysimeters. Retention of nutrients by the soil was increased with time under favourable aerobic canditions. of nutrients by vegetation in the field would minimize leaching losses. Results from this experiment indicated no significant changes in the physical and chemical behaviour of the soils. Proper design of the waste water irrigation system in terms of

loading would maximize the efficiency of renovation without deteriorating the behaviour of the soils.

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ACKNOWLEDGMENTS

The writer is indebted to Dr. Jan de Vries for his support, suggestions and criticisms during the course of the research and preparation of this manuscript. The writer is also deeply indebted to Dr. T. M. Ballard for his assistance and advice during the progress of this study. Special thanks are also extended to Dr. C. A. Rowles and Dr. T. A. Black for their kind assistance.

The financial support from the Department of the Environment, Canada is also gratefully acknowledged.

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INTRODUCTION

The practice of releasing wastewater from domestic, industrial and agricultural sources to receiving waters has contributed significantly to water quality problems. These problems have drawn not only the attention of the public, but also that of the government which considers water pollution as one of the top environmental quality problems. The various methods that are used to cope with this problem involve the reduction of chemical and biological materials, contained in the waste, to environmentally tolerable levels. Chemical, biological and physical means of treatment, separately or in combination, are generally used to remove nutrients, dissolved minerals and organic matter from wastewater.

Three processes of conventional treatments of wastewater with different degrees of purification are currently in practice. They are primary, secondary and tertiary treatments. Primary treatment includes such methods as screening, skimming, sedimentation and lagooning to remove part of the coarse, floatable and suspended solids from the wastewater. Secondary treatment is employed to further remove most of the remaining solids from the primary treated wastewater. Several methods in use involve filtration, activated sludge and aerated stabilization basins. Nutrient removal in these processes is limited. The tertiary treatment is, therefore, an advanced step of the secondary treatment and is designed to remove nutrients and dissolved minerals from the treated water. The methods usually employed are photosynthetic stabilization, chemical precipita-

tion, ion exchange, distillation, electrodialysis, freezing, reverse osmosis and ultrafiltration.

Wastewater filtration with field soils represents a combination of chemical, biological and physical methods for the treatment of wastewater. It is therefore considered as a process of tertiary treatment due to the effectiveness in removal of nutrients and dissolved minerals from the wastewater. Waste water for land irrigation should receive primary or secondary treatment and should be free of any toxic chemicals before being applied to the land. Large amounts of heavy metals such as Cu, Zn, Pb, Ni, Cd and Cr are hazardous to biotic systems and should be removed from the wastewater, prior to application to land, by some means of chemical, biological or physical treatment.

Using soil for wastewater filtration has been a common practice for centuries. It has been used by farmers to maintain and increase soil fertility in many places of the world. Scoble (1905) reported a successful land treatment sewage system in Great Britain. Wastewater from domestic sources combined with trade refuse was treated by screening and filtration through about 6 feet of light loamy soil overlying a porous sandy subsoil at an average application rate of 23,300 gal per acre per year (2.65 cm per day). The drainage water from the cropped soil attained over 90 % purification in terms of chemical, physical and biological qualities.

The use of soil for disposal of wastewater from various industries such as canneries, pulp mills, dairies etc. in the United States since 1930 was reported by Schraufnage (1962).

land through a ridge-and-furrow irrigation system at a rate of 49,000 gal per day per acre (5.57 cm per day) or 238 lb BOD per day per acre (266.6 kg BOD per day per ha) in 1934 at Hampton, U. S. A. No odor was noted. He also reported that municipal waste was disposed of on a deep silt loam underlain by sand at an average rate of 37,000 gal per day per acre (4.21 cm per day) with a BOD of about 8 lb per acre (9.0 kg per ha) in 1959 at Wisconsin, U. S. A. No odor and overflow were reported.

Scott (1962) reported a successful use of cheese whey as a fertilizer and soil conditioner in tests carried out in Wisconsin in 1959. Cheese whey was applied to the sandy soils at a rate of 5,000 - 70,000 lb per day per acre (5610 - 78,540 kg per ha) over a 30-day period. Return yield of oat crop was reported to be 32 bu per acre, despite some vegetation losses on heavy wastewater loaded areas.

Spray irrigation of spent sulfite liquor on land at a maximum rate of 320,000 gal per day (36.4 cm per day) was also reported by Scott. Well tests indicated no trace of liquor in the ground water. The operation cost of the disposal system was estimated to be \$1.39 per ton as compared to \$4.17 per ton of pulp produced.

The economics of land disposal of sludge for soil improvement were statistically evaluated by Thomas and Bendixon (1969). They reported that disposal of sludge on land could reduce the the costs by about 29 %. They indicated the cost of making topsoil with sludge was \$1,600 per acre (\$4,000 per ha), while the comparable cost of improvement with natural topsoil would

have been \$4,500 per acre (\$11,000 per hectare).

Robeck and his colleages (1964), on the basis of tests using 50 lysimeters, suggested that soil system in order to be suitable for wastewater treatment must have a low enough permeability and some adsorptive capacity to allow the suspended and dissolved organic matter to be retained. They pointed out that a soil which has 0.5 - 1.0 % organic matter and effective aggregate size of about 0.3 - 0.1 mm and an application rate from 4 - 10 cm per day can help reduce 90 - 95 % of ABS (Alkyl benzene sulfonate) and COD (Chemical oxygen demand) and also help prevent groundwater contamination from wastewater irrigation. A number of other authors have studied the efficiency of filtration systems in terms of design and operation procedures (Thomas, Warren, and Thomas, 1966; Parizek, 1967; Law, Thomas and Myers, 1970; Laak, 1970; Robeck, Bendixen, Schwartz and Woodward, 1964; de Vries, 1972).

Research on the application of wastewater to forested soil by spray irrigation was carried out at the Pennsylvania State University and New Jersey, U.S.A. (Kardos, 1966; Pennypacker, Sopper and Kardos, 1967; Sopper, 1971; Mather, 1953). In Pennsylvania, hardwood and red pine forest soils of silt loam to silty clay loam texture were subjected to an intermittent application rate of 0.64 cm per hour for a total of 2.5 to 5.0 cm per week. The research was carried out from April to November in 1968 after six years of operation. Renovation of MBAS (detergent residue) in the hardwood plot under a loading of 2.5 cm per week was as high as 70 - 80 % in the upper 120 cm of soil, as compared to 71 - 86 %

with the red pine. Phosphorus removal ranged from 98 - 99 % at the 60 cm soil depth in the hardwood plot and 93 - 97 % in the red pine plot. Nitrate nitrogen removal decreased from 68 - 82 % in the first year to 27 - 70 % six year later. Removals of organic nitrogen were 99 % to 90 % with respect to hardwood and red pine plots. Different degrees of successful removal of other dissolved minerals such as Cl, Na, K, Ca, Mg, Mn and B by soils were also noted. Groundwater recharge amounted to an average of 15.0 thousand cubic metres per hectare or equivalent to 90 % of the wastewater applied at the 5 cm per week rate. Tree growth increased rapidly. No contamination of groundwater or adverse effect on soils was reported. Results of all research showed that the use of soil for wastewater renovation was one of the simplest and most effective methods of wastewater treatment. However, the soil properties and operation procedure are the main factors that determine the suitability and efficiency of the filtration systems.

Since soil filtration of wastewater can be considered as an example of a tertiary treatment process that can be broadly and easily applied in the field, the concept of using forest soil for wastewater reclamation was obvious (Kardos, 1966). Forest soils, unlike crop land or grass land, are often covered with a layer of a carbonaceous forest floor of varying thickness that can serve as an energy source for the activity of microorganisms (Kardos, 1966; Allison, 1966). The relatively high C:N ratio of the forest floor would contribute to the biological immobilization of added inorganic nitrogen to the organic form (Allison 1966). In addition, the high acidity of the mineral soil might

contribute to adsorption of ammonium ions, and to high retentivity of phosphate because of the presence of iron and aluminium oxides and hydroxides (Hemwall, 1957; Parizek, 1967). Municipal wastes originate mainly from domestic sources and may contain such chemicals as detergents, N, Ca, Mg, Na, P and Cl. Application of wastewater to the land will retain such nutrients for vegetation growth. The principal problem may be the possible contamination of groundwater with soluble nitrate nitrogen as is reported by some authors (Pennypacker, Sopper and Kardos 1967).

A study plan was devised, using soil lysimeters in the green house, to characterize the behaviour, over time, of a West Coast forest soil in response to loading with a primary domestic sewage effluent. This study focusses on the 1) nitrogen and phosphorus retention by a forest soil, 2) changes of physical behaviour of soil, 4) optimization of wastewater loading, and 5) suitability and possible problems in field operation.

The general objectives of this research were to investigate 1) the effects resulting from contact between wastewater, bearing nitrogen and phosphorus, and a forest soil, 2) the soils capacity to retain nitrogen and phosphorus, and 3) the means of balancing the amount of addition to soil against the amount of storage while at the same time minimizing leaching loss. This was done by passing wastewater through soil lysimeters. In response to filtration, physical, chemical and biological changes were expected to take place in the soil. Therefore, a research project was devised to determine water and nitrogen balances for the lysimeters as well as to study physical, chemical and biological changes.

RESEARCH METHODS AND MATERIALS

Materials and Sampling

The soil sampling site was located at Loon Lake, University of British Columbia Research Forest, Haney, B. C., at an altitude of about 400 metres. The vegetation consisted of a combination of western red cedar (Thuja plicata Donn) and western hemlock (Tsuga heterophylla (Raf.) Sarg.). The podzolic soil showed well-developed L, F, H, Ae and Bf horizons in the top 60 cm. Undisturbed cores were taken from the forest floor with the same diameter as the inside diameter of the lysimeters. The L, F and H layers of the forest floor were about 2.5, 1.5 and 5.0 cm thick respectively. Mineral soil of loam texture from the A and B horizons was sampled to a depth of about 45 cm. Unchlorinated wastewater was collected from the primary municipal wastewater treatment plant, West Vancouver, B. C., and stored at

a temperature of two degrees Centigrade. Each supply lasted for a period of about 20 days.

Lysimeter Cylinder Apparatus

Six transparent acrylic plastic cylinders of diameter 14 cm and length 80 cm were employed. In order to gauge the energy status (matric potential) of the soil water in the lysimeter, four tensiometer holes were drilled at 20 cm intervals. Sintered glass bead tensiometers were about 5 cm long and 5 mm in diameter and had air intrusion values of about 200 cm of water. The tensiometers were connected to mercury manometers.

Drainage was facilitated through the installation of a porous plate at the bottom of each lysimeter (Fig. 1). The porous plate consisted of a one-cm thick layer of unconsolidated silicon carbide (25 micron particle size) which provided good hydraulic contact between the soil and the drainage system. The porous plate facilitated the maintenance of aerobic conditions in the soil by maintaining the soil water tension at or above the soil air intrusion value. Water from the soil was collected through the drainage system at a tension of 60 cm of water.

The irrigation system was installed at the top of the lysimeter, 6 cm above the forest floor. Polyethylene pressure tubing (1 cm in diameter) with small holes was used so that water could drip from the tubing onto the forest floor. The rate of flow maintained at 0.35 cm per hour by adjusting the head (wastewater surface level) to the appropriate value. The irrigation system was flushed with clean water once a week

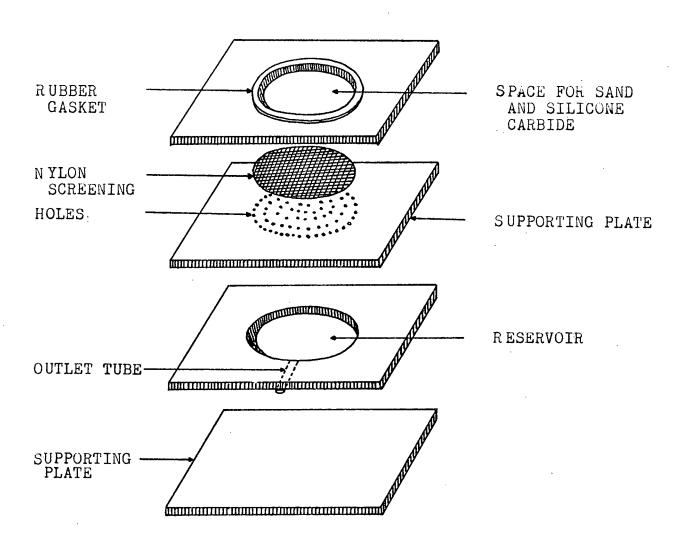


Fig. 1. Porous Plate Assembly

to maintain a constant flow rate (Figure 2).

