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## ABSTRACT

The objective of this study was to quantitatively describe the structure, composition and ecological relationships of old-growth forests of west-central Vancouver Island. Data were obtained by sampling 172 plots, at elevations up to 1000 m , located within thirteen drainage areas. Hypothesized relationships between vegetation and environmental variation were examined using gradient analysis and multivariate methods. Successive reciprocal averaging ordination of the vegetation data led to the recognition of six vegetation groups (floodplain, subalpine, Pinus contorta, Pseudotsuga, Thuja, Abies) and twenty-three community types. Data from the tree, sapling, seedling, shrub, herb and bryophytelichen strata were used. Vegetation groups are differentiated along macro-climatic and soil parent material gradients. The vegetation of the Pseudotsuga group, dominant inland, appears to respond to gradients of elevation and soil moisture. The Thuja group is found only near the coast, and its vegetation varies along gradients of soil nutrients and elevation; soil moisture having little effect. The vegetation patterns of the Abies group are correlated to elevation and soil moisture. Canonical variates analyses revealed a close relationship between vegetational and environmental patterns within most vegetation groups. A preci-pitation-continentality gradient was identified as the major determinant of modal vegetation variation. Along this gradient, alpha and beta diversity increased towards the drier and more continental interior as predicted. Tree size-class distribution data indicate that Pseudotsuga menziesii is a seral species in most community types. The dominance of Thuja plicata near the coast may be maintained because of its longevity
and, possibly, its wind damage resistance. Attention is drawn to the ecological mechanisms operating in coastal forests which have important implications for their successful management.

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## INTRODUCTION

## A. GENERAL INTRODUCTION

Vancouver Island possesses the most productive forest stands in Canada, with the largest total basal area, wood volume per hectare, and the tallest trees. Small protected areas containing examples of such stands can be found in MacMillan Provincial Park, better known as "Cathedral Grove", and in Pacific Rim National Park. Elsewhere on Vancouver Island, "old-growth" stands have been disappearing steadily through continued harvesting by the forest industry. Low-elevation, old-growth forests are almost non-existent in certain parts of Vancouver Island, especially on the east coast and in the Port Alberni area. Ecological studies of the few remaining stands of old-growth forests are urgently needed to provide valuable, or even vital information for future management of Canada's west coast forests on a scientific basis.

The study of old-growth forests will yield information on how these forests maintain themselves, where they attain the best growth, and which site and soil properties are important to their growth. Such information can assist in developing guidelines for harvesting and postharvesting treatments (slashburning, scarification, etc.), as well as provide site-specific 1ists of tree species that are suitable for replanting. The future success of forest management is due, in large part, to proper selection of suitable species for replanting after logging.

This need has already been dramatically demonstrated by the numerous failures of Pseudotsuga menziesii plantation, ten to thirty years after planting (Spiers et al., 1983; Carter et al., 1984). The loss of ten to thirty years in what ideally should be an eighty year rotation period is a potential disaster. Forest management on a sound ecological basis aims at preventing such costly errors.

If the study of old-growth forests can contribute valuable information for forest management purposes, it is essential that such studies be undertaken soon, while nearly all the variation expressed in old-growth forest vegetation, along various environmental gradients, can still be found. In response to these concerns, one of the major objectives of this study is to provide a detailed ecological study of old-growth forest communities on west-central Vancouver Island (Fig. 1).

Numerous ecological studies of forest communities in coastal British Columbia have been carried out by Dr. V.J. Krajina and his students. Most of these studies, however, were done on the east coast of Vancouver Island (Krajina and Spilsbury, 1953; Mueller-Dombois, 1959; McMinn, 1960) or on the adjacent mainland (Orloci, 1961, 1964; Brooke et al, 1970; Klinka, 1976). The relatively few ecological studies of western Vancouver Island have been confined to small areas or special habitats (Wade, 1965; Kuramoto, 1965; Cordes, 1972; Kojima and Krajina, 1975). Other researchers also have studied the vegetation of the east coast of Vancouver Island (Beese, 1981; Roy, 1984; Roehmer, 1972). The vegetation of the Carnation Creek Experimental Watershed on western Vancouver Island was described by Oswald (1973, 1974, 1975). Also, two noteworthy
studies of western Vancouver Island forests above the community level are those of Packee (1976) and Klinka et al. (1979).

## B. OBJECTIVES AND HYPOTHESES

Although the description of the structure, composition and ecological characteristics of the old-growth forest communities of westcentral Vancouver Island is the major objective of this study, several other objectives will be considered concurrently. These secondary objectives may be stated in the form of hypotheses, or predictions from the literature. These working hypotheses are not devised to be examined through the formal hypotheses testing procedure described as "strong inference" by Platt (1964). Quinn and Dunham (1983) have argued that, "strict application of a formal strong inference methodology to elucidating potential causes of patterns in nature is frequently infeasible". A great deal of this difficulty resides in the formulation of appropriate null hypotheses. Also, causal factors of ecological patterns are probably not mutually exclusive, and it becomes impossible to distinguish between alternative hypotheses if these causal factors operate simultaneously (Quinn and Dunham, 1983). Therefore, the hypotheses formulated below should instead serve as reference points for the interpretation and discussion of the results obtained.

## 1. ENVIRONMENTAL GRADIENTS AND VEGETATION PATTERNS

Numerous studies have pointed out the major environmental gradients generally responsible for the largest amounts of variation within vegetation. Nevertheless, it remains interesting to examine for the first
time vegetation-environment relationships in a large, climatically diverse area of western Vancouver Island. Specific questions asked at the outset of the research were : Which environmental gradient, or gradients, will be associated with the largest amount of variation in the vegetation over the whole study area? If certain environmental factors are held constant (through manipulation of field data), do others emerge as having potentially significant control over vegetation patterns? Are the predominant environmental gradients the same in climatically different parts of the study area?

These questions can be investigated through the use of a functional approach to plant community ecology. Austin et al. (1984) summarized this approach and pointed out its similarities to gradient analysis. The functional approach was pioneered in the study of soils by Jenny (1941), in which soils were expressed as being a function of climate, parent material, topography, a biotic factor and time. Jenny (1941) suggested that if all factors except one were held constant, relationships between this one factor and soil properties could be demonstrated and analyzed (Austin et al., 1984). A similar functional, factorial approach to plant ecology was later proposed by Major (1951) and Crocker (1952).

Similarly, in his direct gradient analysis of the Great Smoky Mountains, Whittaker (1956) assumed that vegetation properties were related to meso-climate and topography, when parent material, the biotic factor and time were held constant. The meso-climate gradient was estimated by elevation (approximating temperature), and the topography gradient
was measured by slope aspect and degree of exposure (approximating moisture, Whittaker (1956)). These two gradients served as the axes of the now classic, two-dimensional diagrams of the vegetation of the mountainous: areas studied by Whittaker (Whittaker, 1956; 1960; Whittaker and Niering, 1965). A third factor, such as parent material or latitude (macro-climate) or successional status (time) can be introduced by producing elevation-topography diagrams for areas differing only in the third factor of interest (Whittaker, 1960; Whittaker and Niering, 1968b; Perring, 1960; Peet, 1978; Kessel 1979). Kessel (1979) produced a comprehensive series of two-dimensional diagrams to display major vegetationenvironment relationships for Glacier National Park., Montana. Such graphical methods have permitted the ecological interpretation of complex distribution patterns of individual species and community characteristics, including species diversity and productivity (Whittaker and Niering, 1975).

Within this study area the biotic factor (herbivory, competition) is assumed to be constant, as is the time factor, since mostly old-growth forest stands were sampled. Climate, topography and parent material are the environmental factors showing the greatest variation within the study area.

It is generally accepted that macro-climate (mainly temperature and precipitation) will influence most strongly the vegetation of an area. Elevation represents a major precipitation and temperature gradient within the present study area; as well, precipitation declines markedly along a west to east axis from coastal to more interior parts
of the island. Macro-climate and meso-climate levels can be subjectively distinguished by scale. For example, there is a macro-climatic difference between Tofino and Port Alberni (different total precipitation), while there is only a meso-climatic difference between the north and south facing slopes surrounding Sproat Lake.

In his study of the vegetation of the Siskiyou Mountains of Oregon, Whittaker (1960) documented an increase in alpha and beta diversity along a gradient from the coast to the interior. Alpha diversity represents the species richness, or number of species at a site, while beta diversity refers to the rate of change in species composition (termed "species turnover") along an environmental gradient (Whittaker, 1975). A similar trend was detected in the central Washington Cascade Mountains by Del Moral and Watson (1978), and in Finland by Oksanen (1983).

From the above, the following hypotheses were formulated for the study of the vegetation of west-central Vancouver Island using gradient analysis methods :
a) Vegetation and individual species patterns will be most strongly correlated with macro-climatic factors over the entire study area.
b) Following macro-climate, parent material factors will exert the next strongest influence on vegetation and species patterns.
c) If macro-climate and parent material are held fairly constant, that is if subsections of the entire study area are analysed
separately, meso-climate will be most strongly correlated with vegetation and species patterns.
d) Within the same subsections as in (c) topographical factors related to soil moisture will follow meso-climate in their apparent control over vegetation and species patterns.
e) The macro-climatic gradient of decreasing precipitation and increasing continentality, progressing inland from the coast, should be reflected by increases in both alpha and beta diversity in the vegetation.

## 2. VEGETATION ; PATTERNS VS. ENVIRONMENTAL PATTERNS

A major assumption of indirect gradient analysis, or indirect ordination, is that the vegetation patterns illustrated reflect underlying environmental patterns (Whittaker, 1967; 1978). A further objective of this thesis is to examine the validity of this assumption within the present study area.

The degree to which communities, differentiated by vegetation attributes, can also be independently delineated using environmental variables is a good indication of the relationships between vegetation and environmental patterns. Old-growth forests that have been developing for centuries would seem to represent a system where vegetation and environment are in close harmony. Vegetation variability introduced by most small scale disturbances (eg. deaths of isolated individuals) is minimized in such forests. Disturbance on a larger scale (eg. fire or
storm damage) may have more profound effects on vegetation patterns depending on the type, intensity, and frequency of the disturbance. Counteracting the deterministic relation between environment and vegetation are stochastic events, such as the establishment of different species in newly opened microsites, or the year to year variation in seed production by different species, which also are characteristic of the developing forest. Thus two identical disturbances, but not occurring at the same time or place, may often promote the establishment of a different vegetation due to stochastic events.

These ideas are reformulated in the following statements or hypotheses :
a) The interplay of competitive forces between populations of species in old-growth forests over long periods of time has resulted in species assemblages best suited, or adapted, to the specific site conditions found within each stand; therefore, the vegetation patterns should closely match the environmental patterns.
b) It follows from the preceding statement, that within parts of the study area where large scale natural disturbances (i.e. fire) are, or have been, more frequent, relationships between vegetation patterns and environmental patterns would be expected to be weaker.

## 3. VEGETATION HOMOGENEITY

Another expected characteristic of old-growth forests is homogeneity of vegetation strata within communities under relatively uniform environmental conditions. Again, a decrease in vegetation homogeneity is expected where large scale disturbances have played, or still play a role. These ideas can be reformulated as follows :

Where large scale natural disturbances (i.e. fire) are, or have been, more frequent, vegetation homogeneity will be reduced; therefore, ecologically similar sites will show a greater vegetation variability within these parts of the study area.
4. THE CLIMAX ROLE OF THUJA PLICATA IN COASTAL FORESTS

Thuja plicata is a major forest dominant of the Estevan Coastal Plain. This area, located on the extreme west coast of Vancouver Island, receives over 2000 mm of precipitation annually (Fig. 2). Despite the dominant status of Thuja plicata in this area, there is some doubt that it is the major climax species. Tsuga heterophylla and Abies amabilis both show abundant seedling regeneration and might be predicted to eventually replace Thuja plicata as dominant species. Bellefleur (1981) using a Markovian simulation model of succession (and an admittedly small data set) showed that succession should lead quickly to Tsuga heterophylla dominance. Bellefleur (1981) claimed, however, that the results were an artifact of the data set and agreed with Packee (1976) that Thuja plicata should maintain its dominance over time. Klinka et al. (1979)
include Picea sitchensis, Abies amabilis and Tsuga heterophy11a as dominant trees of their zonal ecosystem (climax association on mesic sites) in the coastal areas where Thuja plicata dominates. Nevertheless, they describe old-growth stands with very large Thuja plicata trees, which they consider to be nearly climatic climax ecosystems, owing to the virtual absence of forest fires. Since there is a concensus that the largest amount of disturbance in these stands comes from the windthrow of individual trees (Klinka et al., 1979; personal observation), the following hypothesis will be investigated :

In the old-growth, Thuja plicata dominated forests of the west coast of Vancouver Island, Thuja plicata can be considered a "climax" species, capable of regenerating and maintaining itself indefinitely.

## C. GRADIENT ANALYSIS

Gradient analysis is the major conceptual approach used to generate and analyse the results of this study. The origin, basic assumptions and premises of this approach are discussed here briefly.

Gradient analysis is a methodology of vegetation study originally developed by R.H. Whittaker in his study of the vegetation of the Great Smoky Mountains (Whittaker, 1956). He later: used the same approach in his studies of the Siskiyou Mountains of Oregon (Whittaker, 1960) and the Santa Catalina Mountains of Arizona (Whittaker and Neiring, 1965; 1968a; 1968b; 1975). Gradient analysis is based on the Gleasonian view of vegetation as a continuum (Gleason, 1926; McIntosh, 1967), and has
been largely responsible for the now general acceptance of this view (Whittaker, 1978). The approach consists of studying the variation in the structure and composition of vegetation along environmental gradients, using variables from three different levels : (1) the abiotic environment, (2) species populations, and (3) community characteristics such as diversity or productivity (Whittaker, 1967). Interrelations between these three levels of variables can be studied through the use of two-dimensional diagrams (Whittaker, 1956; 1960; 1965; 1967; 1978; Kesse1, 1979). In this particular approach, environmental gradients surmised to be important are represented as axes, and the sampled plots are arranged, or ordinated, within the reference space. This technique is now referred to as direct gradient analysis, particularly since the advent of the Wisconsin polar ordination method and other multivariate techniques based solely on vegetation data (Whittaker and Gauch, 1978). Such ordinations are referred to as methods of indirect gradient analysis (thus, indirect ordinations), since they represent diagrammatically patterns of variation in the vegetation which may be interpreted in terms of ecological gradients. It is assumed that the pattern of vegetation variation reflects underlying environmental gradients (Whittaker, 1967; 1978; Whittaker and Gauch, 1978); the strengths of such relationships can be clarified in follow-up analyses by correlating environmental variables with the derived vegetation gradients.

Only recently has gradient analysis been used with a resource management purpose in mind. Kesse1 (1979) applied gradient analysis techniques in the development of a computerized forest fire management program for Glacier National Park, Montana. This thesis uses gradient
analysis in seeking to provide ecological information necessary for forest management decisions and explores some of the theoretical aspects on which this approach is based.

CHAPTER 2.

## STUDY AREA

## A. LOCATION, PHYSIOGRAPHY AND GEOLOGY

## 1. LOCATION AND PHYSIOGRAPHY

The study area is located on the west-central coast of Vancouver Island. The area extends approximately 110 km along the Pacific Ocean coast, from the Cypre River north of Tofino ( $49^{\circ} 20^{\prime} \mathrm{N}, 126^{\circ} \mathrm{W}$ ), to the Nitinat River east of Bamfield ( $48^{\circ} 45^{\prime} \mathrm{N}, 124^{\circ} 35^{\prime} \mathrm{W}$ ). From the coast, the study area extends 60 km inland to Port Alberni (Fig. 1).

The study area lies entirely within the Vancouver Island Mountains physiographic subdivision (Holland, 1964). This unit is further subdivided into the Vancouver Island Ranges, the Estevan Coastal Plain, and the Alberni Basin.

The Vancouver Island Ranges are formed by several small mountain ranges generally following a northwest to southeast axis, separated and dissected by deep, U-shaped valleys (Holland, 1964). One of these valleys, flooded by the sea, is the Alberni Inlet, a classical fjord centrally located in the study area and opening to the Pacific Ocean through the Barkley Sound. Numerous valleys contain large, fjord-like lakes such as Sproat Lake, Nahmint Lake, Henderson Lake, the two arms of Kennedy Lake, and Great Central Lake at the northern boundary of the study area (Fig. 1). Nitinat Lake, at the southeast limit of the study area, has the peculiarity
of being linked directly to the Ocean at high tides. The highest peak within the study area is Mt. Klitsa at 1642 m . The Golden Hinde $(2,200 \mathrm{~m})$, north of the study area, is the highest mountain on Vancouver Island. Pre-Pleistocene uplift and erosion produced a rugged topography which was extensively modified during the Pleistocene glaciations (Holland, 1964). During the most recent glaciation the Vancouver Island ice cap was joined with that of the mainland (Muller et al., 1974). There was a southwest flow of ice across the Island when the general topography allowed it, such as along the Alberni Inlet (Fyles, 1963). Recent botanical discoveries suggest that the ice cover was not complete on Vancouver Island's Brooks Peninsula during the Pleistocene Fraser glaciation (Pojar, 1980). Endemic earthworms found on Vancouver Island would also support the existence of a glacial refugium (Spiers et al., 1984).

The Estevan Coastal Plain is a narrow band, 1.5 to 10 km wide, extending nearly 275 km along the west coast of Vancouver Island (Holland, 1964). This coastal plain reaches its maximum width within the study area between Tofino and Ucluelet (Fig. 1). The topography is generally level to strongly rolling with scattered bedrock knolls. Surface materials consist of thick, unconsolidated Pleistocene and Recent deposits. Drainage on these materials is generally imperfect to very poor (Valentine, 1971). In the portion of Pacific Rim National Park situated between Tofino and Ucluelet wave action on these deposits has created long and wide sand beaches.

The Alberni Basin is a low elevation area (below 300 m ) with relatively level relief at the head of the Alberni Inlet (Holland, 1964). No plots were sampled in this physiographic section since it is mostly agricultural.

There are numerous rivers within the study area. The drainage basins of some of the largest rivers (Kennedy, Taylor, Nitinat, Sarita, Klanawa, and Nahmint) were used to subdivide the study area for sampling purposes (Fig. 3).

## 2. BEDROCK GEOLOGY

The bedrock geology of southwestern British Columbia including Vancouver Island is complex. Several authors have described the heterogeneous geology of the area (Muller, 1971; Muller and Carson, 1969; Muller et al., 1974) and a detailed summary is provided by Packee (1976). Rocks'of the Mesozoïc era predominate. These are mainly faulted and folded sedimentary and volcanic rocks, frequently intruded by igneous batholiths (Muller and Carson, 1969; Muller, 1971). Limestone, chert, argillite, tuff, and greywacke are the most common types of sedimentary rock (Day et al., 1959; Muller and Carson, 1969). Three cycles of volcanism have been described for Vancouver Island (Northcote, 1973). Recent geological discoveries indicate that Vancouver Island and the Wrangell Mountains, near the coastal Yukon-Alaska border, may have drifted north from south of the equator through plate tectonic activity (Jones et al., 1983).

## 3. SURFICIAL GEOLOGY

The main factors influencing the surficial geology of the study area have been the last Pleistocene glaciation, which ended approximately 12,000 to 14,000 years ago according to palynological evidence (Hebda, 1983; Mathewes, 1973), and various post-glacial events. The major types of surficial materials found on the west coast of Vancouver Island are glacial tills, glaciofluvial deposits, and marine sediments. Because of the mountainous topography, colluvial material is frequently encountered (Jungen and Lewis, 1978). The colluvium is formed on slopes from bedrock fragments and slumping morainal material. Marine and fluvioglacial deposits, in the form of sands and clays, are predominant on the Estevan Coastal Plain (Valentine, 1971; Jungen and Lewis, 1978). These deposits originated when the land, depressed by the ice pack, was invaded by the sea following the glacial retreat; rebounding of the land has now raised these sediments above sea level. Recent alluvial deposits are found along all major rivers.

## B. SOILS

British Columbia has been divided into a number of broad soil landscapes defined at the soil great group level (C.S.S.C., 1978). Each is defined as "the total ecosystem with which a soil is associated, with emphasis placed on the soil itself" (Valentine et al., 1978). The study area falls within the Ferro-Humic Podzol and the Humo-Ferric Podzol soil landscapes (Jungen and Lewis, 1978). The Ferro-Humic Podzol soil landscape occurs on the windward side of Vancouver Island. This area is characterized by abundant rainfall and moist, rarely frozen soils throughout most of the year (Jungen and Lewis, 1978). The main soil formation processes are the accumulation of organic matter, iron, and aluminum producing soil with distinct podzolic $B f$ horizons. Continuous seepage is present throughout most of this soil landscape, and is reflected by the high organic matter content of the soils, rather than the typical mottling and gleying (Jungen and Lewis, 1978). Organic matter content often reaches a maximum, sometimes over $30 \%$, near the lower part of the profile (Jungen and Lewis, 1978). The presence of a cemented, indurated pan (Bc horizon) is the major characteristic of morainal soils (Jungen and Lewis, 1978). In these soils the Bhf horizon commonly is most pronounced just above the cemented till which often cannot be broken with a shovel. Morainal soils are mostly imperfectly to poorly drained while colluvial soils, with no cementation, are generally well to moderately well drained (Jungen and Lewis, 1978). Organic horizons between 20 to 40 cm thick frequently were observed on Ferro-Humic Podzols within the study area. Valentine (1971) noted the high organic matter content of
the soil surface layers of the Tofino-Ucluelet lowland. He suggested that dense vegetation coupled with moderate temperatures, allowing nearly continuous biological and chemical activity, forms a "constant source of raw humic material". Most plant nutrients may be derived from this organic matter (in organic horizons and upper mineral horizons) rather than from the mineral solum (Valentine, 1971). Valentine (1971) also noted a generally shallow rooting zone even under dense tree growth. The importance of the organic layers in nutrient cycling in soils of the west coast of Vancouver Island is also supported through the recent discovery of an endemic earthworm (Arctiostrotus simplicigaster vancouverensis) (Spiers et al., 1984). This worm may play a major role in mediating nutrient cycling within the organic layers where it is restricted (Spiers et al., 1984). Chemically the soils of the Ferro-Humic Podzol soil landscape have a very low base saturation, low pH (commonly $<5.0$ ), and high organic carbon, iron, and aluminum contents (Jungen and Lewis, 1978).

The Humo-Ferric Podzol soil landscape occurs farther inland within the study area, especially around Port Alberni. Soil moisture is not as abundant as in the Ferro-Humic Podzol soil landscape owing to the warmer and drier summer climate (Jungen and Lewis, 1978). Colluvial and morainal parent materials are common, the latter usually with a weakly to strongly cemented pan ( $B C$ and $B C c$ horizons). Cementation, when present, is usually strongest near the top of the pan (Jungen and Lewis, 1978). Colluvial soils often are deeply weathered, well to rapidly drained, and contain no signs of cementation (Jungen and Lewis, 1978). Chemically these soils have low pH (4.0 to 5.0 ), moderate to high iron and aluminum content,
and low base saturation (Jungen and Lewis, 1978). In contrast with the soils of the Ferro-Humic Podzol soil landscape, Humo-Ferric Podzol soils have little organic matter accumulation in the upper B horizons (Valentine and Lavkulich, 1978).

Apart from podzols, other soil orders are also encountered within the study area. Folisols, consisting of shallow organic material overlying bedrock, are sometimes found on rock outcrops in high rainfall areas near the coast. Orthic regosols occur on floodplains and on rock outcrops. Gleyed Sombric Brunisols frequently are found on floodplains. Humic Gleysols are most frequent in plots of the Estevan Coastal Plain. Some Dystric Brunisols also occur in the drier inland part of the study area. Other than Folisols, organic soils occasionally are found in bedrock depressions and in areas overlying impervious surficial material; examples of the latter include the bogs on marine clays in the TofinoUcluelet area (Valentine, 1971).

The typical Vancouver Island podzol was reported by Lewis (1976) to be different from the classic podzolic profile. Frequently, no eluviated Ae horizon is found. Despite the strong leaching, the constant addition of abundant organic matter and the constant weathering of iron and aluminum in the upper mineral horizon prevent the net depletion necessary to form an Ae horizon (Lewis, 1976; Valentine and Lavkulich, 1978) ; however, it was also reported that an accumulation of organic matter may sometimes mask the Ae horizon under moist field conditions (Valentine and Lavkulich, 1978). Soils derived from basaltic and andesitic parent materials have no Ae horizons because they contain no silica
to be left behind after weathering (Lewis, 1976). Throughout the study area soil horizon boundaries frequently are very irregular because of the turbic activity associated with windthrow.

Most soils sampled, especially those on colluvial material, were coarse textured and contained a high percentage of large rock fragments. Overall, the pH values of organic horizons varied from 3.0 to $6.2\left(\mathrm{H}_{2} \mathrm{O}\right)$, and the pH values of the upper mineral horizons (mostly $\mathrm{B}_{1}$ ) from 3.8 to $6.3\left(\mathrm{H}_{2} \mathrm{O}\right)$. Total nitrogen in the upper mineral horizons varied from $0.02 \%$ to $0.86 \%$, total carbon from $0.2 \%$ to $24.4 \%$, and C/N ratios from 10 to 94.

## C. CLIMATE

Climatic data are available from several low elevation permanent weather stations within the study area (Anonymous, 1982); high elevation stations are lacking. Most of the study area falls within the humid mesothermal summer-wet climate (Cfb) according to the Köppen system. This is described by Strahler (1965) as a windward, west coast climate with moist maritime polar air masses hitting the coast with frequent eastward-moving cyclonic storms. Precipitation tends to be evenly distributed throughout the year, but there is a winter maximum. The annual temperature range is small for middle latitudes (Strahler, 1965). The air masses collect moisture as they pass over the warm Alaska current and release it as orographic precipitation over the land. A distinctive rain shadow effect is created on leeward mountain slopes and valleys. The driest part of the Port Alberni area as well as all of the southeast coast of Vancouver Island can be classified as a humid mesothermal, summer-dry climate (Csb). This summer-dry, winter-wet climate, predominant farther south along the Pacific Coast, is caused by the replacement of cyclonic, moist maritime polar air masses (Aleutian Low) by a relatively stable, dry maritime tropical air mass (North Pacific High) during the summer (Strahler, 1965). Southern British Columbia is at the northernmost limit of the influence of this system, and rainfall differences between the west and east coasts of Vancouver Island are in a large part due to orographic effects.

Climatic data from Tofino Airport (Fig. 2) are characteristic of the Cfb climate within the study area. Precipitation averages 3288 mm annually, but windward slopes east of Tofino likely receive more. Abundant moisture is always available for plant growth. Dry mineral soil or humus was never observed in summer near the coast, except sometimes on rock outcrops. The temperature is very mild, with the mean daily minimum of the coldest month at $0.8^{\circ} \mathrm{C}$, and the mean daily maximum of the warmest month at $18.3^{\circ} \mathrm{C}$; the mean annual temperature is $8.9^{\circ} \mathrm{C}$ (Fig. 2). The Lupsi Cupsi climatic station near Port Alberni (Fig. 2) is at the wetter limits of a Csb climate. Mean annual precipitation is 1929 mm , and a period of moisture deficit is experienced in mid-summer when the average monthly precipitation reaches 28 mm (Fig. 2). The temperature also is very mild, although a slight continentality effect is noticeable with a lower mean daily minimum temperature of the coldest month ( $-1.1^{\circ} \mathrm{C}$ ), and a higher mean daily maximum temperature of the warmest month $\left(24.6^{\circ} \mathrm{C}\right)$, than at Tofino. The mean annual temperature at Port Alberni is $9.5^{\circ} \mathrm{C}$. Snowfall makes up $5 \%$ of the mean annual precipitation at Port Alberni and less than $2 \%$ at Tofino (Anonymous, 1982). Orloci (1964) considered snow duration and accumulation to be an insignificant ecological factor at low elevation. In contrast, at higher elevations, cooler temperatures (Dfc, microthermal snowy climate) result in a larger percentage of precipitation in the form of snow. Hollyburn Ridge ( 951 m ) near Vancouver receives close to 3000 mm of precipitation annually, of which $28 \%$ falls as snow (Brooke et al., 1970). Heavy snowpacks of moist snow often linger into mid-summer above 1000 m . Snow depths averaged 3 m over several years on April $1^{\text {st }}$ surveys of the north shore mountains near Vancouver (Brooke et al., 1970).

The occurrence of summer fog, particularly in areas nearest to the coast, is another important climatic factor within the study area. Summer fog is formed off the coast of Vancouver Island and moves inland towards a low pressure area created by the daily warming of the land mass. This fog usually covers the Estevan Coastal Plain up tothe first mountain slopes. Azevedo and Morgan (1974) have described a similar phenomenon for north-coastal California. Their data show the predominance of fog at night, dissipating during the day. In the study area, as in northern California, fog could last all day during particularly heavy episodes, and the fog would dissipate last nearest to the coast. Fog of ten appeared in the Tofino-Ucluelet area during summer days which were warm and clear for the rest of the study area. Fog drifting through forest canopies has been shown to cause considerable amounts of precipitation as fog drip (Azevedo and Morgan, 1974). A large portion of this precipitation is probably unrecorded by standard weather stations, but the vegetation should certainly reflect the prevalence of summer fog (Azevedo and Morgan, 1974).

Climatic maps compiled by Colidago (1980) for southern Vancouver Island reveal a complex pattern of decreasing summer precipitation and increasing effective growing degree-days as distance from the coast increases (Fig. 12). The "freeze-free" period (mean daily temperature $>0^{\circ} \mathrm{C}$ ) varies from 240 days at low elevation on the coast near Tofino, to 160 days inland near Port Alberni (Colidago, 1980).

## D. VEGETATION

Most of the study area falls within the Coastal Western Hemlock biogeoclimatic zone of British Columbia (Krajina, 1965; 1969). The vegetation of a small area surrounding Port Alberni has been placed within the wetter subzone of the Coastal Douglas-fir biogeoclimatic zone (Klinka et al., 1979). Klinka et al. (1979) recognize several subzones and variants of these two biogeoclimatic zones within the study area. Subalpine forests, generally above 1000 m elevation, belong to the Mountain Hemlock biogeoclimatic zone (Krajina, 1969; Klinka et al., 1979), or to the Coastal section (SA.3) of the Subalpine forest region (Rowe, 1972).

The lower elevations of the study: area are within the Coast forest region of Rowe (1972). The drier Port Alberni area supports vegetation similar to the Strait of Georgia section (C.1) through the presence of Arbutus menziesii, the only broadleaf evergreen tree in Canada, and the dominance of Pseudotsuga menziesii in the landscape (Rowe, 1972). The adjacent Southern Pacific Coast section (C.2) contains most of the study area and is characterized by stands, often even-aged, dominated in decreasing order by Pseudotsuga menziesii, Tsuga heterophylla, and Thuja plicata on well drained sites. On valley floors or on moist, sheltered slopes Pseudotsuga menziesii is sometimes absent while Tsuga heterophy11a, Thuja plicata, and Abies amabilis increase in importance. These differences indicate, the essentially seral character of Pseudotsuga menziesii in this section (Rowe, 1972). Thuja plicata dominance is associated with seepage areas, while Abies amabilis increases in
abundance with increasing elevation (Rowe, 1972). The Estevan Maritime Coastal Western Hemlock biogeoclimatic variant of Klinka et al. (1979) corresponds approximately to the Vancouver Island portion of Rowe's (1972) Northern Pacific Coast section (C.3). In this section, Tsuga heterophylla and Abies amabilis dominate on rarely occurring well drained sites, while Thuja plicata becomes dominant everywhere else where drainage is deficient (Rowe, 1972). Picea sitchensis is found mostly on alluvial deposits and on the coastal fringe (Cordes, 1972). Because of the very humid climate, forest fires are rare within this section and the major source of forest disturbance is wind (Rowe, 1972; Klinka et al., 1979). Pseudotsuga menziesii is virtually absent within the section (Rowe, 1972). The most productive stands are produced following windthrow (Rowe, 1972; Klinka et al., 1979). Some very productive stands also were observed on ancient avalanche colluvium within this section of the study area.

The Coastal Western Hemlock biogeoclimatic zone (Krajina, 1969), or the Southern Pacific Coast section of the Coastal forest region (Rowe, 1972), have growing conditions suitable for the highest forest productivity in Canada. In some sites of the Drier Coastal Western Hemlock biogeoclimatic subzone, Pseudotsuga menziesii reaches the maximum growth attained by any tree on any site in Canada (Site Index 100 : 54-60 m) (Krajina, 1969).

The vascular floristic patterns and affinities of coastal British Columbia are discussed by Schofield (1969) and Scoggan (1978). The bryoflora has been analysed in more detail by Schofield (1965, 1968a, 1968b,

1969, 1976, 1980, 1984). Many dominant taxa within the study area are restricted to the relatively narrow Coastal or Cordilleran area along western North America. Several Pacific North American taxa such as Arbutus menziesii, Arctostaphylos columbiana, and Oxalis oregana reach the northern extent of their ranges near the study area.

CHAPTER 3.

## METHODS

## A. DATA COLLECTION

## 1. LOCATION OF PLOTS

The selection of sampling sites over a large, mountainous and heterogeneous area, for the purpose of gradient analysis and ordination, requires a minimum of bias, adequate representation of the range of variation in environment and community composition, homogeneity within sampling units, lack of disturbance, and a sufficiently large sample (Whittaker, 1978). Because of the difficulty of selecting plots without bias, random sampling is often recommended for vegetation studies (Smartt and Grainger, 1974). However, formal randomization in large scale studies has been rejected by most researchers (Moore et al., 1970), with some exceptions (Noy-Meir, 1971), because of drawbacks such as inefficiency (for time and sample size) and inadequate representation of variation ranges, because of the high probability of missing many unusual, and often very informative, communities (Whittaker, 1978; Peet, 1981). The random location of plots in the field may be time consuming and yield many unsatisfactory sites (because of heterogeneity or disturbance, especially in an area with active logging such as in this study). Subjective sampling can yield a much broader spectrum of vegetational variation in the same amount of time (Peet, 1981), and the time saved in plot location will permit the collection of a larger sample. In order to make the plot location selection as objective as possible, the study area was subdivided into thirteen
drainage areas (Fig. 3). The number of plots sampled in each area depended on the size of the area and on accessibility. Some drainage areas with very difficult access were not sampled. Within each selected drainage area an effort was made to sample examples of all topographic positions (slopes, ridges, floodplains, etc.) and edaphic conditions present, up to an elevation of about 1000 m , reportedly the lower limit of the Mountain Hemlock biogeoclimatic zone (Brooke et al., 1970; Klinka et al., 1979). Also, only homogeneous (within plot boundaries) old-growth forests with no evidence of major disturbance within the last hundred years were sampled. The maximum ages of stands sampled varied from 150 years to well over 500 years, with a few exceptions. Coastal fringe communities of Picea sitchensis, influenced by ocean spray, and Sphagnum bogs, on deep organic soils, were not sampled. These special plant communities have been studied by Cordes (1972) and Wade (1965), respectively. Sand dune vegetation of Long Beach, Pacific Rim National Park, was studied by Kuramoto (1965).

## 2. VEGETATION SAMPLING

The vegetation was sampled within a circular $500 \mathrm{~m}^{2}$ plot (Pfister and Arno, 1980) at each site. From a centre-point, two tapes were laid out at 90 degrees, and a radius distance of 12.6 m was flagged around the periphery of the plot, using calibrated ropes (Fig. 4). The diameters at breast height ( 1.3 m ) of all stems within the plot were recorded for each species in 10 cm size-classes. Stems over 10 cm DBH are referred to as trees, and stems between 0 and 10 cm DBH are referred to as saplings.

The understory vegetation and tree seedlings (arbitrarily defined as stems less than 1.3 m tall) were recorded in twenty $1 \mathrm{~m}^{2}$ microplots. Two different microplot placement designs were used : a systematic design for plots 1 to 61 (1980) and a stratified random design for plots 62 to 172 (1981) (Fig. 4). Randomization avoided the sampling bias toward the centre of the plot, inherent in the systematic design, and yielded data that were more amenable to statistical interpretation. The stratified random design was obtained by determining from a table of random numbers five microplot locations on a grid of each quarter of the plot surface. This design was repeated in the sampling of plots 62 to 172. Percent coverage was estimated for shrubs, herbs, bryophytes and lichens, in each microplot, using a seven-point scale of coverage ranges similar to that of Daubenmire (1968) : 1 ( $0-1 \%$ ), 2 (1-5 \%), 3 ( $5-25 \%$ ), 4 (25-50 \%), 5 ( $50-75 \%$ ), $6(75-100 \%$ ) and 7 (100 \%). Also, the numbers of tree seedlings were recorded by species within the microplots. Vascular species not encountered within the microplots but found within the larger $500 \mathrm{~m}^{2} \mathrm{plot}$ were recorded as present, and arbitrarily assigned values of 0.01 percent coverage and $1.0 \%$ frequency; non-vascular species outside the microplots were not recorded.

These measurements provide for each plot : basal area (or dominance) of trees, density of trees, density of saplings, density of seedlings, and percent coverage (average of 20 microplots), and frequency (over 20 microplots) for shrubs, herbs, bryophytes and lichens. The heights of at least two dominant trees were measured in each plot using a clinometer, and the maximum height of the shrub and herb strata recorded. Cores of two of the largest trees, of different species, were taken for stand age estimates.

## 3. SOIL AND ENVIRONMENTAL DATA

Within each $500 \mathrm{~m}^{2}$ plot a soil pit was dug to bedrock, to a layer of compacted till, or to a depth of one metre, whichever came first. A soil profile description was written in the field and samples of each organic and mineral horizon were taken for laboratory analyses. The field description included features such as, horizon thickness, percent coarse fragments, field texture (estimated), structure, consistency, charcoal presence, colour, abundance and size of roots, and organic material description. Other site and soil data such as elevation, aspect, percent slope, topographic position, surface shape, soil drainage, estimated soil moisture regime, nature of surficial material, nature of bedrock, evidence of fire and windfall, and presence of earthworms (plots 62 to 172) were also recorded. The distance of each plot from the Pacific Ocean was determined from a map.

The soil analyses were performed by the MacMillan Bloedel Woodlands Services Soil Laboratory, generally following the U.B.C. Pedology Methods Manual (Lavkulich, 1978). All samples were air dried. After drying, organic samples were ground in a Wiley mill to pass through a 20-mesh screen, and mineral samples were passed through a 2 mm sieve and ground to pass a 60 -mesh screen. The pH 's of all samples were determined in both a $1: 1$ water suspension ( $1: 2$ for organics) and $1: 20.01 \mathrm{M} \mathrm{CaCl}_{2}$ suspension (1:4 for organics). Soil texture was determined by the hydrometer method for the top $B$ horizon of plots 1 to 61 . Total organic carbon content was determined by the Walkley-Black method of Wet Oxidation. The total nitrogen content of samples was determined using a Technicon Auto

Analyser II after digestion in sulphuric acid and catalysts (mineral samples), or after digestion in $30 \%$ hydrogen peroxide and sulphuric acid (organic samples). The determinations of pH , total carbon (\%) and total nitrogen (\%) were chosen because these soil properties show the least within-site variability and are therefore more reliable when only one sample per site is taken (Quesnel and Lavkulich, 1980).

## 4. NOMENCLATURE

The taxonomic nomenclature of this study generally follows Scoggan (1978-1979) for vascular plants, Ireland et al. (1980) for mosses, Stotler and Crandall-Stotler (1977) for liverworts, and Hale and Culberson (1970) for lichens.

In a few cases the names used in this study are listed as synonyms by the taxonomic sources. Voucher specimens for most vascular plants and all non-vascular plants are deposited at the University of British Columbia's Botany Department Herbarium (UBC).

## B. DATA ANALYSIS

1. GRADIENT ANALYSIS AND ORDINATIONS
a) Indirect gradient analysis and ordinations

Gradient analysis is an approach to the study of vegetation that seeks to explain the spatial distribution and variation of vegetation in terms of three sets of variables, (1) environmental factors, (2) species populations and (3) community characteristics (Whittaker, 1967). This approach is based on the view of vegetation as a continuum (Gleason, 1926; McIntosh, 1967; Whittaker, 1967) where "vegetation is considered as a continuously varying, stochastic phenomenon wherein plants respond individualistically to environmental conditions" (Peet, 1981). Indirect gradient analysis, or indirect ordination (Whittaker, 1978), is a technique which attemps to identify major environmental gradients underlying the patterns of vegetation variation. Such patterns are graphically illustrated by ordinations of plots obtained by analysing data on species composition. Thus, an important assumption of indirect ordination is that trends in environmental gradients will be reflected by trends in vegetation variation (Whittaker, 1978). Reciprocal averaging (Hill 1973, 1974), a type of standardized principal components analysis was the ordination technique used in this study. In tests, Gauch et al. (1977) have shown it to be one of the best available techniques in exposing environmental gradients using vegetation data where these gradients were already known. Although distortion can present a problem in axis scaling, the method reliably yields a primary axis of variation which is ecologically
interpretable. When the primary axis of variation corresponds to a particularly strong environmental gradient (Fig. 10), the second axis is often correlated with the first, causing an "arch effect" (Gauch et al., 1977). Experience with this study's and other data sets indicates that the ecological significance of the second axis diminishes with increasing strength of the primary environmental gradient, usually identified by the ordination's first axis (Gauch et al., 1977; Peet, 1980). In such cases the third axis often represents more accurately a second major environmental gradient, while the percentages of variation explained by the second and third axes are nearly equal. Detrended correspondance analysis, a recent modification of reciprocal averaging, is reported to overcome this problem (Hill and Gauch, 1980). Random species fluctuations create noise in community data. Gauch (1982) estimates this noise to be on the order of 10 to $50 \%$ of the total variance in the data. Simulation experiments have shown that eigenvector ordination selectively recover meaningful patterns of correlation among several species in the first few ordination axes, while selectively deferring noise to later axes (Gauch, 1982). This would help to explain the observation that, in general, ordinations of field data are frequently useful even when the percentage of variance explained by the first few axes is small (Gauch, 1982). This also explains the common observation that meaningful ecological interpretations of ordinations axes are difficult past the second or third axis, with some exceptions (Noy-Meir, 1971). The goal of ordination has been viewed as accounting for most of the original data variance in the fewest ordination axes, but field data usually contains $\mathrm{x} \%$ "noise variance" and (100-x) \% "structure variance" (Gauch, 1982). Thus the
goal should be to recover (100-x) \% of the variance, preferably only the structure variance to the exclusion of noise variance (Gauch, 1982). Since noise variance has been estimated to be from 10 to $50 \%$, recovery of $100 \%$ of the variance implies that the ordination has failed in noise reduction. A major difficulty with this viewpoint is in deciding what is "structure variance" and what is "noise variance", anything which can be interpreted becomes "structure variance" and whatever cannot be interreted becomes "noise variance". Principal components analysis ordinations based on species covariance and correlation matrices were also tried, but did not produce superior results to reciprocal averaging. An advantage of reciprocal averaging is that it simultaneously produces a species ordination which can be superimposed on the sample ordination (Greenacre, 1981). This can be very helpful in displaying vegetation trends as characterized by major species. The Wisconsin double standardization of data, sometimes recommended for use with RA (Peet, 1981), was not done since the program used included a form of double standardization in the calculation of resemblance coefficients. Species present in less than four or three plots, depending on matrix size, were removed for the analyses. Rare species contribute little information to overall plot similarities, and often cause the plots containing them to be markedly isolated in. reciprocal averaging ordinations. The vegetation data analysed in this study are based on relative importance values of trees (> 10 cm DBH), relative density of saplings ( $0-10 \mathrm{~cm}$ DBH) and seedlings (below breast height), and percent coverage of shrubs, herbs, bryophytes and terricolous lichens. Before this combination of abundance measures and strata was chosen, several trial ordinations were run. Trees were
ordinated alone using three different abundance measures, which, in decreasing order of ordination interpretability they provided, were rated as follows : importance value > relative dominance > relative density. Relative importance value ([relative dominance + relative density $] / 2$ ) is appropriate when tree species occur in a wide range of maximum sizes and densities. In this study, relative dominance overemphasized the importance of a few very large trees, such as Thuja plicata and Pseudotsuga menziesii, while smaller, of ten more numerous, trees, such as Tsuga heterophylla and Abies amabilis, were underrated. Relative density created the reverse problem. Relative density was selected instead of absolute density for saplings and seedings since, especially for seedlings, absolute density values varied enormously between plots. The use of different size-classes of trees (trees, saplings, seedings) offers potential for regrouping samples with similar regeneration trends; thus, differences in canopy dominants, which may have arisen through different disturbance regimes in the past, are offset by similar patterns of regeneration in the understory. Several authors have used this technique for similar purposes (Goff and Zedler, 1972; Peet and Loucks, 1977; Carleton and Maycock, 1978). The different sizeclasses of a tree species are treated as different "pseudo-species" (Carleton and Maycock, 1980) for the purpose of the ordinations. A seedling size-class was used here despite the "highly stochastic nature of establishment and survival (of seedlings) for the first few-years" (Peet and Louck, 1977). Although this was observed in the widely fluctuating absolute densities of seedlings among plots, it was felt that the relative density of seedlings of a particular species, with differences
in reproductive potential and life history patterns taken into account, remains a good indicator of that species' potential role in the future composition of the stand, as well as a good indicator of present ecological conditions. Comparisons of ordinations obtained with and without tree size-class data have shown that better results are obtained using this technique where (1) dominant tree species are numerous, (2) regenerating tree species are few, (3) successional stands are common, and (4) environmental diversity of the study area is small. These conditions were met in an earlier study (Gagnon and Bouchard, 1981). In the present study however, the canopy dominants are few and nearly all are regenerating in some stands. Futhermore, most stands are in late stages of development (although many are dominated by $\underline{P}$. menziesii, a long-lived successional species), and environmental diversity is great. Despite these drawbacks, the use of tree size-classes provided a clearer separation of some ecologically different communities with similar canopies (eg. montane vs lowland Abies forests). The most interpretable ordination results were obtained using data from shrub, herb and bryophytelichen strata along with tree data separated into three size-classes. Peet (1981), studying the vegetation of the Colorado Front Range, and Beese (1981), the vegetation of eastern Vancouver Island, have reported greater success with ordinations of understory data on1y. In both areas the tree layer was not considered the ideal site indicator since it largely reflected past disturbances. To a certain extent this was also the case in this study, but the partitioning of tree data into sizeclasses greatly improved the ordinations.

## b) Successive ordinations

Plots that differ markedly in composition from the majority of plots are usually placed toward the ends of ordination axes. Evidence from tests (Gauch et al.,"1977) and personal experience indicates that reciprocal averaging is particularly sensitive to outlier plots. Outliers are defined as plots of unusual composition relative to the majority of plots in the sample (Gauch et al., 1977). More specifically, outliers may have (a) unusual combinations of species importances, (b) one or a few species dominating strongly, or (c) several species which are uncommon and unimportant elsewhere within the matrix. Outliers of type "a" and "b" are sometimes caused by sampling error, or by the sampling of disturbed or environmentally unusual sites, and are problematic in the interpretation of ordinations (Gauch et al., 1977). Type "c" outliers can be used to advantage in ordination interpretation. In large or complex data sets successive ordinations can permit the segregation of groups or types of communities at the periphery of the ordination field. This progressive fragmentation of the data set superficially resembles classification, but the objective is to understand the environmental relationships between groups of similar plots (Peet, 1980). Ordination is thus used as a classification tool in which distinctive groups are removed successively from the data after the resulting patterns have been examined for environmental correlations. If large enough, the groups removed may also be ordinated to reveal within group patterns and environmental correlations. In this study, from an initial ordination of the total 172 plots sample, three environmentally distinct groups of communities, plus a vegetationally distinct community type (P1), were
segregated from a central cloud of plots. An ordination of the remaining p 1 lots indicated that they could be partitioned again into three environmentally and geographically distinct groups. Ordinations of each of these latter groups allowed several community types to be delineated along distinct environmental gradients. Product moment correlations were calculated between sample scores on ordination axes, environmental variables (Table 1), and community characteristics (Table 2) to help identify gradients underlying the vegetation patterns illustrated by the ordinations. Lack of strong correlations with any single variable may indicate that "complex" environmental gradients (Whittaker, 1978) control the variation of the vegetation. A topographic-moisture gradient is "complex" in the sense that it combines the effects of slope and aspect, as well as topographic position, soil texture and drainage. Thus, complex master environmental gradients might show strong correlations with ordination axes, if they could be expressed quantitatively.

A further ordination analysis was done in order to identify major environmental gradients influencing the vegetation pattern at the level of the entire study area, without the noise introduced by edaphic variations. For this purpose, 105 vegetation plots of modal sites were analysed with a reciprocal averaging ordination. Excluded from this analysis were plots from high elevations, lower.slopes of steep river valleys (cold air drainage or snow accumulation), rock outcrops, very rapidly and very poorly drained sites, and floodplains. Correlations of ordination axes with environmental variables and community characteristics were used to identify the environmental gradients underlying the modal vegetation pattern. The reciprocal averaging and principal components
analysis programs used were developed by Dr. G.E. Bradfield following Orlóci (1978). Product moment correlations were produced using the MIDAS statistical package supported by the University of British Columbia Computing Centre.
c) Direct gradient analysis

Direct gradient analysis, or direct ordination, refers to the arrangement of plots along one or more known, or accepted as given, environmental gradients (Whittaker, 1967, 1978). These gradients may be derived empirically, surmised from observation, or identified through correlation of environmental variables with indirect ordination axes. Direct ordinations were used to display the spatial distribution of communities along topographic-moisture and elevation gradients within three fairly homogeneous vegetation groups. The topographic-moisture gradient used here is similar to gradients utilised by Whittaker (1956), Whittaker and Neiring (1965) and Peet (1981), but particularly resembles that used by Whittaker (1960) in his study of the Siskiyou Mountains of Oregon. The mesic, or moist, end of the gradient is represented by stands found on level, or near level, ground, proceeding to stands found on lower-slopes, where moisture is provided by seepage, but where drainage is better than on level ground. Further towards the xeric or dry end of the gradient are located stands from mid-slopes and upper-slopes. The xeric endpoint is formed of stands situated on crests, ridges or dry summits. Stands situated on sloping ground are arranged in two categories, (a) lower-slopes and (b) mid-slopes and upper-slopes, according to their
aspect. The lower-slope range is shortened because aspect effects are not as important for these stands with increased shelter and moisture availability. The two slope ranges do not overlap as in Whittaker (1960). One Pinus contorta type was included with the Pseudotsuga group, and one with the Thuja group, in the direct ordination figures because of their floristic and geographical affinities with these groups.

The plotting of species abundance or community characteristics along environmental gradients is a variation of direct gradient analysis. In this study geographical coordinates were used as complex environmental gradients combining variations in temperature, precipitation and continentality. This approach is helpful in identifying relationships of vegetation and soils with geographical patterns. This was particularly useful for data obtained from other sources, such as climatological data. In another type of application, the basal area data of tree species from the 105 modal plots were plotted against a geographical gradient defined by the distances of these plots from the Ocean. The SPSS polynomial regression program was used to obtain equations describing the basal area distribution of tree species along this gradient.

## 2. TYPE DELIMITATION

## a) Definition of groups and types

Ordination techniques were used to assist in the classification of the sample plots, first into vegetation groups, and then into community types. Since the classification involved partitioning a continuum, as the ordinations visually illustrate, the groups and types may intergrade and overlap. Thus, it is not classification in a hierarchical sense, but classification in the sense of typification (Noy-Meir and Whittaker, 1978). Vegetation groups are defined as groups of plots that show a general degree of similarity in dominant species and environmental characteristics. Community types are subdivisions of vegetation groups and are defined as assemblages of plots that show a high degree of similarity inispecies composition and abundance, as well as in environmental characteristics. Subdivision into groups or types of the largely continuous pattern seen in the ordinations, was done using the following criteria in order of importance : (1) discontinuities in the ordination scatter diagrams when present, (2) careful inspection of the vegetation data for compositional similarity, (3) similar inspection of the environmental data. Where boundaries between types were drawn, some subjectivity was involved as in any classification. Averages, or noda of the community types are distinct vegetationally, and most are also distinct environmentally (see canonical analyses). Some plots could not be classified and are indicated by single dots in the ordinations. These plots either were unusual compositionally because of edaphic factors or disturbance, or were representative of other, undersampled, communities. Groups and types of communities as defined here do not correspond to any
particular traditional classification system; they are used solely for the purpose of describing useful subdivisions of an otherwise fairly continuous pattern of vegetation variation. The relationships between types is well illustrated by the ordinations, and some could very well be considered as sub-units or variants of other types. Relationships of types are discussed, but no formal hierarchical arrangement of types was attempted.

The community types are similar to dominance-types (Whittaker, 1978) since they are defined primarily on the high similarity of their dominant species. The community types are also similar to habitat-types ${ }^{1}$, defined by Daubenmire (1968) as the "potential climax vegetation" of a site. Daubenmire (1968) views the habitat (soil, macroclimate, topography) as the most durable part of an ecosystem, eventually controlling the final aspect of the vegetation. Successional or preclimax vegetation can take a more varied appearance, on otherwise similar sites, due to various disturbances. Tree regeneration and understory strata (shrubs, herbs and bryophytes) are most important in defining habitat-types, since they are established soon: after disturbance and are likely to persist into the "climax" stage. However, important changes in understory plants have been demonstrated by Alaback (1982) in south-east Alaska forests during later successional stages. The use of sapling and seedling data in the ordinations from which the types were derived allows them to be considered near equivalents of habitattypes, particularly since most plots come from old-growth stands. Habitattypes are usually named by a combination of one, or two, potentially dominant species (climax species) as well as an understory dominant

[^0]
#### Abstract

(Daubenmire \& Daubenmire, 1968). Community types are named informally in this study according to the dominant tree species and, when necessary for differentiation, characteristic understory species are used. General ecological and geographical qualifiers are sometimes added to the names. Community types were also coded by a letter to identify the vegetation group and a number to identify the community type.


b) Vegetation datá summary tables

The presentation of one or a few typical stands per type does not represent the full range of variation encountered, while a large number of stands may obscure underlying patterns (Peet, 1981). As an alternative, the data from each community type were averaged and constancy values, defined as the percent occurrence of species in the sample plots of a type, were calculated. In order to keep table length to a minimum, species had to have $50 \%$ constancy, or more, in at least one of the types represented in the table to be included (or $100 \%$ when the type had only two plots). Of the two Pinus contorta types, one was placed in the Pseudotsuga group and the other in the Thuja group tables, according to their compositional and geographical affinities. Floodplain types were included in the tables of the less diverse Thuja group. Community data summary tables were divided in two, with a table for tree, sapling and seedling data for each group, and a table for understory data for each group. The tables also include community characteristics data, such as mean number of species (species richness or density) and total number of species for trees, shrubs, herbs and bryophytes-lichens. Mean basal area, mean density and mean maximum height are given for trees. Mean
total coverage is given for shrubs, herbs, bryophytes-1ichens and total understory. Two diversity indices, the reciprocal of Simpson's Index (dominance concentration) and the antilog of the Shannon-Weaver Index (equitability) (Peet, 1974), were calculated for the trees ( $>10 \mathrm{~cm} \mathrm{DBH}$ ), the understory vascular plants (shrubs and herbs), and for the bryophytes and lichens. Within the Pseudotsuga group tables the types were arranged, from left to right, in order of increasing soil moisture (except P7 which is drier than P6) and increasing elevation (eg. P3 is drier than P2 but occurs at higher elevations). The Thuja group tables were arranged in order of increasing soil moisture and decreasing drainage (except the F1 type which is moderately well drained). The Abies group tables were organized in order of increasing soil moisture, for high elevation types up to type A4, and in order of decreasing soil moisture for low elevation types (A5 to A7).

## 3. CANONICAL ANALYSES OF COMMUNITY TYPES AND VEGETATION GROUPS BASED ON ENVIRONMENTAL DATA

Canonical variates analysis was used to examine relationships among the vegetation groups and community types, delineated in the ordinations, on the basis of the environmental data. The environmental variables used are listed in Table 1 (distance from the coast, a geographical variable, was not used, as well as pH for A and $\mathrm{B}_{2}$ horizons, which were missing from numerous soil profiles). Canonical analysis accentuates differences among preestablished groups (Seal, 1964), and was used to assess the degree of environmental similarity among what are considered to be vegetationally distinct units. Separate canonical analyses were run for the six vegetation groups, all community types, and the community types within each of the Pseudotsuga, Thuja and Abies groups. To show the results graphically, the means of plots belonging to groups or types were plotted along the first two canonical variate axes for each analysis (Figs. 13 and 14). Seal's (1964) method was used to calculate $90 \%$ confidence circles around the means (radius $=1.645 \div \sqrt{n}$ ). The size of the confidence circles is linked to sample size; groups or types consisting of few plots will have large confidence circles. The generalized distance measure of Mahalanobis (Mahalanobis squared distance, $D^{2}$ ) was used to measure the distance between the type centroids in the environmental space (Goodall, 1978; Orlóci, 1972). As is the case with most ecological data, the assumptions necessary for the statistical interpretation of canonical analysis are violated, therefore the technique becomes a data-exploratory procedure to provide useful insights (Williams, 1983).

Stepwise forward discriminant analysis also was used to analyse environmental relationships among the groups and types. This method selects environmental variables which best discriminate among the vegetation units, and also reassigns the individual plots to units where they share the greatest overall environmental similarity. Thus, the method provided a means to test the vegetation classification using an independent set of environmental variables. The separation of vegetation groups and community types obtained by discriminant analysis was similar to those produced by canonical analysis; therefore, only the latter results are discussed in detail. The tabular results from discriminant analysis are presented in Appendix 4. Canonical and discriminant analysis were performed using programs from the MIDAS statistical package supported by the University of British Columbia Computing Centre.

## 4. VEGETATION STRATA HOMOGENEITY WITHIN TYPES

As a measure of the homogeneity of the vegetation strata within different community types, interplot similarity matrices were calculated using the data from each stratum for individual community types. The mean interplot similarities would indicate the relative homogeneity of the vegetation within each community type, as well as the variations in homogeneity between vegetation strata (Bradfield and Scagel, 1984). The similarity between plots was defined by the cosine function. The value of this function ranges from 0.0 , for plots with no species in common, to 1.0 , for plots with the same species occurring in identical proportions (Bradfield and Scagel, 1984).

The homogeneity of the tree, sapling, seedling, shrub, herb and bryophyte-lichen strata of fourteen community types was compared using this measure (Table 31). Community types with less than five plots were not included in this analysis, except for the two Pinus contorta community types, which otherwise would have left that group unrepresented, and the coastal wet Thuja forests (T5, 4 plots), which appeared unusually homogeneous.

## 5. TREE SIZE-CLASS STRUCTURE OF COMMUNITY TYPES

Graphs showing the size-class distributions of tree species within community types were plotted to provide descriptions of community structure, and to assist in the interpretation of community dynamics. Community types with less than three plots were not analysed. The floodplain forests (F1) community type also was not analysed because of the great heterogeneity of its tree stratum. The data used are the number of stems of tree species in 10 cm DBH size-classes for all plots of each community type. These data were transformed into numbers of stems per size-class per hectare (one plot $=.05 \mathrm{ha}$ ). The number of tree seedlings per hectare also was calculated using density data obtained from the microplots (twenty $1 \mathrm{~m}^{2}$ microplots per plot).

Graphs of tree species stem density per hectare versus size-class were made, using a logarithmic scale for stem density. Hand-fitted and smoothed curves were drawn for the tree species with the highest importance values within the selected community types (Figs. 15, 16 and 17).

## 6. TREE SEEDLING ABUNDANCE ON UNDECOMPOSED WOOD AND FOREST FLOOR SUBSTRATA

The tree seedling density data were analysed to determine whether there was a significant difference between average seedling densities on two broad types of substrata. During the sampling, microplots were recorded as being located on forest floor (including mineral soil, humus or litter) or on undecomposed wood (fallen trees, tree stumps, debris and bark at tree bases). Only the data from plots sampled using a random design of microplot location (Fig. 4) were utilised in the statistical test. To be included, tree species had to be present as seedlings in at least fifty percent of the plots ( $500 \mathrm{~m}^{2}$ ), but not necessarily in $50 \%$ of the microplots within each plot. The mean number of tree seedlings per square metre for each species, on each of the two substratum types, was calculated for community types with sufficient data (at least one hundred randomly selected microplots).

The null hypothesis is that tree seedling abundance, of each species, is equal on both. substratum types. The two sample z-test was used to make the comparisons (Freedman et al., 1978). Compared to the more familiar t-test, the $z$-test provides a good approximation of the true value of $P$ even when the data do not follow the normal curve very well, provided the sample size is large enough for the normal approximation to take over. The t-test requires that the data follow the normal curve closely (Freedman et al., 1978).

CHAPTER 4.

## RESULTS

## A. GRADIENT ANALYSIS OF VEGETATION

## 1. GENERAL VEGETATION PATTERNS

a) 172_p1ots ordination

The reciprocal averaging ordination of the 172 sample plots shows a mass of centrally located plots surrounded by groups of outlier plots (Fig. 5). The data matrix for this ordination consisted of 172 plots and a total of 197 species, or pseudo-species for trees divided into size-classes. Species, or pseudo-species, included in the matrix were present in at least four plots ( 14 trees, 11 saplings, 11 seedings, 24. shrubs, 81 herbs, 56 bryophytes and lichens). The first and second axes explained $11.0 \%$ and $8.8 \%$, respectively, of the total variance. Correlations of environmental variables with the ordination axes are given in Table 4. The strongest correlation with the first axis is with effective rooting depth/soil depth ratio, indicating fuller utilization by tree roots of a decreasing soil layer in plots located towards the positive end of the axis. Pinus contorta has the largest positive eigenvector coefficient on the first axis (Table 3), and is the dominant tree species in a group of shallow soil, rock outcrop communities identified at the extreme right of the ordination (Fig. 5). Two community types, a dry Pinus-Pseudotsuga type (D1) and a coastal dry Pinus type (D2), were recognized within this group by compositional and geographical
differences. Other environmental correlations with the Pinus types are increasingly better drainage, decreasing soil depth, coarser soil material, increasing percent rock fragments, thinner organic layer, increasing fire disturbance, and ridge topographical position (Table 4).

The two other groups identified on this ordination are separated along the second axis. Elevation is the environmental variable most strongly correlated with the second axis (Table 4). A distinct group of subalpine plots characterised by Abies amabilis (saplings, trees, seedlings), Vaccinium alaskaense, Rhytidiopsis robusta and Tsuga mertensiana is identified at the top left of the ordination (Table 3).

Picea sitchensis (trees, seedlings), Rubus spectabilis, Ribes bracteosum and Polystichum munitum have the largest negative eigenvector coefficients on the second axis (Table 3). These species are characteristic of a group of floodplain and river terrace plots identified toward the bottom of the ordination. Correlated with the lower elevation floodplain plots are an increase in organic layer pH , lower topographical position (leve1), finer soil material (alluvial), an increase in $B_{1}$ horizon pH (richer soil), deeper rooting, and increasing tree total basal area and maximum height (both indirect indications of site productivity) (Table 4). Two community types, subsequently referred to as F1 and F2, were identified within the floodplain group based on compositional and quality of drainage differences.

A final community type, termed the dry Pseudotsuga forests (P1), was recognized toward the lower right of the ordination. This type consists of four plots that share a strong dominance by Pseudotsuga menziesii,
contain Arbutus menziesii, but lack Pinus contorta. Gaultheria shallon strongly dominates the shrub layer. Although not distinctive in the ordination, because of compositional similarities to several other plots dominated by Pseudotsuga menziesii, these sites have in common similar soil characteristics and recent fire histories.
b) 140 plots ordination

Following the removal of plots assigned to community types in the first ordination, a second reciprocal averaging ordination was run on the remaining 140 plots (Fig. 6). The data matrix for this ordination consisted of 140 plots and 149 species (or pseudo-species for trees divided into size-classes). Species included were present in at least four plots (12 trees, 7 saplings, 10 seedlings, 20 shrubs, 62 herbs, 38 bryophytes and lichens). The first and second axes explained $13.6 \%$ and $9.8 \%$, respectively, of the total variance. In general, the main vegetation patterns on the first two axes appear to be determined by the interaction of complex environmental gradients associated with distance from the coast and elevation. Fire disturbance has the strongest positive correlation with the first axis (Table 6), while Pseudotsuga menziesii trees and seedlings have the largest positive eigenvector coefficients on this axis (Table 5). Distance from the coast has the strongest correlation with the second axis, decreasing toward the positive pole (Table 6), where Gaultheria shallon, Thuja plicata (seedlings, trees, saplings), Vaccinium ovatum and Blechnum spicant increase in coverage (Table 5). Abies amabilis (trees, saplings, seedlings) has the strongest negative eigenvector coefficients on both the first and second axes (Table 5). This tends to pull plots where it dominates, and regenerates itself, toward the lower left of
the ordination. Based on these results and after careful consideration of the vegetation and environmental data, the 140 plots were subdivided into three broadly defined groups : a Pseudotsuga group at the lower right, a Thuja group at the top, and an Abies group at the lower left. Along the first axis, correlations with several ecological variables help to differentiate the Pseudotsuga group from the others, such as : increasing evidence of fire disturbance, better drainage, thinner organic layer, deeper rooting, deeper rooting in mineral soil, increasing distance from the coast (inland), and decreasing evidence of wind disturbance (Table 6). Similarly, on the second axis, correlations with several ecological variables help to differentiate the Thuja group from the two others, such as : geographical proximity to the coast, increasing evidence of wind disturbance, decreasing elevation (nearer to sea level), decreasing tree height and drainage, and decreasing evidence of fire disturbance (Table 6). General characteristics of the Abies group plots include a tendency to occupy higher elevations, where there is little evidence of disturbance by fire or wind, and having no definite geographical area of maximum occurrence. An aberrant plot with a recent firehistory, situated inland near Port Alberni, and dominated by scattered large P. menziesii which survived the fire, was not assigned to any of the groups (small dot on Fig. 6). The dominance in the understory by a dense cover of Vaccinium ovatum, a shrub most commonly associated with open coastal habitats on poor soils, is probably responsible for the positioning of this plot closer to the Thuja group in the ordination.

## 2. VEGETATION PATTERNS WITHIN THE PSEUDOTSUGA GROUP

The data matrix for the ordination of this group consisted of 59 plots and 119 species (or pseudo-species for trees). Species included were present in at least three plots ( 9 trees, 7 saplings, 9 seedlings, 17 shrubs, 51 herbs, 26 bryophytes and lichens). The first and second axes explained $14.4 \%$ and $9.6 \%$, respectively, of the total variance. The ordination of the plots from the Pseudotsuga group reveals a more detailed pattern within this group (Fig. 7a). The first ordination axis is best correlated with organic layer pH and vascular species richness (Table 8). This results in a separation of species rich plots, with Acer macrophyllum present, at the negative end of the axis, from species poor plots at the positive end, where Tsuga heterophylla (seedlings, saplings, trees), Blechnum spicant, and Polystichum munitum are important. Acer macrophyllum (saplings, seedlings), Cornus nuttallii (seedlings, saplings, trees) and $\underline{P}$. menziesii (seedlings, saplings) have the largest negative eigenvector coefficients on the first axis (Table 7) helping to differentiate the Pseudotsuga-Thuja-Acer forests community type (P2). This is the most floristically rich of the Pseudotsuga types and has the least acidic organic layer, probably because of the litter input from the deciduous trees present. Other characteristics of this community type are a greater understory coverage, furthest distance from the coast (all plots situated very near Port Alberni), shallower soils and thinner organic layer (Table 8).

Variables strongly correlated with the second axis are total shrub coverage and topographical position (Table 8). Plots near the negative
end of the second axis tend to occur in higher topographical positions (ridges, crests, upper slopes), while plots at the positive end tend to occur in lower topographical positions (mid-slopes and lower-slopes). This pattern is also evident in the direct ordination of the Pseudotsuga group (Fig. 7b). For the second axis, a gradient of increasing site productivity, although not directly measured, can:be inferred also from several variables such as : decreasing total shrub coverage and total understory coverage (because of closing canopy), increasing total tree basal area and maximum tree height, increasing percent nitrogen in $B_{1}$ horizons, and decreasing $C / N$ ratios in $B_{1}$ horizons (Table 8). The Tsuga-Pseudotsuga-Polystichum forests community type (P5) occupies the most productive end of this gradient, as well as lower-slopes (Fig. 7a and 7b). The least productive end of the gradient is occupied by two community types, the Pseudotsuga-Linnaea forests (P3) and the montane TsugaGaultheria forests (P7) (Fig. 7a). Floristic differences separate these two types clearly on the reciprocal averaging ordination (Fig. 7a). Ecologically, the two types are differentiated by elevation, with the montane Tsuga-Gaultheria forests occurring. at higher elevations (Fig. 7b).

Two other community types are of intermediate position on the second axis, the Pseudotsuga-Berberis forests (P4) and the montane Tsuga forests (P6). These two types are distinguishable floristically, but intergrade more or less continuously along the elevational gradient (Figs. 7a and 7b). On the second axis, Gau1theria shallon and Hylocomium splendens have the largest negative eigenvector coefficients, corresponding to the poorer and drier sites (Fig. 7a), while Polystichum munitum and Cornus nuttallii have the largest positive eigenvector coefficients,
corresponding to the richer and moister sites (Figs 7a and 7b; Table 7). A gradient of increasing soil moisture availability also can be suggested for the second axis based on relationships indicated in the direct ordination (Fig. 7b). The montane Tsuga-Gaultheria forests community type is situated at the dry end of this gradient on ridges and south-southwest facing slopes, and the Tsuga-Pseudotsuga-Polystichum forests community type is situated at the moist end on lower-slopes, with more seepage, deeper soils and better shelter from drying winds. Communities. of intermediate position on the topographic-moisture gradient of Fig. 7b are also intermediate in position on the second axis of Fig. 7a. The dry PinusPseudotsuga forests (D1) were added to figure 7 b to show their topographical positions. Three unique plots were not assigned to any community type within this group. Plot 112 is from the China Creek area, west of Port Alberni, and has a drier climate and a soil different from that commonly found within the study area. Plot 89 is from an unusual coastal stand dominated by Thuja plicata, but with a high cover of Polystichum munitum which caused this plot to be included with the Pseudotsuga group. Plot 20 represents a relatively recently disturbed site.

## 3. VEGETATION PATTERNS WITHIN THE THUJA GROUP

The data matrix for the ordination of this group consisted of 40 plots and 25 species (or pseudo-species for trees). Species included were present in at least three plots (7 trees, 4 saplings, 6 seedlings, 9 shrubs, 22 herbs and 27 bryophytes.). The first and second axes explained $23.4 \%$ and $13.5 \%$, respectively, of the total variance. Five community types were identified within the Thuja group ordination (Fig. 8a). Based on correlations with ecological variables the first axis is interpreted as a site productivity gradient. The strongest correlations with the first axis are with total shrub coverage, total understory coverage and maximum tree height (Table 10). Other variables correlated with the first axis are also indicators of site productivity, such as increasing soil depth, organic layer percent nitrogen, decreasing organic layer $C / N$ ratio, increasing percent nitrogen and carbon in $B_{1}$ horizons, and increa= sing root restricting depth (Table 10). Abies amabilis (saplings, trees, seedlings), Tsuga heterophylla (seedlings, saplings) and Polystichum munitum have the largest positive eigenvector coefficients on the first axis (Table 9), and characterize a group of productive sites at the right of the ordination; Vaccinium ovatum and Thuja plicata (saplings, seedlings) have the largest negative eigenvector coefficients and characterize the less productive sites to the left. The coastal Tsuga-Blechnum-Polystichum forests community type ( T 2 ) is considered to be the most productive of this group. This community type occurs on upper and mid-slopes only, mostly above 200 m of elevation (Fig. 8 b ), where there is better drainage and less coastal fog influence than at lower elevations. The last two factors seem to greatly influence productivity in coastal forests where moisture is often overabundant. The coastal dry Thuja forests (T1) and
the coastal wet Thuja forests (T5) community types are both considered to occur on the most unproductive sites of this group. The dry type is found on well drained ridges and steep slopes, while the wet type occurs on poorly drained level sites (Fig. 8b). In both cases Vaccinium ovatum dominates the shrub layer, accounting for their close positions on the reciprocal averaging ordination (Fig. 8a).

Environmental gradients underlying the second axis are not as clear. Herb species richness and total herb coverage are strongly corre:lated with the second axis (Table 10), mostly because of plot 85 , a unique sample plot from a rich fen-like coastal Thuja swamp on poorly drained marine clays. The presence of plot 85 (nearly at sea level) also weakens the correlation of the second axis with elevation, which otherwise can be seen on the direct ordination (Fig. 8b). The coastal montane Thuja forests community type (T3), intergrades continuously with the coastal Thuja forests community type (T4) along the elevation gradient (Fig. 8b) although floristic differences (such as higher importance of Abies amabilis in the T3 type), help to separate them in the indirect ordination (Fig. 8a). The coastal Thuja forests form the most common and characteristic community type of the lowland coastal forests of western Vancouver Island.

## 4. VEGETATION PATTERNS WITHIN THE ABIES GROUP

The data matrix for the ordination of this group consisted of 40 plots and 87 species (or pseudo-species for trees). Species included were present in at least three plots (5 trees, 4 saplings, 5 seedlings, 9 shrubs, 36 herbs, 28 bryophytes and lichens). The first, second and third axes explained $16.7 \%, 10.9 \%$ and $10.3 \%$, respectively, of the total variance. The reciprocal averaging ordination of the Abies group is the only case in this study where it was felt that the third axis offered clearer relationships, patterns, and environmental gradient interpretations than the second axis (Fig. 9a). The occasional advantage, for interpretation purposes, of using the third reciprocal averaging axis instead of the second has been noted also by Gauch, et al. (1977) and Peet (1980). Species with the largest positive eigenvector coefficients on the first axis are Tsuga heterophylla (saplings, seedlings, trees), Pseudotsuga menziesii (trees) and Polystichum munitum, while species with the largest negative eigenvector coefficients are Abies amabilis (saplings, seedlings, trees), Rubus pedatus and Streptopus streptopoides (Table 11). Clearly, the first axis separates plots where tree species regeneration is dominated by either Abies amabilis or Tsuga heterophylla. Correlation of environmental variables with the first axis indicate that the plots where Abies regeneration dominates are the furthest from the coast and the highest in elevation (geographically where the highest mountains are found). Percent carbon in the organic layer is highest in these plots while pH of the $\mathrm{B}_{1}$ horizon is lowest (Table 12). At the extremities of the first axis, two distinct community types can be identified, the montane Abies-Streptopus forests (A4), of high elevation,
cool north to north-west facing slopes, and the Tsuga-Blechnum-Polystichum forests (A7), of low elevation mesic sites (Figs. 9a and 9b). The third axis is interpreted as a complex environmental gradient associated with increasing elevation, exposure and fire disturbance, and decreasing productivity inferred from an increase in organic layer $\mathrm{C} / \mathrm{N}$ ratio and a decrease in tree height (Table 12). On the third axis, Gaultheria shallon, Abies amabilis (seedlings, saplings), Rhytidiopsis robusta and Pseudotsuga menziesii have the largest positive eigenvector coefficients. These species characterize the fire prone, most nutrient poor and drier sites within the Abies group. Sphagnum girgensohnii, Abies amabilis (tree), Achlys triphylla and Polystichum munitum have the largest negative eigenvector coefficients on the third axis, and characterize the mesic, most nutrient rich sites (Table 12). The montane Tsuga-AbiesGaultheria forests community type (Al) occurs inland from mid to high elevations on dry slopes where there is visible evidence of fire (Fig. 9b). The opposite end of this gradient is occupied by the lowland Abies forests (A5) and the montane Tsuga-Abies forests (A3). The lowland Abies forests occur at low elevations in valley bottoms but bear close floristic resemblance to the elevationally and topographically different montane Abies-Streptopus forests (Figs. 9a and 9b), montane Tsuga-Abies forests (A3) occur on higher well drained slopes near the coast (Fig. 9a). Other community types are, the montane Abies-Tsuga forests (A2), which occur on high elevation sites with a better drainage than found in sites occupied by the montane Abies-Streptopus forests (A4), and the Tsuga-Gaultheria-Blechnum forests (A6), which occupy possibly less productive sites than the Tsuga-Blechnum-Polystichum forests (A7)(Fig. 9a). Eight
plots were not assigned to any of the types within this group. Plots $28,83,127$ and 168 are strongly influenced by seepage water, making their relationships difficult to evaluate through vegetation data. Plot 151 is a relatively young stand, resulting from a complete blowdown, sampled for comparison purposes. Plot 113 is from a high elevation stand in the drier China Creek area (see Pseudotsuga group). Plot 35 is from a high elevation site ( 865 m ), but lacks the characteristic species which would have placed it within the subalpine group of Fig. 5. Plot 68 is a partially wind disturbed stand most similar to the montane Tsuga-Abies forests.

## 5. VEGETATION AND ENVIRONMENTAL PATTERNS ON A DISTANCE FROM THE COAST GRADIENT

The data matrix for the ordination of this group of modal plots consisted of 105 plots and 147 species (or pseudo-species for trees). Species included were present in at least 4 plots (1l trees, 7 saplings, 10 seedlings, 20 shrubs, 61 herbs, 38 bryophytes). The first and second axes of a reciprocal averaging ordination explained $15.0 \%$ and $8.8 \%$, respectively, of the total variance, and produced a strongly arched scatter of plots (Fig. 10). Species with the largest positive eigenvector coefficients on the first axis are Pseudotsuga menziesii (trees, seedlings), Acer macrophyllum (saplings, seedlings) and Cornus nuttallii (saplings, seedings), while species with the largest negative eigenvector coefficients are Blechnum spicant, Abies amabilis (saplings, trees, seedlings) and Thuja plicata (trees) (Table 13). The first axis separates plots of the Thuja group from plots of the Pseudotsuga group. This is similar to the separation on the first axis of the 140 plots ordination (Fig. 6), except that, in this case, most plots of the Abies group have been removed. The few plots belonging to the Abies group are centrally located on the 105 modal plots ordination. Correlation of environmental variables with the first axis clearly demonstrates the strong effect that distance from the coast has on vegetation variation in the study area. This geographical gradient summarizes the effects of many separate environmental variables including fire and wind disturbance, organic horizons thickness/ effective rooting depth ratio, drainage and organic horizons thickness (Table 14). However, it must be recognized that organic horizons thickness and effective rooting depth are partially a function of the overlying
vegetation. The increasing precipitation and decreasing growing degree-days towards the coast can be seen on isoline maps adapted from climate maps compiled by Colidago (1980) (Fig. 12). Also, the organic horizons thickness/effective rooting depth ratio decreases and vascular species richness increases with increasing distance from the coast (Fig. 12). Correlations of environmental variables with the second axis show relationships similar to those shown on the first axis (Table 14). Polynomial regression curves of the basal area of major tree species show distinct peaks along the distance from the coast gradient (Fig. 11). Thuja plicata reaches a peak in total basal area 13 km from the coast, while Pseudotsuga menziesii reaches its peak at about 48 km from the coast. Tsuga heterophy11a reaches its peak in basal area at 30 km from the coast. The basal area of Abies amabilis increases steadily towards the coast. Thuja plicata shows a marked decrease in basal area when closer than 10 km from the coast. A similar decrease in Tsuga heterophylla basal area occurs at about 50 km from the coast (Fig. 11). The organic horizons thickness/effective rooting depth ratio polynomial regression curve also shows a distinct dip towards the inland part of the island (Fig. 11). The peaks and decreases in basal area of each tree species can be interpreted as responses to climatic variables and disturbance type and regime, which are linked with climate. The variations in the organic horizons thickness/effective rooting depth ratio can be interpreted as the result of climate and vegetation differences. Al1 these relationships are discussed further in chapter 5.