Preparation and Packing of Soil Sample

Sieving of the mineral soil with a 6 mm sieve resulted in the removal of about 47.1 % (weight basis) of coarse fragments from the soil. Thorough mixing of the soil produced a uniform soil ready for packing in the lysimeters. To pack the soil in the lysimeter, a measured amount of soil of known volume (one pint) was poured into the cylinder and compressed uniformly with a wooden packer. Then the soil surface was loosened with a steel bristle brush in order to ensure good continuity between adjacent layers. Soil depth was measured after every ten increments in order to obtain a measure of packing uniformity. The final volume and weight of the soil were recorded. The bulk densities of the lysimeter soils varied between 0.86 and 0.90 gm per cm³.

Incubation of the Soil

The experiment was carried out in the greenhouse from September 1971 until June 1972. In order to minimize temperature variations and to simulate the environmental conditions at the sampling site, temperature of the soil at depths greater than 40.0 cm was maintained at 15.5 degrees Centigrade by placing the lysimeter in an insulated air bath. During the summer, a fan drawing in outside air was also employed to maintain a favourable temperature in the greenhouse. The surface of the soil was shaded to prevent direct contact with sunlight in order to minimize water loss from the forest floor by evaporation. A sketch of a lysimeter system during incubation is shown in Fig.

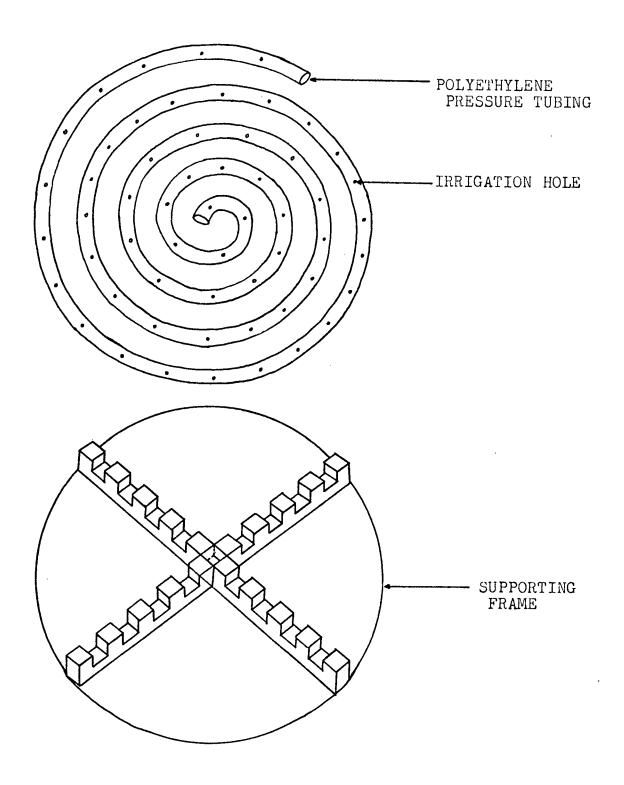


Fig. 2. Irrigation System

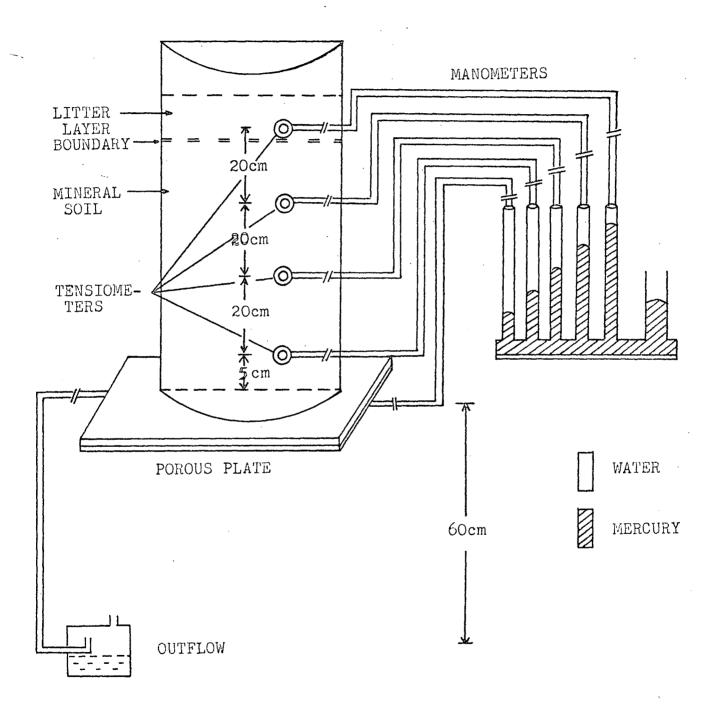


Fig. 3. Measurement of Water Energy Status in the Lysimeter During Incubation.

Wastewater Application and Drainage Water Sampling

The total number of six lysimeters was divided into two groups of three. Wastewater was applied at a rate of 37 cm³ per day (0.23 cm per day) to lysimeters 1 to 3, and at a rate of 75 cm³ per day (0.47 cm per day) to lysimeters 4 to 6. 50 to 100 cm³ of tap water was added at the end of each week. The application flow rate of water was maintained at 0.35 cm per hour. The total amounts of nitrogen applied to the two sets of lysimeters were equivalent to 128 lb N per acre per year (143.4 kg N per hectare per year) and 250 lb N per acre per year (280 kg N per hectare per year) respectively at wastewater nitrogen concentrations varying between 14 ppm and 33 ppm. The total amounts of P applied to the same were equivalent to 27 lb P per acre per year (30.2 kg P per hectare per year) and 53 lb N per acre per year (59.4 kg P per hectare per year) at concentrations between 4.0 and 8.7 ppm.

The volume of drainage water released by the soil lysimeters was measured daily. Water potentials inside the soil lysimeters were also recorded before each wastewater application.

Drainage water sample of sufficient quantity to allow analyses for BOD, nitrogen and phosphorus was collected and stored at a temperature of two degrees Centigrade.

Soil Physical Analyses

Physical properties of both mineral soil and the organic layer were measured to indicate changes due to effluent loading for the one year period.

A. Saturated Hydraulic Conductivity

Hydraulic conductivity is a measure of the ability of a soil to conduct water. It is the flux per unit hydraulic potential gradient and from Darcy's law can be written as

$$K = (Q/At) / (h/L) = v / (h/L)$$

where v is the water flow rate (cm \sec^{-1}), Q the volume of flow (cm³) that passes across the soil cross sectional area A (cm²) in time t (\sec), K is the hydraulic conducitivity (cm \sec^{-1}), and h is the hydraulic head (cm) across a length of flow L (cm).

A steady-state method was employed to measure the saturated hydraulic conductivity in situ (Fig. 4). K was determined by measuring the volume of flow through the soil during a known time interval and hydraulic gradient. The tensiometers were used to measure the hydraulic head drops across the soil layers. In order to minimize air entrapment, the soil was saturated gradually from the bottom up by slowly increasing the elevation of the lysimeter outflow unit, which was connected to a water supply for a period of about 15 hours. Subsequently, steady state was established and maintained by providing constant water levels over the soil surface and at the outlet.

B. Water Retention Characteristics

The measurement of soil water content in conjunction with matric potential yields information about soil water retention characteristics and pore size distribution. Under certain conditions, filter failure of the soil after prolonged periods of loading with wastewater has been found to occur due to the change in biological, chemical and physical conditions inside the soil.

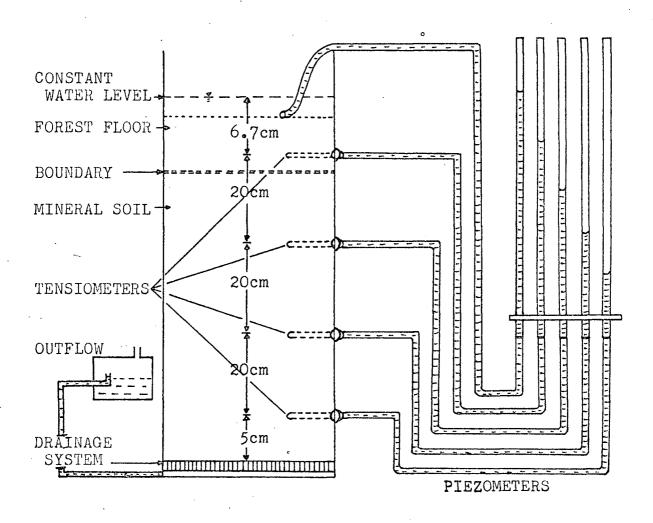


Fig. 4. Steady-State Method of Measuring Saturated Hydraulic Conductivity.

Previous research indicated filter failure occurred at low temperature under aerobic condition or in higher water content under anaerobic conditions (de Vries, 1972). Therefore a technique was employed that allows the simultaneous in situ measurement of the relative soil water content and the corresponding soil water matric potential during drainage after the soil has been saturated (Watson and Whisler, 1968; de Vries, 1969). Water retention curves were obtained by plotting the relative water content, expressed as accumulated outflow, as a function of matric potential. Relative water contents were measured with a gamma radiation attenuation method, and corresponding matric potentials were measured with tensiometer-pressure transducer systems (Chow and de Vries, 1972).

C. Bulk Density

The bulk density of the soil was computed before and after treatment with wastewater. Bulk density of the original soil was determined by placing a known weight of moist soil of known water content in the lysimeter and by measuring the volume. This value represented the average bulk density throughout the soil in the lysimeter. The bulk density of the treated soil was determined by the clod-method at 5-cm intervals (Black, Evans, White, Ensminger and Clark, 1965). This measurement allowed calculation of the porosity of the soil, assuming a particle density of 2.65 gm cm⁻³.

Water and Soil Chemical Analyses

Chemical properties of both wastewater and drainage water were determined periodically in terms of total Kjeldhal N, ni-trate N, ammonium N, total P and BOD, while chemical properties

of both original and treated soils were determined in terms of total Kjeldahl N, nitrate N, and ammonium N. These analyses were carried out to determine the effectiveness of the filtration system and the dynamics of nitrogen and phosphorus retention.

The BOD, which is an indicator of the biodegradable organic matter content of water, was measured periodically by a manometric method (Tool, 1967). Organic nitrogen of both influent and effluent was determined by the macro-Kjeldahl method while the total Kjeldahl nitrogen method was employed to determine ammonia plus organic nitrogen (Standard Methods, 1962). Nitrate nitrogen was measured by the specific ion electrode 1, and total water-soluble phosphorus by the molybdenum blue method (Black, Evans, White, Ensminger and Clark, 1965).

Organic and ammonia nitrogen of both the original and treated soils were determined by the micro-Kjeldahl method (Black, Evans, White, Ensminger and Clark, 1965). Ammonia nitrogen was measured by micro-diffusion followed by colorimetry, and nitrate nitrogen by the chromotropic acid method (West and Ramachandran, 1966). A glass electrode was used to measure the pH of the soil suspension with a water: soil ratio of 1:1. Total carbon was measured by the Leco instrument. All analyses were done in duplicate.

¹⁾ Orion Research Incorporated, nitrate ion electrode model 92-07.

RESULTS AND DISCUSSION

The application of waste water to the soil lysimeters was carried out continuously for a period of 36 weeks. The data were obtained in terms of physical and chemical properties during and after termination of the experiment. Lysimeters 3 and 4 were used for chemical analyses, while lysimeters 1, 2 and 5, 6 were employed to determine the physical properties. All data were expressed on a 7-day basis. This was done by dividing the collected volume of drainage water by the number of days over which this volume was collected, and multiplying the result by seven. All other chemical input and output data were expressed on the same seven day basis.

Water Balance

During the 36 weeks, the two groups of soil lysimeters received average volumes of 8.6 and 16.8 litres of waste water respectively plus an additional volume of 2.2 litres of tap water. The volume of the drainage water indicated that for these two groups about 34.1 % and 20.4 %, respectively, of the total input of liquid was stored or evaporated to the atmosphere. Figure 5 shows the water balance of lysimeters 1, 2, 3 and 4, 5, 6 starting from September 23, 1971, to June 23, 1972. At the beginning, the application of wastewater to the soil lysimeters was not the same, but slightly higher in lysimeters 1, 2, 3 than in lysimeters 4, 5, 6. Wastewater was regularly added to the soil lysimeters six days a week, while fresh water was usually added at the last day of a week. If a certain amount of waste water was not applied to the lysimeters within a scheduled time

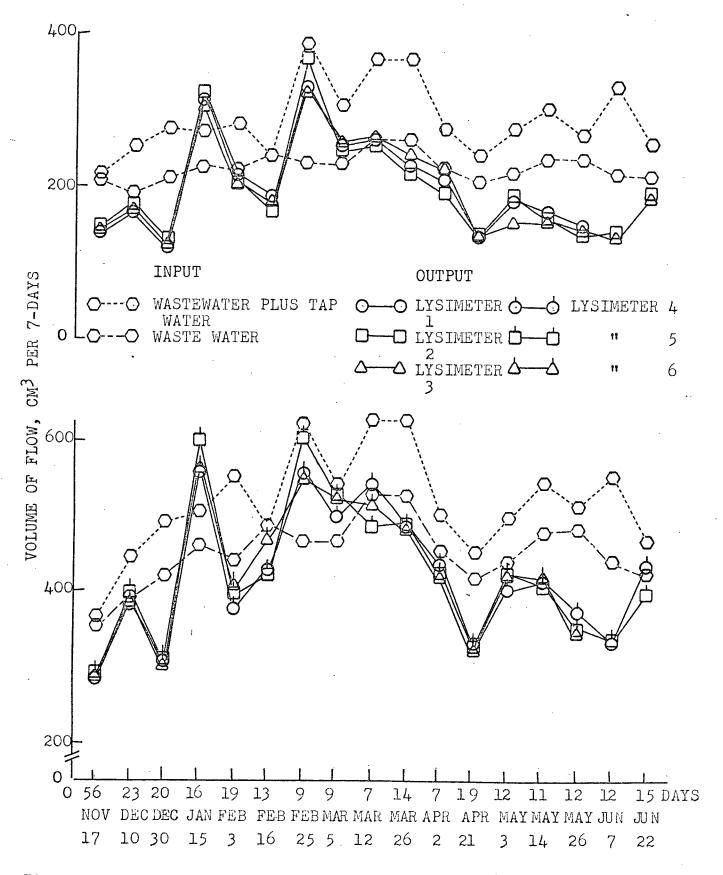


Fig. 5. Partial Water Balances for Lysimeters 1,2,3 (top) and 4,5,6 (bottom).

period, it was added as soon as possible thereafter. This is the reason the amount of wastewater applied expressed on a 7-day basis in both Figures 5 and 6 is not uniform. The volume of drainage water in the 16-day period (January 15) in both Figures 5 and 6 exceeded that of the irrigation water in both soils. This was attributed to the flow from the previous period when the outlet system was closed for a short period of time.