## B. CANONICAL ANALYSES OF VEGETATION GROUPS AND COMMUNITY TYPES BASED ON ENVIRONMENTAL DATA

## 1. VEGETATION GROUPS

The six vegetation group centroids are separated clearly on the first and second canonical variates of the environmental data (Fig. 13). These two axes summarize the main environmental relationships among the six vegetation groups. The Pinus contorta group (D) and the Floodplain group (F) are at opposite ends of the first two canonical variates. This large difference in environmental characteristics also is reflected in the Mahalanobis squared distance ( $\mathrm{D}^{2}$ ) between these two groups (Table 16). Drainage, surficial material, topographic position and coarse fragment content of the $B_{1}$ horizon are the environmental variables most strongly correlated with the first canonical variate (Tab1e 21). Fire and wind disturbance are also correlated, positively and negatively, respectively, with the first axis. The ratio, organic horizons thickness/effective rooting depth, and organic horizons thickness are negatively correlated with the second canonical variate, while organic horizons pH is positively correlated (Tab1e 21). Very rapid drainage, crest or ridge topographic position, and lack of surficial material (rock outcrops) characterize the Pinus contorta group (D). Slower drainage, lower-slope or level topographic positions, and morainal, fluvial or alluvial surficial deposits characterize the Floodplain (F) and Thuja (T) groups. Group positions on the second canonical variate can be best interpreted with the organic horizons thickness/effective rooting depth ratio; the Floodplain group
is characterized by thin organic horizons and deep rooting into mineral soil (ratio closer to zero), while the Thuja group is characterized by thick organic horizons and shallow rooting (ratio closer to one). Although organic horizons are thin in the Pinus contorta group, the rooting is very shallow. The Pseudotsuga group ( P ) is the group with the higher environmental similarity with the Pinus contorta group, based on $\mathrm{D}^{2}$ values (Table 16). The Abies group (A), having several plots situated at high' elevations, is the group most environmentally similar to the Subalpine group (SA) based on $D^{2}$ values. It should be noted that the $D^{2}$ values are calculated over all the dimensions of the canonical analysis while only two dimensions are presented in the figures (Figs. 13 and 14). Comparing only the Pseudotsuga, Thuja and Abies groups, we find that the Pseudotsuga and Thuja groups are the most environmentally different (Table 16).

The canonical analysis results correspond generally to those obtained with reciprocal averaging (Figs. 5, 6 and 13). The correlations of environmental variables with the reciprocal averaging axes also are similar to those with the canonical variate axes (Tables 4,6 and 21). That the results of both analyses conform is interesting since the reciprocal averaging ordinations used vegetation data, and the canonical analysis used environmental data. However, the groups submitted to the canonical analysis were determined using reciprocal averaging ordinations. The results do indicate that environmental patterns correspond to the vegetation patterns.

## 2. PSEUDOTSUGA TYPES

Most of the Pseudotsuga type centroids are separated clearly on the first two canonical variates (Fig. 13). The dry Pseudotsuga forests (P1), the Pseudotsuga-Thuja-Acer forests (P2), and the PseudotsugaLinnaea forests (P3) appear very similar environmentally (Fig. 13;

Table 17). Plots belonging to these three community types are found only in the driest inland part of the study area. The dry Pseudotsuga forests (P1) can be separated from the P2 and P3 types (other two community types) on the basis of its vegetation structure, which is hypothesized to have resulted from a recent, intense fire (see Chapter 4. A, section $1_{a}$, and 4. C, section 2 ). Similarly, the Pseudotsuga-Thuja-Acer forests (P2) show vegetation differences with P1 and P3 which are interpreted to reflect differences in seepage conditions (Chapter 4.C, section 2). Since none of the environmental variables included in the canonical analysis adequately reflected the underlying reasons for the vegetation differences, the three types (P1, P2, P3) were not separated. The Pseudo-tsuga-Thuja-Acer forests (P2) and the Tsuga-Pseudotsuga-Polystichum forests (P5) are at opposite ends of the first canonical axis. These are the most environmentally different types within the Pseudotsuga group (Fig. 13, Table 17). The two most environmentally similar community types are the Pseudotsuga-Berberis forests (P4) and the montane Tsuga forests (P6) which are differentiated vegetationally only along an elevation gradient (Fig. 7a and b). The Pseudotsuga-Linnaea forests (P3) and the montane Tsuga-Gaultheria forests (P7), both occurring in dry sites, show vegetation similarities (fig. 7a) but are environmentally
quite different (Figs. 7b and113), mostly because of differences in elevation. Topographic position, soil depth, effective rooting depth/ soil depth ratio, and organic horizons pH are the environmental variables most strongly correlated with the first canonical axis (Table 21). The Thuja-Pseudotsuga-Polystichum forests (P5), with the largest scores on the first axis, always are found on the lower topographical positions and on the deepest soils. Elevation, organic horizons $\mathrm{pH}, \mathrm{B}_{1}$ horizon \% nitrogen, and topographic position are strongly correlated with the second canonical axis. Montane Tsuga-Gaultheria forests (P7), located toward the top of the second axis, are found at high elevations on ridges and crests. The Pseudotsuga-Thuja-Acer forests (P2), near the bottom of the second axis, have the highest organic horizons pH . All of these environmental variables also were correlated with the reciprocal averaging ordination axes of the vegetation data (Fig. 7a; Table 8).

## 3. THUJA TYPES

The Thuja type centroids are very clearly separated on the first two canonical variates of the environmental data (Fig. 13). The coastal Tsuga-Blechnum-Polystichum forests (T2) and the coastal montane Thuja forests (T3) are at opposite ends of the first axis. These two types appear to be the most environmentally different within the Thuja group (Fig. 13; Table 18). The coastal Thuja forests (T4) and the coastal wet Thuja forests (T5) have the most similar environmental characteristics based on the $\mathrm{D}^{2}$ values (Table 18). The coastal dry Thuja forests (T1) could not be included in the analysis because most of the stands lacked mineral soil, and therefore lacked values for numerous environmental variables. Drainage and $B_{1}$ horizon percent nitrogen are the only variables significantly correlated with the first canonical axis (Table 21). Coastal Tsuga-Blechnum-Polystichum forests (T2), with the largest scores on the first axis, are associated with productive sites, higher $B_{l}$ horizon \% N, and better drainage. Elevation, percent slope, and topographic position are strongly correlated with the second canonical axis (Table 21). The coastal wet Thuja forests (T5) and the coastal Thuja forests (T4), positioned toward the top of the second axis, were found consistently at low elevations on level or moderately sloping terrain. These environmental variables also were strongly correlated with the reciprocal averaging ordination axes of the vegetation data (Fig. 8a; Table 10).

## 4. ABIES TYPES

The Abies type centroids are not as clearly separated on the first two canonical axes as are those of the other groups (Fig. 13). This partly reflects the impression of overlap caused by the larger confidence circles of the centroids, which are generally based on fewer plots than in the other groups, and partly the use of only four environmental variables in the canonical analysis. This was necessary since the computer program used would not perform the analysis with a larger set of variables. The four variables used were preselected with the use of a stepwise discriminant analysis, selecting the environmental variables which permitted the maximum separation of the community types (Appendix 4). The montane Abies-Streptopus forests (A4) and the Tsuga-Blechnum-Polystichum forests (A7) are at opposite ends of the first canonical axis. These are the most environmentally different types within the Abies group (Fig. 13; Table 19). Based on the four variables used, the Abies-Streptopus forests (A4) are markedly different environmentally from all other community types of the Abies group (Fig. 13; Table 19). Of particular interest is the notable environmental difference between this community type and the lowland Abies forests (A5). Despite the environmental difference both community types show strong vegetational similarities (Fig. 9a; Tables 26 and 27). Elevation and organic horizons thickness are strongly correlated with the first canonical axis (Table 21). Stands of the Abies-Streptopus forests (A4), at the positive end of the first axis, occur at the highest elevations within the Abies group. The second axis also is correlated with elevation and \% slope (Table 21);
thus, the lowland Abies forests (A5), the Tsuga-Gaultheria-Blechnum forests (A6), and the Tsuga-Blechnum-Polystichum forests (A7), all found at low elevations on moderate to gentle slopes, are grouped toward the lower end of the second axis. Montane Tsuga-Abies-Gaultheria forests (A1) appear environmentally similar to the montane Abies-Tsuga forests (A2) (Fig. 13; Table 19), but this could be an artifact of the low number of environmental variables used. A better illustration of environmental relationships within the Abies group is obtained through the canonical analysis of all the community types from all the vegetation groups, where all the environmental variables were utilised (Fig. 14; Table 20).

## 5. ALL TYPES AND THE SUBALPINE GROUP

The distribution of community type centroids on the first two canonical variates (Fig. 14) corresponds closely to the general pattern shown in the analysis of the vegetation groups (Fig. 13). Furthermore, the correlations between environmental variables and canonical axes show the same trends in both cases (Table 21). Although the general patterns of both analyses are similar, the canonical analysis of separate types indicates that some community types are environmentally more similar to types belonging to other vegetation groups (Fig. 14; Table 20). For example, the montane Tsuga-Abies-Gaultheria forests (Al) are environmentally more similar to the montane Tsuga forests (P6) and the montane Tsuga-Gaultheria forests (P7), of the Pseudotsuga group, than to any other community type of the Abies group (Fig. 14; Table 20). These community types also are similar vegetationally (Tables $22,23,26$ and 27 ). The coastal Tsuga-Blechnum-Polystichum forests (T2) of the Thuja groups, and the Tsuga-Blechnum-Polystichum forests (A7) of the Abies group are both environmentally (Fig. 14; Table 20) and vegetationally similar (Tables 24, 25, 26 and 27). The two environmentally most similar community types are the Pseudotsuga-Berberis forests (P4) and the montane Tsuga forests (P6), based on the $\mathrm{D}^{\dot{2}}$ value (Table 20). The environmentally most dissimilar community types are the coastal dry Pinus forests (D2) and the Lysichitum variant of the Floodplain forests (F2):.(Table 20). Relationships between community types detected in the canonical analyses of separate vegetation groups generally are maintained in the combined types analysis; however, the coastal Thuja forests (T4) appear environmentally most similar to the montane coastal Thuja forests (T3), than to the coastal wet Thuja
forests (T5), in the combined analysis (Tables 18 and 20). The canonical analysis of all the community types is felt to represent environmental relationships more accurately than the analysis of separate vegetation groups. Possibly because of the greater ranges of environmental variation when all types are analysed together. These results also could help to redefine the community type classification(eg. merging types T2 and A7, as well as A1 and P7), although such was not done in this thesis. These community types (T2, A7, A1, P7) were situated at the boundaries of the three vegetation groups separated in the 140 plots reciprocal averaging ordination (Fig. 6). The two pairs of community types which are differentiated vegetationally along an elevation gradient both show high overall environmental similarities (P4 and P6, T3 and T4). This may indicate that no ecological factor, other than elevation (detected by the direct ordinations, Figs 7 b and 8 b ), is responsible for the vegetational differences observed (Figs. 7a and 8a; Tables 22, 23, 24 and 25).

## C. DESCRIPTION OF COMMUNITY TYPES

The vegetation and ecological characteristics of the community types delineated within the reciprocal averaging ordinations (Figs. 5, 7a, 8a and 9a) are described in this section. The composition, structure and diversity of the various vegetation strata are described briefly. The geographical distribution, topographical characteristics, soil characteristics and disturbance history are also outlined for each community type. Similarities between community types are indicated, as well as similarities with other community types or associations described previously for coastal British Columbia and, when possible, for Washington and Oregon.

The community types within the Pinus contorta vegetation group are described first, followed by community types of the Pseudotsuga group, the Thuja group, the Abies group and the Floodplain group. Last to be described is the Subalpine vegetation group which was not subdivided into community types.

1. PINUS CONTORTA VEGETATION GROUP

Dry Pinus-Pseudotsuga forests (D1)

These open and low stature forests (average maximum height is 18 m ) are strongly dominated by Pinus contorta. This is the only community type within the study area where Arbutus menziesii is always present. Pseudotsuga menziesii seems to be regenerating well (Table 22). At $30.7 \mathrm{~m}^{2} / \mathrm{ha}$, the mean total basal area is the second lowest for the community types described in this study (the lowest is found in coastal dry Pinus forests, of the same vegetation group). The mean tree density (700 trees/ha) is among the highest. The shrub and bryophyte-lichen strata are among the richest found in the study area (Table 23). A large coverage of Vaccinium ovatum and the presence of Arctostaphylos columbiana characterize the shrub stratum (under 1.5 m in height). Several herb species such as Apocynum androsaemifolium, Cryptogramma crispa, Danthonia spicata and Selaginella wallacei are restricted almost entirely to this community type. The bryophyte-lichen stratum is characterized by an abundance of lichens (Cladina rangiferina and many Cladonia species) growing on large bare rock patches representing $28 \%$ of the ground surface (Table 23).

This community type was found only at low elevations in the inland portion of the study area around Port Alberni. It occurs on rock outcrops, predominantly south facing. The soils are very shallow, average 15 cm in depth, and are very rapidly drained. The organic horizons are very thin and roots are abundant down to the bedrock (Appendix 2). Fire is probably responsible for the establishment of Pinus contorta and evidence
of fire is abundant in all stands. Some windthrow also has occurred. The dry Pinus-Pseudotsuga forests are quite similar to the coastal dry Pinus forests (D2), which occupy similar habitats near the coast (Figs. 5 and 14). They also have floristic similarities to the dry Pseudotsuga forests (P1) (Tab1es 22 and 23).

Krajina (1969) lists several biogeocoenoses (numbers 6, 12, 19 and 29) with lists of species similar to those of the dry Pinus-Pseudotsuga forests. Kojima and Krajina (1975) describe a similar Arbutus menziesii stand on a rock outcrop in Strathcona Provincial Park, north of the area studied here. McMinn's (1960) Pseudotsuga-GaultheriaPeltigera association also is similar, although probably is not as xeric.

Coasta1 dry Pinus forests (D2)

Although similar in structure to the dry Pinus-Pseudotsuga forests, the coastal dry Pinus forests have several co-dominant tree species, giving them the richest tree stratum in the study area. Thuja plicata, Chamaecyparis nootkatensis and Tsuga heterophylla are regenerating well on these sites (Table 24). The mean total basal area is the lowest in the study area ( $30.2 \mathrm{~m}^{2} / \mathrm{ha}$ ) and tree density is very high ( 695 trees/ha) as a consequence of numerous small trees (Table 24). The shrub layer, less than 1.5 m in height, is dominated by Gaultheria shallon and Vaccinium ovatum. The herb stratum has a very low total percent coverage and is relatively poor in species. In contrast, the bryophyte-lichen layer has a very high coverage and is the richest of all the community types recognized (Table 25). Some herb species such as Danthonia spicata,

Saxifraga ferruginea and Selaginella wallacei are restricted to this community type near the coast. Numerous bare rock surfaces (26 \% coverage) have a rich assemblage of lichens and mosses including Cladina species, Cladonia species, Pleurozium schreberi, Polytrichum and Rhacomitrium species (Table 25).

This community type is restricted to low elevation, sloping rock outcrops near the coast. These sites are very rapidly drained and have very shallow soils (average soil depth is 11 cm ). The organic horizons are thin and roots extend to the bedrock (Appendix 2). No evidence of fire was found and wind disturbance appears minimal. This community type is most similar to the dry Pinus-Pseudotsuga forests (D1), occupying similar sites inland (Figs. 5 and 14).

No previous descriptions of community types similar to the coastal dry Pinus forests seem to exist. However, this community type represents a coastal variation of the Dry Pinus-Pseudotsuga forests (D1) for which published equivalent descriptions were found.
2. PSEUDOTSUGA VEGETATION GROUP

Dry_Pseudotsuga forests (P1)

This community type has a relatively low canopy (average height is 44 m ), low mean total basal area ( $86.2 \mathrm{~m}^{2} / \mathrm{ha}$ ), and low mean tree density (300 trees/ha). Pseudotsuga menziesii dominates and appears to be the species regenerating best (Table 22). The shrub stratum is rich in species and its coverage is among the largest within the Pseudotsuga group. The herb layer is the richest found in the study area (Table 23). The most conspicuous shrubs are Gaultheria shallon (1 m high), Berberis nervosa and Vaccinium ovatum, while the herbs Chimaphila umbellata, Festuca occidentalis and Pteridium aquilinum are particularly abundant. Boschniakia hookeri, a root parasite of Gaultheria shallon, is always present. The bryophyte-lichen stratum is dominated by Stokesiella oregana and Hylocomium splendens, but otherwise shares many species with the dry Pinus-Pseudotsuga forests (Table 23).

The dry Pseudotsuga forests were found only inland near Port Alberni. They occupy low elevations, strong to extreme slopes .(Canada Soil Survey Committee, 1978) with shallow, rapidly drained soils (average soil depth is 54 cm ). The organic horizons are very thin and roots extend deep into the mineral soil (Appendix 2). This community type often occurs immediately downslope of the dry Pinus-Pseudotsuga forests (D1). Most soils are Orthic Dystric Brunisols developing on colluvial material (Table 29). Fire has occurred fairly recently in all the stands which probably explains why Pseudotsuga menziesii, with its fire resistant bark, dominates. The dense shrub coverage may also be fire induced owing
to the improved conditions for shrub growth following fire. Many of the stands studied almost entirely consist of large, widely spaced Pseudotsuga menziesii trees with charred bark. This type has vegetational and environmental similarities with the Pseudotsuga-Thuja-Acer forests (P2) and the Pseudotsuga-Linnaea forests (P3), although the many species of dry sites it shares with the dry Pinus-Pseudotsuga forests (D1) are an indication of its drier moisture regime (Tables 22 and 23; Figs. 13 and 14).

The Dry Pseudotsuga forests have similarities with numerous communities or associations described for British Columbia, such as the Pseu-dotsuga-Arbutus/Gaultheria habitat type of Beese (1981), the PseudotsugaGaultheria association of MCMinn (1960), and the Gaultheria shallon association of Kojima and Krajina (1975). Also similar, is the Pseudotsuga/ Holodiscus-Gaultheria association described by Franklin and Dyrness (1973) for dry sites within the Oregon Coast Ranges.

## Pseudotsuga-Thuja-Acer forests (P2)

This community type has one of the best developed tree strata in the Pseudotsuga group (mean maximum tree height is 64 m , mean total basal area is $132.5 \mathrm{~m}^{2} /$ ha (Table 22)). The tree stratum is also among the richest found in the study area, and although dominated by Pseudotsuga menziesii, it is characterized by the deciduous trees Acer macrophyllum and Cornus nuttallii. Most tree species seem to be regenerating well (Table 22). The low and sparse shrub layer (under 1 m in height) is dominated by Berberis nervosa, Gaultheria shallon and Rubus ursinus.

The herb stratum is also rich, with high coverages of Achlys triphylla, Festuca subulif1ora, Linnaea borealis, Polystichum munitum and Trientalis 1atifolia. Stokesiella oregana and Hylocomium splendens share dominance in the bryophyte-lichen layer. Leucolepis menziesii, a moss of moist soils (Schofield, 1976), is relatively abundant (Table 23).

The Pseudotsuga-Thuja-Acer forests occur only inland within the study area, close to Port Alberni. They occupy mid-slope positions on strong to extreme, mostly south facing slopes at low elevations (Fig. 7b). The soils are moderately deep (average soil depth is 65 cm ) and well drained, with relatively high pH values (LFH average $=4.8, \mathrm{~B}_{1}$ average $=$ 5.2). The organic horizons are very thin and rooting occurs throughout the mineral soil (Appendix 2). Most soils are Orthic Humo-Ferric Podzols developing on colluvial material (Table 29). The mid-slope topographic position, as well as the vegetation, suggest that seepage probably contributes significant amounts of water and nutrients to the soils. Evidence of fire is abundant, in the form of buried charcoal and charred bark on Pseudotsuga trees. Vegetational and environmental similarities between this community type, the dry Pseudotsuga forests (P1) and the Pseudotsuga-Linnaea forests (P3) are high (Tables 22 and 23; Figs. 13 and 14), although the vegetation is sufficiently different to warrant the distinction made (Fig. 7a). Also, the moisture regime is not as dry as in the other two types.

The Pseudotsuga/Holodiscus/Polystichum habitat type described for eastern Vancouver Island by Beese (1981) is very similar to this community type. Both contain Acer macrophyllum and have similar understories
(although Holodiscus discolor is more abundant in the eastern Vancouver Island type). Beese (1981) found no previous description of his type within British Columbia and suggested that it may have been included in other frequently described types where Polystichum munitum dominates the herb layer. Of these, the Achlys-Polystichum association of Kojima and Krajina (1975) comes closest to resembling the Pseudotsuga-Thuja-Acer forests. This community type probably represents the "classical" Polystichum type in what would be the equivalent of the Coastal wetter Douglas-fir subzone in this study area (Klinka et al., 1979), while the Tsuga-Pseudotsuga-Polystichum forests (P5), represent the Polystichum type in what would be the Coastal drier Western Hemlock subzone in this study area (Klinka et al., 1979).

## Pseudotsuga-Linnaea forests (P3)

Although of similar composition to the tree stratum of the Pseudo-tsuga-Thuja-Acer forests, the tree stratum of this community type is lower (average maximum height is 48 m ) and has a much smaller mean basal area ( $89.4 \mathrm{~m}^{2} / \mathrm{ha}$, Table 22). Tsuga heterophylla becomes the second dominant after Pseudotsuga menziesii, and deciduous trees are often absent. Tsuga heterophylla shows the best regeneration (Table 22). This community type has a shrub stratum characterized by a high coverage of Gaultheria shallon under 1 m in height. The rich herb layer is similar to that of the Pseudotsuga-Thuja-Acer forests (P2), except that Linnaea borealis attains a high coverage and Polystichum munitum is relatively unimportant. Hylocomium splendens dominates the bryophyte-lichen stratum covering most of the forest floor (Table 23).

This community type occurs mostly at mid-elevations, on mid- and upper-slopes (Fig. 7b), inland within the study area near Port Alberni. It is found on strong to extreme slopes with moderately deep (average soil depth is 65 cm ), well drained soils, formed mostly on colluvial material. Organic horizons and mineral horizons have relatively high pH values (LFH average $=4.9 ; \mathrm{B}_{\mathrm{l}}$ average $=5.2$ ). The organic horizons are thin and rooting occurs throughout the mineral soil (Appendix 2). All soils are Orthic Humo-Ferric Podzols (Table 29). The higher upslope position, as well as the absence of certain moisture indicator plant species (Tables 22 and 23 ), suggest that moisture input through seepage is not as pronounced here as in the Pseudotsuga-Thuja-Acer forests. Evidence of fire was present in all stands. Similarities between this type, the dry Pseudotsuga forests (P1) and the Pseudotsuga-Thuja-Acer forests (P2) are evident (Tables 22 and 23; Figs. 13 and 14).

The Pseudotsuga-Linnaea forests resemble the Pseudotsuga/GaultheriaBerberis habitat type described by Beese (1981) for eastern Vancouver Island. Other similarities are with the biogeocoenosis 5 of Krajina (1969) and the Salal-Oregon grape-Douglas-fir biogeocoenotic zonal type of Klinka et al. (1979) for the Drier Maritime Coastal Douglas-fir Subzone (the Pseudotsuga-Linnaea forests are situated on drier sites within the Wetter Subzone found in this study area). Beese (1981) 1ists several other similar community types described for British Columbia. Similar types in Oregon and Washington are summarized by Franklin and Dyrness (1973).

Pseudotsuga-Berberis forests (P4) (Fig. 18b)

The Pseudotsuga-Berberis forests have one of the best developed tree strata within the Pseudotsuga group (mean maximum tree height is 58 m and, mean total basal area is $138.4 \mathrm{~m}^{2} / \mathrm{ha}$ (Table 22) ). Pseudotsuga menziesii dominates, with Tsuga heterophylla as a close second. Tree regeneration is strongly dominated by Tsuga heterophylla (Table 22). Although the shrub layer is similar in most community types of the Pseudotsuga group, Berberis nervosa is notably abundant in the low shrub layer of this type (average height under 1 m ). The low cover, but rich, herb stratum has no particularly characteristic species. Hylocomium splendens and Stokesiella oregana again share dominance in the bryophyte1ichen layer (Table 23).

The Pseudotsuga-Berberis forests are found only inland within the study area. They are found mostly on mid-slope topographic positions at mid-elevations (Fig. 7b). They occur mostly on very strong slopes with deep, rapidly drained soils. The organic horizons are moderately thick (average of 8.6 cm ) and rooting occurs throughout most of the mineral soil (Appendix 2). Most soils belong to the Orthic Humo-Ferric Podzol subgroup (Table 29). The majority of stands show evidence of fire disturbance. This community type has similarities with the Montane Tsuga forests (P6), which are usually found on similar sites but at higher elevations. It also has similarities with the Tsuga-Pseudotsuga-Polystichum forest (P5), often found on adjacent, lower-slope topographic positions (Figs. 7a and b, 13 and 14; Tables 22 and 23).

The Pseudotsuga-Berberis forests are most similar to the "moss" association considered to be the zonal vegetation type for lower elevations in Strathcona Provincial Park by Kojima and Krajina (1975). This association is dominated by Hylocomium splendens and Stokesiella oregana in the bryophyte layer, and Berberis nervosa in the shrub layer. The moss association is interpreted as being intermediate in moisture regime to associationscof drier sites, dominated by Gaultheria shallon, and associations of wetter sites, dominated by Polystichum munitum (Kojima and Krajina, 1975). The same interpretation is reached here for the Pseudotsuga-Berberis forests (Fig. 7a and $b$; Tables 22 and 23). Krajina's (1969) biogeocoenoses 18 and 25, and McMinn's (1960) Pseudotsuga-TsugaHylocomium association also correspond to these forests. The Tsuga/ Gaultheria-Berberis/Achlys habitat type of Beese (1981) is somewhat similar, but has much richer herb and shrub strata. The Tsuga/Rhododendron/ Berberis association described by Franklin and Dyrness (1973) for the Tsuga heterophylla Zone of the western Oregon Cascade Range, is very similar to the Pseudotsuga-Berberis forests, although the latter have no Rhododendron macrophyllum. According to Franklin and Dyrness (1973), this association typifies the climatic climax for the western Oregon Cascades. The Pseudotsuga-Berberis forests may also be considered the climatic climax in the vicinity of Port Alberni.

Tsuga-Pseudotsuga-Polystichum forests (P5) (Fig. 18c).

This community type has the largest mean total basal area (158.4 $\left.m^{2} / h a\right)$, and the second largest mean maximum tree height ( 61 m ) in the Pseudotsuga group (Table 22 ). Tsuga heterophylla and Pseudotsuga menziesii
share dominance nearly equally in the tree stratum, but only Tsuga heterophylla is regenerating well (Table 22). The shrub and bryophytelichen layers have small coverages and are poor in species; the herb stratum has a high coverage of Polystichum munitum (Table 23; Fig. 18c).

The Tsuga-Pseudotsuga-Polystichum forests are located centrally inland within the study area, almost exclusively on lower-slopes (Fig. 7b) where they receive seepage and runoff water. They occur from low elevation up to 500 m , on generally south facing, steep to gentle slopes (Fig. 7b). They are always found on deep soils (averagé soil depth is 100 cm ) formed mostly of very rapidly to rapidly drained colluvial material. Some stands are found on nearly level fluvial material, with slower drainage, but only in the driest part of the study area (plots 1 and 17). The organic horizons are moderately thick (average of 10 cm ) and rooting occurs throughout most of the mineral soil. Most soils of this community type are classified as Orthic Humo-Ferric Podzols (Table 29). Traces of fire are evident in most of the stands, and fire is believed to be at the origin of all the stands. This community type shows similarities to the Pseudotsuga-Berberis forests (P4), a commonly adjacent type on upslope topographical positions (less influenced by seepage water), and also to the Montane Tsuga forests (P6) (Figs 7a and b, 13 and 14; Tables 22 and 23).

The Tsuga-Pseudotsuga-Polystichum forests are undoubtedly very similar to the many Pseudotsuga-Polystichum community types or associations described for coastal British Columbia, Washington and Oregon. For British Columbia, the biogeocoenoses $2 b$ and 24 of Krajina (1969),
the Achlys-Polystichum association of Kojima and Krajina (1975), the Polystichum forest type of Orlóci (1961), the Pseudotsuga-Polystichum association of MCMinn (1960), and the Tsuga-Polystichum habitat type of Beese (1981), are comparable to the Tsuga-Pseudotsuga-Polystichum forests. Franklin and Dyrness (1973) describe a similar Tsuga heterophylla/Polystichum munitum community type for Washington and Oregon based on several published descriptions.

Montane Tsuga forests (P6)

These forests have a high mean total basal area ( $114.5 \mathrm{~m}^{2} / \mathrm{ha}$ ) but a relatively low stature (mean maximum height of 50 m ) (Table 22). Tsuga heterophylla dominates the tree stratum, and is regenerating well, while Pseudotsuga menziesii becomes the second dominant (Table 22). The species poor shrub layer is characterized by a high coverage of Vaccinium parvifolium, averaging 1 m in height. The sparse herb stratum shows no characteristic species. Rhytidiopsis robusta, a moss associated with high elevations (Schofield, 1976), is of ten abundant in the bryophyte-lichen layer (Table 23).

This community type occurs within the central and inland portion of the study area, mostly above 400 m in elevation (Fig. 7b). It is found on moderate to steep mid-slopes and upper-slopes, over deep, rapidly to well drained colluvial material. The organic horizons are moderately thick (average of 8.5 cm ) and rooting occurs throughout most of the mineral soil (Appendix 2). Most soils were classified as Orthic Humo-Ferric Podzols (Table 29). Evidence of fire was found in nearly all stands.

This community type shows similarities to the Pseudotsuga-Berberis forests (P4) with which it intergrades at lower elevations (Figs. 7a and $b, 13$ and 14; Tables 22 and 23). McMinn's (1960) Pseudotsuga-TsugaHylocomium association has a bare forest floor variant which is similar to the Montane Tsuga forests.

Montane Tsuga-Gaultheria forests (P7) (Fig. 18a)

This community type has the smallest mean total basal area ( $86 \mathrm{~m}^{2} / \mathrm{ha}$ ) and mean maximum tree height ( 41 m ) of the Pseudotsuga group. Tsuga heterophylla dominates the tree, sapling, and seedling strata (Table 22). The understory is characterized by a nearly continuous, species poor shrub layer, dominated by low ( $<1 \mathrm{~m}$ ) Gaultheria shallon (Fig. 18a). The herb stratum is species poor and very sparse. Rhytidiadelphus loreus is abundant in the bryopnyte-lichen layer (Table 23).

The montane Tsuga-Gaultheria forests occur inland within the study area, generally above 500 m , on moderate to extreme slopes (Fig. 7b). They are found on upper-slopes or ridges, on deep soils formed mostly by colluvial material. The organic horizons are moderately thick (average of 10 cm ) and rooting occurs throughout most of the mineral soil (Appendix 2). Nearly all soils are rapidly to well drained Orthic Humo-Ferric Podzols (Table 29). Half of the stands show no evidence of fire disturbance, but fire probably remains at the origin of all the stands. This community type has similarities with the Pseudotsuga-Linnaea forests (P3) of which it appears to be a high elevation equivalent with Tsuga regeneration. It also has similarities with the montane Tsuga-Abies-Gaultheria forests (A1) (Figs. 7a and $\mathrm{b}, 9 \mathrm{a}$ and b , and 14 ; Tables 22, 23, 26 and 27).

The montane Tsuga-Gaultheria forests are similar to the Pseudo-tsuga-Tsuga-Gaultheria association of MCMinn (1960) and the TsugaGaultheria habitat type of Beese (1981). This type is also similar to Kojima and Krajina's (1975) Gaultheria shallon association, although Tsuga heterophylla does not regenerate successfully in their association. Del Moral and Long (1977) describe a similar Pseudotsuga-Gaultheria community type for the montane forests of western Washington.

## 3. THUJA VEGETATION GROUP

Coastal dry Thuja forests (T1)

The coastal dry Thuja forests have the lowest mean total basal area ( $86 \mathrm{~m}^{2} / \mathrm{ha}$ ) within the Thuja group; they also are low in stature (mean maximum tree height of 30 m ). Thuja plicata, followed by Tsuga heterophy11a, Pseudotsuga menziesii and Taxus brevifolia, dominate the tree stratum. This is the only community type within the Thuja group with an appreciable amount of Pseudotsuga menziesii. Thuja plicata and Tsuga heterophylla seem to be regenerating very well (Table 24). The dense and continuous shrub layer is dominated by Gaultheria shallon and Vaccinium ovatum, averaging 2 and 3 m high, respectively. The herb stratum is relatively poor and sparse for the Thuja group, and consists nearly entirely of Blechnum spicant. The abundance of Rhytidiadelphus loreus and Hylocomium splendens characterizes the well developed bryo-phyte-lichen layer (Table 25).

This community type is restricted to the coastal part of the study area. It is found at low elevations on very strong to extreme slopes, mostly on ridges (Fig. 8b). Soils are rapidly drained and very shallow (average soil depth is 16 cm ). The soils of most stands are Typic Folisols with organic horizons exceeding 10 cm in thickness and directly overlying the bedrock (Appendix 2; Table 29). No evidence of fire was found in these stands, but all show signs of wind disturbance. The relatively dry habitats occupied by this community type contribute to its vegetational similarities with the coastal dry Pinus forests (D2) (Tables 24 and 25). It also is vegetationally similar to the coastal wet Thuja
forests (T5), although the habitats are entirely different (Fig. 8a and b; Tables 24 and 25).

No references to forest types similar to the coastal dry Thuja forests could be found.

Coasta1 Tsuga-B1echnum-Polystichum forests (T2)

This community type has a large mean total basal area ( $142.3 \mathrm{~m}^{2} / \mathrm{ha}$ ) and excellent tree height (average of 52 m ) (Table 24). Tsuga heterophylla dominates the tree stratum and its regeneration occupies nearly the entire sapling and seedling layers (Table 24). Abies amabilis and Thuja plicata are, respectively, the second and third dominants. The shrub layer is comprised mainly of scattered, 2 m high Vaccinium parvifolium. The herb stratum, although poor in species, is well developed and is characterized by an abundance of Blechnum spicant and Polystichum munitum. The bryophyte-lichen layer is poor and has low coverage (Table 25).

This community type is found in the coastal part of the study area. It occurs from low to mid-elevations on very strong to extreme slopes. All stands are situated on mid-slope topographic positions (Fig. 8b). Soils are deep, well to moderately well drained, developing mostly on colluvial material. Half of the stands are found on ancient rock slide colluvial material. The organic horizons are thick (average of 15 cm ) and nearly half of the rooting occurs in them (Appendix 2). Most soils encountered are Orthic Ferro-Humic Podzols (Table 29). No
evidence of fire was found in any of the stands, but most have been subjected to some wind disturbance. This community type shows some similarities with the Tsuga-Blechnum-Polystichum forests (A7), a community type with less Abies amabilis and Thuja plicata, and one which is not strictly coastal in character (Fig. 14; Tables 24, 25, 26 and 27).

The coastal Tsuga-Blechnum-Polystichum forests generally correspond to the biogeocoenotic types described by Klinka et al. (1979) for their Estevan Submontane and West Vancouver Island Submontane Wetter Coastal Western Hemlock biogeoclimatic subzone variants. The coastal Tsuga-Blechnum-Polystichum forests appear to have better drainage than the more widespread coastal Thuja forests (T4). Half of the stands of the coastal Tsuga-Blechnum-Polystichum forests were found on old landslides, which probably created improved drainage and nutrient conditions on these sites.

Coasta1 Montane Thuja forests (T3)

This community type has the largest mean total basal area (187.3 $\mathrm{m}^{2} / \mathrm{ha}$ ) within the Thuja group; the mean maximum tree height is 42 m (Table 24). These forests are dominated by Thuja plicata followed by Tsuga heterophylla and Abies amabilis. All species are regenerating well, except possibly Thuja plicata (Table 24). The species poor shrub stratum has a high coverage, with large contributions by 1 m high Gaultheria shallon and 2.5 m high Vaccinium species. The herb layer is the richest within the Thuja group, but Blechnum spicant remains the dominant species. The rich bryophyte-lichen layer is characteristic of the Thuja group, but otherwise is not distinctive (Table 25).

The coastal montane Thuja forests are found only near the coast from mid-to high elevations, mostly on mid-slope topographic positions of very strong to steep slopes (Fig. 8b). They occur on deep, well to poorly drained soils. The organic horizons are thick (average of 14 cm ) and rooting is mostly limited to these horizons (Appendix 2). The soils, mostly Gleyed or Orthic Ferro-Humic Podzols, or Humic Gleysols, have developed on a variety of materials (Table 29). Traces of fire disturbance were not found, but many stands show signs of wind disturbance in the form of isolated uprooted trees. This community type is very similar to the coastal Thuja forests (T4) which are found at lower elevations on similar sites (Figs. 8a and b, 13 and 14; Tables 24 and 25). Both types intergrade along an elevation gradient (Fig. 8b) and present similar visual aspects, except for a greater abundance of Abies amabilis (particularly in the sapling and seedling strata) in the coastal montane Thuja forests (Table 24).

The coastal montane Thuja forests do not appear to have been described previously.

Coastal Thuja forests (T4) (Fig. 18d)

The coastal Thuja forests have an impressive mean total basal area of $180.4 \mathrm{~m}^{2} /$ ha (Table 24). Thuja plicata dominates, with Tsuga heterophylla and Abies amabilis as second and third dominants, respectively. These species are regenerating well, although not abundantly in the case of Thuja plicata (Table 24). The shrub stratum is well developed, with a large coverage of Gaultheria sha1lon close to 2 m in height, and several

Vaccinium species over 3 m in height (Fig. 18d). The herb layer, one of the poorest in the Thuja group, is strongly dominated by Blechnum spicant (Fig. 18d). The bryophyte-lichen stratum is relatively rich and is characteristic of the Thuja group (Table 25).

The coastal Thuja forests are found near the coast from low to mid-elevations on a variety of topographical positions except ridges (Fig. 8b). This community type occurs on deep, generally imperfectly drained soils, situated on level to strongly sloping terrain. The organic horizons are very thick (average of 20 cm ) and most of the root mass is restricted to them (Appendix 2). The soils almost invariably show signs of $B$ horizon gleying and cementation. They are mostly Humic Gleysols, with some Ferro-Humic Podzols, Ortstein and Duric Humo-Ferric Podzols (Table 29). Because of the frequent cementation of the mineral horizons, most seepage occurs in the organic horizons (this was observed once during a rainstorm). Plot 24 , although not coastal, receives abundant seepage water and supports vegetation typical of coastal sites. Very few stands showed traces of fire, but most had evidence of wind disturbance by uprooted individual trees. This community type is similar to the coastal montane Thuja forests (T3), found at higher elevations on similar sites (Figs. 8a and b, 13 and 14; Tables 24 and 25).

No detailed description of the coastal Thuja forests of the west coast of Vancouver Island seems to have been published. Klinka et al. (1979) list the major species and ecological characteristics of forests described as stands of "decadent" old-growth trees, with very dense and tall shrub layers. In the Estevan Submontane Wetter Maritime Coastal

Western Hemlock biogeoclimatic subzone variant these stands are nearly at climatic climax because of the virtual absence of forest fires (Klinka et al., 1979). These authors also note the considerable relief of the forest floor in these forests caused by the continual windthrow of individual trees. Forests of a similar nature are probably found all along the coast of British Columbia and Washington. Hines (1971) describes a Tsuga-Picea/Gaultheria/Blechnum community type for north coastal Oregon which appears similar to the Coastal Thuja forests of Vancouver Island, especially in the shrub and herb strata, where the dominants are identical. The more southerly location (and thus drier climate) of the Oregoncstands may explain why they contain so little Thuja plicata as compared to the Vancouver Island plots.

Coastal wet Thuja forests (T5)

This community type has one of the lowest mean total basal areas of the Thuja group ( $87.7 \mathrm{~m}^{2} / \mathrm{ha}$ ), and it also has the lowest mean maximum tree height ( 24 m ). The mean tree density ( 855 trees $/ \mathrm{ha}$ ) is the highest in the study area and results from a large number of small trees (Table 24). Thuja plicata dominates the tree stratum with Tsuga heterophy11a and Pinus contorta as co-dominants. Taxus brevifolia is always present. Thuja plicata and Tsuga heterophylla are regenerating well (Table 24). A nearly impenetrable and continuous shrub layer is dominated by 2 m high Gaultheria shallon and Vaccinium ovatum. Pyrus fusca, essentially restricted to this community type, of ten reaches 4 to 6 m in height. The relatively rich herb stratum has a large coverage and is dominated by Blechnum spicant. The rich bryophyte-lichen stratum is characteristic of the Thuja group (Table 25).

The coastal wet Thuja forests occur very near the coast, close to sea level, always on level sites (Fig. 8b). They are found on deep, poorly drained soils on morainal or fluvial deposits. The organic horizons are very thick (average of 20 cm ) and contain most of the root mass (Appendix 2). Most soils are Humic Gleysols (Table 29). Evidence of disturbance by fire or wind was found in some stands. This community type has close vegetational similarities with the coastal dry Thuja forests (Tl), but these two types occur in markedly different habitats (Fig... 8a and b). The coastal wet Thuja forests also have similarities with the coastal Thuja forests (Figs. 8a and b, 13 and 14; Tables 24 and 25).

Communities very similar in appearance to the Coastal wet Thuja forests are described as Coastal forested swamps by Franklin and Dyrness (1973) for western Washington's coastal plain. Klinka et al. (1979) report that, on the west coast of Vancouver Island, Thuja plicata and Pinus contorta are the major species on flat areas and on the lower parts of gentle slopes when there is a large water surplus.

## 4. ABIES VEGETATION GROUP

Montane Tsuga-Abies-Gaultheria forests (A1)

This community type has a large mean total basal area ( $121 \mathrm{~m}^{2} / \mathrm{ha}$ ) and a high mean tree density ( 700 trees/ha), but the smallest mean maximum tree height ( 44 m ) within the Abies group (Table 26). The tree stratum, the richest within the Abies group, is dominated by Tsuga heterophy11a, Pseudotsuga menziesii and Abies amabilis, of which only Pseudotsuga menziesii is not regenerating (Table 26). A continuous shrub layer over 1 m high, the richest within the Abies group, is dominated by Vaccinium alaskaense, Gaultheria shallon and $\underline{V}$. parvifolium. The herb stratum is the poorest in species and the smallest in total coverage within the study area. The bryophyte-lichen layer is well developed and is dominated by Rhytidiade1phus loreus (Table 27).

The montane Tsuga-Abies-Gaultheria forests are found mostly within the central part of the study area. They occur on generally south facing, strong to extreme slopes, from mid- to high elevations, mostly on upperslope topographical positions (Fig. 9b). The deep, rapidly to well drained soils are formed mostly of colluvial material. The organic horizons are moderately thick (average of 9.3 cm ) and rooting is shallow (mean effective rooting depth of 19 cm ) (Appendix 2). The soils are Orthic Humo-Ferric Podzols (Table 29). All the stands show evidence of fire disturbance, some only as reflected by the abundance of Pseudotsuga menziesii, and some by charcoal in the soil. This community type has environmental similarities to the montane Tsuga-Gaultheria forests (P7) and the montane Tsuga forests (P6) (Figs. 7b, 9 b and 14), but it differs
vegetationally from them in the abundance of Abies amabilis and Vaccinium alaskaense. Within the Abies group it is not particularly similar to other types, except for the presence of Abies amabilis regeneration (Figs. 9a and b, 13 and 14; Tables 26 and 27).

The montane Tsuga-Abies-Gaultheria forests are similar to the Chamaecyparis/Gaultheria habitat type described for eastern Vancouver Island by Beese (1981). In this habitat type Abies amabilis is rare in the tree stratum, but seedlings are relatively abundant and Gaultheria shallon dominates the shrub layer. The Abies amabilis/Gaultheria shallon association, described by Franklin and Dyrness (1973) for the Abies amabilis Zone in southern Washington, is somewhat related with a high G. shallon coverage and a very poorly developed herb stratum.

Montane Abies-Tsuga forests (A2)

These forests have a high mean total basal area ( $146.5 \mathrm{~m}^{2} / \mathrm{ha}$ ) and the lowest mean tree density ( 280 trees/ha) in the study area (Table 26). Small trees (not including saplings) are scarce and most trees are found within a narrow range of size-classes. Tsuga heterophylla, Abies amabilis and Thuja plicata dominate the tree stratum with the former two species having equal dominance in sapling and seedling layers (Table 26). The shrub stratum has a large coverage and is dominated by Vaccinium alaskaense over 1 m in height. The herb layer, relatively rich and of high coverage for the Abies group, is characterized by Blechnum spicant. The bryophyte-1ichen stratum has a small coverage and is made up of species generally found within the Abies group (Table 27).

This community type is found above 500 m in elevation in the central part of the study area (Fig. 9b). It occurs on upper-slope positions of extreme slopes, over deep deposits of colluvial or morainal material. The soils are moderately well to imperfectly drained Gleyed FerroHumic Podzols (Table 29). The organic horizons are moderately thick (average of 10.5 cm ) and contain almost all of the roots (Appendix 2). A disturbance origin (fire or wind) of the stands is suspected because of the even-sized structure of the tree stratum. This community type has similarities to the montane Tsuga-Abies forests (A3), which are found on better drained sites, but in wetter coastal areas (Figs. 9a and b, and 14; Tables 26 and 27).

The montane Abies-Tsuga forests correspond to the Rhytidiadelphus-Oval-leaved \& Alaskan Blueberry-Amabilis Fir-Western Hemlock biogeocoenotic type of the West Vancouver Island Montane Wetter Maritime CWH subzone variant (Klinka et al., 1979). Klinka et al. (1979) report that Rhytidiopsis robusta, Gaultheria shallon and Vaccinium parvifolium are more common in stands of drier sites at the upper limit of the variant, and in stands situated the farthest inland. Both stands sampled are found at the eastern limit of the subzone variant mentioned.

Montane Tsuga-Abies forests (A3)

This community type has the lowest mean total basal area (79.5 $\mathrm{m}^{2} / \mathrm{ha}$ ) within the Abies group (Table 26). The species poor tree stratum consists entirely of Tsuga heterophylla and Abies amabilis, with Tsuga heterophylla showing the most abundant regeneration (Table 26). The
sparse and species poor shrub layer is dominated by Vaccinium parvi=: folium. The herb stratum is equally poor and sparse, with Blechnum spicant accounting for most of the coverage. The bryophyte-lichen layer is tẏpical of the Abies group (Table 27).

The montane Tsuga-Abies forests are most of ten found at mid to high elevations near the coast (only once in the central part of the study area). They occur mostly on upper-slope topographic positions of generally north facing, very strong to steep slopes (Fig. 9b). The deep and well drained soils are mostly formed of colluvial material. The organic horizons are thick (average of 15 cm ) and contain most of the roots (Appendix 2). The soils are classified as Orthic Humo-Ferric Podzols (Table 29). Most stands show evidence of wind disturbance, but only one shows evidence of fire. Although this community type is found at, or near, the well drained crests of mountains nearest to the coast, it has no similarities with the community types of the Thuja group, found on adjacent lower topographical positions (montane coastal Thuja forests (T3) or coastal Thuja forests (T4). This type has some similarities to the more coastal montane Abies-Tsuga forests (A2), and to the cooler and wetter montane Abies-Streptopus forests (A4) (Figs. 9a and b, and 14; Tables 26 and 27).

The montane Tsuga-Abies forests have similarities to the Rhyti-diadelphus-Red Huckleberry \& Alaskan Blueberry-Amabilis Fir-Western Hemlock biogeocoenotic type of the West Vancouver Island Submontane Wetter Maritime CWH subzone variant (Klinka et al., 1979). Klinka et al. (1979) report that the most frequent disturbance in these communities is caused by southerly, or westerly, gale force winds. Such disturbance,
particularly on exposed upper-slopes and crests, is responsible for the development of nearly even-aged stands of Tsuga-heterophylla and Abies amabilis (Klinka et al., 1979). In the Montane Tsuga-Abies forests such a structure is frequently observed, giving the impression of a twotiered forest comprising an upper tier of mature trees and a lower tier of saplings.

Montane Abies-Streptopus forests (A4)

These forests have a high mean total basal area ( $122 \mathrm{~m}^{2} / \mathrm{ha}$ ) as well as a high mean maximum tree height ( 53 m (Table 26)). Only Abies amabilis, the dominant species, and Tsuga heterophylla form the tree stratum. Abies amabilis shows the best regeneration (Table 26). The coverage of the shrub stratum, the poorest in species within the study area, is almost entirely made up by Vaccinium alaskaense. Oplopanax horridus is nearly always present. The herb layer, with the largest coverage and number of species within the Abies group, is characterised by Rubus pedatus, Streptopus roseus and S. streptopoides. Several fern species, indicative of moist and:nutrient-rich soils, are present (Adiantum pedatum, Athyrium filix-femina and Gymnocarpium dryopteris). The sparse bryophyte-lichen layer is characteristic of the Abies group, except for the only occurrence within the study area of Eurhynchium pulchellum in two stands on limestone bedrock (Table 27). This moss is usually found on calcareous substrata (Schofield, 1976).

This community type is found in the central part of the study area above 600 m . It occurs on mid-slope and upper-slope topographic positions
of very strong, generally north facing slopes (Fig. 9b). Soils are deep, moderately well to imperfectly drained, and have formed from colluvial material. The organic horizons are very thick (average of 21 cm ) and contain most of the roots (Appendix 2). The soils belong to various types (Table 29), but most are gleyed as a result of constant seepage (as the presence of Oplopanax horridus indicates). No evidence of fire disturbance was found, but wind disturbance was noticed in the two stands nearest to the ocean. This community type is environmentally quite different from other community types, but is perhaps closest to the montane Abies-Tsuga forests (A2) or to the montane Tsuga-Abies forests (A3) (Figs. 9a and b, 13 and 14; Tables 26 and 27). The montane Abies-Streptopus forests are quite similar in composition to the lowland Abies forests (A5), although they occur at totally different elevations (Fig. 9a and b).

The Abies/Vaccinium alaskaense/Streptopus habitat type described for eastern Vancouver Island by Beese (1981) corresponds nearly exactly to the Montane Abies-Streptopus forests. These forests also are similar to the Streptopo-Abietum association described by Brooke et al. (1970) for the Coastal Subalpine Mountain Hemlock Zone. This association is characterized by constant seepage (Brooke et al., 1970). Kojima and Krajina (1975) pointed out the resemblance of the Streptopo-Abietum association with their Vaccinium alaskaense association situated at lower elevations. In this study area, the Abies-Streptopus forests and the lowland Abies forests (comparable to the $\underline{V}$. alaskaense association) also are floristically similar (Fig. 9a and b; Tables 26 and 27). The Abies amabilis/Streptopus roseus association described by Franklin and Dyrness (1973) for the Abies amabilis zone of Washington also is related.

Lowland Abies forests (A5) (Fig. 18f)

The lowland Abies forests have a low mean total basal area ( $107.7 \mathrm{~m}^{2} / \mathrm{ha}$ ) but an excellent mean maximum tree height ( 54 m ) compared to other community types of the Abies group (Table 26). Abies amabilis achieves its highest dominance in the study area (Fig. 18f), and Tsuga heterophylla is the second dominant in a two species tree stratum. Both species are regenerating (Table 26). The moderately developed shrub layer is made up of Vaccinium alaskaense and V. parvifolium averaging 2 m in height. The herb stratum is marked by the abundance of Blechnum spicant, Dryopteris austriaca and Tiarella trifoliata. The well developed bryophyte-lichen stratum, although the richest within the Abies group, has no particularly characteristic species.

The lowland Abies forests occur throughout the study area, except for the coastal plain and the driest inland areas. They occur at low to mid-elevations, on the lower-slopes or terraces of narrow river valleys; aspects most often are north facing (Fig. 9b). The topographic location suggests that the sites are subjected to cold air drainage (Miller et al., 1983) or snow accumulation (Kojima and Krajina, 1975). The lowland Abies forests are generally found on strong slopes, with deep, moderately well drained soils formed from a variety of materials (colluvial, morainal, fluvial). The organic horizons are thick (average of 15 cm ) and half of the root mass is found within them (Appendix 2). Evidence of wind disturbance was found in less than half of the stands, and fire disturbance was found in only two stands. This community type shows environmental similarities with the Tsuga-Blechnum-Polystichum forests (A7) and the
coastal Tsuga-Blechnum-Polystichum forests (T2) (Figs. 13 and 14). Vegetationally, it is most similar to the montane Abies-Streptopus forests (A4), although this type occurs at a much higher elevation (Figs. 9a and b, 13 and 14; Tables 26 and 27).

The lowland Abies forests fit closely the description given by Kojima and Krajina (1975) of the Vaccinium alaskaense association found in Strathcona Provincial Park. These authors indicate that most of their stands were found on terraces close to the bottom of protected valleys, or on the gentle slopes of hillsides. They suggest that snow accumulation may explain the presence of this association which usually occurs at higher elevations. In the present study area, stands of the lowland Abies forests were found in very similar sites to those described by Kojima and Krajina (1975). On eastern Vancouver Island, the Abies/ Vaccinium alaskaense-V. parvifolium habitat type described by Beese (1981) has similarities with the lowland Abies forests. Franklin and Dyrness (1973) report that the climatic climax community for the Abies amabilis Zone ( $600-1300 \mathrm{~m}$ ) of the northern Washington Cascade Range is an Abies amabilis/Vaccinium alaskaense association with an abundance of mesic herbs and Rubus pedatus. However, the lowland Abies forests described here are not considered zonal plant communities because of the particular topographic factors influencing their microc1imate.

Tsuga-Gaultheria-Blechnum forests (A6)

This community type has a low mean total basal area ( $116.5 \mathrm{~m}^{2} \% \mathrm{ha}$ ) and a low mean maximum tree height ( 46 m ), relative to the other types
within the Abies group (Table 26). Tsuga heterophylla strongly dominates the tree stratum and is the major regenerating species. Abies amabilis is the second dominant, but is not regenerating abundantly (Table 26). The species poor shrub stratum has a large coverage, dominated by Gaultheria shallon (over 1 m high) and Vaccinium parvifolium (2 to 3 m high). The herb layer is the most species-poor within the study area and is entirely dominated by Blechnum spicant. The bryophyte-lichen stratum, with the highest coverage within the Abies group, is characterized by an abundance of Stokesiella oregana (Table 27).

The Tsuga-Gaultheria-Blechnum forests were found on the coast and in the central part of the study area, on low elevation mid- and lower-slope topographic positions (Fig. 9b). The soils are deep, moderately well drained, Gleyed Humo-Ferric Podzols formed from morainal material (Table 29). The organic horizons are moderately thick (average of 11.5 cm ), and roots are abundant in the upper mineral horizons (Appendix 2). No evidence of fire was found, but one stand did show signs of wind disturbance. This community type has environmental similarities with several other community types, (Figs. 13 and 14), but is relatively distinct vegetationally, except for some similarities with some types of the Thuja group (Fig. 9a; Tables 24, 25, 26 and 27).

Tsuga-Blechnum-Polystichum forests (A7) (Fig. 18e)

This community type has the highest mean total basal area. ( $185.2 \mathrm{~m}^{2} / \mathrm{ha}$ ) and the highest mean maximum tree height ( 64 m ) of the Abies group (Table 26). Tsuga heterophylla dominates the tree stratum
in all stands and is the only species regenerating. Pseudotsuga menziesii, Thuja plicata and Picea sitchensis are the second dominants depending on the stand (Table 26). The shrub layer is the most species-poor within the study area. Dominance of the herb stratum is shared equally by Blechnum spicant and Polystichum munitum (Fig. 18e). The bryophytelichen layer is sparse, but generally characteristic of the Abies group (Table 26).

This community type is found from the coast to the central part of the study area, but is absent from the dry inland sector. These forests occur on the lower and mid-slope topographic positions of very strong slopes, mostly at low elevations (Fig. 9b). The soils are deep and well to moderately well drained. In most cases soils have formed from morainal material, except those of two coastal stands which have developed from colluvial material originating from ancient land slides. Better drainage on the colluvium may account for the vegetational similarities between the coastal and the inland stands of this type. The organic horizons are thick (average of 13 cm ) and contain most of the root mass (Appendix 2). Most soils are Orthic Humo-Ferric Podzols; also represented are a Gleyed Humo-Ferric $P o d z o l$ and a Gleyed Ferro-Humic Podzol (Table 29). Evidence of fire disturbance was found in two of the central stands, and land slides were at the origin of the two coastal stands. This community type is most similar to the coastal Tsuga-Blechnum-Polystichum forests (T2) (Fig. 14; Tables 24, 25, 26 and 27). The difference is mostly one of geographical location, resulting in a wetter soil moisture regime for the coastal type because of higher amounts of precipitation. This is reflected by a lesser amount of Pseudotsuga
menziesii and Polystichum munitum, and a higher amount of Abies amabilis and Blechnum spicant in the coastal Tsuga-Blechnum-Polystichum forests (T2), than in the Tsuga-Blechnum-Polystichum forests.

The Tsuga-Blechnum-Polystichum forests are very similar to the better drained and most productive stands of the Rhytidiadelphus-Red Huckleberry \& Alaskan Blueberry-Amabilis Fir-Western Hemlock biogeocoenotic type within the West Vancouver Island Submontane Wetter Maritime CWH Subzone variant (Klinka et al., 1979) ; however the Tsuga-BlechnumPolystichum forests described here probably have a drier soil moisture regime as indicated by the predominance of Tsuga heterophylla and Pseudotsuga menziesii over Abies amabilis (Table 26).

## 5. FLOODPLAIN VEGETATION GROUP

Floodplain forests (F1)

Floodplain forests possess the largest mean total basal area in the study area ( $246.2 \mathrm{~m}^{2} / \mathrm{ha}$ ), as well as a high mean maximum tree height ( 60 m (Table 24)). The dominant tree is most often Picea sitchensis, but Thuja plicata and even Pseudotsuga menziesii dominate in some of the plots. Tsuga heterophylla is, on average, the second dominant and is the only species regenerating well (Table 24). Rubus spectabilis, Ribes bracteosum and Vaccinium species often form a continuous shrub layer over 2 m in height. The herb stratum, the richest in the study area, is dominated by Polystichum munitum and Athyrium filix-femina which form a continuous, one metre high layer in some stands. Many herb species such as Trautvetteria caroliniensis, Melica subulata, Luzula parviflora and Aruncus sylvester are restricted to this community type. Leucolepis menziesii, Plagiomnium insigne and Stokesiella praelonga characterize the bryophyte-lichen stratum (Table 25).

This community type is found at low elevations throughout the study area on the floodplains of major rivers. The shrub stratum is often absent or weakly developed in younger stands, or stands situated on the most active floodplains. In contrast, such stands have a well developed herb stratum with up to $85 \%$ coverage of Polystichum munitum. Older stands, or stands on less active floodplains, have a dense shrub layer (mainly Rubus spectabilis) and usually have a few very large Picea sitchensis. The soils are deep, well drained loams of alluvial or fluvial nature and contain very few coarse fragments; the organic horizons
are thin and tree roots penetrate deeply into the mineral soil (Appendix 2). The water table was not encountered in any of the soil pits; however, the soils remain moist owing to their fine texture and the level terrain. The soils are mostly Gleyed Sombric Brunisols (Table 29). Signs of wind disturbance, mostly in the form of uprooted large Picea sitchensis, were found in half of the stands; a few stands contained evidence of fire. Floodplain forests have few similarities to other types described for the study area. Their closest affinity is with the Thuja group (Figs. 5, 13 and 14; Tables 24 and 25).

Floodplain forests correspond closely in vegetation composition and ecological characteristics to the Picea sitchensis-Polystichum munitum-Leucolepis menziesii forest type described by Cordes (1972) for floodplains of the west coast of Vancouver Island. First and second river terrace communities described by Fonda (1974) for the Olympic National Park, Washington State, also have similarities with the floodplain forests. The former differ from the latter by having a lesser coverage of Rubus spectabilis (probably because of elk browsing) and an abundance of Oxalis oregana in the understory. Phytogeographically, the Picea sitchensis dominated floodplain forests range along the coast from southern Alaska to southern Oregon where they merge with the Redwood forests (Fonda, 1974). Oxalis oregana is a characteristic element of these forests in Washington, Oregon, and California, but it was found only near the Klanawa River (plots 170 et 171) within the study area. The phytogeography of this species in British Columbia has recently been discussed by Ogilvie et al. (1984).

## Floodplain forests (Lysichitum variant) (F2)

This community type has a structure similar to the typical floodplain forests (F1), with a very large mean total basal area ( $236.5 \mathrm{~m}^{2} / \mathrm{ha}$ ) and a high mean tree height (56 m (Table 24)). Picea sitchensis and Thuja plicata share dominance equally. Tsuga heterophylla is the third dominant and is the only species regenerating well (Table 24). The dense and almost continuous shrub stratum (over 3 m high) is dominated by Gaultheria shallon, Rubus spectabilis and Vaccinium species. The high coverage herb layer is dominated nearly equally by Blechnum spicant, Lysichitum americanum and Polystichum munitum. The bryophyte-1ichen stratum is similar in composition to that of the typical floodplain forests (F1).

The Lysichitum variant of the floodplain forests was found in only one area near the coast, on the alluvial plain of a small river. The soils are very poorly drained Humic Gleysols formed from alluvial deposits (Table 29). The organic horizons are thin, and the effective rooting depth (averaging 80 cm ) is restricted by a shallow water table (Appendix 2). Evidence of fire and wind disturbance was found. Apart from the poorer drainage and heavier soil texture, this community type is similar to the typical floodplain forests (Figs. 5 and 14; Tables 24 and 25).

The Lysichitum variant of the floodplain forests is very similar, floristically and ecologically, to the Picea sitchensis-Lysichitum americanum forest type described by Cordes (1972) for the west coast of Vancouver Island.

## 6. SUBALPINE VEGETATION GROUP ( SA)

The subalpine vegetation group is formed by an heterogeneous assemblage of eleven plots ranging in elevation from 485 m to 1050 m (mean $=789 \mathrm{~m}$ ). Most plots of this group probably represent low elevation extensions of vegetation types more common above the upper elevation limit set for this study; therefore, the description of this group is general.

Tsuga heterophy11a dominates the tree stratum; Abies amabilis is the second dominant and also is the most abundantly regenerating species (Table 28). Tsuga mertensiana and Chamaecyparis nootkatensis also are important in the tree stratum. Both of these species are characteristic of high elevation coastal forests (Krajina, 1969; Brooke et al., 1970; Klinka et al., 1979). Pseudotsuga menziesii is relatively abundant in many stands, presumably because of past fire disturbance. A general pattern also can; be described for the understory strata. Most stands have a shrub layer strongly dominated by Vaccinium alaskaense (up to $50 \%$ coverage). The herb stratum is generally sparse and is characterized by species such as Clintonia unifolia, Rubus pedatus, Tiarella unifolia and Veratrum viride. The bryophyte-lichen layer is usually well developed, with total coverages of up to $75 \%$, and is always strongly dominated by Rhytidiopsis robusta. Other species, such as Vaccinium membranaceum, Gaultheria ovatifolia, Phyllodoce empetriformis and Viola orbiculata, were found most often within this group. Rhododendron albiflorum and Fritillaria camschatcensis were found only in plots of the subalpine vegetation group.

The environmental conditions (mainly the soil moisture and nutrient status) appear to vary widely in this group (Appendix 2). Associations within the subalpine Mountain Hemlock Zone of coastal British Columbia have been described by Brooke et al. (1970). Klinka et al. (1979) describe eastern and western variants withị a Maritime Forested Mountain Hemlock biogeoclimatic subzone for Vancouver Island. Fonda and Bliss (1969) discuss an Abies amabilis-Tsuga mertensiana community type for the Olympic Mountains of Washington. Franklin and Dyrness (1973) also discuss the communities of the Tsuga mertensiana Zone of the Pacific Northwest States.

## D. VEGETATION STRATA HOMOGENEITY AND SPECIES RICHNESS WITHIN TYPES

The average homogeneity coefficients and the average species richness values of vegetation strata in fourteen community types are summarized in Table 31. The seedling stratum, followed closely by the tree and sapling strata, are, on average over all types, the most homogeneous in the study area. Most community types are composed of oldgrowth stands with closed canopies where seedling establishment is restricted almost entirely to a few shade tolerant species; thus, a high degree of uniformity within the seedling stratum is to be expected. Community types with open canopies, such as the dry Pinus-Pseudotsuga forests (D1), the coastal dry Pinus forests (D2), and some stands within the Floodplain forests (F1) show a much lower homogeneity in their seedling strata. The lower homogeneity of the tree stratum may reflect variation in the type and intensity of disturbances at the origin of the stands. The tree stratum of ten contains large proportions of long-lived, shade intolerant species (eg. Pseudotsuga menziesii) which are usually absent in seedling strata.

The shrub stratum is less homogeneous than the tree, sapling, and seedling strata, but is, on average, more homogeneous than the herb and bryophyte-1ichen layers (Table 31). The coastal wet Thuja forests (T5) have the most homogeneous and one of the most species-rich shrub strata. The least homogeneous shrub stratum is found in the Floodplain forests (F1) and may reflect variations in flooding regime or stand age (see Chapter 4. C, section 5).

The most homogeneous herb strata occur in the Thuja group community types, because of the strong dominance of Blechnum spicant growing profusely on the thick organic horizons to the exclusion of nearly all other herbs. The least homogeneous herb strata are found in the montane Tsuga forests (P6) and the montane Tsuga-Gaultheria forests (P7). The herb strata of these two types (P6 and P7) have very low total coverages and, although average species richness is low, the total number of species encountered is relatively high (Table 23), resulting in herb stratum heterogeneity within the types.

The bryophyte-1ichen layer is overall the least homogeneous of all the vegetation strata, possibly because it reflects varying microsite conditions from stand to stand (microtopography, number of fallen logs, area of bare rock surfaces, etc.).

The average homogeneities of community types over all strata are also shown in Table 31. The coastal Thuja forests (T4) and the coastal wet Thuja forests (T5) have the most homogeneous vegetation; the Floodplain forests (F1) have the least homogeneous vegetation. Thuja types T4 and T5 occur at low elevation very near to the coast where the climate and other environmental conditions are more uniform (no dry summer period, no freezing, abundant seepage, thick organic layers, etc.), and where fire disturbances are unusual (see fire index for Thuja types in Table 31) and wind disturbances not as frequent as at high elevations. The combination of these factors may be responsible for the extremely homogeneous vegetation. By contrast, numerous tree species can grow very successfully on floodplains where nutrients and moisture are abundant. The
establishment of seedlings following a major disturbance on a floodplain is probably mostly a result of stochastic events and proximyty of seed sources, rather than environmental factors, leading to the lack of homogeneity in the tree stratum. Further heterogeneity is introduced by the flooding regime which may favor or hinder the development of a particular understory species or stratum.

There does not appear to be a clear, general relationship between the mean richness of a stratum and its homogeneity. The shrub layer tends to be slightly more homogeneous with increasing richness ( $\mathrm{x}=.349$ ), but the herb layer tends to be slightly less homogeneous with increasing richness $(r=-.349)$. Herb and shrub layers also tend to be less homogeneous with increasing fire index values (herbs, $r=-.589$; shrubs, $r=-.425$ ), while the tree stratum tends to be more homogeneous ( $r=.358$ ). The interpretation of the fire index values is limited because they do not include information on fire intensity or frequency. The fire index values do indicate that fires have occurred predominantly in dry PinusPseudotsuga forests (D1) and in all Pseudotsuga types (P3 to P7 in table 31). Fire may be said to be a rare occurrence in types where the fire index value is smaller than its standard deviation (an estimate of the variability).

## E. TREE SIZE-CLASS STRUCTURE OF COMMUNITY TYPES

The size-class distributions of tree species can offer some insight into community dynamics so long as certain limitations of this approach are borne in mind (Harper, 1977). Individuals of a single tree species which become established on a site following a disturbance usually will not have the same size, even when the time of establishment is identical. Within site differences in nutrient or water availability, and presence of competing neighbours may cause size differences between trees of identical age. In addition, the establishment of trees following a disturbance is often spread over several years, during which time conditions of resource availability and competition may also vary. For these reasons, even typically seral species will show a wide range of sizes (although a peak number of stems is usually found in one size-class) in stands that owe their origin to a single disturbance in the past. This effect is amplified here because data from several plots were compiled for each community type; thus apart from the likely environmental differences between plots, the time and intensity of the disturbance also may vary. Nevertheless, the graphs (Figs. 15-17) do illustrate differences in size class structure between seral tree species, whose presence results from; past disturbance, and shade tolerant species, whose regeneration is not strictly linked to disturbance. Shade tolerant or "climax" species, will be referred to as "primary" species in the terminology of Brokaw (1980). Stem numbers in the size-class distribution figures (Figs. 15, 16 and 17) are on a logarithmic scale. It is noted that the classical, reversed "J"-shaped curve remains characteristic of primary species, while a bellshaped distribution of sizes is associated with seral or pioneer species.

## 1. PSEUDOTSUGA TYPES

The size-class distribution curves of major tree species in different community types of the Pseudotsuga group are generally similar (Fig. 15). Pseudotsuga menziesii, with most of its stems in the larger size-classes, is characteristic of a seral species; however, this trend is less obvious in community types restricted to the driest, low elevation, inland part of the study area (types D1, P1, P2 and P3). Only in the dry Pinus-Pseudotsuga forests (D1), on rock outcrops, does the size-class distribution of Pseudotsuga menziesii appear characteristic of a primary species. In all types where they are important, Tsuga heterophylla and Thija plicata have size-class distributions characteristic of primary species. In community types with progressively greater soil moisture supply (types P3 to P5, and the higher elevation type P6), Tsuga heterophy11a is increasingly found in larger size-classes. In the montane Tsuga forests (P6), the size-class containing most Pseudotsuga menziesii stems also contains an equivalent number of Tsuga heterophylla stems. This contrasts with the drier, lower elevation PseudotsugaBerberis forests ( P 4 ), where the size-class with the most Pseudotsuga menziesii stems has very few Tsuga heterophylla stems (Fig. 15). This difference could be caused by a cooler and wetter climate at higher elevations allowing Tsuga heterophylla to grow as rapidly as Pseudotsuga menziesii after a disturbance. Double peaks can be seen in the size-class distribution curves of Pseudotsuga menziesii in types P2 and P7. Because the plots of the Pseudotsuga-Thuja-Acer forests (P2) are very similar environmentally (Appendix 2) and are spatially close (within a 2 km
radius of the Dog Mountain peninsula on Sproat Lake, Fig. 1), it is proposed that the two peaks correspond to two distinct fire disturbances in the past. The largest Pseudotsuga menziesii trees all have fire charred bark. The double Pseudotsuga menziesii peaks in type $P 7$ cannot be interpreted similarly with equal confidence since the plots are scattered spatially.

## 2. THUJA TYPES

Pinus contorta, in the coastal wet Thuja forests (T5), is the only example of a seral species size-class distribution within the Thuja group (Fig. 16). In the coastal dry Pinus forests (D2) Pinus contorta appears to be a primary species. Abies amabilis, Tsuga heterophylla, and Thuja plicata all have size-class distributions characteristic of primary species (Fig. 16). In the coastal montane Thuja forests (T3) and the coastal Thuja forests (T4), the curves for Thuja plicata are strongly skewed toward the larger size-classes (Fig. 16). This particular sizeclass distribution may reflect the very large sizes attained by Thuja plicata as well as its impressive longevity of approximately 1000 years, which is twice that of Tsuga heterophylla and Abies amabilis (Waring and Franklin, 1979). Thuja plicata also shows low mortality when mature because of its high resistance to fungal and insect attack (Minore, 1979).

## 3. ABIES TYPES

In most of the community types presented in Fig. 17, Abies amabilis and Tsuga heterophylla are considered primary species able to regenerate in the understory; however, wind disturbance may cause the occasional pulse-1ike establishment of trees, indicated by peaks in the larger size-classes of some community types (A3, A4, A5 and T2).

Abies amabilis presents a size-class distribution curve characteristic of a primary species in the A1, A4, and A5 community types. Tsuga heterophy11a may also be considered a primary species in the Al , A7, and T2 types; however its distribution curves have peaks in the larger size-classes of the A3, A4 and A5 types, suggesting that disturbance may partly explain its presence in these types. The montane Tsuga-Abies forests (A3), situated on upper mountain slopes and ridges near the coast, are particularly susceptible to wind disturbance causing partial or total blowdowns. Stands of this community type have been observed in various stages of recovery following wind disturbance. A twotiered structure caused by wind disturbance in coastal mountain Abies amabilis and Tsuga heterophylla forests has been described by Klinka et al. (1979). The upper layer consists of mostly even-sized, dominant trees; numerous small trees and saplings characterize the understory. Pseudotsuga menziesii's size-class distribution curve in the montane Tsuga-Abies-Gaultheria forests (A1) is characteristically seral (Fig. 17).
F. TREE SEEDLING ABUNDANCE ON UNDECOMPOSED

WOOD AND FOREST FLOOR SUBSTRATA

The abundance patterns of seedlings of the major tree species on undecomposed wood and forest floor substrata vary greatly within the study area (Table 32). The abundance of Tsuga heterophylla seedlings on undecomposed wood is almost always significantly higher than on the forest floor in the community types studied (Table 32). In community types of the Thuja group, Thuja plicata seedlings are always significantly more abundant on undecomposed wood (Table 32); however, in the drier, inland types (P4 and P6), Thuja plicata seedlings, although of low densities, are equally abundant on both substratum classes (Table 32). Abies amabilis seedlings are, in general, equally abundant on undecomposed wood and forest floor substrata. Pseudotsuga menziesii seedlings occur in equal amounts on both substratum classes in the two community types ( P 4 and P 6 ) where there were sufficient data for analysis (Table 32). Pseudotsuga menziesii is a long-lived, seral species in these closed canopy, old-growth forests, and is absent from the sapling size-class (Fig. 15). Seedling establishment conditions for this shade-intolerant species are poor, as reflected by low seedling densities (Table 32). Patterns of Pseudotsuga menziesii seedling abundance on organic or mineral soil substrata are likely to be different in a more open environment following a forest fire.

## DISCUSSION

## A. VEGETATION ANALYSIS

## 1. GENERAL VEGETATION PATTERNS

The interpretation of the 172 and 140 plots ordinations (Figs. 5 and 6) supports the first two hypotheses formulated in the Introduction. These hypotheses proposed that, in order of decreasing importance, macroclimatic and soil parent material factors would correlate most strongly with the vegetation patterns. In the 172 plots ordination, an elevation macro-climatic gradient is reflected in the vegetation pattern expressed along the first and second axes (Fig. 5), leading to the isolation of a group of subalpine vegetation plots. Low elevation plots, belonging to the floodplain vegetation group and the Pinus contorta vegetation group of rock outcrops, also are isolated on the first axis of this ordination, reflecting a soil parent material gradient, secondary in importance to the macro-climatic gradient which is reflected on two axes (Fig. 5).

The highest elevations represent the cooler end of the macroclimatic gradient, where continuous snow accumulation occurs during winter months. This, in turn, has a marked effect on organic horizons structure, and on tree seedling establishment and survival (Brooke et al., 1970; Klinka et al., 1979). Abies amabilis seedlings are reportedly superior to Tsuga heterophylla seedlings at resisting mechanical damage caused by litter debris accumulating in winter snow packs (Thornburg, 1969).

The larger size of the Abies amabilis seedling (Schopmeyer, 1974) is probably very important in this aspect. Abies amabilis is undoubtedly regenerating the most successfully of all the tree species within the subalpine vegetation group (Table 28).

The parent material gradient separating the floodplain vegetation group from the Pinus contorta group contrasts the droughty, nutrient poor residual soils of rock outcrops, against the moist, nutrient rich alluvial soils of floodplains. Species such as Pinus contorta, Arbutus menziesii, Rhacomitrium lanuginosum and Vaccinium ovatum characterize the rock outcrops (Table 3), and are either tolerant of drought and poor soil nutrient conditions, or they are shade intolerant (Minore, 1979). Arbutus menziesii and Arctostaphylos columbiana are found only on the dry and hot microclimates of rock outcrops near the northern boundaries of their ranges. Rock outcrops of the interior of the study area also represent the only habitats where Pseudotsuga menziesii regenerates within the study area (Table 22).

The characteristic species of floodplains, such as Picea sitchensis, Rubus spectabilis, Ribes bracteosum, Polystichum munitum and Trautvetteria carolinensis, likely have high edaphic requirements, such as abundant moisture and high soil nutrient levels, combined with good drainage. Polystichum munitum probably requires relatively high levels of soil nutrients based on the high potassium (2 \%) content of its leaves (Klinka, 1974). Rubus spectabilis and Ribes bracteosum, along with other floodplain species such as Adenocaulon bicolor, Athyrium filix-femina and Melica subulata (Table 25) are known as "nitrate accumulators" from the easily
detectable nitrates in their leaves (Krajina et al., 1982, p. 57). Picea sitchensis is also beleived to require relatively high soil nutrient levels (Krajina et al., 1982; Minore, 1979).

The Pseudotsuga, Thuja and Abies vegetation groups were differentiated within an ordination of 140 plots after the removal of plots from previously identified groups (Fig. 6). Strong correlations with distance from the coast, on both the first and second axes (Table 6), indicate that a macro-climatic gradient is again linked to the vegetation patterns expressed in this ordination. Plots of the Pseudotsuga group are found in the part of the study area farthest from the coast, while plots of the Thuja group are found only near the coast. Coastal areas receive $50 \%$ more annual precipitation than the Port Alberni surroundings (Fig. 2). The effect of this steep rainfall gradient on the vegetation is explored further in section 6 (The climatic master gradient). Plots of the Abies group do not have particular geographical affinities, and although a few are intermediate in geographical location between plots of the Pseudotsuga and Thuja groups, most are associated with high elevations or otherwise micro-climatically cooler sites, such as the bottom of steep-walled valleys subjected to cold:air drainage or delayed snowmelt, or to the base of steep north-facing slopes (Fig. 9b).