Nutrient Concentrations in Waters

Nutrient concentrations in both wastewater and drainage water were measured periodically in terms of nitrogen, phosphorus and BOD. Figure 6 shows that the concentrations of nitrogen in the drainage water from both groups of soil lysimeters increased with time. The concentration of nitrogen in the drainage water from lysimeters 4, 5 and 6 was about twice than that of the lysimeters 1, 2 and 3. The data of Fig. 6 indicate that the concentration of nitrogen in the drainage water from lysimeters 4, 5 and 6 increased to values higher than that of the wastewater after 23 weeks of loading (January 15). This suggests that all of the nitrogen applied was being leached from the soil. However, soil analyses for nitrogen concentration carried out on both groups of lysimeters did not indicate significant changes in the soil nitrogen content during the application period as shown in Figure 8. It is interesting to note that despite a decrease in the nitrogen concentration of the influent wastewater during the application period, the concentration of the drainage water continued to increase after 23 weeks of treatment.

Concentrations of total phosphorus of both wastewater and drainage water are shown in Figure 11. The data of Figure 11 in-

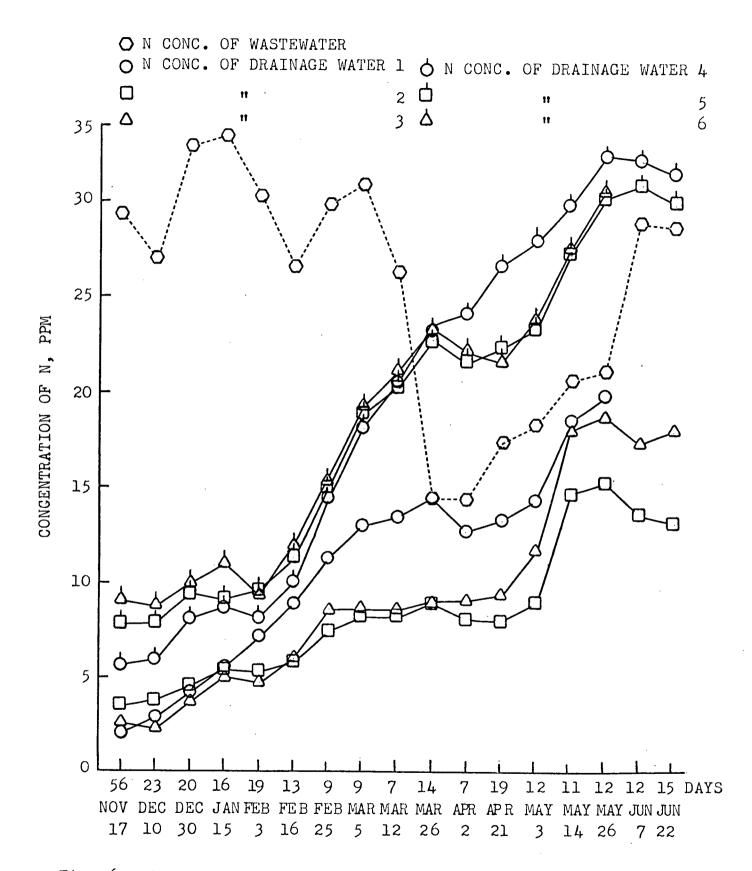


Fig. 6. Concentrations of N in Wastewater and Drainage Water

dicate that the concentration of phosphorus in wastewater varied between 4 ppm and 8.7 ppm with an average concentration of 6.0 ppm. A sharp drop of the concentration in wastewater after 25 weeks of application occurred in conjunction with a drop of the nitrogen content mainly due to the low nutrient concentrations of the wastewater from the treatment plant. The concentration of the drainage water was very low, within the lowest limit of detection, indicating a high retention of added phosphorus by the soil.

Nitrogen Balance

The final calculation showed that the total inputs of nitrogen for the two groups of lysimeters were 223.7 mg and 436.9 mg, or equivalent to 1.4 % and 2.7 % of the total amount of nitrogen present in the original soils (the experimental error was 5%). Data on the nitrogen balance showed that lysimeters 4, 5 and 6 attained an effluent renovation of 43 % in terms of nitrogen as compared to 75 % for lysimeters 1, 2, and 3. Fig. 7 indicates the rate of total nitrogen output from both soil lysimeters increased with time. This increase in nitrogen output from lysimeters 4, 5, and 6 was initially three times higher than that of lysimeters 1, 2, and 3 and the difference increased to five times after 25 weeks (March 26) of wastewater application. Figure 7 shows that the increase of the total nitrogen output of lysimeters 4, 5, and 6 exceeded that of input after 25 weeks (March 26) of application. This indicates that the high nitrogen loading exceeded the soil's capability for biological immobilization, so the added nitrogen and some of the retained nitrogen were biologically converted to nitrate which in

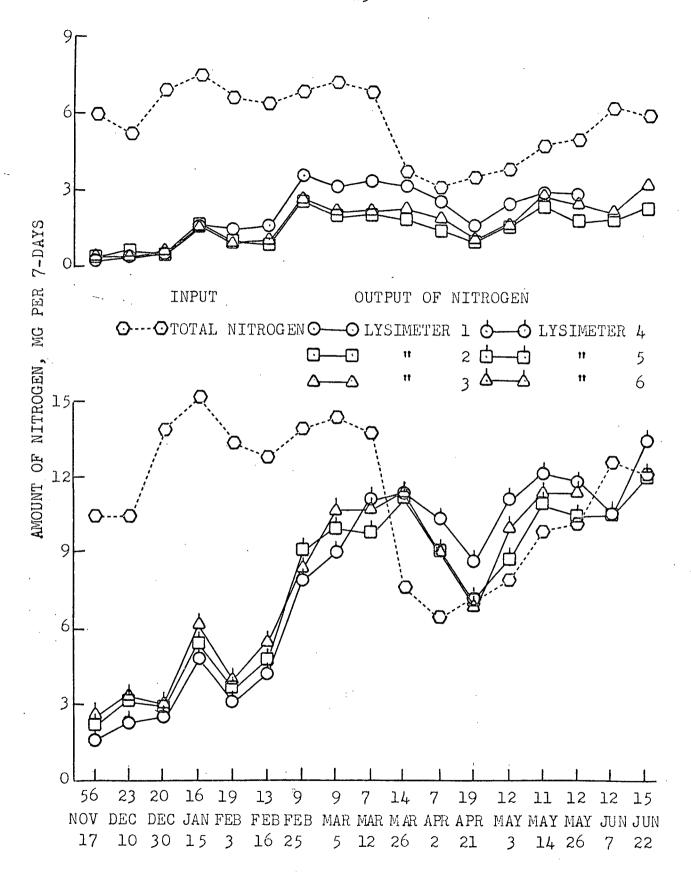


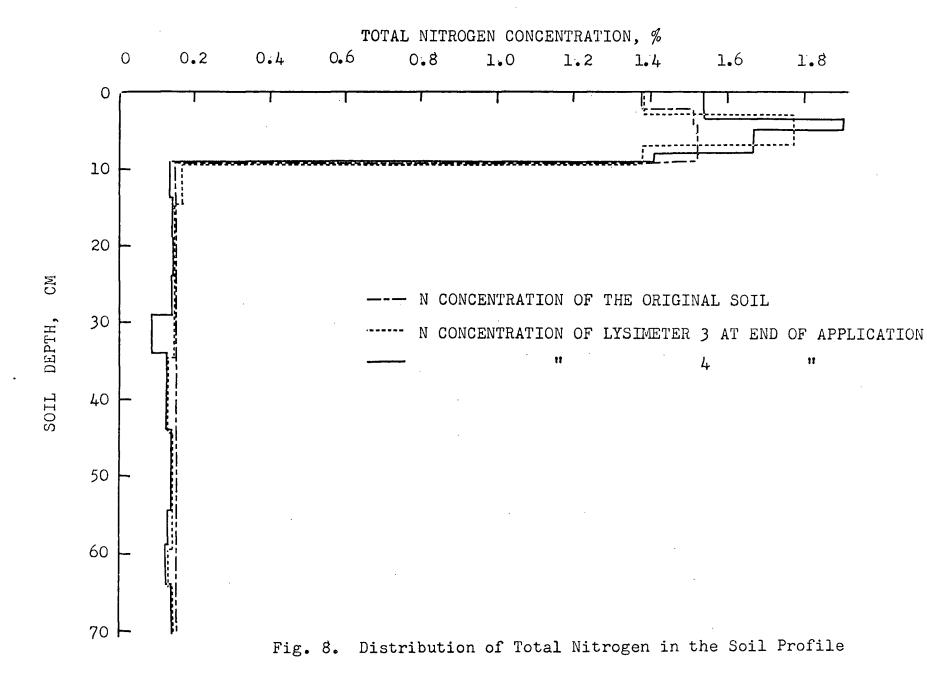
Fig. 7. Total Nitrogen Balances for Lysimeters 1,2,3 (top) and 4,5,6 (bottom).

turn was leached from the soil (Tables 3 and 4). In addition, the residence time of the wastewater in the soil was relatively short. Daily recorded data of wastewater input and effluent output show that about 80 to 90 % of the input water was drained from the soil within 24 hours. Winsor and Pollard (1956) in one of their experiments found a maximum nitrogen immobilization of 56 % after 2 days of incubation at 23.5 degrees Centigrade and 80 % moisture equivalent when 100 ml of solution containing 15 mg of inorganic nitrogen (C:N = 5:1) was added to a marketgarden soil (C:N = 9.5 : 1). Under field conditions, nitrogen immobilization could be enhanced by increasing the contact time between wastewater and soil. Removal of water from the soil by forest vegetation, resulting in lower antecedent soil water contents, would contribute to the desired increase in contact time. This could also be done by reducing the application rate of wastewater to the soil. The total nitrogen output of lysimeters 1, 2, and 3 was 56.5 mg, well below that of the total input 223.7 mg (Tables 1 and 2). This suggested the residence time of wastewater in the soil was more favourable for biological immobilization for lysimeters 1, 2, and 3 than for lysimeters 4, 5, and 6.

Carbon Balance

Total soil carbon was determined before and after treatment of wastewater with the soils. This organic matter served as the main energy source for the heterotrophic organisms in the conversion of inorganic nitrogen to organic form in the soil. The importance of C:N ratio for the immobilization of nitrogen was demonstrated by Allison (1966) and Winsor and Pollard (1956).