A canonical analysis of the six vegetation groups reveals a clear separation of the groups based on environmental data (Fig. 13). However, the floodplain group, the Pinus contorta group and the subalpine group are not as well separated as in the reciprocal averaging vegetation ordination (Fig. 5). This is probably results from the abundance of edaphic variables and the lack of direct climatic variables in the environmental
data matrix used in the canonical analysis (Table 1). In this case, the ordination based on vegetation data is believed to reflect relationships more accurately (Fig. 5). Predictably, the canonical analysis reveals that the vegetation group most environmentally similar to the subalpine group is the Abies group (Table 16). Within the three larger vegetation groups, the Pseudotsuga and Thuja groups are the least environmentally similar (Table 16). Important macro-climatic differences between coastal areas (Thuja group) and more inland areas (Pseudotsuga group) are reflected in these results based on non-climatic variables. Several edaphic variables, such as organic horizons thickness, rooting characteristics, percent carbon and nitrogen in $B$ horizons, and type of disturbance vary along the distance from the coast gradient (Table 6), and are directly or indirectly related to climate. A gradual change occurs from a fire dominated disturbance regime inland, towards the coast where fire is virtually absent and the main disturbance factor is wind. In several plots of the Thuja group, windthrow of a few individual trees was the most frequently observed disturbance, although a few large scale wind disturbances were seen (e.g. plots 72 and 151). All of the dominant tree species in the Thuja group, such as Thuja plicata, Tsuga heterophy11a and Abies amabilis are characteristically shallow rooting (Minore, 1979); conversely, Pseudotsuga menziesii is a deep rooting species (Minore, 1979). The root distribution recorded in soil profiles (Appendix 2) reflects forest composition, which is linked in turn to disturbance type, and both are related to climate. The shallow rooting habit of trees near the coast, where the strongest winds occur, may increase the incidence of windthrow. Thuja plicata, the dominant tree species in forests near the coast, is regarded
as being more wind-resistant than Abies amabilis and Tsuga heterophylla (Minore, 1979). Pseudotsuga menziesii, the dominant tree species inland, is considered the most fire resistant of the coastal tree species, and Thuja plicata the least (Minore, 1979). Some community types within the Abies group, such as the montane Abies-Streptopusforests (A4) and the lowland Abies forests (A5), show little disturbance by either fire or wind. Montane Abies-Streptopus forests (A4) occur on steep, but moist, high elevation sites away from the coast, where strong winds and forest fires are infrequent. The montane Tsuga-Abies forests (A3), however, are situated in relatively high elevation sites near the coast and show abundant windthrow.

Soil organic matter accumulation is greatest in forests near the coast, where plant production is nearly continuous owing to the mild climate and plentiful soil moisture (Valentine, 1971). Nutrient cycling may occur predominantly in the thick $H$ horizon of these forests through the intermediary of a recently discovered indigenous earthworm (Spiers et al., 1984). This phenomenon seems closely linked to the presence of most of the tree root mass within the organic horizons of these forests (Spiers et al., 1984). Thuja plicata roots are apparently more numerous in organic layers, when these are thick, than in the underlying soil (Ross, 1932).

Percent carbon is also higher in the mineral $B_{1}$ horizons of stands closest to the coast. This reflects a climatically controlled soil gradient from a predominance of Humo-Ferric Podzols inland to a predominance of Ferro-Humic Podzols near the coast (Jungen and Lewis, 1978).

Other environmental variables, such as elevation, percent slope and drainage quality decrease towards the coast, and parent material is more frequently morainal than colluvial (Table 6). These environmental factors are likely associated with changing topography and not related to climate. An exception might be drainage, because it is partially defined on annual duration of soil saturation (correlated with amount of soil mottling), which could be different for identical sites depending on the total amount of precipitation and evapotranspiration. Morainal surficial material, predominant near the coast, has slower drainage than colluvial material, prevalent inland.

## 2. THE PSEUDOTSUGA GROUP

Within the area where plots of the Pseudotsuga group are found, macro-climate and soil parent material are fairly homogeneous. The environmental factors most closely associated with vegetation patterns appear to be at the scale of meso-climate and soil moisture (Figs. 7a and b, Table 8). These observations support hypotheses lc and 1d formulated in the Introduction. Meso-climate and soil moisture have been found repeatedly to play a major role in the distribution of vegetation in mountainous areas when macro-climate and parent material were uniform (Whittaker, 1956; 1960; Whittaker and Niering, 1965). The direct ordination of the Pseudotsuga group shows a clear relationship between vegetation patterns and elevation (= meso-climate) and topographicmoisture gradients (Fig. 7b). Low elevation sites with abundant soil moisture, often at the base of mountain slopes, are most frequently occupied by Tsuga-Pseudotsuga-Polystichum forests (P5). Dry sites at higher elevations ( $500-800 \mathrm{~m}$ ), such as crests, ridges and steep upperslopes, are occupied by the montane Tsuga-Gaultheria forests (P7). On sites of intermediate moisture regime, an altitudinal gradient can be followed through the low elevation, warmest Pseudotsuga-Thuja-Acer forests (P2), to the mid-elevation Pseudotsuga-Berberis forests (P4), and to the cooler montane Tsuga forests (P6) (Fig. 7a and b).

Dry Pseudotsuga forests (P1), Pseudotsuga-Linnaea forests (P3) and Pseudotsuga-Thuja-Acer forests (P2) are clearly differentiated vegetationally (Fig. 7a, Tables 22 and 23) but appear similar environmentally in the canonical analyses (Figs. 13 and 14). This discrepancy
may be explained by a combination of fire disturbance and soil moisture differences, unaccounted for by the environmental variables used in the canonical analyses, but nevertheless reflected by the vegetation. The dry Pseudotsuga forests (P1), because of the almost exclusive presence of large, scattered Pseudotsuga menziesii trees with charred bark, appear to have had the most recent fires. Pseudotsuga-Thuja-Acer forests (P2) appear to be environmentally very similar to the Pseudotsuga-Linnaea forests (P3) according to the canonical analysis results (Table 17). However, these two community types are distinctly separated on the ordination based on vegetation data (Fig. 7a). The Pseudotsuga-Linnaea forests (P3) contain numerous species associated with dry sites, such as Gaultheria shallon and Linnaea borealis, whereas the Pseudotsuga-ThujaAcer forests (P2) have a higher abundance of species more characteristic of mesic habitats, such as Polystichum munitum and Tiarella trifoliata (Table 23). Increased soil moisture, through seepage, is probably responsible for the observed vegetational differences between the two community types, although fire history also may be important. The environmental variables recorded (Table 1) do not measure soil seepage and would contribute little to the differentiation of the two community types in the canonical analyses (Figs. 13 and 14). Also, only the occurrence of forest fire was recorded, and not its intensity or actual date of occurrence.

The dry Pinus-Pseudotsuga forests (D1) form the dry end of the soil moisture gradient on low elevation rock outcrops (Fig. 7b).

Environmentally, the two most dissimilar community types of the Pseudotsuga group are the Pseudotsuga-Thuja-Acer forests (P2) and the Tsuga-Pseudotsuga-Polystichum forests (P5) (Table 17). The environmental variables most strongly correlated with this difference are topographical position, maximum soil depth and LFH pH (Table 21). The higher soil moisture conditions associated with lower-slopes and level topography, as well as deep soils, are characteristic of the Tsuga-PseudotsugaPolystichum forests (P5). Polystichum munitum is a dominant understory component in these forests, and Tsuga heterophy11a is regenerating abundantly (Tables 7, 22 and 23). The Pseudotsuga-Thuja-Acer forests (P2) are found only in the warmest and driest sector of the study area, and are characterized by a relative abundance of Thuja plicata, Cornus nuttallii and Acer macrophyllum (Tables 7 and 22). The abundance of these tree species appears linked to soil seepage conditions. The higher pH of the organic layer of these forests probably can be explained by the calcium rich litter of Thuja plicata (Minore, 1979), as well as that from the deciduous species Acer macrophyllum and Cornus nuttallii ${ }^{(1)}$. The low LFH pH in the Tsuga-Pseudotsuga-Polystichum forests (P5) is probably linked to the abundance of Tsuga heterophylla and the acidifying effects of its litter.

From its central position in both the reciprocal averaging ordination and the direct ordination of the Pseudotsuga group, the PseudotsugaBerberis forests (P4) appear to represent the characteristic mesic,
(1) Cornus florida, closely related to $\underline{C}$. nuttallii, has high concentrations of calcium in its leaves (Thomas, 1969).
mid-slope community type of the inland part of the study area (Fig. 7a and b).

The hypotheses put forward in the Introduction concerning the close relationship between vegetation and environmental patterns in old-growth forests, and the possible role of major disturbances (hypotheses 2a and 2b), are well illustrated in the Pseudotsuga group. In general, vegetation patterns closely match the environmental patterns, except for community types P1, P2 and P3 where the relationship is weak (Figs. 13 and 14). These three community types are found in the warmest and driest areas where the Pseudotsuga group occurs, and where forest fire recurrence is probably highest. The strong dominance of Pseudotsuga menziesii in most stands, the abundance of charred bark on trees and charcoal in the soil, all present direct evidence in support of this assumption. Thus, different fire histories, rather than edaphic factors, may be the cause of vegetational differences between some of the community types within the Pseudotsuga group.
3. THE THUJA GROUP

While macro-climate is relatively uniform within the coastal sector where most of the plots from the Thuja vegetation group are found, the same cannot be said of soil parent material. Surficial deposits vary from poorly drained marine clays or sands, to imperfectly drained cemented tills and to well drained colluvium. Considering the variety of parent materials, a soil nutrient gradient very likely influences the vegetation pattern within the Thuja group. This is supported by the numerous correlations between the first reciprocal averaging ordination axis of the Thuja group and soil nutrient variables, such as percent nitrogen in LFH and $B_{1}$ horizons, and $C / N$ ratio in the LFH (Table 10). This soil nutrient gradient appears to be linked to a site productivity gradient which, in the direction of increasing productivity, is reflected in the vegetation by increasing maximum tree height, decreasing tree species richness and decreasing total coverage of the shrub and bryophyte strata (Table 10). Species richness is expected to decrease towards more productive environments, through increased competitive interaction between species (Del Mora1, 1983). The increased amount of light'resulting from the sparse overstory of poor sites, seems especially favourable to the development of a very dense shrub layer, where Gaultheria shallon and Vaccinium ovatum are particularly important. Two community types of the Thuja group, the coastal dry Thuja forests (T1) and the coastal wet Thuja forests (T5), although at opposite ends of the moisture gradient (Fig. 8b) are grouped together at the nutrient-poor end of the soil nutrient gradient reflected on the first axis of the vegetation ordination (Fig. 8a). Coastal Tsuga-Blechnum-Polystichum forests (T2) have the best soil nutrient and
drainage characteristics, and are probably the most productive forest communities within the Thuja group. Half of the plots of this community type occurred on ancient landslides, which may have improved soil drainage and nutrient conditions. Coastal Tsuga-Blechnum-Polystichum forests are also found at mid-elevations on mountain slopes (Fig. 8b), several kilometers away from the coast. Summer fogs rarely occur at these mid-elevations, and this may permit greater productivity through increased solar radiation. The effects of summer sea fogs on solar radiation and moisture have been described for the coast of California (Azevedo and Morgan, 1974).

An altitudinal gradient was also detected in the Thuja vegetation group. The low elevation, widespread coastal Thuja forests (T4) intergrade at higher elevations with the coastal montane Thuja forests (T3), characterized by an increased importance of Abies amabilis in the tree, sapling and seedling layers (Table 24). Along a soil moisture gradient, the driest sites are rock outcrops where only the coastal dry Pinus forests (D2) are found (Fig. 8b).

Although a direct ordination separated the various community types or the Thuja group on altitudinal and topography-moisture gradients (Fig. 8b), these same gradients are not distinctly reflected in the indirect ordination based on vegetation data alone (Fig. 8a). Moreover, elevation does not seem to exert the strongest influence on vegetation, as was hypothesized in the Introduction (hypothesis 1c). Instead a soil nutrient gradient appears to be most closely linked with the main vegetation variation (Table 10). Soil moisture appears to have little influence
on vegetation patterns, probably because precipitation exceeds 3000 mm annually and soil water deficits are non-existent (Fig. 2).

Elevation is clearly the second most important environmental gradient in the canonical analysis of the Thuja group (Fig. 13, Table 21), but an outlier plot (no. 85) tends to obscure the relationship with elevation in the indirect ordination (Fig. 8a, Table 10). Also strongly correlated with the second canonical axis is percent slope (Fig. 13, Table 21), suggesting a link between vegetation patterns and a gradient in soil moisture, from saturated soils at low elevations to better drained soils at higher elevations. This could occur through better drainage (increased slope) and absence of fog at higher elevations.

It should be noted also that previous studies demonstrating the strong correlation of elevation (hypothesis lc) and moisture (hypothesis 1d) gradients with the vegetation patterns of mountainous areas were carried out in drier climates, where summer soil moisture deficits occur frequently (Whittaker, 1956; 1960; Whittaker and Niering, 1965; Peet, 1981). Thus slope aspect and topographical position are expected to influence vegetation patterns wherever soil moisture is limited. In the coastal part of this study area, where soil moisture is probably abundant year round on all slope aspects and most topographical positions, the influence of soil moisture on vegetation patterns may be greatly reduced or nullified.

Large scale disturbance by fire is absent from areas near the coast, large scale windthrow is infrequent and natural landslides rarely occur. Thus, it would appear than in this relatively stable environment, old-growth
forests have become closely attuned to their environments and vegetation accurately reflects environmental gradients (hypothesis 2a), based on the results obtained from canonical analyses (Figs. 13 and 14, Tables 18 and 20).
4. THE ABIES GROUP

The Abies vegetation group is not associated with a specific geographical area; therefore, macro-climate (mostly precipitation) is less homogeneous than in the Pseudotsuga and Thuja groups. The Abies group is characterized by the dominance of Abies amabilis or Tsuga heterophylla, or both (Table 26). Environmentally, the majority of community types recognized within the group are found in sites with cool micro-climates. The lowland Abies forests (A5) are generally found at the bottom of steep valleys, often on north-facing slopes or on the upper terraces near rivers. In the drier parts of the study area, lowland Abies forests (A5) were, encountered most frequently in river valleys at the base of'steep north-facing slopes, while Tsuga-PseudotsugaPolystichum forests (P5) occupied the opposite south-facing slopes (plots 29 and 27, Figs. 1, 7b and 9b). Such a distribution may be explained by a cooler micro-climate on the northern aspect, where a lesser amount of solar radiation leads to lower evapotranspiration rates, higher soil moisture levels, and delayed snow melt in the spring. Cold air drainage also may be involved in the case of some narrow valleys.

In northern Washington, Abies amabilis forests predominate between 600 to 1300 m in elevation, where temperatures are cool, precipitation is high and snowpack is deep (Teskey et al., 1984). Low summer water deficits and low air temperatures have been reported as characteristic of areas where Abies amabilis is dominant (Waring et al., 1972). The montane Abies-Streptopus forests (A4) are all found above 600 m in elevation on north-facing slopes, inland within the study area (Fig. 9b).

This community type probably has the coldest environmental conditions of all those studied, except for the subalpine vegetation group. At lower elevations, but near the summit of small mountains near the coast, are found the montane Tsuga-Abies forests (A3). This community type usually occurs immediately above stands of the coastal montane Thuja forests (T3), on steeper slopes with faster drainage, and higher probabilities of wind disturbance. Plot 151 was sampled in a dense stand of Tsuga heterophylla and Abies amabilis originating from a complete windthrow of the previous forest (Fig. 9a and b). The montane Abies-Tsuga forests (A2) occur in similar topographical situations as the montane Tsuga-Abies forests (A3), but at higher average elevation inland, where snowpack is probably deeper and of longer duration. The higher relative densities of Abies amabilis seedlings and saplings in type A2, as compared to type A3, indirectly support this suggestion.

The Tsuga-Blechnum-Polystichum forests (A7) are one of the two community types within the Abies group to be almost entirely dominated by Tsuga heterophylla. This is strictly a low elevation community type, where relatively recent disturbances (100-200 years) may have played an important role in the strong Tsuga heterophylla dominance. However, the Tsuga-Gaultheria-Blechnum forests (A6), also dominated by Tsuga heterophy11a, appear to be climatically controlled and probably represent an intermediate community type along a precipitation gradient beginning with the coastal Thuja forests (T4) and ending in the driest inland sector around Port Alberni, with community types of the Pseudotsuga group. Finally, the montane Tsuga-Abies-Gaultheria forests (A1) occupy the driest habitats within the Abies vegetation group (Fig. 9b).

In terms of the major environmental gradients correlated with vegetation patterns in the Abies group, hypotheses formulated earlier appear to be confirmed ( 1 c and 1 d ), although the precondition that macroclimate be uniform is not held. It is possible that strong micro-climatic effects override macro-climate in the case of the Abies group. The Abies community types found in the interior of the study area, may avoid soil moisture deficits and high temperatures because they occupy habitats with characteristically cool and moist micro-c1imates (types A2, A4 and A5). A climatic gradient, from the cool climates where community types A4, A5 and A2 are found to the milder climates where the A7, A6 and A3 community types are found, is reflected in the first axis of the reciprocal averaging ordination (Fig. 9a). The third axis separates the only community type of dry habitats (A1) from the other types. The direct ordination of the plots also illustrates these gradients of mesoclimate (linked to elevation) and soil moisture (linked to slope aspect and topographical position). a finer topographical position scale would probably resolve the overlap between types A'7 and A5 (Fig. 9b).

The interpretation of the canonical analysis of the Abies group is limited because only four environmental variables were used. Nevertheless, low elevation community types (A5, A6, A7) are separated from higher elevation community types (A4, A2, A3), and those occurring on steep slopes (A2, A3) are separated from those occurring on gentle slopes or level terrain (A5, A6) (Fig. 13, Table 21). Increasing organic horizons thickness, positively correlated with the first canonical axis (Table 21), is characteristic of the montane Abies-Streptopus forests (A4), where cooler temperatures and long snowpack duration are expected
to impede organic matter decomposition. It is surprising to note how similar vegetationably the montane Abies-Streptopus forests (A4) are to the lowland Abies forests (A5), but how dissimilar they are environmentally (Figs. 9a and 14, Tables 19, 20, 26 and 27). Environmental similarity would undoubtedly increase if climatic variables were available to include in the analyses, allowing micro-climatic similarities between the two types to surface.

Classification of individual plots into broad vegetation groups, as well as into more narrowly defined community types was acheived using successive ordinations of the vegetation data (Peet, 1980). Overall, the community types and vegetation groups defined following this approach also differ environmentally. This is demonstrated by the canonical analysis of all community types and the subalpine vegetation group based solely on environmental data (Fig. 14). These results generally indicate that differences at the plant community level are paralleled by differences at the environmental level within the study area (hypothesis 2 a). Such a close vegetation - environment correspondance was expected at the outset of this study because of the sampling of "old-growth" forests.

The effects of fire disturbance are probably at least partially responsible for the weaker matching between vegetational and environmental differences encountered in some community types within the Pseudotsuga group: (types P1, P2 and P3). Differing fire disturbance histories are hypothesized to be the cause of the difference in vegetation composition between these particular community types. Soil seepage, unaccounted for in the set of measured environmental variables, also may explain why the Pseudotsuga-Thuja-Acer forests (P2), with a characteristically mesic vegetation, are grouped environmentally with the Pseudotsuga-Linnaea forests (P3) of drier habitats in Fig. 14. However, these results (Fig. 14) represent only the first two canonical axes. When Mahalanobis squared distances accounting for all dimensions are inspected, we find that the dry Pseudotsuga forests (P1) are much more environmentally
similar to the Pseudotsuga-Linnaea forests ( P 3 ), than this latter community type is to the Pseudotsuga-Thuja-Acer forests (P2) (Tab1e 20). Therefore, community types P1 and P3 are the most likely to have vegetational differences based on differing fire histories because they are the most environmentally similar . The Pseudotsuga-Thuja-Acer forests (P2) are thus more likely to show vegetational differences from types P1 and P3 on the basis of the presence of unrecorded soil seepage.

Evidence from discriminant analyses also suggests a weaker relationship between environmental characteristics and vegetation composition in the Pseudotsuga group. Using discriminant functions based on environmental data to re-classify plots into community types (originally defined by composition) it was found that a correct re-classification was obtained for 62.5 \% of the plots from the Pseudotsuga group. In the Thuja vegetation group the re-classification success was $69.4 \%$. Reclassification success for all community types analyzed together was 72.5 \% (Appendix 4). This shows a lower concordance between environmental characteristics and vegetation composition in the Pseudotsuga group, subjected to large scale fire disturbances, as compared to the Thuja group where disturbances are less prevalent, or to the entire study area. Thus, the hypothesis formulated earlier regarding the effect of disturbance seems to hold (Introduction, 2b).

High environmental similarities between community types of different vegetation groups is observable between the T2 type of the Thuja group and the A7 type of the Abies group, as well as between the A1 type of the Abies group and the P6 and P7 types of the Pseudotsuga group (Fig. 14,

Table 20). In both cases these community types are environmentally more similar to types of another vegetation group, than to community types within their own vegetation group. This resulted from the separation of plots for further analyses through successive ordinations; these particular community types being near the edges of separated plot clusters on the ordination diagrams. This situation can be seen as a reflection of the continuous nature of vegetation, and an excellent example of the difficulty encountered in attempts at partitionning this continuum for classification purposes. The vegetational and environmental differences between these community types are sufficient to maintain their separate status. However, the coastal Tsuga-Blechnum-Polystichum forests (T2) might fit just as well with the Abies vegetation group; similarly, the montane Tsuga-Abies-Gaultheria forests (Al) could be included with the Pseudotsuga group (Fig. 14, Table 20).

Environmental differences detected through the canonical analyses are sometimes slight between two community types which are differentiated vegetationally along only one major environmental gradient, such as elevation. This is particularly evident in the case of the PseudotsugaBerberis forests (P4) and the montane Tsuga forests (P6), adjacent community types along an elevation gradient of mesic sites on inland mountain slopes (Figs. 7a, 7b and 14, Table 20).

Environmental relationships between community types of the three larger vegetation groups were analyzed individually within each group (Fig. 13). The results are similar to those obtained in a global analysis of all community types from all vegetation groups (Fig. 14). The latter
analysis has the advantage of showing the relationships between community types of different vegetation groups, as well as possibly improving the characterization of within group relationships through increased total variation in the environmental variables employed.
6. THE CLIMATIC MASTER GRADIENT

The successive vegetation ordination approach has revealed a variety of environmental gradients correlated with vegetation variation in each set of plots analysed. A macro-climatic gradient from low elevation vegetation to subalpine forests was detected in the first ordination, as well as a parent material gradient from floodplains to rock outcrops (Fig. 5). A subsequent ordination detected what appeared to be a general gradient of increasing distance from the coast, linked with macro-climate (Fig. 6). In other ordination of smaller groups of plots, gradients of elevation (meso-climate), soil moisture (topography) and soil nutrient factors were correlated with vegetation patterns. The major macro-climatic gradient of precipitation occurring in the study area had not been clearly identified in the vegetation ordinations, except perhaps in the one using 140 plots (Fig. 6). Thus, an ordination of modal plots was done in order to assess the importance of this macroclimatic gradient on the general vegetation patterns within the entire study area. The modal plots were selected from all the sampled plots with the objective of producing a data set in which edaphic and mesoclimatic variations would be minimized. Such a technique has been called the "functional approach to plant community ecology" (Austin et al., 1984) and assumes that if certain factors known to influence vegetation are held constant, in this case through data manipulation, the relationship between vegetation and the factor allowed to vary can be analyzed. Plots located at both extremities of the edaphic gradients of soil moisture and soil nutrients were eliminated, as well as plots at the cooler end of meso-climatic or micro-climatic gradients. Modal plots represent low-
to mid-elevation sites of intermediate edaphic conditions throughout the study area. Because of the selection criteria, very few plots were included from the Abies vegetation group. The majority of modal plots come from the Pseudotsuga and Thuja vegetation groups, both occurring in geographically distinct areas.

The results of the reciprocal averaging ordination of the modal plots show that the vegetation pattern expressed on the first two axes is strongly correlated with the distance from the coast (Fig. 10, Table 14). It is assumed that the distance from the coast gradient is closely linked to a steep precipitation gradient. Evidence for this major climatic gradient is easily obtained (Figs. 2 and 12). Precipitation decreases from an average of over 3000 mm annually to less than 2000 mm within a 60 km distance from the coast (Fig. 2). The same trend can be seen during the growing season (fig. 12), when soil water deficits are most likely to occur inland (Fig. 2). The steepness of this precipitation gradient is a result of orographic precipitation and rain shadow effects caused by the interception of moisture laden air masses by high mountains parallel to the coast. Even though the annual mean temperature is only slightly higher inland than on the coast (Fig. 2), this difference translates into 200-300 extra effective growing degree-days annually for the inland areas (Fig. 12). Therefore, more heat is available inland for the growth of plants. However, soil moisture deficits may be encountered inland during July or August (Fig. 2).

The precipitation gradient directly influences a disturbance type gradient. Fire disturbance is undoubtedly predominant in its scale. and its effect on vegetation at the low precipitation end of the gradient
(Table 14). Towards the coast, with increasing precipitation and the absence of soil moisture deficits, forest fires rarely occur and the main disturbances are caused by wind (Klinka et al., 1979). Improving drainage away from the coast reflects a natural change in topography to steeper slopes and more abundant colluvial material, as well as a much shorter annual period of soil saturation (Table 14). Numerous other soil attributes vary along the distance from the coast-precipitation gradient (Table 14). The strongest correlation is with the organic horizons thickness/effective rooting depth ratio. This ratio decreases inland, where trees are more deeply rooted into the mineral soil, and increases towards a value of 1 near the coast, where the effective rooting depth often coincides with the thickness of the organic horizons (Table 14, Fig. 12). The thickness of the organic horizons is, in turn, probably related to climate. Plentiful moisture and mild temperatures lead to abundant and nearly continuous plant growth on the coast, and to the accumulation of thick organic soil horizons (Valentine, 1971). Decomposition may be slowed by a heat deficit, compared to inland areas. Plant productivity is probably reduced inland by summer soil moisture deficits and by colder winter temperatures, leading to a lesser accumulation of litter. Decomposition of this litter probably proceeds faster than on the coast because of higher summer temperatures. Organic horizons thickness is also partially related to the type of vegetation and the type of litter produced, influencing decomposition rates and by-products (acidity, nutrients). However, vegetation itself is also closely related to the precipitation gradient. Whether the climate or the vegetation exerts the strongest influence on organic horizons thickness may be difficult to assess.

Different tree species are known to have distinct rooting depth patterns (Minore, 1979; Eis, 1974; McMinn, 1963; Strong and La Roi, 1983). Pseudotsuga menziesii, dominant inland, is known to be a deep rooting species (Minore, 1979). Thuja plicata, Tsuga heterophylla and Abies amabilis, dominants near the coast, are known to be shallow rooting (Minore, 1979). Shallow effective rooting near the coast also may result partially from increased waterlogging of the soil as a product of high precipitation. Whatever the cause, it remains that the tree root mass is often restricted to the organic horizons in natural forest communities near the coast. As distance from the coast increases, the tree root mass occupies more and more of the mineral soil. This tendency is correlated wịth increasing dominance by Pseudotsuga menziesii and decreasing precipitation (Figs. 11 and 12). Total tree basal area tends to increase towards the coast and may indicate an increase in forest productivity linked with the precipitation gradient (Table 14). Maximum tree height decreases near the coast, possibly reflecting the change in dominant tree species (Waring and Frank1in, 1979) or increasing wind disturbance, or both (Table 14).

The vegetation pattern and changing environmental variables along the precipitation gradient are documented by the ordination of modal plots (Fig. 10) and by correlations between site variables and ordination axes (Table 14). Also of interest is the graph showing the distribution patterns of major tree species along this same gradient (Fig. 11). Pseudotsuga menziesii and Thuja plicata each attain their maximum and minimum basal areas, at opposite ends of the gradient (Fig. 11). Although not indicated in the graph, Thuja plicata is present in small amounts at
the dry end of the gradient, but the polynomial curve follows the best fit of the more abundant data near the wetter end of the gradient. The presence of a Pseudotsuga menziesii peak at the dry end of the gradient is not surprising in light of this species' well known adaptation to fire (Franklin and Dyrness, 1973; Minore, 1979). Other species, such as Tsuga heterophylla, Thuja plicata and Abies amabilis, possess thin bark and are usually killed by fire. These species are also more tolerant of cooler temperatures (or less heat available for growth) as can be inferred from their northern distributions along the coast, whereas Pseudotsuga menziesii reaches its northern coastal distribution limit on Vancouver Island (Krajina et al., 1982). The slight decrease in Pseudotsuga menziesii and increase in Tsuga heterophylla basal areas past the 50 km mark may be a response to increasing orographic precipitation caused by a second major ridge of mountains just west of Port Alberni (Fig. 11).

Towards the wet end of the gradient Thuja plicata reaches an average basal area of $120 \mathrm{~m}^{2} /$ hectare in modal vegetation $10-15 \mathrm{~km}$ from the coast, then declines to less than $80 \mathrm{~m}^{2} / \mathrm{ha}$ within 5 km of the coast. This decline could reflect a lowered forest productivity near the coast due to the high frequency of summer fogs. These fogs reduce the amount of solar radiation reaching the forest canopy, thus reducing photosynthesis and impeding evapotranspiration rates. Although low productivity sometimes results from low soil nutrient levels, this seems unlikely here since percent soil nitrogen increases towards the coast, in both the organic LFH and mineral B1 horizons (Table 14). Abies amabilis basal area increases linearly in modal vegetation towards the coast (Fig. 11), in agreement with the general autecological characteristics ascribed to this species (Minore, 1979; Krajina et al., 1982).

The relationship between basal area and distance from the coast is more complex in Tsuga heterophylla than in the other species examined (the polynomial equation for $T$. heterophylla has the lowest $r^{2}$ value, Fig. 11). Toward the dry end of the gradient, the observed drop in basal area may, reflect y combination of fire disturbance and suboptimal moisture conditions. Toward the coast, the decline and slight rise in Tsuga heterophy1la basal area coincide with an opposite trend in Thuja plicata. This is suggestive of a comptetitive interaction between the two species (possibly for space which is occupied for greater periods by the longer lived Thuja plicata (Waring and Franklin, 1979)) since abiotic conditions near the coast are unlikely to impair the growth of Tsuga heterophylla. In on1y a small segment of the gradient does Tsuga heterophy11a become the dominant tree in terms of basal area (29 to 31 km from the coast). This region appears to represent a transition zone between coastal and inland forest types where, perhaps, decreased competition from the other dominants permits better growth in Tsuga heterophylla.

The organic horizons thickness/effective rooting depth ratio also varies with distance from the coast (Figs. 11 and 12). The ratio is lowest toward the dry end of the gradient, in the region of Pseudotsuga menziesii's peak in basal area, and increases towards the wetter end of the gradient. This ratio reflects the different utilization of the soil horizons (organic vs. mineral) by the roots of the changing assemblage of tree species along this major climatic gradient (Pseudotsuga menziesii vs. Thuja plicata).

The results illustrated by Figure 11 have major implications for forest management. Because of its economic desirability, Pseudotsuga menziesii has been for many years the preferred species for replanting after
logging in this area. Yet the data presented here show that Pseudotsuga menziesii is not an important species within 25 km of the coast, and that it is totally absent within 15 km of the coast on modal sites (Fig. 11). Pseudotsuga menziesii is present near the coast, however, in a few nonmodal sites such as rock outcrops (e.g. the coastal dry Pinus forests, Table 24), although such sites are unproductive and are not usually logged. Other more productive sites, which may be considered modal, have been replanted with Pseudotsuga menziesii after logging. These sites, often very near to the coast, previously would have'supported mature stands of Thuja plicata, Tsuga heterophylla and Abies amabilis. The absence of Pseudotsuga menziesii from the natural modal vegetation could be attributed simply to the lack of forest fires, although other types of disturbances, such as blowdowns and landslides, occasionally occur which would create openings for this species. A few Pseudotsuga menziesii trees have indeed been found near the coast on steep slopes of old landslide colluvium in the Cypre River valley and near Kennedy Lake. Nevertheless, the question remains why Pseudotsuga menziesii is not a more widespread and important species in the immediate vicinity of the coast.

The answer to this question may have been found recently by Spiers et al. (1983) studying Pseudotsuga menziesii plantations near the coast. The trees in these plantations are starting to show serious growth defects, 20 to 30 years after planting. Abnormally high levels of arsenic have been detected in the leaders of planted Pseudotsuga menziesii, while levels in adjacent, naturally regenerating species were near background levels (Spiers et al., 1983). These authors have suggested that arsenic is found in an arsenate form (Sadiq et al., 1983), analogous to a form of phosphate
absorbed by plants, in the frequently waterlogged mineral soil. Anaerobic and reducing conditions encountered in saturated soils may hinder also the natural physiological processes of selective soil nutrient uptake by Pseudotsuga menziesii roots, leading to the uptake of potentially toxic arsenate along with the nutrient phosphate. High arsenic concentrations in the meristems of the tree could possibly cause growth defects by interfering with the synthesis of plant growth hormones (Spiers et al., 1983). The delay in the appearance of symptoms probably is related to the time required by the deep rooting Pseudotsuga menziesii to reach the satured lower horizons where arsenic will occur in the arsenate form.. The delay also could be related to a slow accumulation of arsenic up to a critical point when toxicity occurs. Soil saturation is probably highest in the spring when active growth is taking place (Spiers et al., 1983). Waterlogging is probably increased on the plantation sites following the removal of the original vegetation which removes large amounts of soil moisture through evapotranspiration. These excessive soil moisture conditions at greater depths are avoided by Thuja plicata, Tsuga heterophylla and Abies amabilis because of their shallow rooting habits. Avoidance of saturated soil horizons probably occurs also for Pseudotsuga menziesii on the rapidly drained sites, such as rock outcrops and old landslide colluvium, where it is found near the coast. Carter et al. (1984) have described growth abnormalities, identical to those reported by Spiers et al. (1983), in other coastal plantations of Pseudotsuga menziesii. They tentatively diagnosed a boron deficiency from tissue and soil analyses, although arsenic concentrations were not reported.

An increase in alpha diversity of vascular plants (species richness) with increasing continentality, or distance from the coast, was observed by Whittaker (1960) in the forest vegetation of south-coastal Oregon. Del Moral and Watson (1978) reported similar findings for the Washington Cascades. Oksanen (1983) describes an increase in alpha diversity of lichens and vascular plants with increasing continentality in Finland. These trends were observed also in western Vancouver Island, supporting hypothesis 1 e formulated in the Introduction. The alpha diversity gradient is particularly steep in the case of vascular plants, with fewer than 16 species occurring on average in the 0.05 ha plots near the coast, increasing to an average of more than 26 species in inland plots (Fig. 12). Bryophyte species richness appears to increase towards the wetter coastal areas, while vascular species richness increases towards the more continental and drier areas, particularly the herb layer (Table 19).

Numerous factors have been proposed to account for gradients in vascular plant diversity, but a synthesis of the causes underlying the patterns at different scales has not yet been acheived. At the microscale level, Del Moral's (1983) experimental results, based on the theoretical and conceptual frameworks of Grime (1980), Tilman (1980) and Huston (1979), demonstrate the effects of a combined interaction of site productivity, disturbance level and moisture stress on species diversity in subalpine meadows. On a larger regional scale, Whittaker (1975) suggested that site productivity and moisture levels were not the major controls of vascular plant alpha diversity, but that heat (possibly measured by growing degree-days) may represent the key factor. Recent experimental results indicate that the most productive sites have lower
alpha diversity than sites with intermediate productivity (De1 Mora1, 1983). The explanation offered in this case is that competitive interactions between species tend to reduce diversity in productive sites. Extreme physical stress also tends to reduce diversity, but it enhances it at intermediate levels by again preventing or reducing competitive interactions (Del Moral, 1983). Disturbance plays a major role in increasing diversity, and the more productive the site the more frequent the disturbances must be in order to prevent the competitive exclusion of several species by one or a few dominants (Del Moral, 1983). Thus, it may be that the more frequent large scale forest fire disturbances of the interior part of the study area enhance vascular plant alpha diversity. On the other hand, the relatively stable environment of the coastal sector may allow strong competitive interactions to take place and reduce diversity. Dominance concentration (Whittaker, 1975; Peet, 1974) is also much higher in the shrub and herb strata of the community types of the Thuja group than of the Pseudotsuga group (Tables 23 and 25). High dominance concentration (= low equitability) implies that a stratum, or community, is strongly dominated by one or a few species (Whittaker, 1975).

Beta diversity also has been observed to increase from coastal to more continental areas (Whittaker, 1960; Del Moral and Watson, 1978). As used here, beta diversity ${ }^{1}$, refers to the number of half-changes in

[^1]compositional similarity that occur along distinct environmental gradients (Whittaker and Woodwe11, 1978). This measure was useful for comparing vegetation-environment relationships in two geographically distinct groups of plots along the major rainfall gradient : (1) the Pseudotsuga vegetation group representing the drier, more continental interior sector, and (2) the Thuja group representing the very humid, coastal sector. Beta diversity along a soil moisture gradient (at low elevation) was 4.8 in the Pseudotsuga group (endpoints : plots 110 and 17 , Fig. 7b); in the Thuja group (endpoints : plots 53 and 50, Fig. 8 b ) beta diversity was 0.7 . Along an elevation gradient, values of 4.7 in the Pseudotsuga group (endpoints : plots 13 and 139, Fig. 7b) and 0.3 in the Thuja group (endpoints : 50 and 152, Fig. 8b) were obtained. Difference in total lengths of the elevation gradients concerned ( 800 m in the Pseudotsuga group and 600 m in the Thuja group) are considered insufficient to explain the large discrepancy in the beta diversity values calculated. The decline in temperatures with increasing elevation probably leads to greater moisture availability through reduced evapotranspiration, thus superimposing a soil moisture gradient on elevation. This could explain the high beta diversity of the Pseudotsuga group along the elevation gradient, since soil moisture conditions would be expected to vary more widely with elevation here than in the Thuja group. Evapotranspiration would not be expected to differ significantly between high and low elevations in the Thuja group, because of the frequent occurrence of summer fogs at low elevation.

The higher beta diversity of the Pseudotsuga group along the soil moisture gradient at low elevation probably reflects the greater length of this gradient in the drier interior sector. Differences between dry
and wet habitats are greater in absolute terms in dry areas (vegetation growing on rock outcrops experiences longer periods of drought in dry areas than in areas of high precipitation). On the other hand, wet habitats would be similar in terms of absolute soil moisture availability, in wet or dry areas. Slope aspect effects on soil moisture and on the amount of heat available for growth are also likely to be stronger in the drier, interior sector.

Thus, the trends in beta diversity identified here are in general agreement with results from other studies on coastal forests (Whittaker, 1960; Del Moral and Watson, 1978). The same trend app1ies along both the environmental gradients of soil moisture and elevation.
7. HOMOGENEITY AND SPECIES RICHNESS OF STRATA

The investigation of homogeneity in separate strata of fourteen community types supports the third hypothesis formulated in the Introduction (Table 31). The community types of the Pseudotsuga vegetation group have, on average, a vegetation homogeneity of 0.72 , while community types of the Thuja vegetation group have, on average, a vegetation homogeneity of 0.84. The high mean fire indices within the Pseudotsuga group indicate that fire is a common form of disturbance (Table 31); the opposite is true for the community types of the Thuja group (Table 31). Thus, it appears that large scale disturbances such as fire may tend to reduce vegetation homogeneity. If the vegetation of the two groups is compared on a stratum by stratum basis, few differences are seen for the tree, seedling and bryophyte-1ichen layers (Table 31). The largest difference occurs at the level of the herb stratum, with an average homogeneity of 0.96 in the Thuja group and 0.47 in the Pseudotsuga group. The shrub and sapling layers are also markedly more homogeneous in the Thuja vegetation group. Interestingly, a significant positive correlation exists, over all types, between increasing fire index values and decreasing homogeneity of the herb and shrub strata ( $\mathrm{r}=-.589$ and -.425 ). The reverse occurs in the tree stratum, where homogeneity increases with increasing fire index values ( $\mathrm{r}=.358$ ). This result reflects the abundance of homogeneous post-fire stands strongly dominated by Pseudotsuga menziesii, the only major tree species of the study area well adapted to forest fire disturbances.

The seedling stratum is generally the most homogeneous within the entire study area (Table 31). The explanation for this may be that most communities have closed canopies permitting only shade tolerant tree seedlings to germinate and become established. In support of this hypothesis, it can be seen that rock outcrop communities, with open canopies and mixtures of shade tolerant and intolerant seedlings, have lower seedling layer homogeneities. Floodplain forests (F1), often relatively open and with ideal germination and early growth conditions (ample moisture and soil nutrients), have the lowest seedling stratum homogeneity of the community types studied (Table 31). The least homogeneous vegetation layer is by far the bryophyte-lichen layer. This stratum is strongly influenced by the micro-heterogeneity of the forest floor. An exception is found in the very homogeneous bryophyte-lichen layer of the PseudotsugaLinnaea forests (P3). This community type is characterized by a nearly complete cover of Hylocomium splendens giving it the highest dominance concentration (= lowest ${ }^{l} / \lambda$ value or highest $\lambda$ value) for this layer in all the community types described (Tables 23,25 and 27). In fact, as a general rule it appears that any vegetation stratum with a high homogeneity is 1ikely to have a strong dominance concentration, which means that one species has a much higher coverage, or relative density, than other species in the layer. The extremely homogeneous shrub and herb layers of the coastal Thuja forests (T4) are a good example (Tables 25 and 31), as are the sapling and seedling layers of the Tsuga-Blechnum-Polystichum forests (A7) (Tables 26 and 31).

The two most homogeneous community types are the coastal wet Thuja forests (T5) and the coastal Thuja forests (T4) (Table 31). The coastal
wet Thuja forests (T5) have extremely homogeneous vegetation strata, all above 0.9 except for the bryophyte layer. This community type is found only within a narrow range of environmental conditions, disturbances are rare, and it is not very widespread (Fig. 8b). The coastal Thuja forests (T4), on the other hand, occupy a wider range of environments near the coast (Fig. 8b). This, and the fact that the 19 plots used to calculate the homogeneity values came from a relatively large area, amplifies the extraordinary homogeneity of these forests. The herb stratum is remarkable with its homogeneity of 0.98 . This undoubtedly results from the nearly exclusive dominance of Blechnum spicant, growing profusely on the thick, forest floor organic horizons characteristic of this community type (Table 25). The reasons for such high homogeneity may lie in the relatively uniform climate, with no extreme temperatures, abundant moisture and a very low frequency of major disturbances. Fire is virtually absent, and major windthrow is unusual at the low elevations where these forests occur. Occasionaly, individual trees are blown down, but such occurrences do not appear to initiate significant changes in understory conditions because of the naturally open nature of the canopy of very large and widely spaced Thuja plicata trees. This combination of minimal disturbance and optimal plant growth conditions seems, in large part, responsible for the low species richness and diversity, and the concentration of dominance in a small number of species in each stratum. In the coastal Thuja forests (T4) the increasing homogeneity through seedling, sapling, and tree layers, suggests that a process of elimination (through competition?) takes place during the development of the forest canopy (Table 31). This may be related to the longevity of Thuja plicata, whose life span is twice that of
co-dominant tree species (Waring and Franklin, 1979). This inference of a monopolization of space by a long-lived species corresponds to the inhibition model of Connell and Slatyer (1977).

The least homogeneous community type are the floodplain forests (F1) (Table 31). Colonization of floodplains after disturbance is more likely linked to stochastic events and available seed sources than to the ecological tolerances of species. The differing ages of formation of the floodplains sampled, as. well as their differing flooding regimes, add to the observed heterogeneity. Compared with other community types, the surficial materials of floodplains are much younger and are subject to a higher disturbance frequency in the form of floodings (mechanical damage or new sediment deposition). The high species richness of the herb stratum of the floodplain forests (F1) appears largely linked to this more or less regular and frequent disturbance regime as well, because a low species richness would be predicted for such productive sites (Table 3l). De1 Moral (1983) has shown that with increasing site productivity, disturbance regime had to increase as well to maintain species diversity by reducing the occurrence of competitive interactions. Floodplain forests (F1) have the highest species richness of community types of mesic habitats within the study area (Table 30).

No distinct relationship was found between species richness and homogeneity of vegetation strata when all community types and all strata were considered, possibly because the data were too heterogeneous for any clear trends to emerge.

## B. COMMUNITY DYNAMICS

Since the publication of a seminal paper by Henry and Swan (1974) there has been a growing recognition of the importance of natural disturbances in the composition, structure and dynamics of nearly all natural vegetation (White, 1979). Even though the coastal forests of Vancouver Island are uniquely imposing in stature and age, they share a common feature with many other vegetation types (e.g. Grimm, 1984) in that disturbance has played a major role in their development. In fact, continuous small or large scale disturbances may be essential to their maintenance.

## 1. PSEUDOTSUGA COMMUNITY TYPES

Fire is undoubtedly the most noticeable form of disturbance within the study area. It has permitted Pseudotsuga menziesii to remain the dominant tree species in most community types of the drier interior sector of the study area. Because of its thick bark, Pseudotsuga menziesii is considered the most fire resistant of all coastal tree species (Minore, 1979). However, its seedlings are incapable of establishing themselves under the shade of its canopy, except perhaps in the driest parts of coastal British Columbia (Krajina, 1969; Krajina et al., 1982). Thus, the bell-shaped distribution curves of Pseudotsuga menziesii stems in most community types are characteristic of a seral species (Fig. 15) which depends entirely on the occurrence of a major disturbance, fire in this case, for its establishment. In the dry Pinus-Pseudotsuga forests (D1) of rock outcrops, the dry Pseudotsuga forests (P1), and the Pseudo-tsuga-Linnaea forests (P3), the seral role of Pseudotsuga menziesii is not as obvious because some regeneration appears to occur (Fig. 15). However, only on the rock outcrop communities (type D1) of the interior of the study area does Pseudotsuga's size-class distribution curve appear characteristic of a primary species (Fig. 15).

It has been suggested that Pseudotsuga menziesii cannot regenerate in the moist coastal forests because its seedlings or saplings cannot transpire sufficiently in the shade to rid themselves of excess moisture (Krajina, 1969; Krajina et al., 1982). Thus, Krajina (1969) argues that P. menziesii becomes "shade tolerant", or has the ability to regenerate under a canopy, only on dry sites near the Coast. Conversely, Tsuga
heterophylla requires shade to germinate on dry sites, and even then the saplings or young trees will likely die following a drought (Krajina, 1969). Viewed differently, it may be that Pseudotsuga menziesii is the most shade tolerant of the tree species that can grow in climatically dry areas or edaphically dry sites on Vancouver Island. Tsuga heterophy11a is more shade tolerant, in an absolute sense, than Pseudotsuga menziesii, but it is probably incapable of surviving on the driest sites, as observed by Krajina (1969). On such dry sites (or areas), the relatively shade intolerant Pseudotsuga menziesii will then become the dominant tree species, because it is the most shade tolerant of all the other species capable of growing there (i.e. Arbutus menziesii, Pinus contorta). This concept has been proposed by Daubenmire in his habitat type approach to vegetation (Daubenmire and Daubenmire, 1968). In all community types of the Pseudotsuga group, except those of the driest environments (types D1 and P1), Tsuga heterophy11a and Thuja plicata are clearly primary, or climax, species (Fig. 15). Both are easily killed by fires (Minore, 1979).

Tsuga heterophylla seedlings are commonly found growing on undecomposed wood substrata on the forest floor of Pseudotsuga community types (Table 32). In the community type where the relationship was not significant, the mean seedling density was still the highest on the undecomposed wood substrata (Table 32). Christy and Mack (1984) have shown that Tsuga heterophy1la "juveniles" are almost exclusively restricted to decaying logs, predominantly those in intermediate stages of decomposition. These partially decomposed "nurse logs" are regarded as presenting an optimal compromise of litter-shedding characteristics (best in youngest
logs) and substratum conditions (best in oldest logs) that permits the successful germination and establishment of tree seedlings (Christy and Mack, 1984). It is assumed that litter accumulation represents an important impediment to the establishment of Tsuga heterophylla seedlings, and that the nurse logs provide the necessary elevated "safe sites" (sensu Harper et al., 1965) within the community. Nurse logs with hundreds of Tsuga heterophylla seedlings and saplings were frequently observed within the Pseudotsuga group. In Tsuga-Pseudotsuga-Polystichum forests (P5), seedlings, saplings and small Tsuga heterophylla trees were often seen growing on or very near the bases of the large Pseudotsuga menziesii dominants. This location may be the only place free from litter in forests where logs are rare. Pseudotsuga menziesii seedlings are not statistically associated with a particular substratum (Table 32), but this could be an artifact of the very low seedling densities encountered under the dense canopy of the community types analyzed.

Tsuga heterophylla occurs in increasingly larger size-classes along a moisture gradient within the Pseudotsuga group (community types P3 < P4< P5 = P6, Fig. 15). In the cool and moist conditions of the montane Tsuga forests (P6), Tsuga heterophylla probably grows as fast as Pseudotsuga menziesii following a disturbance, and may thus be a good species for reforestation, either alone or in mixture with Pseudotsuga menziesii. This may be particularly advantageous if a relatively short planting to harvest rotation is planned ( 80 to 100 years); otherwise, the superior, long-term size potential of Pseudotsuga menziesii negates the use of Tsuga heterophylla. On the poor and shallow soils of the montane TsugaGaultheria forests (P7), Tsuga heterophylla may also be the most appropriate
choice for reforestation. In all other community types at lower elevations (P1, P2, P3, P4 and P5), Pseudotsuga menziesii wou1d appear to be the most ideally suited species for reforestation. The frequency of droughts causing mortality would preclude any use of Tsuga heterophylla as a viable reforestation species in most of these community types, particularly those with the driest moisture regimes (P1 and P3). The Pseudo-tsuga-Thuja-Acer forests (P2) are the only community type within the Pseudotsuga group which has more Thuja plicata stems per hectare than Tsuga heterophylla stems (Fig. 15). The hypothesized presence of soil seepage would also explain this discrepancy. Tsuga heterophylla regeneration is reportedly poor in these nutrient rich sites, occurring only on decaying wood; conversely, Thuja plicata does well in the same sites (Krajina, 1969).

The double peaks in the size-class distribution curve of Pseudotsuga menziesii within the Pseudotsuga-Thuja-Acer forests (P2) are interpreted as resulting from two periods of establishment, following two distinct forest fires (Fig. 15). The close geographical proximity of all the plots permits such an explanation, since their fire disturbance history may be assumed to be identical. Fires usually occur on a relatively large scale in coastal forests, but with a low frequency of once every few hundred years. Indirect evidence of this is found in the dominance of vast areas by Pseudotsuga menziesii, a species which can establish quickly in openings cleared by fire, as well as avoid damage from surface fires because of the thick, fire resistant bark of mature trees (Minore, 1979). Historically, large conflagrations were likely in the heavy fuel loads of coastal forests following periods of unusually
hot and dry weather. A recent forest fire, fought without much success with modern equipment and techniques, burned almost the entire southfacing slopes of the Sproat Lake valley. This example provides an indication of the minimum areal extent of forest fires which have occurred in the drier coastal areas of British Columbia.

An alternative to the two fires hypotheses is that all Pseudotsuga menziesii trees belong to the same cohort in which a size hierarchy has developed through intraspecific competition (Harper, 1977). When such a hierarchy of sizes is created through competition however, the smaller individuals are always more numerous. This is not supported by the data in Fig. 15 and the competition hypothesis is therefore rejected. More likely, the second fire following which the smaller (i.e. younger) Pseudotsuga trees got established, was of a lighter intensity than the first burn. Many already established Pseudotsuga trees would have survived the second fire, while almost all trees of other species would have been killed. All the larger Pseudotsuga menziesii trees found in plots of the Pseudotsuga-Thuja-Acer forests (P2) have fire charred bark. A sufficiently large sample of increment cores from the Pseudotsuga menziesii population of the area would permit the validation of the two fires hypothesis.
2. THUJA COMMUNITY TYPES

Wind disturbance, causing major blowdowns over large areas or the isolated falling of single trees, is regarded as the second most prevalent type of forest disturbance within the study area. Wind related effects increase towards the coast where the forests are more directly exposed to storms. Large scale blowdown is probably the major factor initiating secondary forest succession on the coast, as forest fires are believed to be extremely rare. The prevalence and extent of wind disturbance also appears to be linked with elevation. Extensive blowdowns are most frequently observed on the ridges or summits of mountains nearest to the coast (e.g. plot 151, Figs. 1 and 9 ), where the montane Tsuga-Abies forests (A3) are usually found. In valleys or on plains closer to sea level, where the coastal Thuja forests (T4) predominate, isolated tree falls triggered by wind tend to be the most frequent form of disturbance. Landslides also influence the development of some coastal community types, but their frequency and areal extent are small. The bare areas with improved drainage created by landslides would be rapidly colonized by light demanding or fast growing species such as Pseudotsuga menziesii and Picea sitchensis. Apart from rock outcrops, old landslides appear to be the only suitable environment for Pseudotsuga menziesii near the coast. Plot 87 on the upper-slope of the Cypre River valley and plot 155 on upper-slopes near Kennedy Lake are examples of such sites. Picea sitchensis also occurs on old landslides near the coast, but only at the base of slopes where moisture and nutrient levels are probably higher (plots 91 and 69).

Daubenmire and Daubenmire (1968; p. 55) have summarized an observation common to all who have studied forests dominated by Thuja plicata : "Thuja plicata is distinctive from the other trees in that younger ageclasses often seem inadequate to guarantee replacement of larger individuals...". This can be seen in the coastal montane Thuja forests (T3) and the coastal Thuja forests (T4), where the size-class distribution curve of Thuja plicata dips far below the curves of Tsuga heterophylla and Abies amabilis in the smaller size-classes, but extends much further into the larger size-classes (Fig. 16). Daubenmire and Daubenmire (1968) have suggested that the longevity of individuals and the layering habit of Thuja plicata were probably a key to understanding its persistence. Each individual needs only to leave one successfull offspring to maintain the population density, "thus, the longer it lives, the more sparse the reproduction can be and yet suffice" (Daubenmire and Daubenmire, 1968; p. 55). Indeed, Thuja plicata has a potential longevity of over 1,000 years, at least double that of the co-dominants Tsuga heterophy11a and Abies amabilis (Waring and Franklin, 1979). If layering does occur, the longevity of the "genetic" individual may be much longer. The coastal Thuja forests (T4) may thus represent an ideal example of Connell and Slatyer's (1977) inhibition model, where the largest and longest-lived species eventually achieves dominance as succession proceeds. Young coastal forests developing after extensive wind damage, however, are strongly dominated by Tsuga heterophylla and Abies amabilis (e.g. plot 151). Also, the coastal Tsuga-Blechnum-Polystichum forests (T2), which are suspected of having had a large disturbance at their origin, have very little Thuja plicata (Table 24). In most plots of the coastal Thuja
forests (T4), where Thuja plicata is the first dominant, no important disturbance could be detected, and it is possible that such stands have had no major disturbance in several hundreds, if not thousands of years.

Thuja plicata's dominance in the low elevation coastal areas would be threatened if its trees, otherwise long-lived, were frequently felled or fatally damaged by wind, béfore the time necessary for their successful regeneration. Thus, two hypotheses can be put forward to explain the presence of apparently stable Thuja plicata communities near the coast : the first states that strong winds very rarely occur in areas where Thuja plicata is the present dominant; the second states that Thuja plicata is less susceptible than other tree species to wind damage. The first hypotehsis appears untenable, because strong winds occasionally do occur even at low elevations near the coast. On the other hand, several lines of circumstantial evidence appear to support the second hypothesis. Thuja plicata is indeed considered the most wind resistant coastal tree, after Pseudotsuga menziesii, possibly because of its very dense and extensive root system (Minore, 1979; Klinka and Feller, 1984). Some mechanical resistance to wind-toppling may also be gained by the fluted and buttressed bases of large Thuja plicata trees (Putz et al., 1983). Also, mechanical resistance to bole snapping by wind may be increased by the fact that the trunks of old Thuja plicata trees are almost always hollow, perhaps conferring the enhanced stress resisting characteristics of hollow cylinders. Some evidence for a superior resistance to wind by large Thuja plicata trees was found in several plots. For example in plot 72, large individuals of Thuja plicata were observed still standing while almost all Tsuga heterophylla and Abies amabilis trees had been blown down in
approximately the same direction, apparently during the same storm. Large Thuja plicata trees may be able to lose their leader and uppermost branches during storms without fatal consequences, partly because of a high resistance to rot-causing fungi and to insect attack (Minore, 1979). The so called "candelabra" appearance of large Thuja trees near the coast, seems to be caused by the death of the leader and upper-crown branches, and by the shared apical dominance of several large lateral branches. This particular candelabra shape has not been noticed in Thuja plicata trees elsewhere than in the wettest areas nearest to the coast. This shape may develop with increasing age of the individuals; however, ancient Thuja trees can be found, in moist pockets even in the driest and most fire prone areas, that do not possess the characteristic candelabra shape. The possibility of a distinct genetic race, restricted to a narrow coastal band, seems to be discounted by the high genetic uniformity of Thuja plicata (Copes, 1981). The most likely explanation is that a particular set of environmental factors (abundant moisture, mild temperatures, occasional strong winds, lack of fire) combine with the genetic characteristics of the species (resistance to rot and insects, weak apical dominance, longevity) to produce the observed candelabra shape in old Thuja plicata trees near the coast.

Germination and establishment sites for seedlings appear to be almost entirely restricted to undecomposed wood substrata, mostly large logs, in the Thuja group (T2, T3 and T4, Table 32). These logs represent "safe sites" with particular combinations of ecological factors (abiotic and biotic) which permit the successful germination of seeds and establishment of seedlings (Harper et al., 1965). It has been frequently observed
that Tsuga heterophylla and Thuja plicata regenerate on fallen trees, or "nurse logs", in coastal forests (Franklin and Dyrness, 1973). Both of these species have small seeds, producing small and fragile seedlings that are likely to be susceptible to mechanical damage by burial. Abies amabilis, on the other hand, has larger seeds and produces large, robust seedlings (Schopmeyer, 1974), which are not expected to be as strongly affected by litter accumulation. This could explain why the distribution of Abies amabilis seedlings is unrelated to the occurrence of undecomposed wood in most of the community types analyzed (Table 32). The importance of undecomposed wood for the germination and establishment of Thuja plicata seedlings can be seen in the eighteen-fold increase in the number of seedlings found on undecomposed wood compared to forest floor sites in Thuja forests (T3 and T4, Table 32). Thuja plicata logs may provide the "safest" sites for seedling establishment and development to maturity. The decay rate of Thuja plicata logs appears to be extremely slow, compared to that of Abies amabilis and Tsuga heterophylla logs (Foster and Lang, 1982; Graham, 1981). Abies amabilis appeared to have the fastest decay rate in the field; it is probably comparable to the decay rate of Abies balsamea (Foster and Lang, 1982). The discovery, in plot 73, of a large Thuja plicata tree, approximately 400 years old by a growth-ring count, growing on top of a Thuja $\log$ of similar size illustrates the slow decay rate of Thuja plicata. Indeed, this log was still sound and not in contact with the soil along some of its length. Where systematic observations were made (some plots of the coastal Thuja forests), it was found that almost all seedlings, saplings and young trees of Thuja plicata and Tsuga heterophylla were rooted on decaying fallen trees, stumps or even
on the bases of living trees. This was not as obvious for larger and older trees, whose nurse logs may have had eventually rotted away.

The fourth hypothesis stated in the Introduction, that Thuja plicata is able to maintain itself in all of the coastal forests it presently dominates, appears very plausible. The extent to which vegetative regeneration of Thuja plicata contributes towards its total regeneration remains to be assessed. Schmidt (1955) reported that vegetative regeneration might be as important as regeneration from seed in high density stands. Vegetative regeneration may occur through the layering of low branches pinned under litter, from the rooting of broken live branches, or fallen live boles (Schmidt, 1955). Such occurrences were not observed in the coastal montane Thuja forests (T3), or in the coastal Thuja forests (T4).

Reforestation within the community types of the Thuja group can be effectively carried out using several species. For short rotations on productive sites (T2 > T3 > T4), both Tsuga heterophylla and Abies amabilis could be recommended, with perhaps some Picea sitchensis only on the best sites. Pseudotsuga menziesii cannot be recommended as a viable reforestation species in the area occupied by the Thuja vegetation group (Spiers et al., 1983; Carter et al., 1984). The sites near the coast where this species appears to grow successfully are very limited. On the poorest sites (T1 and T5), the best growth might be acheived by Thuja plicata, or by the faster growing Pinus contorta. Thuja plicata represents the climax species in dry (D2, T1) as well as wet (T5) nutrientpoor sites (Fig. 16). Definitely seral in the coastal wet Thuja forests
(T5), Pinus contorta may be capable of self-regeneration in the very open, coastal dry Pinus forests (D2) (Fig. 16). The presence of understory fern species could serve as useful indicators of site productivity near the coast. Polystichum munitum was most abundant in productive sites where access to mineral soil was not restricted by thick organic horizons. Conversely, Blechnum spicant dominates on thick organic horizons found on the less productive sites.

## 3. ABIES COMMUNITY TYPES

Because of its high shade tolerance, large seed size, and capability to withstand long periods of suppression under a forest canopy, Abies amabilis is considered a climax or primary species. It has this role in all of the community types within the Abies group (except for type A7) and in many within the Thuja group (Figs. 16 and 17).

The importance of disturbance in producing the tree size-class structures of community types within the Abies group can be inferred from the irregular shapes of the distribution curves shown in Fig. 17. For example, community type $A 3$ is found on mountain ridges near the coast, where wind disturbance appears to maintain a two-tiered arborescent structure. This is characterized by a lower layer of suppressed trees and saplings and a high1y discontinous upper layer consisting of trees released from competition after the last wind disturbance. This particular forest structure was also observed by Klinka et al. (1979), and its origin may be similar to the wind driven wave-regeneration phenomenon described for high elevation Abies balsamea forests of the northeastern United States (Sprugel and Bormann, 1981). A similar interpretation could be made for the montane Abies-Tsuga forests (A2).

The coastal Tsuga-Blechnum-Polystichum forests (T2) have a sizeclass structure which is also heavily skewed towards the larger sizeclasses (Fig. 17). Landslide disturbance is suspected to have been at the origin of at least half of the stands of this community type, and evidence of wind disturbance was found in the remaining stands. It could be argued that the coastal Tsuga-Blechnum-Polystichum forests (T2) are
actually long-duration seral communities which occur near the coast after major disturbances have improved seedling establishment conditions, soil drainage or soil nutrient availability. The slow accumulation of organic matter on the forest floor of these communities may eventually direct their development towards community types dominated by Thuja plicata (T3 or T4). This hypothetical successional sequence may never be entirely completed on old landslides because of the profound modification of the site drainage.

The Tsuga-Blechnum-Polystichum forests (A7) show evidence of past disturbance in the form of landslides in the two coastal plots, and fire in the inland plots. The shape of the size-class distribution curve of Tsuga heterophylla, characteristic of a climax species (Fig. 17), and the small importance of seral species (Table 26), may suggest that the original disturbances are very old. Alternatively, it is possible that Tsuga heterophylla, because of particular environmental conditions, established itself with more success than the usual seral species following the disturbance.

The montane Abies-Streptopus forests (A4) and the lowland Abies forests (A5) not only have floristic similarities, but are also similar in tree size-class structure (Fig. 17). Abies amabilis has a size-class distribution curve characteristic of a primary species in both community types, whereas Tsuga heterophylla's size-class distribution curve has a peak in the larger size- classes (Fig. 17). The occurrence of past disturbances was not frequently recorded in these community types, except for the occasional tree blown down by wind. In fact, partly decayed,
standing dead tree boles were often observed. The role of Tsuga heterophylla, in these two community types, may be that of an opportunistic, gap-regenerator which invades openings following the removal of a large canopy tree.

In the montane Tsuga-Abies-Gaultheria forests (Al), fire has been the major type of disturbance, as confirmed by the presence of Pseudotsuga menziesii with its characteristically seral size-class distribution curve (Fig. 17).

For reforestation purposes within the Abies group, Abies amabilis and Tsuga heterophylla may be equally appropriate in most of the montane community types (A2, A3 and A4), and in the lowland Abies forests (A5). Pseudotsuga menziesii could represent a viable species for the montane Tsuga-Abies-Gaultheria forests (A1) and some inland stands of the Tsuga-Blechnum-Polystichum forests (A7). Coastal stands of the same community type would be excellent sites for Picea sitchensis.

The dynamics of the floodplain communities sampled in this study appear to correspond generally to previous accounts by Cordes (1972) for Vancouver Island and by Fonda (1974) for the Olympic Peninsula. Very young floodplains dominated by Alnus rubra (Fonda, 1974) were not sampled, but they were frequently observed along all major rivers. The youngest stand sampled is probably plot 171, near the Klanawa River. This plot consisted of a dense grove of Picea sitchensis with a shrubless understory nearly completely covered by Polystichum munitum. Further away from the edge of the Klanawa River, plot 170 occupies an older, less frequently inundated floodplain. Here, Tsuga heterophylla was more abundant than Picea sitchensis in the tree stratum. In increasingly older floodplains, only a few large Picea sitchensis individuals remain. Tree species regeneration occurs almost exclusively on Picea logs, and is dominated by Tsuga heterophylla (Tables 24 and 32). Again, the fallen logs provide safe sites against burial by litter and mechanical damage during floods. The canopy of the older stands is always sparse, which probably explains the presence of an extremely dense and tall shrub layer, dominated by Rubus spectabilis and Ribes bracteosum. In the Olympic Peninsula such an extensive shrub layer never develops (Fonda, 1974), possibly because of a strong browsing pressure by elk.

The large sizes of trees of different species growing on floodplains indicates the high growth potential of these habitats (Table 24). Some of the largest Pseudotsuga menziesii trees encountered during the study ( $181 \mathrm{~cm} \cdot \mathrm{DBH}$ ) were found on an old floodplain situated near Nahmint

Lake (plot 122). Thus, since environmental factors on floodplains may be considered to be non-limiting (except perhaps tolerance to flooding), the original tree species composition is likely to depend mainly on stochastic events, such as seed dispersal or the availability of local seed sources, when a major disturbance releases a floodplain for colonization.

Fast growing trees which can exploit fully the ideal growth conditions of floodplains should be selected for reforestation. Picea sitchensis probably remains the best suited species on all sites, but Pseudotsuga menziesii may be an alternative choice on older floodplain terraces in the interior of the study area.

CHAPTER 6.

## CONCLUSIONS

The analysis of vegetation-environment relationships in old-growth forests of a large sector of the west coast of Vancouver Island was the prime objective of this study. Macro-climate appears to have the strongest influence on vegetation over the whole study area; variation in soil parent material is ranked second in importance. Within areas of relatively uniform macro-climate and soil parent material, stronger relationships with other environmental factors were found.

In the Pseudotsuga group, characteristic of the drier inland section of the study area, vegetation is correlated with meso-climate (elevation) and soil moisture gradients. Large scale fire disturbances have played a major role in the determination of the vegetation composition and structure in this group. A1so, homogeneity of the vegetation, between environmentally similar sites, was generally the highest in areas where fire disturbance was absent or infrequent, as in the Thuja group. However, different trends were observed for individual vegetation strata. The herb and shrub strata increased in homogeneity with decreasing fire disturbance, but the opposite trend was observed in the tree layer. This trend is linked to the presence of very homogeneous, almost monospecific, post-fire stands dominated by Pseudotsuga menziesii.

The Thuja group, found exclusively near the coast, displays variation mainly along gradients of soil nutrients and meso-climate
(elevation). The importance of soil nutrients is probably related to the large variability in parent material found in the Thuja group. Moreover, the extremely abundant precipitation probably explains the absence of a major soil moisture gradient. The longevity of Thuja plicata, and its apparently high resistance to wind damage, are features thought to be important in maintaining the high dominance of this species in forests nearest to the coast.

The Abies group was found over a range of coastal and more inland sites; thus, affinities with macro-climate were difficult to deduce. Vegetation appeared mainly related to meso-climate (elevation) and soil moisture gradients. The cool, moist micro-climates associated with several community types within the Abies group may have nullified the influence of macro-climate.

Alpha and beta diversity were found to increase towards the interior of the study area. These diversity increases may be caused by the increasing amount of heat available for plant growth, by the decreasing productivity brought on by moisture deficits, by the increasing frequency and severity of large scale fire disturbances, or, most likely, by a combination of all of these factors.

Analyses of tree species size-class distributions confirm the essentially seral role of Pseudotsuga menziesii in most community types, while Tsuga heterophylla, Abies amabilis and Thuja plicata are the major potential "climax" species. The eventual dominance of a particular species, or combination of species, is linked to a complex interplay of disturbance regime and ecological site characteristics.

Finally, a gradient analysis approach to resource inventory and management may represent an advantage over more traditional methods, in its relative freedom from resource mapping and complex integration of diverse resource maps. The vegetation patterns of a sector can instead be modelled through multiple regression equations using a few ecological factors, previously identified as strongly linked to vegetation variation. This information would then form a useful basis for forest management decisions bearing on harvesting, post-harvesting treatments, and species selection for reforestation.

The selection of appropriate tree species for reforestation is one of the most important steps in forest management. Within the area studied, it appears that Pseudotsuga menziesii constitutes the most appropriate choice in many situations. Possible exceptions are high elevation and nutrient poor sites where other species, such as Tsuga heterophylla, may grow as fast or faster. However, reforestation with Pseudotsuga menziesii on coastal sites, within the area occupied by the Thuja group, should be strictly avoided because of severe growth problems, likely caused by arsenic accumulation.

Integral conservation of particular sites or areas also should be part of a comprehensive and ecological forest management program. As forest management techniques develop, intensive management will be increasingly directed towards the most productive sites, with easiest access and gentle terrain. This represents a desirable trend if it allows forests to be used as a truly renewable resource, with better reforestation, control of soil erosion and minimal nutrient loss. As a
result, less logging pressure should be felt by less productive sites. Already, sites characterized as unproductive, such as rock outcrops, very steep slopes and some high elevation sites, are neglected in most logging operations. These sites are particularly suited for integral conservation; they are rich in species, usually occur at the extremeties of ecological gradients, and therefore represent ideal sites for the conservation of genetic variation (e.g. community types D1, D2, P7, T1, T5 and A1). The maintenance of genetic diversity within populations of economically valuable tree species is a duty of the forest industry and of the relevant governmental agencies. Thus, through the conservation of particular habitats, or entire areas, the forest industry could contribute towards this goal.

Another ecological aspect of importance to forest management is the prevalence of natural fire disturbance in the interior sector of the study area, and its virtual absence in the coastal sector. Therefore, the use of fire as a forest management tool may recreate naturally occurring phenomena in the Pseudotsuga forests, to which the biota is adapted, but unexpected ecological problems may be created in Thuja forests near the coast, where nutrient cycling appears to occur mostly within organic soil horizons, which may be partly or totally destroyed during burns.

It is hoped that the information contained in this thesis will contribute to improved forest management, and will not acquire an historical value too soon.

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Table 1 : List of environmental variables.

## I. TOPOGRAPHIC

1. elevation (m)
2. aspects $\left(0^{\circ}-180^{\circ}\right.$, NNE to SSW)
3. slope (\%)
4. position : 1 - crest

2 - upper-slope
3 - mid-slope
4 - lower-slope
5-1evel
6 - depression
II. EDAPHIC
(a) physical
5. drainage : 1 - very rapid

2 - rapid
3 - well
4 - moderately well
5 - imperfect
6 - poor
7 - very poor
6. effective rooting depth (cm)
7. root restricting depth (cm)
8. soil depth (cm)
9. material : 0-rock

1 - colluvial
2 - morainal
3 - fluvial
4 - alluvial
10. LFH tickness (cm)
11. $\mathrm{B}_{1} \%$ coarse fragments
12. $B_{1}$ texture: 1 - sand

2 - loamy sand
3 - sandy loam
4-1oam
5 - sandy clay loam
6 - silt loam
7 - silt
8 - sandy clay
9 - clay loam
10 - silty clay loam
11 - silty clay
12 - clay
13. V6/V7
14. V6/V8
15. V10/V6
II. EDAPHIC
(b) chemical
16. LFH pH ( $\left.\mathrm{H}_{2} \mathrm{O}\right)$
17. LFH $\mathrm{pH}\left(\mathrm{CaCl}_{2}\right)$
18. LFH \% C
19. LFH \% N
20. LFH C/N
21. A pH $\left(\mathrm{H}_{2} \mathrm{O}\right)$
22. A pH $\left(\mathrm{CaCl}_{2}\right)$
23. $\mathrm{B}_{1} \mathrm{pH}\left(\mathrm{H}_{2} \mathrm{O}\right)$
24. $\mathrm{B}_{1} \mathrm{pH}\left(\mathrm{CaCl}_{2}\right)$
25. $\mathrm{B}_{1} \% \mathrm{C}$
26. $\mathrm{B}_{1} \% \mathrm{~N}$
27. $\mathrm{B}_{1} \mathrm{C} / \mathrm{N}$
28. $\mathrm{B}_{2} \mathrm{pH}\left(\mathrm{H}_{2} \mathrm{O}\right)$
29. $\mathrm{B}_{2} \mathrm{pH}\left(\mathrm{CaCl}_{2}\right)$
III. GEOGRAPHIC
30. distance from coast
IV. DISTURBANCE
31. fire : 0 - no evidence 1 - charcoal or scars
32. wind : 0-no evidence 1 - blowdowns

Table 2 : List of community characteristics.

## I. RICHNESS

1. tree species
2. shrub species
3. herb species
4. bryophyte and lichen species
5. understory vascular species
6. vascular species
7. total species
II. COVERAGE (\%)
8. understory strata
9. shrub stratum
10. herb stratum
11. bryophyte and lichen stratum
III. TREE STRATUM
12. tree basal area ( $\mathrm{m}^{2} / \mathrm{ha}$ )
13. tree density (stems/ha)
14. maximum tree height (m)

Table 3 : Species with the ten largest positive and negative eigenvector coefficients on axes one and two of the reciprocal averaging ordination of the 172 plots.

## Axis 1 (\% variance $=11.0$ )

|  |  |  |  |
| :--- | :--- | :--- | :--- |
| Pinus contorta (sap.) | .387 | Abies amabilis (tree) | -.168 |
| P. contorta (tree) | .384 | A. amabilis (sap.) | -.138 |
| Pseudotsuga menziesii (seed.) | .330 | A. amabilis (seed.) | -.126 |
| P. menziesii (sap.) | .217 | Tsuga heterophy11a (seed.) | -.123 |
| Acer macrophy1lum (seed.) | .179 | T. heterophy1la (tree) | -.110 |
| P. menziesii (tree) | .177 | Vaccinium alaskaense | -.106 |
| Arbutus menziesii (tree) | .163 | T. heterophylla (sap.) | -.099 |
| P. contorta (seed.) | .162 | Blechnum spicant | -.098 |
| Rhacomitrium lanuginosum | .157 | Rubus pedatus | -.065 |
| Vaccinium ovatum | .155 | Picea sitchensis (tree) | -.055 |

## Axis 2 (\% variance $=8.8$ )

| Abies amabilis (sap.) | .301 | Picea sitchensis (tree) | -.351 |
| :--- | :--- | :--- | :--- |
| A. amabilis (tree) | .192 | P. sitchensis (seed.) | -.346 |
| A. amabilis (seed.) | .190 | Rubus spectabilis | -.276 |
| Vaccinium alaskaense | .172 | Ribes bracteosum | -.222 |
| Rhytidiopsis robusta | .157 | Polystichum munitum | -.212 |
| Tsuga mertensiana (tree) | .143 | Athyrium filix-femina | -.158 |
| Chamaecyparis nootkatensis (sap.) | .127 | Rubus parviflorus | -.154 |
| C. nootkatensis (tree) | .110 | Trautvetteria carolinensis | -.149 |
| P. contorta (tree) | .104 | Sambucus racemosa | -.142 |
| Rubus pedatus | .094 | A. macrophyllum (sap.) | -.131 |

Table 4 : Product moment correlations between environmental variables, community characteristics, and reciprocal averaging axes of the 172 plots ( $n=172 ; * *=p \leqslant .01$; * $=.01<\mathrm{p} \leqslant .05$ ).

|  | axis 1 | axis 2 |
| :---: | :---: | :---: |
| axis 2 | -.279** | - |
| elevation | -. 347 ** | .656** |
| aspect | . 299** | -. 100 |
| position | -. 269** | -. 299** |
| drainage | -. 386 ** | -. 014 |
| soil depth | -. 279** | -. 036 |
| material | -. 213** | -. 205** |
| LFH $\mathrm{pH}\left(\mathrm{H}_{2} \mathrm{O}\right)$ | . $292 \%$ \% | $\underline{-.413 * *}$ |
| LFH $\mathrm{pH}\left(\mathrm{CaCl}_{2}\right)$ | . $331 \% *$ | -. $432 * *$ |
| LFH thickness | -.293** | .180* |
| LFH \% N | -. $231 * *$ | . 051 |
| LFH C/N | .244** | . 007 |
| $\mathrm{B}_{1} \mathrm{pH}\left(\mathrm{H}_{2} \mathrm{O}\right)$ | . 039 | -. 215** |
| $\mathrm{B}_{1} \mathrm{pH}\left(\mathrm{CaCl}_{2}\right)$ | . 023 | -. 202 ** |
| $\mathrm{B}_{1} \%$ coarse fragments | . 235** | . 201 |
| effective rooting depth | .158* | -. 199** |
| eff. r. d./soil depth | . $453 * *$ | -. 123 |
| LFH thick./eff. r. d. | -. 257 ** | . 212 ** |
| fire disturbance | . $301 * *$ | -. 087 |
| tree spp. richness | . 304 ** | . 005 |
| shrub spp. richness | . 247 ** | -. 219** |
| understory coverage | . 129 | -. 218** |
| herb coverage | -.150* | -. 254** |
| bryo. coverage | . $408 * *$ | -. 102 |
| tree basal area | -. 205** | -. $264 * *$ |
| tree height | -.191* | -. 240 ** |

Table 5 : Species with the ten largest positive and negative eigenvector coefficients on axes one and two of the reciprocal averaging ordination of the 140 plots.

Axis 1 (\% variance $=13.6$ )

| Pseudotsuga menziesii (tree) | .339 | Abies amabilis (sap.) | -.361 |
| :--- | :--- | :--- | :--- |
| P. menziesii (seed.) | .238 | A. amabilis (tree) | -.336 |
| Acer macrophyllum (sap.) | .221 | A. amabilis (seed.) | -.266 |
| Cornus nuttallii (sap.) | .188 | Vaccinium alaskaense | -.190 |
| Hylocomium splendens | .187 | Rubus pedatus | -.169 |
| C. nuttallii (seed.) | .173 | Blechnum spicant | -.139 |
| A. macrophyllum (seed.) | .169 | Plagiothecium undulatum | -.089 |
| Berberis nervosa | .150 | Streptopus streptopoides | -.087 |
| Linnaea borealis | .146 | Rhizomnium glabrescens | -.072 |
| Thuja plicata (sap.) | .133 | Rhytidiadelphus loreus | -.072 |

Axis 2 (\% variance $=9.8$ )

Gaultheria shallon
T. plicata (seed.)