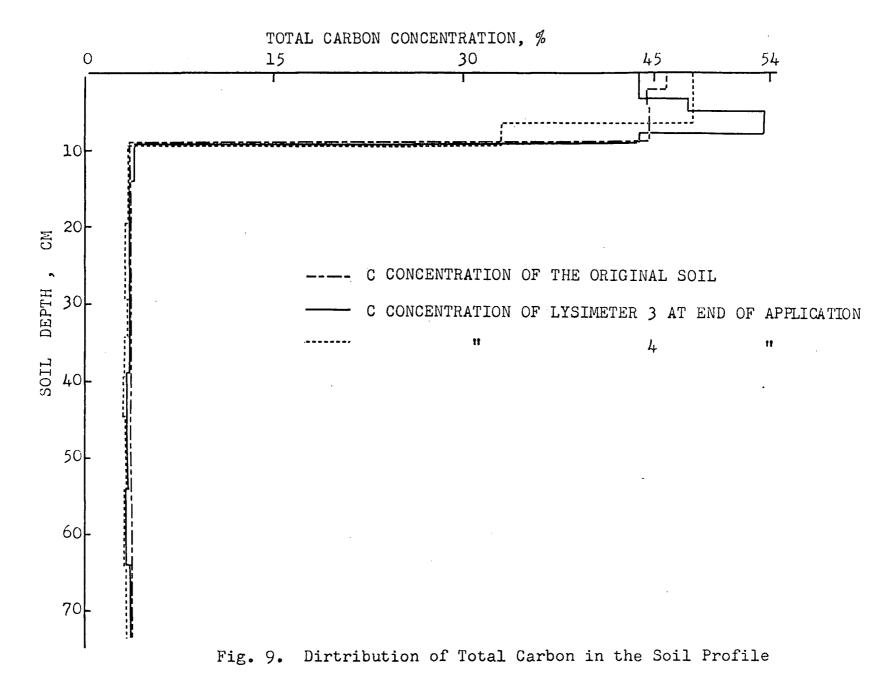
Allison observed that the maximum nitrogen immobilization and minimum carbon dioxide production was reached at about 19 to 21 days of incubation when wheat straw and sodium nitrate had been added to a sandy loam. Immediately after the peak, nitrogen mineralization became dominant and carbon dioxide production closely paralleled nitrogen immobilization. This result was comparable with that of Winsor and Pollard, who found that the nitrogen immobilization peak was at two days instead of 20 days. In the conclusion of his review, Allison (1966) pointed out that this difference in maximum nitrogen immobilization is related to the ease of decomposition of organic materials. The decomposition of organic matter was dependent upon its composition. Lignin, oils, fats and resins are resistant to decompostion, while cellulose, starches, sugars, proteins, amono acids, amides, alcohols and aldehydes etc. are readily decomposable. Since sugar is easily decomposable and thus available to microorganisms, a rapid maximum nitrogen immobilization is expected due to the high carbon and energy supply. On the other hand, wheat straw contains lignin and hemicellulose matter that are more resistant to decomposition, so a longer time is needed for the same maximum nitrogen immobilization (Allison, 1966; Buckman and Brady, In the present experiment, the overall treatment process did not result in any apparent changes in nitrogen and carbon contents of both forest floor and mineral soil (Figures 8 and 9) As can be seen from Table 8, the C:N ratios at the end of the treatment period vary between 25 to 31 and 24 to 35 in the forest floors and 25 to 31 and 22 to 24 in the mineral soils in lysimeters 3 and 4 respectively. The C:N ratios of the original soil

varied from 29 to 33 in the forest floor and was 24 in the minerel soils. This apparent absence of change in the C:N ratios in response to treatment is probably due to the fact that the C:N ratios of both the original soils and added wastewater were not high enough to favor a significant change in biological immobilization. Winsor and Pollard (1956) found that in the glasshouse and market-garden soils, the ratio of carbon added to nitrogen immobilized by microorganisms was 8.3 to 10.8 i.e. 8.3 to 10.8 parts of added carbon are necessary to immobilize one part of nitrogen. Of course, the contact time between wastewater and soil is of prime importance in the process of immobilization as discussed before.

In the present study of forest soil, the C:N ratio of the original mineral soil was 24, and the C:N ratio of the treated soil was similar to that of the original soil in spite of waste water application. The C:N ratio of added wastewater was about 2.5: 1 based on average concentrations of a BOD of 110 ppm and total nitrogen 26 ppm. It is also important to note that immobilization and mineralization occur together in the soil (Allison, 1966).

Since the total amounts of nitrogen added to the two gruops of soils were only 1.4 % and 2.7 % of the original soil nitrogen as compared to about 5 % experimental error in the analyses, apparent changes in nitrogen content, based on soil analysis before and after treatment, are not likely to be significant. Since the soil was highly aerobic, conditions promoted the oxidation of nitrogen to the nitrate form which was readily subject to leaching. The analyses of drainage water showed that almost





all of the nitrogen coming from the soil was in nitrate form (Table 1).

Phosphorus Removal

Satisfactory retention of phosphorus in the forest soil (Table 4) was probably associated with relatively high amounts of reactive iron and aluminium oxides and hydroxides. total water soluble phosphorus input and output showed that the retention of added phosphorus was more than 99 % in both soils. Sopper (1971) found that the renovations of phosphorus in a hardwood plot subject to respective application rates of 2.5 cm and 10 cm per week were 99.9 % and 99.3 %, while in a red pine plot they were 97.0 and 98.7 % with respect to weekly application rates of 2.5 and 5.0 cm. Both soil depths (silt loam to silty clay loam) were reported as 60 cm and average phosphorus concentration of wastewater was 8.5 mg/l over a period of six months. Hemwall (1957) reported that fixation of phosphorus mainly occurred as a result of chemical precipitation and physicochemical sorption rather than by microbiological retention. and Jackson (1950) studied the solubility equilibrium constants of dihydroxy aluminium dihydrogen phosphate , Al(OH)2 H2PO4 variscite crystal species, and dihydroxy iron dihydrogen phosphate, Fe(OH)2 H2PO1 - strengite crystal species, and found that they related the equilibrium concentration of phosphorus in the soil solution directly to the aluminium and iron activity of the However, phosphorus is fixed either by precipitation or sorption by aluminium and iron oxides and hydroxides under acid conditions to form Al(H_2O)₃ (OH)₂ H_2PO_4 or $Fe(H_2O)_3$ (OH)₂ H_2PO_4 (Hemwall, 1957; Russell, 1961; Tisdale and Nelson, 1966).

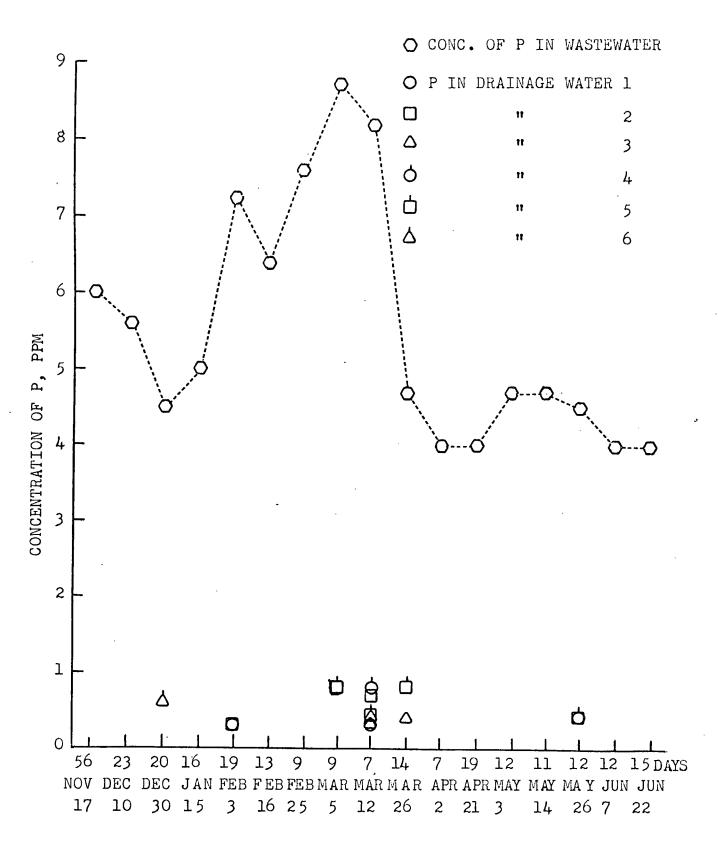


Fig. 10. Concentrations of P in Wastewater and Drainage Water

Physical Properties of the Soil System

Water Retention Properties

Physical properties of the soil are among the important factors in determining the long term suitability of the soil system for wastewater renovation. The results of this research show that the physical behaviour of the soil did not change significantly with time, depending on degree of loading . Figures 11 and 12 show that the water retention characteristics of lysimeter 1 were not changed as compared to the original soil, except in the forest floor where the aeration porosity was relatively reduced. This reduction in aeration porosity of the forest floor was probably due to settling and deposition of organic matter although Figures 8 and 9 did not show significant changes of nitrogen and carbon in lysimeter 1 after wastewater treatment. In the case of lysimeter 6 (Figure 13), the relative total amount of water released by the mineral soil as the matric potential was decreased from 0 to -60 cm of water was higher than that of the original soil, indicating a higher aeration porosity. No specific data were available to account for this result, although the bulk densities of both the original and treated soils in lysimeter 4 (subject to same loadings as in lysimeter 6) were not changed (Table 8). In the forest floor of lysimeter 6, the aeration porosity was relatively lower than that of lysimeter 1 and the original soil. The possible reason may be a higher deposition of organic matter in lysimeter 6.

Saturated Hydraulic Conductivities

Saturated conductivity of the soil was measured to determine

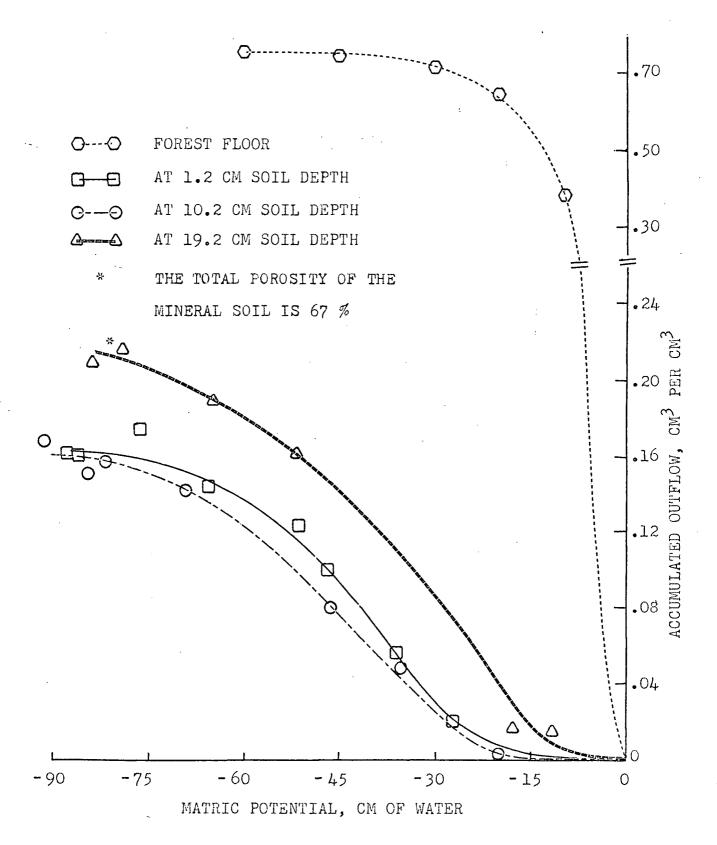


Fig. 11. Water Retention Curve of the Original Soil.

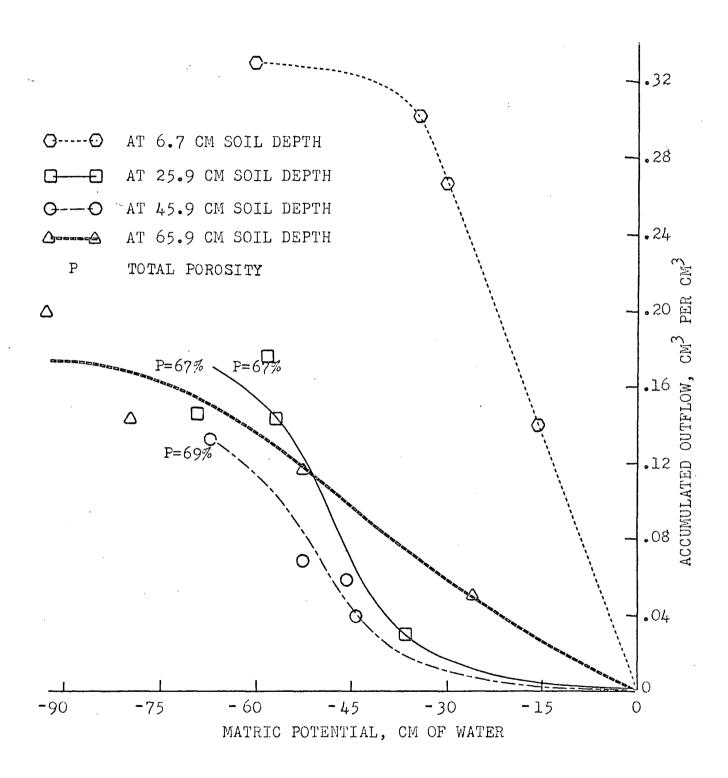


Fig. 12. Water Retention Curve of Lysimeter 1.