Vaccinium ovatum
Blechnum spicant
T. plicata (tree)
T. plicata (sap.)

Pinus contorta (tree)
Carex obnupta
Pyrus fusca
Cornus canadensis

| .333 | Abies amabilis (seed.) | -.223 |
| :--- | :--- | :--- |
| .310 | A. amabilis (tree) | -.216 |
| .285 | A. amabilis (sap.) | -.190 |
| .277 | P. menziesii (tree) | -.181 |
| .275 | Rubus pedatus | -.157 |
| .239 | A. macrophyllum | -.145 |
| .142 | C. nuttallii (sap.) | -.136 |
| .138 | C. nuttallii (seed.) | -.113 |
| .114 | Polystichum munitum | -.110 |
| .102 | A. macrophyllum (seed.) | -.110 |

Table 6 : Product moment correlations between environmental variables, community characteristics, and reciprocal averaging axes of the 140 plots ( $n=140$; ** $=p \leqslant .01$; * $=$. $01<\mathrm{p} \leqslant .05$ ).

|  | axis 1 | axis 2 |
| :---: | :---: | :---: |
| axis 2 | -. 408 * | - |
| elevation | -. 005 | $\underline{-.496 * *}$ |
| aspect | . 290 ** | -. 140 |
| slope | .182* | -. 307** |
| drainage | -. $532 \% *$ | . $469 * *$ |
| material | -. 300 ** | . 356 ** |
| LFH $\mathrm{pH}\left(\mathrm{H}_{2} \mathrm{O}\right)$ | . 240 ** | . 116 |
| LFH $\mathrm{pH}\left(\mathrm{CaCl}_{2}\right)$ | . 311 ** | . 084 |
| LFH thickness | -. 483 ** | . $358 * *$ |
| LFH \% N | -. $314 * *$ | . 061 |
| LFH C/N | . 252 ** | -. 135 |
| $\mathrm{B}_{1} \%$ coarse fragments | . 372 ** | -. 345 ** |
| $\mathrm{B}_{1} \% \mathrm{C}$ | -. 252 ** | . 112 |
| $\mathrm{B}_{1} \% \mathrm{~N}$ | -. $316 * *$ | . 118 |
| effective rooting depth | . 395 ** | -. 285 ** |
| root restricting depth | . 213 * | -. 264 ** |
| eff. r. d./soil depth | . $513 * *$ | -. 230** |
| LFH thick./eff. r. d. | $\underline{-.537 * *}$ | . 388 ** |
| fire disturbance | . 578 ** | -. $441 * *$ |
| wind disturbance | -. $437 * *$ | .591** |
| distance from coast | . 501 ** | -. $728 \% \%$ |
| tree spp. richness | . 268 ** | -. 020 |
| shrub spp. richness | -. 050 | . 223 ** |
| herb spp. richness | . 217 ** | -. 436** |
| bryo. spp. richness | -. 198** | .231** |
| vascular spp. richness | .208* | -. $310 \% *$ |
| understory coverage | -. 153 | . 456 ** |
| shrub coverage | -. 236 ** | .463** |
| herb coverage | -. $254 * *$ | . $363 * *$ |
| bryo. coverage | . 360 ** | -. 075 |
| tree height | . 225 ** | $\underline{-.472 * *}$ |

Table 7 : Species with the ten largest positive and negative eigenvector coefficients on axes one and two of the reciprocal averaging ordination of the Pseudotsuga group.

Axis 1 (\% variance $=14.4$ )

| Tsuga heterophylla (sap.) | .265 | Acer macrophyllum (sap.) | -.378 |
| :--- | :--- | :--- | :--- |
| T. heterophylla (seed.) | .253 | Cornus nuttallii (seed.) | -.274 |
| T. heterophylla (tree) | .225 | Pseudotsuga menziesii (seed.) | -.270 |
| Blechnum spicant | .112 | A. macrophyllum (seed.) | -.255 |
| Polystichum munitum | .089 | C. nuttallii (sap.) | -.236 |
| Vaccinium parvifolium | .075 | Thuja plicata (sap.) | -.187 |
| Scapania bolanderi | .072 | P. menziesii (sap.) | -.179 |
| Rhytidiopsis robusta | .071 | Rhytidiadelphus triquetrus | -.174 |
| Abies amabilis (sap.) | .059 | C. nuttallii (tree) | -.172 |
| Plagiothecium undulatum | .054 | Linnaea borealis | -.153 |

Axis 2 (\% variance $=9.6$ )

Polystichum munitum
C. nuttallii (seed.)
C. nuttallii (sap.)
T. plicata (seed.)
A. macrophyllum (sap.)

Blechnum spicant
A. macrophyllum (seed.)

Taxus brevifolia (sap.)
Rubus ursinus
Isothecium stoloniferum
. 379 Gaultheria shallon -. 489
. 188 Hylocomium splendens -. 355
.184 P. menziesii (seed.) -. 174
. 178 Pinus monticola (seed.) -. 117
. 176 Rhytidiopsis robusta -. 114
. 160 Vaccinium alaskaense -. 110
. 156 Rhytidiadelphus loreus -. 092
. 134 Vaccinium ovatum -. 090
. 120 Linnaea borealis -. 090
. 111 Chimaphila umbellata -. 084

Table 8 : Product moment correlations between environmental variables, community characteristics, and reciprocal averaging axes of the Pseudotsuga group ( $n=59$; ** $=\mathrm{p} \leqslant .01 ; \quad *=.01<\mathrm{p} \leqslant .05)$.

|  | axis 1 | axis 2 |
| :---: | :---: | :---: |
| axis 2 | .283* | - |
| elevation | . 197 | -. 372 ** |
| slope | . 185 | . 268 ** |
| position | . 238 | .640** |
| soil depth | . 371 ** | . 227 |
| LFH $\mathrm{pH}\left(\mathrm{H}_{2} \mathrm{O}\right)$ | -. 483 ** | -. 070 |
| LFH $\mathrm{pH}\left(\mathrm{CaCl}_{2}\right)$ | -. $534 \% \%$ | -. 099 |
| LFH thickness | . 336 ** | -. 059 |
| $\mathrm{B}_{1} \% \mathrm{~N}$ | . 186 | . 364 ** |
| $\mathrm{B}_{1} \mathrm{C} / \mathrm{N}$ | -. 104 | -. $434 * *$ |
| eff. r. d./soil depth | -. 277 * | -. 238 |
| distance from coast | -.431** | -. 373 ** |
| tree spp. richness | -. $463 * *$ | -. 117. |
| shrub spp. richness | -.423** | . 034 |
| herb spp. richness | -. 432 ** | . 071 |
| vascular spp. richness | -. $512 * *$ | . 045 |
| understory coverage | -.454** | $\underline{-.527 * *}$ |
| shrub coverage | -. 289* | -. $726 * *$ |
| herb coverage | -. 136 | . 390 ** |
| bryo. coverage | -. 408 ** | -.528** |
| tree basal area | . 069 | .622** |
| tree height | . 049 | . $588 * *$ |

Table 9 : Species with the ten largest positive and negative eigenvector coefficients on axes one and two of the reciprocal averaging ordination of the Thuja group.

## Axis 1 (\% variance $=23.4$ )

| Abies amabilis (sap.) | .309 | Vaccinum ovatum | -.401 |
| :--- | :--- | :--- | :--- |
| A. amabilis (tree) | .288 | Thuja plicata (sap.) | -.350 |
| Tsuga heterophylla (seed.) | .283 | T. plicata (seed.) | -.288 |
| Polystichum munitum | .146 | Carex obnupta | -.191 |
| T. heterophylla (sap.) | .130 | Pinus contorta (tree) | -.183 |
| A. amabilis (seed.) | .127 | Linnaea borealis | -.179 |
| Scapania bolanderi | .088 | Gaultheria shallon | -.155 |
| Vaccinium alaskaense | .078 | Pyrus fusca | -.136 |
| Blechnum spicant | .073 | Sphagnum girgensohnii | -.119 |
| T. heterophylla (tree) | .068 | Hylocomium splendens | -.118 |

Axis 2 (\%variance $=13.5$ )

| Linnaea borealis | .402 | Vaccinium ovatum | -.252 |
| :--- | :--- | :--- | :--- |
| Carex obnupta | .378 | Pseudotsuga menziesii (tree) | -.175 |
| A. amabilis (sap.) | .351 | T. heterophylla (sap.) | -.146 |
| Coptis asplenifolia | .295 | Gaultheria shallon | -.125 |
| Calamagrostis nutkaensis | .255 | T. plicata (sap.) | -.118 |
| Maianthemum dilatatum | .204 | T. heterophylla (tree) | -.109 |
| Lysichitum americanum | .169 | P. menziesii (seed.) | -.092 |
| Rubus spectabilis | .161 | Polystichum munitum | -.085 |
| Vaccinium ovalifolium | .141 | Taxus brevifolia (sap.) | -.080 |
| Cornus canadensis | .130 | Isothecium stoloniferum | -.072 |

Table 10 : Product moment correlations between environmental variables, community characteristics, and reciprocal averaging axes of the Thuja group ( $\mathrm{n}=40 ; * *=\mathrm{p} \leqslant .01$; * $=.01<\mathrm{p} \leqslant .05)$.

|  | axis 1 | axis 2 |
| :--- | :---: | :---: |
| axis 2 | .210 | - |
| elevation | $.453 * *$ | $.393 *$ |
| drainage | -.013 | $.385 *$ |
| soil depth | $.428 * *$ | .283 |
| LFH pH (H20) | .050 | $.361 *$ |
| LFH \% N | $-.473 * *$ | . .195 |
| LFH C/N | $.449 * *$ | .078 |
| B1 \% C | $.516 * *$ | -.061 |
| B1 \% N | .220 | $.448 * *$ |
| Bl C/N | $.356 *$ | $.383 *$ |
| root restricting depth | $-.581 * *$ | .180 |
| tree spp. richness | $-.448 * *$ | .133 |
| shrub spp. richness | .018 | $.640 * *$ |
| herb spp. richness | -.187 | $.568 * *$ |
| vascular spp. richness | $-.803 * *$ | .091 |
| understory coverage | $-.808 * *$ | -.246 |
| shrub coverage | -.159 | $.483 * *$ |
| herb coverage | $-.512 * *$ | .011 |
| bryo. coverage | $.647 * *$ | -.083 |
| tree height |  |  |

Table 11 : Species with the ten largest positive and negative eigenvector coefficients on axes one and three of the reciprocal averaging ordination of the Abies group.

Axis 1 (\% variance $=16.7$ )

| Tsuga heterophylla (sap.) | .339 | Abies amabilis (sap.) | -.356 |
| :--- | :--- | :--- | :--- |
| T. heterophylla (seed.) | .257 | Rubus pedatus | -.347 |
| Pseudotsuga menziesii (tree) | .222 | A. amabilis (seed.) | -.344 |
| Polystichum munitum | .213 | A. amabilis (tree) | -.285 |
| T. heterophylla (tree) | .186 | Streptopus streptopoides | -.188 |
| Gaultheria shallon | .131 | Vaccinium alaskaense | -.180 |
| Thuja plicata (tree) | .112 | Rhytididelphus loreus | -.155 |
| Vaccinium parvifolium | .110 | Streptopus roseus | -.134 |
| Blechnum spicant | .099 | Athyrium filix-femina | -.102 |
| Stokesiella oregana | .087 | Tiarella trifoliata | -.095 |

Axis 3 (\% variance $=10.3$ )

|  |  |  |  |
| :--- | :--- | :--- | :--- |
| Gaultheria shallon | .359 | Sphagnum girgensohnii | -.258 |
| A. amabilis (seed.) | .257 | A. amabilis (tree) | -.242 |
| Rhytidiopsis robusta | .237 | Achlys triphylla | -.231 |
| A. amabilis (sap.) | .218 | Polystichum munitum | -.219 |
| P. menziesii (tree) | .206 | Tiarella trifoliata | -.213 |
| T. heterophylla (tree) | .164 | Blechnum spicant | -.200 |
| Vaccinium alaskaense | .162 | T. heterophylla (sap.) | -.178 |
| Taxus brevifolia (sap.) | .139 | Rubus pedatus | -.152 |
| Chamaecyparis nootkatensis (tree) | .118 | T. heterophylla (seed.) | -.149 |
| Vaccinium parvifolium | .104 | Dryopteris austriaca | -.133 |

Table 12 : Product moment correlations between environmental variables, community characteristics, and reciprocal averaging axes of the Abies group $(n=40 ; * *=p \leqslant .01$; * $=.01<\mathrm{p} \leqslant .05$ ) .

| . | axis 1 | axis 3 |
| :---: | :---: | :---: |
| axis 3 | $\underline{-.499 * *}$ | - |
| elevation | $\underline{-.421 * *}$ | . $595 * *$ |
| position | . 071 | -. $500 * *$ |
| LFH \% C | -. $436 * *$ | . $314 * *$ |
| LFH C/N | -. 169 | . 470 ** |
| $\mathrm{B}_{1} \mathrm{pH}\left(\mathrm{H}_{2} \mathrm{O}\right)$ | . 349 * | -. 166 |
| $\mathrm{B}_{1} \mathrm{pH}\left(\mathrm{CaCl}_{2}\right)$ | . $364 *$ | -. 180 |
| $\mathrm{B}_{1} \mathrm{C} / \mathrm{N}$ | -. 065 | . $369 *$ |
| fire disturbance | . 098 | .411** |
| distance from coast | -. 488 ** | . 345 * |
| tree spp. richness | -. 197 | . 509 ** |
| herb spp. richness | -. 330* | . 024 |
| bryo. spp. richness | -. 073 | -. 312* |
| vascular spp. richness | -.331* | . 117 |
| Herb coverage | -. 067 | -. 360* |
| tree height | . 210 | $\underline{-.409 * *}$ |

Table 13 : Species with the ten largest positive and negative eigenvector coefficients on axes one and two of the reciprocal averaging ordination of the 105 modal plots.

Axis 1. (\% variance $=15.0$ )

| Pseudotsuga menziesii (tree) | .353 | Blechnum spicant | -.292 |
| :--- | :--- | :--- | :--- |
| P. menziesii (seed.) | .263 | Abies amabilis (sap.) | -.283 |
| Acer macrophyllum (sap.) | .263 | A. amabilis (tree) | -.206 |
| Cornus nuttallii (sap.) | .214 | Thuja plicata (tree) | -.168 |
| C. nuttallii (seed.) | .201 | A. amabilis (seed.) | -.140 |
| A. macrophyllum (seed.) | .195 | Vaccinium alaskaense | -.094 |
| Hylocomium splendens | .193 | Gaultheria shallon | -.093 |
| Linnaea borealis | .174 | Rhizomnium glabrescens | -.081 |
| Berberis nervosa | .161 | Tsuga heterophylla (tree) | -.070 |
| Achlys triphylla | .143 | Plagiothecium undulatum | -.067 |

Axis 2 (\% variance $=8.8$ )

|  |  |  |  |
| :--- | :--- | :--- | :--- |
| Tsuga heterophylla (sap.) | .333 | Gaultheria shallon | -.299 |
| T. heterophylla (seed.) | .280 | T. plicata (tree) | -.257 |
| T. heterophylla (tree) | .235 | Blechnum spicant | -.235 |
| Polystichum munitum | .213 | A. macrophyllum (sap.) | -.227 |
| P. menziesii (tree) | .128 | T. plicata (seed.) | -.224 |
| Picea sitchensis (tree) | .128 | A. amabilis (sap.) | -.223 |
| Rhytidiopsis robusta | .086 | T. plicata (sap.) | -.189 |
| Hypnum circinale | .080 | C. nuttaliii (seed.) | -.159 |
| Isothecium stoloniferum | .071 | A. macrophyllum (seed.) | -.136 |
| Polypodium glycyrrhiza | .058 | C. nuttallii (sap.) | -.113 |

Table 14 : Product moment correlations between environmental variables, community characteristics, and reciprocal averaging axes of the 105 modal plots ( $n=105$; ** $=\mathrm{p} \leqslant .01 ; *=.01<\mathrm{p} \leqslant .05)$.
axis 1
axis 2
axis 2

| $.670 * *$ | - |
| :--- | :---: |
| $.222 *$ | $.296 *$ |
| .150 | $.459 * *$ |
| $-.621 * *$ | $-.699 * *$ |
| $-.378 * *$ | $-.452 * *$ |
| $-.528 * *$ | $-.493 * *$ |
| $-.296 * *$ | $-.196 *$ |
| $.294 * *$ | .179 |
| $.372 * *$ | $.294 * *$ |
| $-.359 * *$ | $-.208 *$ |
| $-.419 * *$ | -.134 |
| $.393 * *$ | $.267 * *$ |
| $.201 *$ | $.267 * *$ |
| $.586 * *$ | $.280 * *$ |
| $-.608 * *$ | $-.468 * *$ |
| $.717 * *$ | $.543 * *$ |
| $-.711 * *$ | $-.652 * *$ |
| $.805 * *$ | . $.621 * *$ |
| $.318 * *$ | -.101 |
| -.030 | $-.284 * *$ |
| $.559 * *$ | $.324 * *$ |
| $-.266 * *$ | $-.207 * *$ |
| $.488 * *$ | .171 |
| $-.170 * *$ | $-.638 * *$ |
| $-.254 * *$ | $-.584 * *$ |
| $-.416 * *$ | $-.415 * *$ |
| $-.463 * *$ | -.176 |
| $-.390 * *$ | $-.265 *$ |
| $.291 * *$ | $.492 * *$ |

Table 15 : Names of community types and vegetation groups.

| Vegetation groups (code) | Community types (code) |
| :---: | :---: |
| Pinus contorta (D) | Dry Pinus-Pseudotsuga forests (D1) |
|  | Coastal dry Pinus forests (D2) |
| Floodplain (F) | Floodplain forests (F1) |
|  | Floodplain forests (Lysichitum variant) (F2) |
| Pseudotsuga (P) | Dry Pseudotsuga forests (P1) |
|  | Pseudotsuga-Thuja-Acer forests (P2) |
|  | Pseudotsuga-Linnaea forests (P3) |
|  | Pseudotsuga-Berberis forests (P4) |
|  | Tsuga-Pseudotsuga-Polystichum forests (P5) |
|  | Montane Tsuga forests (P6) |
|  | Montane Tsuga-Gaultheria forests (P7) |
| Thuja (T) | Coastal dry Thuja forests (T1) |
|  | Coastal Tsuga-Blechnum-Polystichum forests (T2) |
|  | Coastal montane Thuja forests (T3) |
|  | Coastal Thuja forests (T4) |
|  | Coastal wet Thuja forests (T5) |
| Abies (A) | Montane Tsuga-Abies-Gaultheria forests (A1) |
|  | Montane Abies-Tsuga forests (A2) |
|  | Montane Tsuga-Abies forests (A3) |
|  | Montane Abies-Streptopus forests (A4) |
|  | Lowland Abies forests (A5) |
|  | Tsuga-Gaultheria-Blechnum forests (A6) |
|  | Tsuga-Blechnum-Polystichum forests (A7) |
| Subalpine (SA) | (no community types differentiated) |

Table 16 : Canonical analysis results of vegetation groups based on environmental data.

Mahalanobis squared distances between groups :

Subalpine
Floodplain 41.1
Pinus contorta $67.7 \quad 106.9$

Pseudotsuga
16.536 .6
51.2

Thuja
23.533 .9
$66.0 \quad 16.1$

Abies
12.5
38.2
54.5
9.6
5.7
$\begin{array}{llllll}\text { SA } & \text { F } & \text { D } & \text { P } & \text { T } & \text { A }\end{array}$

Table 17 : Canonical analysis results of Pseudotsuga group community types based on environmental data (community type codes are listed in table 15).

Mahalanobis squared distances between types :

| P1 | - |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| P2 | 15.9 | - |  |  |  |  |
| P3 | 23.5 | 25.6 | - |  |  |  |
| P4 | 19.4 | 20.7 | 26.1 | - |  |  |
| P5 | 69.4 | 70.7 | 62.5 | 30.1 | - |  |
| P6 | 32.9 | 39.9 | 32.8 | 11.7 | 19.1 | - |
| P7 | 34.2 | 36.6 | 38.4 | 16.5 | 49.7 | 18.4 |

Table 18 : Canonical analysis results of Thuja group community types based on environmental data (community type codes are listed in table 15).

Mahalanobis squared distances between types :

| T2 | - |  |  |  |
| :---: | :---: | :---: | :---: | :---: |
| T3 | 262.8 | - |  |  |
| T4 | 140.3 | 82.0 | - |  |
| T5 | 206.1 | 196.1 | 53.1 | - |
|  | T2 | T3 | T4 | T5 |
|  |  |  |  |  |

Table 19 : Canonical analysis results of Abies group community types based on environmental data (community type codes are listed in table 15).

Mahalanobis squared distances between types :

| A1 | - |  |  |  |  |  |  |
| :--- | :--- | :--- | :--- | :--- | :--- | :--- | :--- |
| A2 | 1.8 | - |  |  |  |  |  |
| A3 | 5.3 | 6.2 | - |  |  |  |  |
| A4 | 20.9 | 20.2 | 36.8 | - |  |  |  |
| A5 | 7.0 | 14.7 | 11.6 | 26.0 | - |  |  |
| A6 | 5.5 | 11.5 | 9.6 | 31.7 | 2.1 | - |  |
| A7 | 7.8 | 13.6 | 4.2 | 44.5 | 5.4 | 3.5 | - |
|  | A1 | A2 | A3 | A4 | A5 | A6 | A7 |

Table 20 : Canonical analysis results of all community types based on environmental data (community type codes are listed in table 15, coastal dry Thuja forests (T1) are not included in this analysis).

Mahalanobis squared distances between types :

| SA | - |  |  |  |  |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| F1 | 73.2 | - |  |  |  |  |  |  |  |  |
| F2 | 89.3 | 38.3 | - |  |  |  |  |  |  |  |
| D1 | 82.0 | 139.7 | 156.0 | - |  |  |  |  |  |  |
| D2 | 124.3 | 154.9 | 185.8 | 54.4 | - |  |  |  |  |  |
| P1 | 51.8 | 77.0 | 82.5 | 58.9 | 81.0 | - |  |  |  |  |
| P2 | 58.4 | 56.7 | 70.5 | 57.5 | 69.5 | 15.2 | - |  |  |  |
| P3 | 43.1 | 57.3 | 68.8 | 72.9 | 103.8 | 9.9 | 16.4 | - |  |  |
| P4 | 33.8 | 52.3 | 73.3 | 55.3 | 81.5 | 12.4 | 12.6 | 17.3 | - |  |
| P5 | 36.4 | 61.6 | 85.0 | 85.1 | 108.5 | 30.6 | 28.7 | 36.0 | 12.7 | - |
| P6 | 19.7 | 59.3 | 82.2 | 65.7 | 92.2 | 19.4 | 23.6 | 23.1 | 7.9 | 11.5 |
| P7 | 18.4 | 64.4 | 82.7 | 51.6 | 79.0 | 27.9 | 29.0 | 30.8 | 12.2 | 27.2 |
| T2 | 44.7 | 68.7 | 75.7 | 82.2 | 88.0 | 51.3 | 45.6 | 55.0 | 31.4 | 20.1 |
| T3 | 34.4 | 69.4 | 59.2 | 103.6 | 124.2 | 52.2 | 52.3 | 47.9 | 36.7 | 34.0 |
| T4 | 45.3 | 52.8 | 50.4 | 94.0 | 111.1 | 46.1 | 40.4 | 43.5 | 29.0 | 31.3 |
| T5 | 52.8 | 45.3 | 38.9 | 114.7 | 132.2 | 50.4 | 44.4 | 45.9 | 38.3 | 47.0 |
| A1 | 25.6 | 82.8 | 94.2 | 55.5 | 93.0 | . 27.2 | 33.2 | 34.1 | 21.1 | 28.6 |
| A2 | 34.1 | 110.1 | 96.5 | 108.9 | 139.6 | 61.6 | 65.3 | 60.0 | 49.5 | 41.5 |
| A3 | 46.5 | 107.5 | 118.6 | 92.1 | 106.5 | 70.5 | 72.6 | 78.8 | 44.0 | 36.2 |
| A4 | 27.4 | 109.5 | 113.1 | 115.8 | 121.1 | 82.8 | 83.0 | 80.1 | 58.2 | 50.7 |
| A5 | 33.8 | 50.2 | 68.2 | 85.8 | 103.8 | 44.5 | 43.9 | 47.0 | 22.8 | 17.6 |
| A6 | 49.3 | 78.0 | 85.5 | 104.8 | 108.2 | 50.9 | 53.3 | 61.8 | 35.4 | 25.6 |
| A7 | 41.7 | 69.1 | 85.9 | 87.3 | 107.7 | 40.7 | 44.3 | 47.7 | 21.7 | 15.5 |
|  | SA | Fl | F2 | D1 | D2 | P1 | P2 | P3 | P4 | P5 |
| P7 | 9.3 | - |  |  |  |  |  |  |  |  |
| T2 | 33.1 | 35.2 | - |  |  |  |  |  |  |  |
| T3 | 35.3 | 33.5 | 22.1 | - |  |  |  |  |  |  |
| T4 | 36.6 | 33.5 | 16.5 | 11.2 | - |  |  |  |  |  |
| T5 | 44.9 | 45.9 | 46.9 | 28.6 | 14.6 | - |  |  |  |  |
| A1 | 12.5 | 18.4 | 42.4 | 50.3 | 44.1 | 53.1 | - |  |  |  |
| A2 | 38.8 | 48.1 | 35.3 | 26.0 | 35.1 | 57.7 | 35.8 | - |  |  |
| A3 | 41.9 | 39.8 | 28.9 | 24.8 | 37.7 | 63.6 | 61.4 | 47.0 | - |  |
| A4 | 46.9 | 37.8 | 36.0 | 23.1 | 36.5 | 52.5 | 58.1 | 35.9 | 25.3 |  |
| A5 | 25.4 | 29.1 | 16.4 | 19.7 | 14.1 | 27.6 | 35.6 | 40.1 | 29.3 |  |
| A6 | 35.8 | 42.3 | 21.7 | 22.9 | 21.3 | 36.5 | 46.5 | 40.6 | 30.4 |  |
| A7 | 23.8 | 33.9 | 15.4 | 26.4 | 21.5 | 42.8 | 32.4 | 29.5 | 29.9 |  |
|  | P6 | P7 | T2 | T3 | T4 | T5 | A1 | A2 | A3 |  |
| A5 | 34.9 | - |  |  |  |  |  |  |  |  |
| A6 | 33.0 | 11.9 | - |  |  |  |  |  |  |  |
| A7 | 43.7 | 11.7 | 18.7 | - |  |  |  |  |  |  |
|  | A4 | A5 | A6 | A7 |  |  |  |  |  |  |

Table 21 : Correlations between canonical variates and environmental variables (see Figs. 13 and 14).

| Environmentalvariables | Vegetation groups | All types | $\underline{\text { Pseudotsuga }}$ | Thuja | Abies |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | CV1 CV2 | CV1 CV2 | CV1 CV2 | CV1 CV2 | CV1 | CV2 |
| Elevation | . $32-.02$ | . $28-.56$ | . 12.60 | -. $25-.76$ | . 74 | . 63 |
| Aspect | . $41-.00$ | . $41-.14$ | .15 .09 | -. $01-.17$ |  |  |
| Slope | . $43-.13$ | . $34-.51$ | . $30-.07$ | . $06-.73$ | . 05 | . 91 |
| Position | -. 63 . 34 | -. 56 . 52 | . $58-.47$ | -. 12.48 |  |  |
| Drainage | -. $83-.10$ | -. 82.21 | .03 .05 | -. 35.41 |  |  |
| Eff. root. depth | .15 .43 | .19 .19 | . $03-.03$ | -. $08-.19$ |  |  |
| Root rest. depth | .05 .36 | . 02.02 | . 41.14 | -. $12-.31$ |  |  |
| Soil depth | -. 30.11 | -. $37-.08$ | .48 .12 | -. $06-.19$ |  |  |
| Material | -. 71.25 | -. 59.52 | -. 05.06 | -. 25.35 |  |  |
| LFH thickness | -. $36-.47$ | -. $53-.32$ | .38 . 23 | -. 02.28 |  | -. 18 |
| $\mathrm{B}_{1} \%$ coarse frag. | . $52-.18$ | . $43-.31$ | -. $01-.14$ | . $01-.21$ |  |  |
| Texture | -. $26-.08$ | -. 26 . 03 | -. 06 . 10 | -. 01.04 | -. 03 | . 19 |
| Eff. r. d./r rest. d. | .18 . 19 | . 29.27 | -. $42-.22$ | .04 .15 |  |  |
| Eff. r. d./soil d. | .51 . 28 | . 60.21 | -. $46-.20$ | . $05-.03$ |  |  |
| LFH thick./eff. r. d. | -. $34-.58$ | -. $50-.38$ | . 24.23 | -. 19.26 |  |  |
| LFH $\mathrm{pH}\left(\mathrm{H}_{2} \mathrm{O}\right.$ ) | -. 10.42 | $.10 \quad .67$ | -. $38-.52$ | -. $28-.01$ |  |  |
| LFH $\mathrm{pH}\left(\mathrm{CaCl}_{2}\right)$ | -. 07.49 | $.16 \quad .70$ | -. $44-.51$ | -. 22.07 |  |  |
| LFH \% C | . $23-.21$ | . $12-.38$ | . 16.20 | . $33-.10$ |  |  |
| LFH \% N | -. $25-.38$ | -. $44-.35$ | . $39-.36$ | -. $03-.26$ |  |  |
| LFH C/N | .41 .22 | . 54.11 | -. $25-.46$ | .23 .12 |  |  |
| $\mathrm{B}_{1} \mathrm{pH}\left(\mathrm{H}_{2} \mathrm{O}\right)$ | .06 .34 | . 17.46 | -. $04-.54$ | -. 02.31 |  |  |
| $\mathrm{B}_{1} \mathrm{pH}\left(\mathrm{CaCl}_{2}\right)$ | . 05.24 | .13 .34 | . $02-.55$ | -. 02.21 |  |  |
| $\mathrm{B}_{1} \% \mathrm{C}$ | -. $08-.34$ | -. $09-.17$ | -. $01-.45$ | . $24-.42$ |  |  |
| $\mathrm{B}_{1} \% \mathrm{~N}$ | -. $22-.28$ | -. $27-.09$ | . $42-.43$ | . $39-.42$ |  |  |
| $\mathrm{B}_{1} \% \mathrm{C} / \mathrm{N}$ | . $25-.01$ | . $34-.08$ | -. $30-.00$ | -. $27-.11$ |  |  |
| Fire disturbance | . 53.39 | . 65.14 | -. $07-.38$ | -. 01.42 |  |  |
| Wind disturbance | -. $58-.39$ | -. $62-.05$ | -. $08 \quad .34$ | -. $18-.09$ |  |  |

Vegetation groups : $\mathrm{n}=157$, r at $.01=.21$;
Pseudotsuga $: n=56, r$ at $.01=.34 ;$ Thuja $: n=36, r$ at $.01=.34$;
Abies : $\mathrm{n}=32$, r at $.01=.45$
All types : $\mathrm{n}=149$, r at $.01=.21$;

Table 22 : Pseudotsuga group and community type Dl tree strata summary table (see table 15 for community type codes).

| Community types | D1 | P1 | P2 | P3 | P4 | P5 | P6 | P7 | D1 | P1 | P2 | P3 | P4 | P5 | P6 | P7 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Number of: plots | 3 | 4 | 4 | 5 | 11 | 17 | 12 | 7 |  |  |  |  |  |  |  |  |
| Trees (> 10 cm DBH) | Mean relative importance value (\%) |  |  |  |  |  |  |  |  |  | Constancy (\%) |  |  |  |  |  |
| Acer macrophyllum |  |  | 6 | + | $+$ | + |  |  |  |  | 100 | 20 | 9 | 5 |  |  |
| Arbutus menziesii | 13 | 4 |  |  |  |  |  |  | 100 | 50 |  |  |  |  |  |  |
| Cornus nuttallii |  |  | 6 | 1 | 1 |  |  |  |  |  | 100 | 20 | 27 |  |  |  |
| Pinus contorta | 66 | 1 |  |  |  |  |  |  | 100 | 25 |  |  |  |  |  |  |
| Pseudotsuga menziesii | 20 | 93 | 67 | 69 | 50 | 42 | 33 | 30 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 |
| Thuja plicata | 1 | 1 | 12 | 9 | 12 | 11 | 12 | 14 | 33 | 25 | 50 | 60 | 72 | 65 | 75 | 71 |
| Tsuga heterophylla |  | 1 | 8 | 17 | 34 | 44 | 54 | 55 |  | 25 | 75 | 100 | 100 | 100 | 100 | 100 |
| Saplings (0-10 cm DBH) |  | Mean | rel | ativ | e de | , | ( |  |  |  |  |  |  |  |  |  |
| Acer macrophyllum |  |  | 29 | 3 | $+$ |  |  |  |  |  | 100 | 20 | 9 |  |  |  |
| Arbutus menziesii | 25 | 13 |  |  |  |  |  |  | 66 | 25 |  |  |  |  |  |  |
| Cornus nuttallii |  | 5 | 17 | 1 | 15 |  |  |  |  | 50 | 75 | 20 | 36 |  |  |  |
| Pinus contorta | 53 | 31 |  |  |  |  |  |  | 100 | 25 |  |  |  |  |  |  |
| Pseudotsuga menziesii | 22 | 35 | 10 | 5 |  | $+$ |  |  | 100 | 75 | 75 | 20 |  | 5 |  |  |
| Thuja plicata |  | 1 | 24 | 12 | 13 | 1 | 10 | 12 |  | 25 | 100 | 40 | 27 | 17 | 33 | 71 |
| Tsuga heterophylla |  | 13 | 17 | 68 | 66 | 94 | 82 | 73 |  | 25 | 100 | 100 | 90 | 100 | 100 | 100 |
| Seedlings (below BH) |  | Mean | rel | ativ | e de | nsi | y (\%) |  |  |  |  |  |  |  |  |  |
| Abies amabilis |  |  |  | 5 | 1 | 1 | 4 | $+$ |  |  |  | 20 | 18 | 17 | 58 | 14 |
| Acer macrophyllum | 4 | 2 | 18 |  | + | 2 | + |  | 100 | 50 | 100 |  | 36 | 11 | 8 |  |
| Arbutus menziesii | 18 | 26 |  |  |  |  |  |  | 33 | 50 |  |  |  |  |  |  |
| Cornus nuttallii |  | 2 | 20 | 1 | 2 | + | $+$ |  |  | 50 | 100 | 20 | 45 | 5 | 16 |  |
| Pinus contorta | 9 |  |  |  |  |  |  |  | 100 |  |  |  |  |  |  |  |
| Pseudotsuga menziesii | 67 | 64 | 26 | 41 | 10 | 1 | 3 | 7 | 100 | 100 | 100 | 100 | 100 | 52 | 75 | 71 |
| Thuja plicata |  | 2 | 15 | 3 | 8 | 12 | 6 | 7 |  | 50 | 75 | 40 | 72 | 52 | 75 | 85 |
| Tsuga heterophylla | 2 | 3 | 19 | 48 | 77 | 83 | 85 | 83 | 33 | 25 | 100 | 100 | 100 | 100 | 100 | 100 |

Table 22 (continued)

| Community types | D1 | Pl | P2 | P3 | P4 | P5 | P6 | P7 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Species richness ${ }^{1}$ (.05 ha) | $\begin{gathered} 4.3 \\ (0.6)^{2} \end{gathered}$ | $\begin{gathered} 4.2 \\ (2.2) \end{gathered}$ | $\begin{gathered} 5.7 \\ (0.9) \end{gathered}$ | $\begin{gathered} 4.2 \\ (0.8) \end{gathered}$ | $\begin{gathered} 4.4 \\ (1.1) \end{gathered}$ | $\begin{gathered} 3.7 \\ (0.9) \end{gathered}$ | $\begin{gathered} 3.9 \\ (0.9) \end{gathered}$ | $\begin{gathered} 3.8 \\ (1.4) \end{gathered}$ |
| Total species ${ }^{1}$ | 6 | 8 | 8 | 10 | 9 | 8 | 9 | 7 |
| Species diversity ${ }^{3}: \operatorname{Exp}_{1 / \lambda^{6}}\left(H^{\prime}\right)^{5}$ | $\begin{aligned} & 2.5 \\ & 2.0 \end{aligned}$ | $\begin{aligned} & 1.4 \\ & 1.1 \end{aligned}$ | $\begin{aligned} & 2.9 \\ & 2.1 \end{aligned}$ | $\begin{aligned} & 2.7 \\ & 1.9 \end{aligned}$ | $\begin{aligned} & 3.1 \\ & 2.6 \end{aligned}$ | $\begin{aligned} & 3.0 \\ & 2.6 \end{aligned}$ | $\begin{aligned} & 2.7 \\ & 2.4 \end{aligned}$ | $\begin{aligned} & 2.8 \\ & 2.4 \end{aligned}$ |
| Mean basal area ( $\left.\mathrm{m}^{2} / \mathrm{ha}\right)^{4}$ | $\begin{aligned} & 30.7 \\ & (6.5) \end{aligned}$ | $\begin{gathered} 86.2 \\ (24.5) \end{gathered}$ | $\begin{aligned} & 132.5 \\ & (20.5) \end{aligned}$ | $\begin{gathered} 89.4 \\ (20.3) \end{gathered}$ | $\begin{aligned} & 138.4 \\ & (43.5) \end{aligned}$ | $\begin{aligned} & 158.4 \\ & (52.9) \end{aligned}$ | $\begin{aligned} & 114.5 \\ & (31.1) \end{aligned}$ | $\begin{gathered} 86.0 \\ (13.4) \end{gathered}$ |
| Mean density(trees/ha) ${ }^{3}$ | $\begin{aligned} & 700 \\ & (87) \end{aligned}$ | $\begin{aligned} & 300 \\ & (73) \end{aligned}$ | $\begin{gathered} 340 \\ (157) \end{gathered}$ | $\begin{gathered} 420 \\ (136) \end{gathered}$ | $\begin{gathered} 485 \\ (104) \end{gathered}$ | $\begin{gathered} 414 \\ (118) \end{gathered}$ | $\begin{gathered} 388 \\ (122) \end{gathered}$ | $\begin{gathered} 634 \\ (288) \end{gathered}$ |
| Mean max. height (m) | $\begin{gathered} 18 \\ (7) \end{gathered}$ | $\begin{array}{r} 44 \\ (7) \end{array}$ | $\begin{array}{r} 64 \\ (8) \end{array}$ | $\begin{gathered} 48 \\ (10) \end{gathered}$ | $\begin{gathered} 58 \\ (10) \end{gathered}$ | $\begin{gathered} 61 \\ (8) \end{gathered}$ | $\begin{array}{r} 50 \\ (8) \end{array}$ | $\begin{gathered} 41 \\ (16) \end{gathered}$ |

[^2]Table 23 : Pseudotsuga group and community type D1 understory strata summary table (see table 15 for community type codes).

| Community types | D1 | P1 | P2 | P3 | P4 | P5 | P6 | P7 | D1 | P1 | P2 | P3 | P4 | P5 | P6 | P7 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Number of plots | 3 | 4 | 4 | 5 | 11 | 17 | 12 | 7 |  |  |  |  |  |  |  |  |