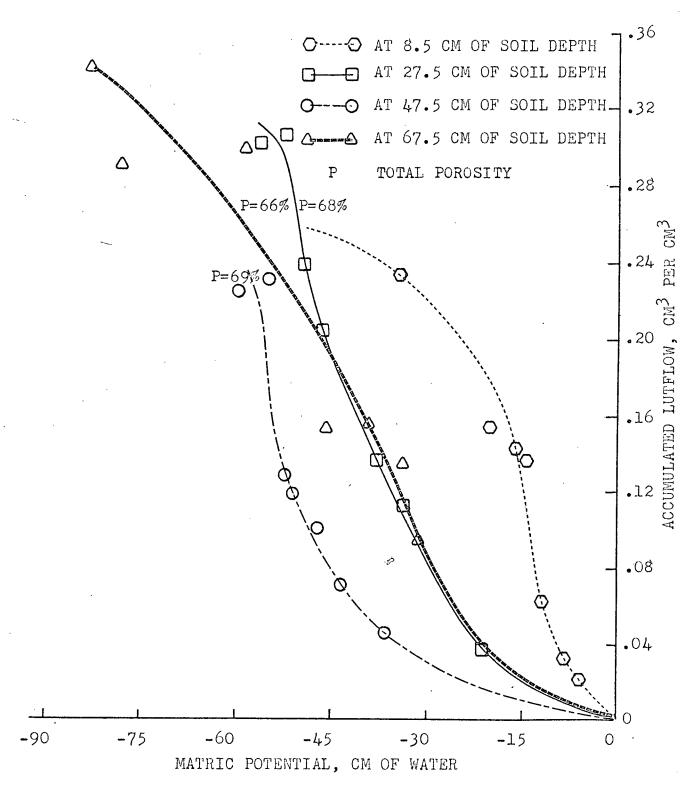


Fig. 13. Water Retention Curve of Lysimeter 6

the effect of wastewater treatment on the soil's ability to transmit water. Figure 14 indicates that the hydraulic conductivities of both soils (lysimeters 1 and 6) were lower than that of the original soil. This result is difficult to explain in view of the fact that the aeration porosity of the mineral soil in lysimeter 6 was higher than that of the original soil (Figures 11 and 13). This might be due to introduction of by-products from microorganisms that might interfere with water movement. McCalla (1950) found that when sucrose was added to soil, the percolation rate dropped rapidly, but when the soil was kept in a refrigerator, the percolation rate did not change. This indicated that the percolation rate had a close relationship with the activity of microorganism. Lysimeters 2 and 5 received the same conditions of treatment with wastewater as lysimeters 1 and 6, but were subjected to a resting and drying of about a month. Results of measurement show that the saturated conductivities of both soil were higher than that of the original soil. The biological activity may have been responsible for the increase of the conductivities.

Since it is likely that the suspended solids in the waste-water, including organic matter, were filtered out by the forest floor, the activity of microorganisms in the forest floor probably was very high. This can be seen from the water retention and hydraulic conductivity characteristics where the aeration porosity of the treated forest floor is much lower than that of the original sample.

Physical Changes Occuring During Incubation

Physical changes in the soil profile during incubation were

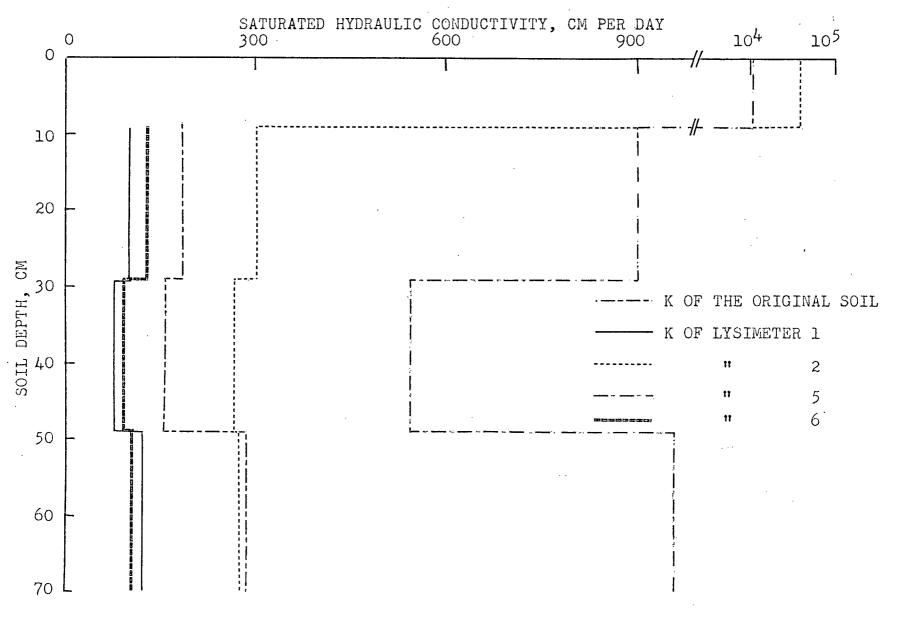


Fig. 14. Saturated Hydraulic Conductivity of The Soils

evaluated during wastewater application. Figures 15 and 16 show that both the matric and total potentials decrease with depth. This implies a downward movement of water in the soil profile. The top 20 cm of lysimeter 1 ($d\Psi/dZ = 3.4$) had a steeper potential gradient than that below 20 cm indicating a lower conductivity across the 0 to 20 cm depth interval, assuming a constant flux with depth (Darcy's Law). A transmission zone existed in the depth intervals from 24 cm to 44 cm. The water content and matric potential were approximately constant across the zone and the only driving force was the gravitational potential gradient. The total potential gradient was unity, and the hydraulic conductivity was equal to the flux. Calculations based on Darcy's Law in Figures 15 and 16 show that the unsaturated conductivities within this zone in lysimeters 1 and 4 are 0.16 cm per day and 0.27 cm per day at matric potentials -50 and -40 cm of water respectively.

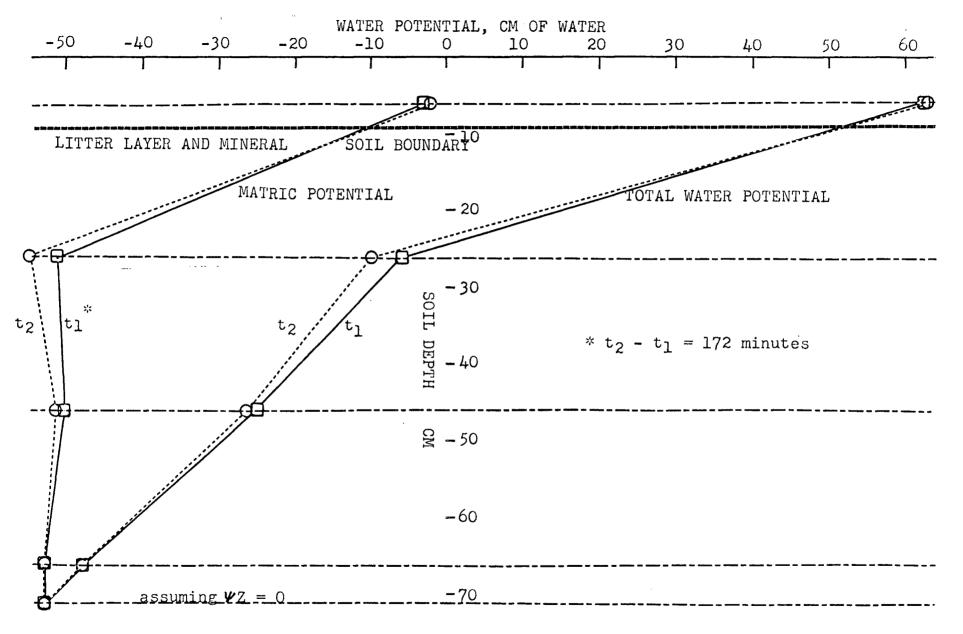


Fig. 15. Changes of Total Water Potential with Time and Depth for the Forest Soil During Incubation in Lysimeter 1.

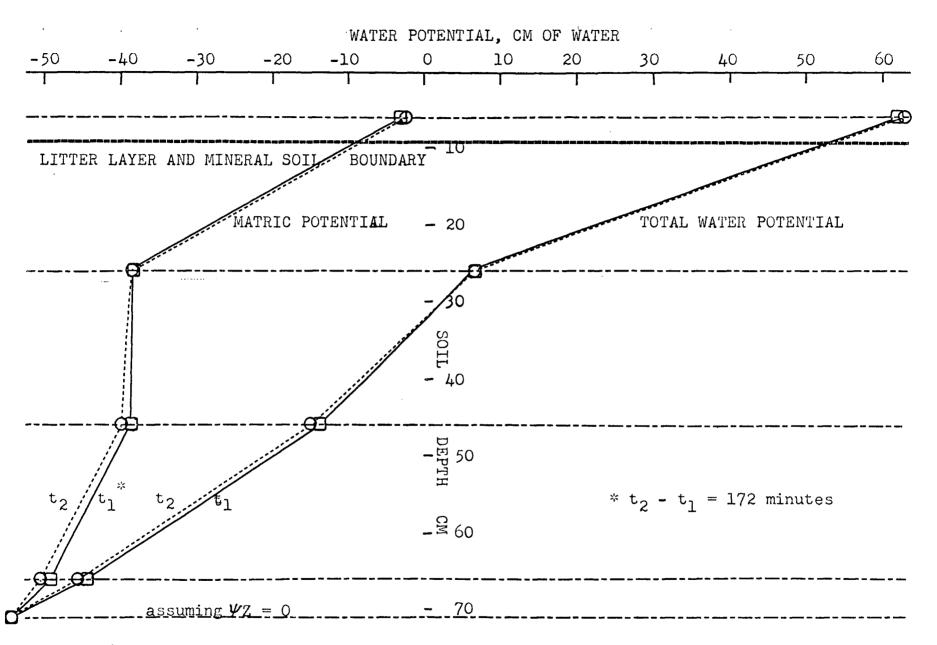


Fig. 16. Changes of Total Water Potential with Time and Depth for the Forest Floor During Incubation in Lysimeter 4.

CONCLUSION

It is of scientific and practical importance to characterize the nature of the forest soil with respect to chemical, physical and possible biological changes. Results of this research suggest that a large scale field operation is feasible. Most of the nutrients lost by leaching in this experiment can be taken up by trees in the field. It has been reported that a coniferous forest has a maximum annual uptake of 50 - 60 kg Nper hectare (45 - 54 lb N per acre) and 6 - 12 kg P per hectare (5 - 11 1b P per acre) (Cole, Gessel and Dice, 1967). The total N and P applied to both groups of soil lysimeters in our study were 143 - 280 kg N per hectare (128 - 250 lb N per acre) and 30 - 59 kg P per hectare (27 - 53 lb P per acre). This indicated the total available nutrients applied exceeded the maximum demand of forest trees. For field operation, a smaller wastewater application rate is suggested than that used in the present experiment in order to increase soil wastewater contact time, reduce leaching loss and maximize nitrogen uptake by forest trees. Therefore, care should be exercised in loading, both in terms of quantity and duration. A project that can be looked upon as a follow-up of this study will be carried out in the University of British Columbia Research Forest in Haney, B. С.

Brief conclusions are therefore drawn from the results reported herein with regard to disposal of wastewater on the forest soil:

1. About 73 % of the total wastewater applied was leached from

- the soil, suggesting that a large quantity of water could be recharged as ground water or taken up by vegetation under field conditions.
- 2. Satisfactory renovation of wastewater with respect to phosphorus and nitrogen was achieved at an application rate of 0.23 cm per day during the period September, 1971, to June, 1972.
- 3. Nitrogen concentration of drainage water increased with time in both groups of lysimeters.
- 4. The output of total nitrogen from lysimeters 4,5,6 (with application rate of 0.46 cm per day) exceeded that of input after 25 weeks of loading indicating low degree of biological immobilization.
- 5. Nitrogen balance data showed that renovation of the waste water with respect to nitrogen in lysimeters 1, 2, and 3 (with application rate of 0.23 cm per day) attained a value as high as 75 %, but renovation in lysimeters 4, 5, and 6 was only 43 %. The renovation would be higher under vegetation growth.
- 6. The C:N ratios were quite constant in both groups of soils, probably due to the balanced immobilization and mineralization of nitrogen. The low C:N ratio of organic matter added, as compared to the original soil, and ease of organic decomposition in the forest floor were important factor for this constancy.
- 7. Renovation of the wastewater with respect to phosphorus by the forest soil was as high as 99.4 % and 99.0 % in lysimeters 3 and 4 due to the high contents of reactive aluminium

- and iron oxides and their hydroxides in the acid mineral soil.
- 8. Renovation of the wastewater with respect to BOD (biochemical oxygen demand) was 100 % in both groups of soils.
- 9. The physical properties of the soils were not greatly altered by a prolonged period of wastewater applications, except in the forest floor where aeration porosity was reduced.
- 10. It is suggested that further research might be carried out on biological effects of alternate wetting and drying, where wastewater is applied to the forest soil. In addition, retention of other nutrients from wastewater in forest soil should be evaluated.
- 11. Further research on contamination of soil with heavy metals from wastewater disposal should also be carried out.

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APPENDIX

i

Table 1. Concentrations and Amount of N in Wastewater Applied to Lysimeters 1, 2, and 3.

| Pe | eriods | .] | Days | cm^3 | cm ³ | NO ₃ -N | N | N | | Water | Water |
|-----|-----------------|-----|------|--------|-----------------|--------------------|------|--------|--------------|-------|---------------|
| | | | | | per 7-day | ppm | ppm | mg | mg/ 7-day | cm3 | cm3/ 7-day |
| | | | • | | | | | | | | |
| Sep | 23-Nov | 17 | 56 | 1633 | 204.1 | 1.5 | 29.3 | 47.87 | 5.98 | 77 | 9.6 |
| Nov | 18-Dec | 10 | 23 | 630 | 191.7 | 1.6 | 26.9 | 16.95 | 5.16 | 200 | 60.9 |
| Dec | ll-Dec | 30 | 20 | 597 | 210.0 | 1.0 | 32.8 | 19.7 | 6.90 | 200 | 70.0 |
| Dec | 31 - Jan | 15 | 16 | 516 | 225.8 | 1.0 | 33.4 | 17.26 | 7.55 | 100 | 43.8 |
| Jan | 16-Feb | 3 | 19 | 599 | 220.7 | 0.5 | 30.2 | 18.11 | 6.67 | 300 | 110.5 |
| Feb | 4-Feb | 16 | 13 | 450 | 242.3 | 0.3 | 26.4 | 11.89 | 6.40 | 0 | 0 |
| Feb | 17-Feb | 25 | 9 | 300 | 233.3 | 0.3 | 29.7 | 8.91 | 6.93 | 200 | 155.6 |
| Feb | 26-Mar | 5 | 9 | 298 | 231.8 | 1.1 | 30.8 | 9.18 | 7.14 | 100 | 77.8 |
| Mar | 6-Mar | 12 | 7 | 260 | 262.0 | 0.6 | 26.2 | 686 | 6.86 | 100 | 100.0 |
| Mar | 13-Mar | 26 | 14 | 522 | 261.0 | 0.6 | 14.6 | 7.61 | 3.81 | 200 | 100.0 |
| Mar | 27-Apr | 2 | . 7 | 224 | 224.0 | 0.6 | 14.3 | 3.20 | 3.20 | 50 | 50.0 |
| Apr | 3-Apr | 21 | 19 | 557 | 205.2 | 0.6 | 17.3 | 9.62 | 3.54 | 100 | 36.8 |
| Apr | 22-May | 3 | 12 | 372 | 217.0 | 1.0 | 18.2 | 6.76 | 3.94 | 100 | 58.3 |
| May | 4-May | 14 | 11 | 371 | 236.1 | 1.2 | 20.5 | 7.61 | 4.84 | 100 | 63.6 |
| May | 15-May | 26 | 12 | 408 | 238.0 | 0.7 | 21.0 | 8.58 | 5.00 | 50 | 29.2 |
| May | 27-Jun | 7 | 12 | 372 | 217.0 | 0.5 | 28.8 | 10.73 | 6.26 | 200 | 116.7 |
| Jun | 8-Jun | 22 | 15 | 450 | 210.0 | 0.4 | 28.6 | 12.88 | 6.01 | 100 | 46.7 |
| • | n . 4 3 | | 051 | 0.5/5 | 1 | | _ | | | 03.55 | |
| '1 | otal | | 274 | 8561 | | | 2 | 223.72 | | 2177 | |

Table 2A. Concentrations and Amount of N in Drainage Water from Lysimeter 1.

| | Periods | 5 | Days | cm ³ | cm ³ per 7-day | NO ₃ -N | N ppm | N mg | N mg/ 7-day |
|----------|--------------|----|------|-----------------|---------------------------------|--------------------|----------|----------------|-------------------|
| Sep | 23-Nov | 18 | 56 | 1100.0 | 137.5 | 2.0 | 2.0 | 2.20 | 0.28 |
| Nov | 19-Dec | 11 | 23 | 544.0 | 165.8 | 2.7 | 2.8 | 1.50 | 0.46 |
| Dec | 12-Dec | 31 | 20 | 341.5 | 119.8 | 4.0 | 4.1 | 1.39 | 0.49 |
| Jan | l-Jan | 16 | 16 | 729.6 | 319.0 | 5.4 | 5.5 | 4.02 | 1.76 |
| Jan | 17-Feb | 4 | 19 | 571.3 | 211.5 | 6.8 | 7.0 | 3.99 | 1.48 |
| Feb | 5-Feb | 17 | 13 | 343.2 | 184.8 | 8.0 | 8.8 | 3.05 | 1.64 |
| Feb | 18-Feb | 26 | 9 | 421.6 | 328.0 | 10.8 | 11.2 | 4.73 | 3.67 |
| Feb | 27-Mar | 6 | 9 | 324.0 | 252.0 | 12.0 | 12.9 | 4.18 | 3.25 |
| Mar | 7-Mar | 13 | 7 | 261.1 | 261.1 | 12.8 | 13.3 | 3.48 | 3.48 |
| Mar | 14-Mar | 27 | 14 | 451.3 | 225.7 | 13.5 | 14.4 | 6.51 | 3.26 |
| Mar | 28-Apr | 3 | 7 | 207.6 | 207.6 | 12.0 | 12.6 | 2.61 | 2.61 |
| Apr | 4-Apr | 22 | 19 | 351.7 | 129.6 | 12.8 | 13.2 | 4.64 | 1.72 |
| Apr | 23-May | 4 | 12 | 311.0 | 181.4 | 13.7 | 14.2 | 4.41 | 2.57 |
| May | 5-May | 15 | 11 | 262.1 | 166.2 | 17.8 | 18.4 | 4.80 | 3.05 |
| May | 16-May | 27 | 12 | 253.8 | 147.8 | 19.2 | 19.8 | 5.01 | 2.92 |
| May | 28-Jun | 8 | 12 | - | - | - | - | - | - |
| Jun | 9-Jun | 23 | 15 | : | - | - | - | - | - |
| <u>.</u> | Total | | 247 | 6473.8 | | | | 56 . 52 | |

Table 2B. Concentrations and Amount of N in Drainage Water from Lysimeter 2.

| | Periods | 5 | Days | cm ³ | cm ³ per 7-day | NO ₃ -N | N ppm | N mg | N mg/ 7-day |
|-----|---------|----|------|-----------------|---------------------------------|--------------------|----------|---------|-------------------|
| Sep | 23-Nov | 18 | 56 | 1134.0 | 142.0 | 3.2 | 3.4 | 3.86 | 0.36 |
| Nov | 19-Dec | 11 | 23 | 575.5 | 175.0 | 3.5 | 3.7 | 2.14 | 0.65 |
| Dec | 12-Dec | 31 | 20 | 374.5 | 130.8 | 4.3 | 4.4 | 1.64 | 0.57 |
| Jan | 1-Jan | 16 | 16 | 717.8 | 313.2 | 4.7 | 5.3 | 3.80 | 1.66 |
| Jan | 17-Feb | 4 | 19 | 548.2 | 202.4 | 4.7 | 5.2 | 2.84 | 1.04 |
| Feb | 5-Feb | 17 | 13 | 307.4 | 165.8 | 4.8 | 5.7 | 1.76 | 0.95 |
| Feb | 18-Feb | 26 | 9 | 464.6 | 360.8 | 7.0 | 7.3 | 3.40 | 2.64 |
| Feb | 27-Mar | 6 | 9 | 313.7 | 244.0 | 7.6 | 8.2 | 2.57 | 2.00 |
| Mar | 7-Mar | 13 | 7 | 251.3 | 251.3 | 7.0 | 8.2 | 2.06 | 2.06 |
| Mar | 14-Mar | 27 | 14 | 431.5 | 215.8 | 8.4 | 8.8 | 3.82 | 1.91 |
| Mar | 28-Apr | 3 | 7 | 191.6 | 191.6 | 7.8 | 8.0 | 1.53 | 1.53 |
| Apr | 4-Apr | 22 | 19 | 370.3 | 136.2 | 7.3 | 7.8 | 2.86 | 1.06 |
| Apr | 23-May | 4 | 12 | 315.1 | 183.8 | 8.5 | 8.8 | 2.76 | 1.62 |
| May | 5-May | 15 | 11 | 250.5 | 159.0 | 14.2 | 14.6 | 3.73 | 2.37 |
| May | 16-May | 27 | 12 | 231.7 | 135.0 | 14.9 | 15.2 | 3.53 | 2.06 |
| May | 28-Jun | 8 | 12 | 241.5 | 140.8 | 13.0 | 13.5 | 3.26 | 1.91 |
| Jun | 9-Jun | 23 | 15 | 403.2 | 188.0 | 13.0 | 13.0 | 5.21 | 2.43 |
| J | otal [| | 274 | 7132.4 | | | | 50.77 | |

Table 2C. Concentrations and Amount of N in Drainage
Water from Lysimeter 3.

| | Periods | 5 | Days | cm ³ | cm ³ per 7-day | NO ₃ -N | N | N mg | N mg/ 7-day |
|-----|---------|----|------|-----------------|---------------------------------|--------------------|-------|---------|-------------------|
| Sep | 23-Nov | 18 | 56 | 1165.0 | 145.6 | 2.5 | 2.50 | 2.91 | 0.36 |
| Nov | 19-Dec | 11 | 23 | 551.0 | 167.7 | 2.2 | 2.23 | 1.23 | 0.37 |
| Dec | 12-Dec | 31 | 20 | 342.0 | 119.7 | 3.6 | 3.60 | 1.23 | 0.43 |
| Jan | 1-Jan | 16 | 16 | 688.7 | 301.3 | 4.9 | 4.94 | 3.40 | 1.48 |
| Jan | 17-Feb | 4 | 19 | 550.6 | 202.8 | 4.0 | 4.70 | 2.59 | 0.95 |
| Feb | 5-Feb | 17 | 13 | 344.8 | 185.7 | 5.5 | 5.94 | 2.05 | 1.10 |
| Feb | 18-Feb | 26 | 9 | 415.6 | 323.2 | 8.0 | 8.40 | 3.49 | 2.71 |
| Feb | 27-Mar | 6 | 9 | 329.5 | 256.3 | 8.0 | 8.62 | 2.84 | 2.21 |
| Mar | 7-Mar | 13 | 7 | 259.3 | 259.3 | 8.0 | 8.52 | 2.21 | 2.21 |
| Mar | 14-Mar | 27 | 14 | 475.9 | 238.0 | 9.0 | 9.79 | 4.66 | 2.33 |
| Mar | 28-Apr | 3 | 7 | 221.7 | 221.7 | 8.1 | 8.93 | 1.98 | 1.98 |
| Apr | 14-Apr | 22 | 19 | 369.5 | 136.1 | 8.5 | 9.17 | 3.39 | 1.25 |
| Apr | 23-May | 4 | 12 | 259.7 | 151.5 | 11.0 | 11.63 | 3.02 | 1.76 |
| May | 5-May | 15 | 11 | 247.0 | 157.2 | 17.1 | 17.85 | 4.41 | 2.81 |
| May | 16-May | 27 | 12 | 236.3 | 137.8 | 17.9 | 18.70 | 4.42 | 2.58 |
| May | 28-Jun | 8 | 12 | 226.0 | 131.8 | 16.5 | 17.30 | 3.91 | 2.28 |
| Jun | 9-Jun | 23 | 15 | 397.9 | 185.7 | 17.5 | 17.89 | 7.12 | 3.32 |
| 7 | [otal | | 274 | 7080.5 | | | | 54.86 | |

Table 3. Concentrations and Amount of N in Wastewater Applied to Lysimeters 4, 5, and 6.

| P | eriods | | Da y s | cm3 | cm3 | NO ₃ -1 | 1 N | N | | Water | Water |
|-----|--------|----|---------------|-------|--------------|--------------------|------|--------|--------------|--------|----------------------------|
| | | | | | per 7-day | ppm | ppm | mg | mg/ 7-day | cm^3 | cm ³ / 7-day |
| | | | | | | | | | | | |
| Sep | 23-Nov | 17 | 56 | 2842 | 355.3 | 1.5 | 29.3 | 83.27 | 10.44 | 95 | 11.9 |
| Nov | 18-Dec | 10 | 23 | 1275 | 388.0 | 1.6 | 26.9 | 34.29 | 10.44 | 200 | 60.9 |
| Dec | 11-Dec | 30 | 20 | 1200 | 420.0 | 1.0 | 32.8 | 39.60 | 13.86 | 200 | 70.0 |
| Dec | 31-Jan | 15 | 16 | 1050 | 459.4 | 1.0 | 33.4 | 34.65 | 15.16 | 100 | 43.8 |
| Jan | 16-Feb | 3 | 19 | 1200 | 442.1 | 0.5 | 30.2 | 36.27 | 13.36 | 300 | 110.5 |
| Feb | 4-Feb | 16 | 13 | 900 | 484.6 | 0.3 | 26.4 | 23.76 | 12.79 | 0 | 0 |
| Feb | 17-Feb | 25 | 9 | 600 | 466.7 | 0.3 | 29.7 | 17.82 | 13.86 | 200 | 155.6 |
| Feb | 26-Mar | 5 | 9 | 600 | 466.7 | 1.1 | 30.8 | 18.48 | 14.37 | 100 | 77.8 |
| Mar | 6-Mar | 12 | 7 | 525 | 525.0 | 0.6 | 26.2 | 13.74 | 13.74 | 100 | 100.0 |
| Mar | 13-Mar | 26 | 14 | 1050 | 525.0 | 0.6 | 14.6 | 15.31 | 7.66 | 200 | 100.0 |
| Mar | 27-Apr | 2 | 7 | 450 | 450.0 | 0.6 | 14.3 | 6.44 | 6.44 | 50 | 50.0 |
| Apr | 3-Apr | 21 | 19 | 1125 | 414.5 | 0.6 | 17.3 | 19.29 | 7.11 | 100 | 36.8 |
| Apr | 22-May | 3 | 12 | 750 | 437.5 | 1.0 | 18.2 | 13.63 | 7.95 | 100 | 58.3 |
| May | 4-May | 14 | 11 | 750 | 477.3 | 1.2 | 20.5 | 15.40 | 9.80 | 100 | 63.6 |
| May | 15-May | 26 | 12 | 825 | 481.2 | 0.7 | 21.0 | 17.36 | 10.13 | 50 | 29.2 |
| May | 27-Jun | 7 | 12 | 750 | 437.5 | 0.5 | 28.8 | 21.60 | 12.60 | 200 | 116.7 |
| Jun | 8-Jun | 22 | 15 | 900 | 420.0 | 0.4 | 28.6 | 25.97 | 12.12 | 100 | 46.7 |
| | _ | | | _ | | | | | | | |
| 7 | Cotal | | 274 | 16792 | 2 | | L | +36.88 | | 2195 | |

Table 4A. Concentrations and Amount of N in Drainage Water from Lysimeter 4.

| | Periods | 3 | Days | cm ³ | cm ³ per 7-day | NO ₃ -N | N | N mg | N mg/ 7-day |
|-----|--------------|----|------|-----------------|---------------------------------|--------------------|------|---------|-------------------|
| Sep | 24-Nov | 18 | 56 | 2272.0 | 284.0 | 5.6 | 5.6 | 12.72 | 1.59 |
| Nov | 19-Dec | 11 | 23 | 1251.9 | 381.0 | 5.7 | 5.9 | 7.40 | 2.25 |
| Dec | 12-Dec | 31 | 20 | 870.0 | 304.5 | 7.8 | 8.0 | 6.95 | 2.43 |
| Jan | 1-Jan | 16 | 16 | 1270.1 | 555.7 | 8.3 | 8.6 | 10.96 | 4.79 |
| Jan | 17-Feb | 4 | 19 | 1027.2 | 378.4 | 7.5 | 8.1 | 8.36 | 3.08 |
| Feb | 5-Feb | 17 | 13 | 794•9 | 428.0 | 9•3 | 9.9 | 7.85 | 4.23 |
| Feb | 18-Feb | 26 | 9 | 714.5 | 555.7 | 14.0 | 14.4 | 10.28 | 8.00 |
| Feb | 27-Mar | 6 | 9 | 639.9 | 497.7 | 17.9 | 18.1 | 11.58 | 9.01 |
| Mar | 7-Mar | 13 | 7 | 540.0 | 540.0 | 20.0 | 20.4 | 11.02 | 11.02 |
| Mar | 14-Mar | 27 | 14 | 968.9 | 484.5 | 23.0 | 23.4 | 22.72 | 11.36 |
| Mar | 28-Apr | 3 | 7 | 431.7 | 431.7 | 23.7 | 24.0 | 10.38 | 10.38 |
| Apr | 4-Apr | 22 | 19 | 894.7 | 329.6 | 26.0 | 26.5 | 23.69 | 8.73 |
| Apr | 23-May | 4 | 12 | 684.4 | 399.2 | 27.5 | 27.8 | 19.04 | 11.11 |
| May | 5-May | 15 | 11 | 643.0 | 409.2 | 29.2 | 29.7 | 19.12 | 12.17 |
| May | 16-May | 27 | 12 | 632.3 | 368.8 | 31.9 | 32.2 | 20.33 | 11.86 |
| May | 28-Jun | 8 | 12 | 567.2 | 330.9 | 31.7 | 32.0 | 18.17 | 10.60 |
| Jun | 9-Jun | 23 | 15 | 919.6 | 429.1 | 31.2 | 31.3 | 28.77 | 13.43 |
| J | Total | | 274 | 15122.3 | | | | 249.34 | |

Table 4B. Concentrations and Amount of N in Drainage Water from Lysimeter 5.

| I | eriods | | Days | cm ³ | cm ³ per 7-day | NO ₃ -N | N | N mg | N mg/ 7-day |
|-----|--------------|----|------|-----------------|---------------------------------|--------------------|------|---------|-------------------|
| Sep | 23-Nov | 18 | 56 | 2269.0 | 287.0 | 7.7 | 7.7 | 17.68 | 2.21 |
| Nov | 19-Dec | 11 | 23 | 1309.5 | 397.0 | 7.8 | 7.8 | 10.26 | 3.13 |
| Dec | 12-Dec | 31 | 20 | 871.5 | 305.8 | 9.3 | 9.3 | 8.12 | 2.85 |
| Jan | 1-Jan | 16 | 16 | 1374.5 | 598.0 | 8.5 | 9.0 | 12.42 | 5.42 |
| Jan | 17-Feb | 4 | 19 | 1065.3 | 393.5 | 9.1 | 9.4 | 9.97 | 3.67 |
| Feb | 5-Feb | 17 | 13 | 780.6 | 420.2 | 10.6 | 11.2 | 8.62 | 4.64 |
| Feb | 18-Feb | 26 | 9 | 773.6 | 603.0 | 14.6 | 14.9 | 11.54 | 8.95 |
| Feb | 27-Mar | 6 | 9 | 673.5 | 525.0 | 18.3 | 18.9 | 12.74 | 9.90 |
| Mar | 7-Mar | 13 | 7 | 483.7 | 483.7 | 19.7 | 20.2 | 9.72 | 9.72 |
| Mar | 14-Mar | 27 | 14 | 974.8 | 487.4 | 22.2 | 22.6 | 22.14 | 11.07 |
| Mar | 28-Apr | 3 | 7 | 421.3 | 421.3 | 21.1 | 21.5 | 9.08 | 9.08 |
| Apr | 4-Apr | 22 | 19 | 880.5 | 324.5 | 21.7 | 22.2 | 19.53 | 7.18 |
| Apr | 23-May | 4 | 12 | 719.6 | 420.0 | 22.9 | 23.2 | 16.72 | 9.75 |
| May | 5-May | 15 | 11 | 635.1 | 404.0 | 26.9 | 27.2 | 17.30 | 11.00 |
| May | 16-May | 27 | 12 | 594.7 | 347.0 | 29.7 | 30.0 | 17.85 | 10.40 |
| May | 28-Jun | 8 | 12 | 571.2 | 334.0 | 30.3 | 30.7 | 17.47 | 10.20 |
| Jun | 9-Jun | 23 | 15 | 846.9 | 395.0 | 29.4 | 29.8 | 25.30 | 11.82 |
| 7 | Total | | 274 | 15245.3 | | | | 246.48 | |

Table 4C. Concentrations and Amount of N in Drainage
Water from Lysimeter 6.

| | Periods | 5 | Days | cm ³ | cm ³ per 7-day | NO ₃ -N | N ppm | N mg | N mg/ 7-day |
|----------|--------------|----|------|-----------------|---------------------------------|--------------------|----------|---------|-------------------|
| Sep | 23-Nov | 18 | 56 | 2270.0 | 287.0 | 9.0 | 9.0 | 20.42 | 2.55 |
| Nov | 19-Dec | 11 | 23 | 1248.5 | 379.0 | 8.7 | 8.7 | 10.94 | 3.31 |
| Dec | 12-Dec | 31 | 20 | 865.0 | 303.0 | 9.9 | 9.9 | 8.54 | 2.98 |
| Jan | l-Jan | 16 | 16 | 1283.9 | 561.5 | 10.4 | 10.9 | 13.99 | 6.12 |
| Jan | 17-Feb | 4 | 19 | 1106.1 | 407.0 | 8.7 | 9.2 | 10.86 | 4.00 |
| Feb | 5-Feb | 17 | 13 | 861.9 | 464.5 | 11.2 | 11.8 | 10.17 | 5.47 |
| Feb | 18-Feb | 26 | 9 | 702.5 | 547.0 | 15.0 | 15.2 | 10.72 | 8.32 |
| Feb | 27-Mar | 6 | 9 | 669.5 | 522.0 | 19.0 | 19.2 | 12.94 | 10.70 |
| Mar | 7-Mar | 13 | 7 | 512.6 | 512.6 | 20.8 | 21.0 | 10.75 | 10.75 |
| Mar | 14-Mar | 27 | 14 | 963.2 | 481.6 | 22.9 | 23.2 | 22.36 | 11.18 |
| Mar | 28-Apr | 3 | 7 | 416.5 | 416.5 | 21.8 | 22.1 | 9.19 | 9.19 |
| Apr | 4-Apr | 22 | 19 | 866.1 | 318.0 | 21.2 | 21.5 | 18.63 | 6.83 |
| Apr | 23-May | 4 | 12 | 719.8 | 420.0 | 23.4 | 23.7 | 17.13 | 9.96 |
| May | 5-May | 15 | 11 | 655.6 | 417.0 | 26.9 | 27.4 | 17.96 | 11.40 |
| May | 16-May | 27 | 12 | 583.6 | 341.0 | 30.2 | 30.4 | 17.86 | 10.42 |
| May | 28-Jun | 8 | 12 | - | - | - | - | - | - |
| Jun | 9-Jun | 23 | 15 | | - | - | - | *** | - |
| <u>,</u> | Total | | 247 | 13723.8 | | | | 212.46 | |

Table 5. Concentrations and Amount of Total Soluble P in Wastewater and Drainage Water in Lysimeters 1, 2, and 3.

| Time | W | astewa | ater | Drainage Water | | | | | | | | |
|------|-----|--------|--------------|----------------|-----|-----|------|------|------|------|-------------|------|
| | P | P | P ma/ | | P | | | P | | | P / | |
| days | ppm | mg | mg/ 7-day | | ppm | | | mg | | 7- | mg/ -day | |
| | | | | 1 | 2 | 3 | 1 | 2 | 3 | 1 | 2 | 3 |
| 56 | 6.0 | 9.80 | 1.22 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| 23 | 5.6 | 3.53 | 1.07 | 0 | 0 | 0 | 0 | 0 | 0 | О | 0 | · O |
| 20 | 4.5 | 2.69 | 0.94 | 0 | 0 | 0 | 0 , | 0 | 0 | 0 | 0 | 0 |
| 16 | 5.0 | 2.58 | 1.13 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| 19 | 7.2 | 4.31 | 1.60 | 0 | 0.3 | 0 | 0 | 0.16 | 0 | 0 | 0.06 | 0 |
| 13 | 6.4 | 2.84 | 1.53 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| 9 | 7.6 | 2.28 | 1.77 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| 9 | 8.7 | 2.59 | 2.02 | 0 | 0 | 0 | 0 | 0 | · O | 0 | 0 | 0 |
| 7 | 8.2 | 2.15 | 2.15 | 0.3 | 0.7 | 0.3 | 0.08 | 0.18 | 0.08 | 0.08 | 0.18 | 0.08 |
| 14 | 4.7 | 2.45 | 1.23 | 0 | 0 | 0.4 | 0 | 0 | 0.19 | 0 | 0 | 0.10 |
| 7 | 4.0 | 0.90 | 0.90 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| 19 | 4.0 | 2.22 | 0.82 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| 12 | 4.7 | 1.75 | 1.02 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| 11 | 4.7 | 1.74 | 1.11 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | O | 0 |
| 12 | 4.5 | 1.83 | 1.07 | 0.4 | 0.4 | 0 | 0.10 | 0.09 | 0 | 0.06 | 0.05 | 0 |
| 12 | 4.0 | 1.49 | 0.87 | - | 0 | 0 | - | 0 | 0 | - | 0 | 0 |
| 15 | 4.0 | 1.80 | 0.84 | - | 0 | 0 | - | 0 | 0 | - | 0 | 0 |
| 274 | | 46.95 | | | | | 0.18 | 0.43 | 0.27 | ı | (Tota | al) |

Table 6. Concentrations and Amount of Total Soluble P in Wastewater and Drainage Water in Lysimeters 4, 5, and 6.

| Time | I | <i>N</i> astewa | ater | Drainage Water | | | | | | | | |
|------|-----|-----------------|--------------|----------------|-----|-----|------|------|------|------|-------------|------|
| | P | P | P / | | P | | | P | | | P / | |
| days | ppm | mg | mg/ 7-day | | ppm | | | mg | | 7- | mg/ -day | |
| | | | | 4 | 5 | 6 | 4 | 5 | 6 | 4 | 5 | 6 |
| 56 | 6.0 | 17.20 | 2.15 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| 23 | 5.6 | 7.10 | 2.17 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| 20 | 4.5 | 5.40 | 1.89 | 0 | 0.6 | 0 | 0 | 0.52 | 0 | 0 | 0.18 | 0 |
| 16 | 5.0 | 5.25 | 2.30 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| 19 | 7.2 | 8.64 | 2.45 | 0.3 | 0 | 0 | 0.31 | 0 | 0 | 0.11 | 0 | 0 |
| 13 | 6.4 | 5.76 | 3.10 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| 9 | 7.6 | 4.56 | 3.55 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| 9 | 8.7 | 5.22 | 4.06 | 0 | 0 | 0.8 | 0 | 0 | 0.54 | 0 | 0 | 0.42 |
| 7 | 8.2 | 4.31 | 4.31 | 0.8 | 0.4 | 0.4 | 0.43 | 0.19 | 0.21 | 0.43 | 0.19 | 0.21 |
| 14 | 4.7 | 4.94 | 2.47 | 0 | 0 | 0.8 | 0 | 0 | 0.77 | 0 | 0 | 0.39 |
| 7 | 4.0 | 1.80 | 1.80 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| 19 | 4.0 | 4.50 | 1.66 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| 12 | 4.7 | 3.53 | 2.06 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| 11 | 4.7 | 3.53 | 2.25 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| 12 | 4.5 | 3.71 | 2.17 | 0.4 | 0 | 0.4 | 0.25 | 0 | 0.23 | 0.15 | 0 | 0.14 |
| 12 | 4.0 | 3.00 | 1.75 | 0 | 0 | _ | 0 | 0 | - | 0 | 0 | - |
| 15 | 4.0 | 3.60 | 1.68 | 0 | 0 | | 0 | 0 | - | 0 | 0 | - |
| 274 | | 92.05 | | | | | 0.99 | 1.51 | 1.52 | | (Tota | al) |

Table 7. Chemical and Physical Properties of the Original Soil and the Treated Soil in Lysimeter 3.

| Soil Depth cm | Total Kjeldhal N % | NH ₄ -N | Total N % | C % | C:N | рН | Water Content gm/gm | Bulk Density gm/cm ³ |
|---------------------|-----------------------------|--------------------|-----------------|--------|-------|------|---------------------------|---------------------------------------|
| 0 -3.5 | 1.53 | 296.2 | 1.53 | 43.72 | 28.56 | - | 3.31 | - |
| 3.5- 5 | 1.89 | 602.2 | 1.90 | 47.50 | 25.00 | ~ | 3.32 | - |
| 5 - 8 | 1.66 | 676.3 | 1.66 | 53.50 | 32.23 | - | 4.18 | - |
| 8 - 9 | 1.40 | 431.7 | 1.41 | 43.70 | 30.99 | - | 4.07 | - |
| 9 -14 | 0.14 | 24.9 | 0.14 | 3.90 | 27.86 | 4.04 | 0.34 | 0.81 |
| 14 -19 | 0.14 | 3.8 | 0.14 | 3.61 | 25.79 | 4.03 | 0.44 | 0.84 |
| 19 -24 | 0.14 | 3.4 | 0.14 | 3.52 | 25.14 | 4.06 | 0.44 | 0.86 |
| 24 -29 | 0.13 | 2.7 | 0.13 | 3.42 | 26.31 | 4.25 | 0.44 | 0.88 |
| 29 -34 | 0.10 | 1.2 | 0.11 | 3.40 | 30.90 | 4.44 | 0.44 | 0.90 |
| 34 -39 | 0.12 | 1.4 | 0.12 | 3.52 | 29.33 | 4.57 | 0.45 | 0.88 |
| 39 -44 | 0.12 | 1.6 | 0.12 | 3.27 | 27.25 | 4.60 | 0.45 | 0.88 |
| 44 -49 | 0.12 | 1.4 | 0.12 | 3.41 | 28.42 | 4.72 | 0.46 | 0.87 |
| 49 -54 | 0.13 | 1.2 | 0.13 | 3.46 | 26.62 | 4.68 | 0.46 | 0.89 |
| 54 -59 | 0.13 | 1.2 | 0.13 | 3.22 | 24.77 | 4.64 | 0.46 | 0.88 |
| 59 -64 | 0.12 | 1.2 | 0.13 | 3.23 | 24.85 | 4.65 | 0.45 | 0.83 |
| 64 -69 | 0.13 | 8.7 | 0.13 | 3.51 | 27.00 | 4.65 | 0.46 | 0.82 |
| 69 -71 | 0.13 | 16.6 | 0.13 | 3.51 | 27.00 | 4.65 | - | ~ |
| Original | Soil | | | | | | | |
| L (0 -2 | .2)1.38 | - | 1.38 | 45.89 | 33.33 | - | - | - |
| F (2.2-4 | .2)1.51 | _ | 1.51 | 44.35 | 29.45 | - | - | - |
| Н (4.2-9 | .0)1.56 | - | 1.56 | 44.44 | 28.56 | | - | |
| Soil Lay | er 0.15 | | 0.15 | 3.46 | 23.72 | 4.65 | 0.45 | 0.88 |

Table 8. Chemical and Physical Properties of the Original Soil and the Treated Soil in Lysimeter 4.

| Soil | Total | | Total | | | | Water | Bulk |
|------------|----------|-------|-------|-------|-------|------|---------|--------------------|
| Depth | Kjeldhal | NH4-N | | С | C:N | рН | Content | Density |
| | N | | N | | | | | _ |
| cm | % | ppm | % | % | | | gm/gm | gm/cm ³ |
| 0 - 3 | 1.38 | 128.6 | 1.38 | 48.10 | 34.86 | _ | 2.92 | _ |
| 3 - 7 | 1.76 | 481.3 | | 48.20 | | | 2.93 | _ |
| | | | | | | | | - |
| 7 - 9.5 | 1.37 | 408.7 | 1.38 | 32.89 | 23.83 | - | 3.73 | - |
| 9.5-14.5 | 0.15 | 17.7 | 0.15 | 3.47 | 23.13 | 4.51 | 0.47 | 0.85 |
| 14.5-19.5 | 0.14 | 16.2 | 0.16 | 3.42 | 21.97 | 4.03 | 0.49 | 0.84 |
| 19.5-24.5 | 0.14 | 13.3 | 0.14 | 3.33 | 23.78 | 4.09 | 0.49 | 0.86 |
| 24.5-29.5 | 0.14 | 13.0 | 0.13 | 3.19 | 23.78 | 4.13 | 0.49 | 0.85 |
| 29.5-34.5 | 0.14 | 13.1 | 0.14 | 3.33 | 23.78 | 4.16 | 0.49 | 0.85 |
| 34.5-39.5 | 0.13 | 20.1 | 0.13 | 3.14 | 23.58 | 4.07 | 0.50 | 0.85 |
| 39.5-44.5 | 0.12 | 37.9 | 0.13 | 3.04 | 23.38 | 4.17 | 0.50 | 0.85 |
| 44.5-49.5 | 0.13 | 39.8 | 0.14 | 3.23 | 22.33 | 4.40 | 0.50 | 0.91 |
| 49.5-54.5 | 0.15 | 45.1 | 0.15 | 3.19 | 21.27 | 4.58 | 0.49 | 0.88 |
| 54.5-59.5 | 0.14 | 45.3 | 0.14 | 3.14 | 22.52 | 4.59 | 0.50 | 0.86 |
| 59.5-64.5 | 0.13 | 45.9 | 0.13 | 3.09 | 23.77 | 4.58 | 0.52 | 0.84 |
| 64.5-69.5 | 0.14 | 43.8 | 0.14 | 3.12 | 22.82 | 4.64 | 0.52 | 0.81 |
| 69.5-72.0 | 0.14 | 41.0 | 0.14 | 3.28 | 21.87 | - | 0.52 | 0.74 |
| Original S | Boil | | | | | | | |
| Soil Layer | 0.15 | - | 0.15 | 3.46 | 23.72 | 4.65 | 0.45 | 0.86 |

Table 9. Matric Potential vs. Volumetric Water Content in Lysimeter 1.

| Parameters | Matric Potential cm of water | | | | | | |
|--------------|--|-------|-------|-------|--|--|--|
| Soil Depth | Volumetric Water Content cm ³ per cm ³ | | | | | | |
| 6 . 7 | 14.5 | 30.5 | 34.2 | 59•9 | | | |
| | 0.140 | 0.266 | 0.302 | 0.328 | | | |
| 25.9 | 35•9 | 56.4 | 57•9 | 69.1 | | | |
| | 0.029 | 0.143 | 0.175 | 0.145 | | | |
| 45.9 | 43.8 | 45.4 | 52.2 | 66.8 | | | |
| | 0.039 | 0.058 | 0.068 | 0.132 | | | |
| 65.9 | 25.6 | 52.3 | 79•3 | 92.7 | | | |
| | 0.050 | 0.116 | 0.142 | 0.198 | | | |

Table 10. Matric Potential vs. Volumetric Water Content in Lysimeter 6.

| Parameters Soil Depth cm | Matric Potential Volumetric Water Content | | | | cm of water | | |
|--------------------------|--|---------|----------|-----------|-------------|-------|-------|
| Cini | VOLUM | ecric w | Tacer co | cm per cm | | | |
| 8.5 | 5•3 | 8.0 | 11.3 | 13.7 | 15.4 | 19.5 | 33.6 |
| | 0.021 | 0.031 | 0.062 | 0.136 | 0.142 | 0.153 | 0.233 |
| 27.5 | 20.4 | 33.0 | 37.2 | 45•5 | 48.7 | 51.3 | 55•4 |
| | 0.035 | 0.112 | 0.136 | 0.204 | 0.238 | 0.306 | 0.302 |
| 47.5 | 36.0 | 43.0 | 46.5 | 50.3 | 51.8 | 54.2 | 58.8 |
| 47.0 | 0.045 | 0.070 | 0.101 | 0.118 | 0.128 | 0.231 | 0.225 |
| 67.5 | 30.9 | 33.1 | 38.4 | 45.0 | 57.6 | 77.1 | 83.0 |
| | 0.093 | 0.134 | 0.155 | 0.153 | 0.299 | 0.291 | 0.341 |

Table 11. Matric Potential vs. Volumetric Water Content of the Original Soil.

| Parameters Soil Depth cm | Matric Potential cm of water Volumetric Water Content cm ³ per cm ³ | | | | | •••• | | | |
|--------------------------|--|-------|-------|----|-------------|---------------|------|-----|-------|
| 1.2 | 26.9 | 35.9 | 46. | 9 | 51.3 | 65.4 | 77 | •3 | 86.0 |
| 1.62 | 0.002 | 0.056 | 0.099 | | 0.125 | 0.144 | 0.: | 176 | 0.160 |
| 10.2 | 19.8 | 35•1 | 46.3 | | 69.2 | 81.5 | 84. | .4 | 91,6 |
| | 0.002 | 0.048 | 0.080 | | 0.142 | 0.157 | 0.3 | 151 | 0.168 |
| 19.2 | 11.5 | 17.7 | 51.7 | | 64.8 | 78.9 | 83.7 | | 113.3 |
| | 0.014 | 0.017 | 0.1 | 61 | 0.189 | 0.216 | 0.7 | 209 | 0.221 |
| 2) F. F. | 10.0 | 20.6 | | | 0.0 .710 | 45.0 0.745 | | | 755 |

^{1).} Bulk density = 0.82 gm per cm^3 , water content = 0.44 gm per gm.

^{2).} Forest floor: Matric potential vs gravimetric water content.

Table 12. Saturated Hydraulic Conductivity of Both the Original and Treated Soils.

| Lysimeters Soil Depth cm | Treated l | Treated 2 | Treated 5 | Treated 6 | Original Soil |
|--------------------------|--------------|--------------|--------------|--------------|------------------|
| 0 - 9 | - | 63750.9 | 11962.9 | 1 | - |
| 9 - 29 | 102.2 | 302.9 | 902.4 | 129.6 | 184.3 |
| 29 - 49 | 78.1 | 266.4 | 544•5 | 92.8 | 157.7 |
| 49 - 69 | 123.0 | 274•3 | 958.6 | 105.6 | 285.3 |

Notes: 1) Both lysimeters 1 and 6 were measured in May, 1972.

- 2) Both lysimeters 2 and 5 were measured in June, 1972, after subject to period of drying.
- 3) The original soil was lysimeter 2 which was measured in August, 1971.
- 4) The unit of conductivity is cm per day.