## Herbs

| Achillea millefolium | + |  |  |  |  |  |  |  | 66 |  |  |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Achlys triphylla | + | 2 | 11 | 12 | 4 | 3 | 4 | $+$ | 33 | 100 | 100 | 100 | 100 | 100 | 75 | 42 |
| Adenocaulon bicolor |  |  | + | + | $+$ | + |  |  |  |  | 50 | 20 | 18 | 17 |  |  |
| Allotropa virgata |  | + |  | + | + |  | $+$ | $+$ |  | 50 |  | 40 | 27 |  | 16 | 14 |
| Apocynum androsaemifolium | + | + |  |  |  |  | $+$ |  | 66 | 50 |  |  |  |  | 8 |  |
| Arenaria macrophylla | + | $+$ |  | + | + |  |  |  | 33 | 50 |  | 20 | 9 |  |  |  |
| Blechnum spicant |  |  |  |  | + | 3 | 2 | $+$ |  |  |  |  | 9 | 52 | 58 | 28 |
| Boschniakia hookeri |  | + |  | + | + |  |  | $+$ |  | 100 |  | 60 | 9 |  |  | 42 |
| Bromus vulgaris |  |  | + | + |  | + |  |  |  |  | 50 | 20 |  | 5 |  |  |
| Calypso bulbosa |  |  | + | + | + |  |  |  |  |  | 50 | 40 | 27 |  |  |  |
| Campanula scouleri |  | $+$ | + | + | $+$ |  | $+$ |  |  | 75 | 50 | 40 | 9 |  | 16 |  |
| Chimaphila menziesii |  | $+$ | $+$ | + | $+$ | $+$ | + |  |  | 50 | 75 | 80 | 63 | 17 | 50 |  |

Table 23 (continued)

| Community types | D1 | P1 | P2 | P3 | P4 | P5 | P6 | P7 | D1 | P1 | P2 | P3 | P4 | P5 | P6 | P7 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Herbs (continued) |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |
| Chimaphila umbellata | + | 6 | $+$ | 2 | + | + | + | + | 33 | 100 | 50 | 100 | 54 | 5 | 41 | 42 |
| Corallorhiza maculata |  |  | + | + |  |  | + | + |  |  | 25 | 60 |  |  | 8 | 14 |
| Corallorhiza mertensiana |  |  |  | + | $+$ | $+$ | + | + |  |  |  | 60 | 27 | 11 | 50 | 28 |
| Cryptogramma crispa | + |  |  |  |  |  |  |  | 66 |  |  |  |  |  |  |  |
| Danthonia spicata | 4 | 1 |  |  |  |  |  |  | 100 | 50 |  |  |  |  |  |  |
| Festuca occidentalis | + | 3 | 2 | 1 | + |  | + | + | 66 | 100 | 50 | 40 | 9 |  | 8 | 14 |
| Festuca ovina | 1 |  |  |  |  |  |  |  | 66 |  |  |  |  |  |  |  |
| Festuca subulif1ora |  | $+$ | 3 | 2 | + | + | + | 1 |  | 50 | 100 | 100 | 36 | 35 | 16 | 28 |
| Fragaria virginiana | + | $+$ |  |  |  |  |  |  | 66 | 50 |  |  |  |  |  |  |
| Galium triflorum |  |  | + |  |  | $+$ |  |  |  |  | 50 |  |  | 29 |  |  |
| Goodyera oblongifolia | + | + | $+$ | + | $+$ | $+$ | + | + | 100 | 100 | 100 | 100 | 81 | 35 | 33 | 42 |
| Hieracium albiflorum | $+$ | 1 | + | + |  |  | + | + | 100 | 75 | 25 | 20 |  |  | 16 | 14 |
| Hypochaeris radicata | + | + |  |  |  |  |  |  | 66 | 50 |  |  |  |  |  |  |
| Lathyrus nevadensis |  |  | + | 1 |  |  |  |  |  |  | 25 | 60 |  |  |  |  |
| Linnaea borealis | 1 | 8 | 10 | 20 | 8 | 2 | 1 | 3 | 33 | 100 | 100 | 100 | 81 | 29 | 33 | 28 |
| Listera cordata | + | $+$ | + | + | + | $+$ | + | + | 66 | 75 | 75 | 80 | 54 | 17 | 50 | 85 |
| Madia madioides |  | 1 |  |  |  |  |  |  |  | 50 |  |  |  |  |  |  |
| Montia parviflora |  |  | + |  | + | $+$ |  |  |  |  | 50 |  | 9 | 17 |  |  |
| Polypodium glycyrrhiza | + | + | 1 | + | + | $+$ | + | + | 66 | 100 | 100 | 40 | 36 | 35 | 25 | 14 |
| Polystichum munitum | + | $+$ | 18 | 4 | 8 | 29 | 3 | + | 66 | 100 | 100 | 100 | 100 | 100 | 75 | 57 |
| Pteridium aquilinum |  | 6 | + | + | + | + | + | + |  | 100 | 75 | 60 | 63 | 23 | 8 | 14 |
| Saxifraga ferruginea | + |  |  |  |  |  |  |  | $66^{\prime}$ |  |  |  |  |  |  |  |
| Selaginella wallacei | 5 | $+$ |  | + |  | + |  |  | 100 | 25 |  | 20 |  | 5 |  |  |
| Tiarella laciniata |  |  | + | + | + | + | + |  |  |  | 25 | 20 | 54 | 41 | 41 |  |
| Tiarella trifoliata |  |  | 1 | + | 1 | 2 | 1 |  |  |  | 50 | 20 | 63 | 88 | 33 |  |
| Trientalis latifolia | + | 2 | 4 | 2 | + | + | + | + | 66 | 100 | 100 | 80 | 81 | 17 | 33 | 28 |
| Trillium ovatum |  |  | + |  | + | + | + |  |  |  | 25 |  | 54 | 76 | 58 |  |
| Viola sempervirens | + | + | 1 | 2 | 2 | $+$ | + |  | 33 | 75 | 75 | 80 | 54 | 17 | 25 |  |

Table 23 (continued)

| Community types | D1 | P1 | P2 | P3 | P4 | P5 | P6 | P7 | D1 | P1 | P2 | P3 | P4 | P5 | P6 | P7 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Bryophytes \& lichens |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |
| Cladina impexa | 1 |  |  |  |  |  |  |  | 66 |  |  |  |  |  |  |  |
| Cladina rangiferina | 5 | + |  |  |  |  |  | + | 100 | 50 |  |  |  |  |  | 14 |
| Cladonia bellidiflora | 1 | + |  |  |  |  |  | + | 100 | 25 |  |  |  |  |  | 14 |
| Cladonia multiformis | 3 | + |  |  |  |  |  |  | 100 | 50 |  |  |  |  |  |  |
| Cladonia gracilis | 1 | + |  |  |  |  |  |  | 66 | 25 |  |  |  |  |  |  |
| Dicranum fuscescens | + | 2 | 1 | 1 | + | + | 1 | 2 | 33 | 100 | 100 | 100 | 90 | 58 | 58 | 100 |
| Dicranum scoparium | 10 | 2 | + | + |  |  |  |  | 66 | 50 | 25 | 20 |  |  |  |  |
| Heterocladium macounii |  |  |  |  | 1 | 1 | + |  |  |  |  |  | 27 | 52 | 33 |  |
| Hylocomium splendens | 2 | 21 | 22 | 52 | 23 | 2 | 14 | 35 | 100 | 100 | 100 | 100 | 100 | 76 | 91 | 100 |
| Hypnum circinale |  | + | + | + | 1 | 3 | 3 | 1 |  | 50 | 50 | 40 | 81 | 94 | 100 | 85 |
| Isopterygium elegans | + |  |  | + | + | + | + |  | 33 |  |  | 20 | 27 | 76 | 58 |  |
| Isothecium stoloniferum | 2 | 3 | 3 | + | 2 | 5 | 4 | $+$ | 33 | 75 | 75 | 20 | 81 | 94 | 83 | 42 |
| Leucolepis menziesii |  |  | 3 | + | + | + | + |  |  |  | 75 | 60 | 18 | 11 | 8 |  |
| Peltigera leucophlebia | 1 | + |  |  |  |  |  | + | 66 | 75 |  |  |  |  |  | 28 |
| Peltigera membranacea | + | 1 |  |  | $+$ | $+$ | + | $+$ | 33 | 75 |  |  | 18 | 5 | 16 | 14 |
| Peltigera polydactyla |  |  |  |  | + | + | + | $+$ |  |  |  |  | 18 | 17 | 50 | 14 |
| Plagiothecium undulatum | $+$ | + | $+$ | 1 | $+$ | . 2 | 2 | $+$ | 33 | 25 | 50 | 60 | 45 | 70 | 50 | 57 |
| Pogonatum contortum |  |  | 1 | 3 | $+$ | $+$ | + | + |  |  | 50 | 100 | 9 | 29 | 50 | 28 |
| Polytrichum commune | 1 | + |  |  |  |  |  |  | 100 | 25 |  |  |  |  |  |  |
| Polytrichum juniperinum | 4 | 2 |  | + |  |  |  |  | 100 | 100 |  | 20 |  |  |  |  |
| Rhacomitrium canescens | 3 | 2 |  |  |  |  |  |  | 100 | 75 |  |  |  |  |  |  |
| Rhizomnium glabrescens |  |  | + | + | + | $+$ | + | + |  |  | 25 | 20 | 54 | 65 | 41 | 14 |
| Rhytidiadelphus loreus | + | $+$ | 2 | 4 | 6 | 4 | 4 | 15 | 66 | 100 | 75 | 100 | 90 | 76 | 83 | 85 |
| Rhytidiadelphus triquetrus | 1 | + | 6 | 1 | + | $+$ |  | 1 | 66 | 75 | 75 | 80 | 18 | 5 |  | 28 |
| Rhytidiopsis robusta | + |  |  | 1 | 2 | + | 9 | 5 | 33 |  |  | 40 | 63 | 17 | 66 | 71 |
| Scapania americana | + |  |  |  |  |  |  |  | 66 |  |  |  |  |  |  |  |
| Scapania bolanderi |  |  | $+$ | 1 | 1 | 2 | 4 | 2 |  |  | 50 | 100 | 81 | 94 | 100 | 85 |
| Stereocaulon tomentosum | 1 |  |  |  |  |  |  |  | 66 |  |  |  |  |  |  |  |
| Stokesiella oregana | 4 | 28 | 24 | 6 | 20 | 13 | 10 | 10 | 100 | 100 | 100 | 100 | 100 | 94 | 91 | 71 |
| Trachybryum megaptilum | 1 | 1 |  | 1 | $+$ | $+$ | 1 | 1 | 33 | 100 |  | 40 | 45 | 5 | 50 | 71 |
| Rock | 28 | 2 | 9 | 1 | 2 | 10 | 8 | 2 | 100 | 50 | 50 | 20 | 54 | 88 | 75 | 28 |

Table 23 (continued)

| Community types | D1 | P1 | P2 | P3 | P4 | P5 | P6 | P7 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Strata mean coverage (\%) |  |  |  |  |  |  |  |  |
| Shrubs | $\begin{aligned} & 45 \\ & (7)^{1} \end{aligned}$ | $\begin{gathered} 98 \\ (35) \end{gathered}$ | $\begin{gathered} 38 \\ (12) \end{gathered}$ | $\begin{gathered} 72 \\ (46) \end{gathered}$ | $\begin{gathered} 39 \\ (24) \end{gathered}$ | $\begin{gathered} 12 \\ (11) \end{gathered}$ | $\begin{gathered} 36 \\ (32) \end{gathered}$ | $\begin{aligned} & 102 \\ & (12) \end{aligned}$ |
| Herbs | $\begin{gathered} 24 \\ (26) \end{gathered}$ | $\begin{gathered} 35 \\ (10) \end{gathered}$ | $\begin{gathered} 54 \\ (28) \end{gathered}$ | $\begin{gathered} 49 \\ (28) \end{gathered}$ | $\begin{gathered} 26 \\ (25) \end{gathered}$ | $\begin{gathered} 44 \\ (29) \end{gathered}$ | $\begin{gathered} 13 \\ (15) \end{gathered}$ | $\begin{gathered} 7 \\ (8) \end{gathered}$ |
| Bryo. \& lichens | 54 <br> (5) | $\begin{gathered} 64 \\ (14) \end{gathered}$ | 64 <br> (9) | $\begin{gathered} 73 \\ (31) \end{gathered}$ | $\begin{gathered} 59 \\ (25) \end{gathered}$ | $\begin{gathered} 35 \\ (19) \end{gathered}$ | $\begin{gathered} 55 \\ (24) \end{gathered}$ | $\begin{gathered} 73 \\ (38) \end{gathered}$ |
| Total understory <br> Species richness (. 05 ha ) | $\begin{aligned} & 123 \\ & (22) \end{aligned}$ | $\begin{aligned} & 197 \\ & (24) \end{aligned}$ | $\begin{aligned} & 157 \\ & (36) \end{aligned}$ | $\begin{aligned} & 195 \\ & (54) \end{aligned}$ | $\begin{aligned} & 125 \\ & (48) \end{aligned}$ | $\begin{gathered} 91 \\ (37) \end{gathered}$ | $\begin{aligned} & 105 \\ & (46) \end{aligned}$ | $\begin{aligned} & 182 \\ & (51) \end{aligned}$ |
| Shrubs | $\begin{gathered} 8.7 \\ (1.5) \end{gathered}$ | $\begin{gathered} 7.8 \\ (0.3) \end{gathered}$ | $\begin{gathered} 6.8 \\ (1.5) \end{gathered}$ | $\begin{gathered} 6.4 \\ (1.5) \end{gathered}$ | $\begin{gathered} 5.7 \\ (2.4) \end{gathered}$ | $\begin{gathered} 4.5 \\ (2.5) \end{gathered}$ | $\begin{gathered} 3.8 \\ (1.4) \end{gathered}$ | $\begin{gathered} 3.6 \\ (1.3) \end{gathered}$ |
| Herbs | 17.3 <br> (7) | $\begin{aligned} & 20.3 \\ & (0.9) \end{aligned}$ | $\begin{aligned} & 18.5 \\ & (4.4) \end{aligned}$ | $\begin{aligned} & 17.4 \\ & (6.2) \end{aligned}$ | $\begin{aligned} & 14.3 \\ & (2.8) \end{aligned}$ | $\begin{aligned} & 11.7 \\ & (6.2) \end{aligned}$ | $\begin{aligned} & 10.4 \\ & (5.5) \end{aligned}$ | $\begin{gathered} 6.9 \\ (5.7) \end{gathered}$ |
| Bryo. \& 1ichens | $\begin{aligned} & 18.7 \\ & (3.2) \end{aligned}$ | $\begin{aligned} & 13.8 \\ & (3.4) \end{aligned}$ | $\begin{gathered} 9.5 \\ (3.3) \end{gathered}$ | $\begin{aligned} & 11.0 \\ & (2.0) \end{aligned}$ | $\begin{aligned} & 10.8 \\ & (2.4) \end{aligned}$ | $\begin{aligned} & 11.0 \\ & (2.5) \end{aligned}$ | $\begin{aligned} & 12.0 \\ & (2.3) \end{aligned}$ | $\begin{gathered} 9.9 \\ (2.3) \end{gathered}$ |
| Total species : Shrubs | 12 | 14 | 12 | 12 | 15 | 17 | 12 | 9 |
| Herbs | 32 | 33 | 36 | 35 | 41 | 54 | 42 | 25 |
| Bryo. \& lichens | 31 | 23 | 17 | 20 | 26 | 27 | 29 | 21 |
| Species diversity |  |  |  |  |  |  |  |  |
| Shrubs \& herbs $\operatorname{Exp}_{1 / \lambda^{3}}\left(H^{\prime}\right)^{2}$ | $\begin{array}{r} 9.6 \\ 18.1 \end{array}$ | $\begin{array}{r} 15.5 \\ 8.7 \end{array}$ | 15.2 13.8 | $\begin{array}{r} 15.0 \\ 9.1 \end{array}$ | $\begin{aligned} & 14.7 \\ & 11.5 \end{aligned}$ | $\begin{aligned} & 8.7 \\ & 6.0 \end{aligned}$ | $\begin{aligned} & 8.9 \\ & 8.8 \end{aligned}$ | $\begin{aligned} & 5.7 \\ & 2.9 \end{aligned}$ |
| Bryo. \& lichens $\operatorname{Exp}_{I / \lambda^{3}}\left(H^{\prime}\right)^{2}$ | $\begin{aligned} & 20.6 \\ & 16.2 \end{aligned}$ | 9.0 5.4 | 6.6 4.5 | 7.1 3.9 | 8.4 5.6 | 10.7 7.5 | $\begin{aligned} & 12.8 \\ & 10.0 \end{aligned}$ | $\begin{aligned} & 8.3 \\ & 6.3 \end{aligned}$ |

[^3]Table 24 : Thuja group and community types D2, F1 and F2 tree strata summary table (see table 15 for community type codes).

| Community tÿpes | D2 | T1 | T2 | T3 | T4 | T5 | F2 | F1 | D2 | T1 | T2 | T3 | T4 | T5 | F2 | F1 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Number of plots | 4 | 3 | 6 | 7 | 19 | 4 | 2 | 8 |  |  |  |  |  |  |  |  |
| Trees (> 10 cm DBH ) | Mean relative importance value (\%) |  |  |  |  |  |  |  | Constancy (\%) |  |  |  |  |  |  |  |
| Abies amabilis |  | 1 | 28 | 18 | 12 |  | 4 | 9 |  | 33 | 100 | 100 | 89 |  | 50 | 12 |
| Chamaecyparis nootkatensis | 15 | 4 |  | 5 |  |  |  |  | 100 | 33 |  | 28 |  |  |  |  |
| Picea sitchensis |  | 1 | 5 |  | + |  | 37 | 35 |  | 33 | 16 |  | 5 |  | 100 | 62 |
| Pinus contorta | 28 |  |  |  |  | 11 |  |  | 100 |  |  |  |  | 75 |  |  |
| Pinus monticola | 4 |  |  |  | + | 1 |  |  | 75 |  |  |  | 5 | 25 |  |  |
| Pseudotsuga menziesii | 17 | 11 | 3 |  |  |  |  | 5 | 100 | 66 | 16 |  |  |  |  | 12 |
| Taxus brevifolia |  | 3 |  | 1 | + | 6 | 1 |  |  | 100 |  | 42 | 15 | 100 | 50 |  |
| Thuja plicata | 17 | 47 | 18 | 51 | 54 | 53 | 37 | 21 | 75 | 100 | 50 | 100 | 100 | 100 | 100 | 62 |
| Tsuga heterophylla | 19 | 34 | 46 | 25 | 33 | 29 | 21 | 26 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 87 |
| Saplings ( $0-10 \mathrm{~cm} \mathrm{DBH}$ ) | Mean relative density (\%) |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |
| Abies amabilis |  |  | 4 | 64 | 13 | + | 14 | 1 |  |  | 83 | 100 | 57 | 25 | 50 | 25 |
| Chamaecyparis nootkatensis | 35 |  |  | + |  |  |  |  | 100 |  |  | 14 |  |  |  |  |
| Pinus contorta | 14 |  |  |  |  | 1 |  |  | 100 |  |  |  |  | 50 |  |  |
| Pinus monticola | 1 |  |  |  |  | 1 |  |  | 75 |  |  |  |  | 25 |  |  |
| Taxus brevifolia | 1 | 4 |  |  | 8 | 6 |  |  | 25 | 33 |  |  | 57 | 75 |  |  |
| Thuja plicata | 27 | 54 |  | 6 | 7 | 49 |  | 2 | 100 | 100 |  | 57 | 63 | 100 |  | 12 |
| Tsuga heterophylla | 21 | 42 | 96 | 30 | 72 | 42 | 86 | 85 | 100 | 100 | 100 | 85 | 100 | 100 | 100 | 87 |
| Seedlings (below BH) | Mean relative density (\%) |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |
| Abies amabilis |  |  | 5 | 11 | 5 |  |  | 3 |  |  | 83 | 100 | 774 |  |  | 25 |
| Chamaecyparis nootkatensis | 24 |  |  | 2 |  |  |  |  | 100 |  |  | 14 |  |  |  |  |
| Picea sitchensis |  |  |  |  | + |  | 15 | 23 |  |  |  |  | 15 |  | 100 | 62 |
| Pinus contorta | 12 |  |  |  |  |  |  |  | 100 |  |  |  |  |  |  |  |
| Pinus monticola | 1 |  |  |  |  | 1 |  |  | 75 |  |  |  |  | 25 |  |  |
| Pseudotsuga menziesii | 4 | 5 |  |  |  |  |  | 1 | 100 | 66 |  |  |  |  |  | 12 |
| Thuja plicata | 29 | 50 | 6 | 24 | 27 | 70 | 30 | 10 | 100 | 100 | 50 | 100 | 94 | 100 | 100 | 25 |
| Tsuga heterophylla | 28 | 41 | 89 | 63 | 67 | 29 | 55 | 49 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 |

Table 24 (continued)

| Community types | D2 | T1 | T2 | T3 | T4 | T5 | F2 | F1 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Species richness ${ }^{1}$ | $\begin{gathered} 6.5 \\ (1.0)^{2} \end{gathered}$ | $\begin{gathered} 4.7 \\ (0.5) \end{gathered}$ | $\begin{gathered} 3.0 \\ (0.6) \end{gathered}$ | $\begin{gathered} 3.8 \\ (0.7) \end{gathered}$ | $\begin{gathered} 3.8 \\ (0.7) \end{gathered}$ | $\begin{gathered} 4.2 \\ (0.5) \end{gathered}$ | $\begin{aligned} & 4.0 \\ & (-) \end{aligned}$ | $\begin{gathered} 3.9 \\ (1.8) \end{gathered}$ |
| Total species ${ }^{1}$ | 8 | 7 | 5 | 5 | 6 | 6 | 5 | 8 |
| Species diversity ${ }^{3}: \operatorname{Exp}_{1 / \lambda^{6}}\left(H^{\prime}\right)^{5}$ | $\begin{aligned} & 5.4 \\ & 5.1 \end{aligned}$ | $\begin{aligned} & 3.6 \\ & 2.8 \end{aligned}$ | $\begin{aligned} & 3.6 \\ & 3.1 \end{aligned}$ | $\begin{aligned} & 3.3 \\ & 2.8 \end{aligned}$ | $\begin{aligned} & 2.7 \\ & 2.4 \end{aligned}$ | $\begin{aligned} & 3.2 \\ & 2.6 \end{aligned}$ | $\begin{aligned} & 3.4 \\ & 3.1 \end{aligned}$ | $\begin{aligned} & 4.8 \\ & 4.1 \end{aligned}$ |
| Mean basal area ( $\left.\mathrm{m}^{2} / \mathrm{ha}\right)^{4}$ | $\begin{gathered} 30.2 \\ (10.2) \end{gathered}$ | $\begin{aligned} & 86.0 \\ & (9.5) \end{aligned}$ | $\begin{aligned} & 142.3 \\ & (47.4) \end{aligned}$ | $\begin{aligned} & 187.3 \\ & (81.5) \end{aligned}$ | $\begin{aligned} & 180.4 \\ & (54.8) \end{aligned}$ | $\begin{gathered} 87.7 \\ (21.6) \end{gathered}$ | $\begin{aligned} & 236.5 \\ & (14.8) \end{aligned}$ | $\begin{aligned} & 246.2 \\ & 124.9) \end{aligned}$ |
| Mean density (trees/ha) ${ }^{3}$ | $\begin{gathered} 695 \\ (213) \end{gathered}$ | $\begin{gathered} 740 \\ (250) \end{gathered}$ | $\begin{array}{r} 407 \\ (93) \end{array}$ | $\begin{gathered} 583 \\ (195) \end{gathered}$ | $\begin{gathered} 455 \\ (148) \end{gathered}$ | $\begin{gathered} 855 \\ (209) \end{gathered}$ | $\begin{array}{r} 330 \\ (19) \end{array}$ | $\begin{gathered} 315 \\ (124) \end{gathered}$ |
| Mean max. height (m) | $\begin{gathered} 17 \\ (10) \end{gathered}$ | $\begin{aligned} & 30 \\ & (4) \end{aligned}$ | $\begin{gathered} 52 \\ (7) \end{gathered}$ | $\begin{aligned} & 42 \\ & (9) \end{aligned}$ | $\begin{aligned} & 43 \\ & (6) \end{aligned}$ | 24 <br> (1) | $\begin{aligned} & 56 \\ & (-) \end{aligned}$ | 60 <br> (9) |

1 Includes tree, sapling and seedling strata
2 (standard deviation)
3 Includes tree stratum only
4 Includes trees and saplings
5 Antilog of Shannon's Index
${ }^{6}$ Reciprocal of Simpson's Index

Table 25 : Thuja group and community types D2, F1 et F2 understory strata summary table (see table 15 for community type codes).

| Community types | D2 | T1 | T2 | T3 | T4 | T5 | F2 | FI | D2 | T1 | T2 | T3 | T4 | T5 | F2 | F1 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Number of plots | 4 | 3 | 6 | 7 | 19 | 4 | 2 | 8 |  |  |  |  |  |  |  |  |


| Shrubs | Mean coverage (\%) |  |  |  |  |  |  |  |  |  | Constancy (\%) |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Gaultheria shallon | 45 | 64 | 9 | 45 | 57 | 75 | 33 | 1 | 100 | 100 | 83 | 100 | 100 | 100 | 100 | 37 |
| Menziesia ferruginea | 1 | 2 | $+$ | 4 | 2 | 5 | 5 | + | 50 | 100 | 33 | 85 | 94 | 100 | 100 | 12 |
| Pyrus fusca | $+$ |  |  |  | + | 8 |  | $+$ | 25 |  |  |  | 5 | 100 |  | 12 |
| Ribes bracteosum |  |  |  |  |  |  |  | 5 |  |  |  |  |  |  |  | 62 |
| Rubus spectabilis |  | + | + | $+$ | 2 | 2 | 21 | 23 |  | 33 | 83 | 57 | 78 | 75 | 100 | 100 |
| Vaccinium alaskaense | 2 | + | 5 | 15 | 11 | 6 | 7 | 1 | 50 | 66 | 83 | 100 | 100 | 100 | 100 | 75 |
| Vaccinium ovalifolium | + |  | + | 2 | 1 | + | 4 | 1 | 25 |  | 66 | 71 | 57 | 25 | 100 | 37 |
| Vaccinium ovatum | 25 | 50 | + |  | 1 | 34 | 2 |  | 75 | 100 | 16 |  | 57 | 100 | 100 |  |
| Vaccinium parvifolium | 6 | 7 | 13 | 14 | 13 | 13 | 13 | 5 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 |

## Herbs

| Achlys triphylla |  |  |  |  | + |  |  | 5 |  |  |  |  | 5 |  |  | 50 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Adenocaulon bicolor |  |  |  |  |  |  | 1 | + |  |  |  |  |  |  | 50 | 50 |
| Adiantum pedatum |  |  |  |  |  |  |  | + |  |  |  |  |  |  |  | 50 |
| Agrostis scabra | $+$ |  |  |  |  |  |  |  | 75 |  |  |  |  |  |  |  |
| Aruncus sylvester |  |  |  |  |  |  |  | + |  |  |  |  |  |  |  | 62 |
| Athyrium filix-femina |  |  | + |  | $+$ |  | 1 | 13 |  |  | 33 |  | 10 |  | 100 | 100 |
| Blechnum spicant | $+$ | 32 | 46 | 48 | 61 | 65 | 29 | 6 | 26 | 100 | 100 | 100 | 100 | 100 | 100 | 62 |
| Boschniakia hookeri | $+$ | + |  |  | + | + |  |  | 75 | 66 |  |  | 5 | 50 |  |  |
| Boykinia elata |  |  |  | + |  |  | + | $+$ |  |  |  | 28 |  |  | 100 | 25 |
| Bromus vulgaris |  |  |  |  |  |  |  | + |  |  |  |  |  |  |  | 50 |
| Calamagrostis nutkaensis |  |  |  |  |  | 1 |  |  |  |  |  |  |  | 50 |  |  |
| Carex obnupta |  |  |  |  | + | 5 | 3 | + |  |  |  |  | 10 | 75 | 100 | 12 |
| Coptis asplenifolia |  |  |  | 3 | + |  |  | + |  |  |  | 57 | 5 |  |  | 12 |

Table 25 (continued)

| Community types | D2 | T1 | T2 | T3 | T4 | T5 | F2 | F1 | D2 | T1 | T2 | T3 | T4 | T5 | F2 | F1 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Herbs (continued) |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |
| Danthonia spicata | + |  |  |  |  |  |  |  | 75 |  |  |  |  |  |  |  |
| Dryopteris austriaca |  |  | + |  |  |  |  | + |  |  | 50 |  |  |  |  | 50 |
| Galium triflorum |  |  |  |  | + |  | + | 1 |  |  |  |  | 5 |  | 50 | 87 |
| Goodyera oblongifolia | + |  |  | + | + |  |  | + | 75 |  |  | 28 | 15 |  |  | 12 |
| Linnaea borealis | 4 | 1 |  | 1 | + | 5 |  |  | 100 | 66 |  | 42 | 10 | 100 |  |  |
| Listera caurina |  |  | + | + | + |  |  |  |  |  | 33 | 71 | 5 |  |  |  |
| Listera cordata |  | + | + | + | + | + |  |  |  | 33 | 16 | 85 | 47 | 25 |  |  |
| Luzula parviflora |  |  | + |  |  |  |  | + |  |  | 16 |  |  |  |  | 75 |
| Lysichitum americanum |  |  |  |  | 2 | + | 26 | + |  |  |  |  | 10 | 25 | 100 | 12 |
| Maianthemum dilatatum | + | + |  | 2 | + | 2 | 1 | 3 | 50 | 33 |  | 71 | 57 | 100 | 100 | 100 |
| Melica subulata |  |  |  |  |  |  |  | 1 |  |  |  |  |  |  |  | 62 |
| Polystichum munitum |  |  | 12 | 1 | + |  | 23 | 45 |  |  | 100 | 42 | 26 |  | 100 | 100 |
| Rubus pedatus |  |  |  | 3 |  |  |  |  |  |  |  | 57 |  |  |  |  |
| Saxifraga ferruginea | + |  |  |  |  |  |  |  | 75 |  |  |  |  |  |  |  |
| Selaginella wallacei | $+$ |  |  |  |  |  |  |  | 50 |  |  |  |  |  |  |  |
| Streptopus amplexifolius |  |  | + | + | + | + | + | + |  |  | 33 | 85 | 10 | 25 | 100 | 75 |
| Tiarella laciniata |  |  | + | 1 | + |  | 2 | + |  |  | 66 | 85 | 21 |  | 100 | 62 |
| Tiarella trifoliata |  |  | + | 1 | $+$ |  | 5 | 9 |  |  | 66 | 85 | 26 |  | 100 | 100 |
| Trautvetteria caroliniensis |  |  |  |  |  |  |  | 10 |  |  |  |  |  |  |  | 100 |
| Trisetum cernuum |  |  |  |  |  |  | + | + |  |  |  |  |  |  | 100 | 25 |
| Trillium ovatum |  |  | + | + | + |  |  | 1 |  |  | 50 | 42 | 36 |  |  | 75 |
| Veratrum viride |  |  |  | + | + | + | + | + |  |  |  | 28 | 5 | 50 | 50 | 37 |
| Viola glabella |  |  |  | + |  |  | + | + |  |  |  | 14 |  |  | 100 | 50 |

Bryophytes \& lichens


Table 25 (continued)

| Community types | D2 | T1 | T2 | T3 | T4 | T5 | F2 | F1 | D2 | T1 | T2 | T3 | T4 | T5 | F2 | F1 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| $\frac{\text { Bryophytes \& lichens }}{\text { (continued) }}$ |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |
|  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |
| Campylopus atrovirens | + |  |  |  |  |  |  |  | 50 |  |  |  |  |  |  |  |
| Cephalozia bicuspidata |  | 1 | 2 | 2 | 2 | 2 | + | + |  | 33 | 66 | 71 | 57 | 75 | 50 | 12 |
| Cladina impexa | 3 |  |  |  |  |  |  |  | 100 |  |  |  |  |  |  |  |
| Cladina rangiferina | 9 |  |  |  |  |  |  |  | 100 |  |  |  |  |  |  |  |
| Cladonia bellidiflora | + |  |  |  |  |  |  |  | 100 |  |  |  |  |  |  |  |
| Cladonia multiformis | + |  |  |  |  |  |  |  | 50 |  |  |  |  |  |  |  |
| Cladonia gracilis | 1 |  |  |  |  |  |  |  | 100 |  |  |  |  |  |  |  |
| Cladonia uncialis | 1 |  |  |  |  |  |  |  | 100 |  |  |  |  |  |  |  |
| Dicranum fuscescens | + | + | + | + | + | + |  | + | 25 | 100 | 16 | 42 | 36 | 25 |  | 12 |
| Dicranum scoparium | 7 |  |  |  |  | + |  | + | 100 |  |  |  |  | 75 |  | 12 |
| Diplophyllum albicans | + | 1 |  | + | + | + |  | + | 25 | 33 |  | 14 | 26 | 75 |  | 12 |
| Diplophyllum plicatum |  | 1 |  |  | + | 1 |  |  |  | 33 |  |  | 10 | 50 |  |  |
| Hebertus aduncus | + |  |  |  | + | 1 |  |  | 75 |  |  |  | 15 | 75 |  |  |
| Heterocladium macounii |  | + | + |  |  |  |  | + |  | 66 | 33 |  |  |  |  | 25 |
| Hookeria lucens |  | + | + | + | + | + | 1 | + |  | 100 | 66 | 100 | 84 | 100 | 100 | 62 |
| Hylocomium splendens | 7 | 14 | + | 4 | 7 | 8 | 2 | 2 | 100 | 100 | 16 | 85 | 100 | 100 | 100 | 87 |
| Hypnum circinale |  | + | 1 | + | + | + |  | + |  | 66 | 83 | 85 | 26 | 25 |  | 37 |
| Isopterygium elegans |  | + | 1 | + | + |  | + | 1 |  | 33 | 100 | 57 | 10 |  | 50 | 50 |
| Isothecium stoloniferum | + | 5 | 3 | 3 | 1 |  | 1 | 1 | 25 | 100 | 83 | 100 | 89 |  | 50 | 75 |
| Leucolepis menziesii |  |  |  |  | + |  | 10 | 6 |  |  |  |  | 10 |  | 100 | 75 |
| Mylia taylorii | + |  | + | + | + | + |  |  | 25 |  | 16 | 14 | 5 | 50 |  |  |
| Pellia neesiana |  |  | + | + | + |  | 3 | + |  |  | 33 | 57 | 15 |  | 100 | 50 |
| Plagiochila porelloides |  | + | + | + | + | + | 3 | + |  | 33 | 33 | 42 | 63 | 75 | 100 | 62 |
| Plagiomnium insigne |  |  |  |  |  |  | + | 2 |  |  |  |  |  |  | 100 | 75 |
| Plagiothecium undulatum | + | 2 | 6 | 4 | 4 | 4 | 4 | , | 100 | 100 | 83 | 100 | 100 | 100 | 100 | 62 |
| Pleurozium schreberi | 1 |  |  |  |  |  |  |  | 75 |  |  |  |  |  |  |  |
| Polytrichum commune | + |  |  |  |  |  |  |  | 75 |  |  |  |  |  |  |  |
| Polytrichum juniperinum | + |  |  |  |  |  |  |  | 50 |  |  |  |  |  |  |  |
| Polytrichum piliferum | + |  |  |  |  |  |  |  | 50 |  |  |  |  |  |  |  |

Table 25 (continued)

| Community types | D2 | T1 | T2 | T3 | T4 | T5 | F2 | F1 | D2 | T1 | T2 | T3 | T4 | T5 | F2 | F1 |  |
| :--- | ---: | ---: | ---: | ---: | ---: | ---: | ---: | ---: | ---: | ---: | ---: | ---: | ---: | ---: | ---: | ---: | ---: |
| Bryophytes \& lichens |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |
| (continued) |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |


| Table 25 (continued) |  |  |  |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Community types | D2 | T1 | T2 | T3 | T4 | T5 | F2 | F1 |
| Strata mean coverage (\%) |  |  |  |  |  |  |  |  |
| Shrubs -- | $\begin{gathered} 79 \\ (30)^{1} \end{gathered}$ | $\begin{aligned} & 125 \\ & (31) \end{aligned}$ | $\begin{gathered} 29 \\ (28) \end{gathered}$ | $\begin{gathered} 80 \\ (35) \end{gathered}$ | $\begin{gathered} 87 \\ (30) \end{gathered}$ | $\begin{aligned} & 143 \\ & (9) \end{aligned}$ | $\begin{gathered} 86 \\ (25) \end{gathered}$ | $\begin{gathered} 46 \\ (44) \end{gathered}$ |
| Herbs | $\begin{gathered} 9 \\ (7) \end{gathered}$ | $\begin{gathered} 34 \\ (32) \end{gathered}$ | $\begin{gathered} 60 \\ (27) \end{gathered}$ | $\begin{gathered} 66 \\ (19) \end{gathered}$ | $\begin{gathered} 65 \\ (14) \end{gathered}$ | $\begin{gathered} 88 \\ (14) \end{gathered}$ | $\begin{gathered} 94 \\ (28) \end{gathered}$ | $\begin{aligned} & 119 \\ & (34) \end{aligned}$ |
| Bryo. \& 1ichens | $\begin{gathered} 62 \\ (22) \end{gathered}$ | $\begin{gathered} 63 \\ (26) \end{gathered}$ | 27 <br> (9) | $\begin{gathered} 42 \\ (20) \end{gathered}$ | $\begin{gathered} 47 \\ (20) \end{gathered}$ | $\begin{gathered} 58 \\ (21) \end{gathered}$ | $\begin{aligned} & 62 \\ & (-) \end{aligned}$ | $\begin{gathered} 39 \\ (21) \end{gathered}$ |
| Total understory Species richness (.05 ha) | $\begin{aligned} & 150 \\ & (47) \end{aligned}$ | $\begin{aligned} & 221 \\ & (45) \end{aligned}$ | $\begin{aligned} & 116 \\ & (23) \end{aligned}$ | $\begin{aligned} & 188 \\ & (45) \end{aligned}$ | $\begin{aligned} & 199 \\ & (42) \end{aligned}$ | $\begin{aligned} & 289 \\ & (15) \end{aligned}$ | $242$ <br> (3) | $\begin{aligned} & 204 \\ & (52) \end{aligned}$ |
| Shrubs | $\begin{gathered} 5.3 \\ (0.9) \end{gathered}$ | $\begin{gathered} 5.0 \\ (1.0) \end{gathered}$ | $\begin{gathered} 4.7 \\ (1.4) \end{gathered}$ | $\begin{gathered} 5.3 \\ (1.1) \end{gathered}$ | $\begin{gathered} 6.1 \\ (1.3) \end{gathered}$ | $\begin{gathered} 7.3 \\ (0.5) \end{gathered}$ | $\begin{aligned} & 7.0 \\ & (-) \end{aligned}$ | $\begin{gathered} 5.9 \\ (1.8) \end{gathered}$ |
| Herbs | $\begin{gathered} 9.0 \\ (1.8) \end{gathered}$ | $\begin{gathered} 3.7 \\ (0.6) \end{gathered}$ | $\begin{gathered} 6.2 \\ (3.0) \end{gathered}$ | $\begin{aligned} & 11.3 \\ & (4.5) \end{aligned}$ | $\begin{gathered} 4.8 \\ (3.1) \end{gathered}$ | $\begin{gathered} 7.5 \\ (1.9) \end{gathered}$ | $\begin{aligned} & 14.5 \\ & (3.5) \end{aligned}$ | $\begin{aligned} & 20.1 \\ & (5.2) \end{aligned}$ |
| Bryo. \& Iichens | $\begin{aligned} & 20.5 \\ & (1.3) \end{aligned}$ | $\begin{aligned} & 13.0 \\ & (1.0) \end{aligned}$ | $\begin{aligned} & 11.7 \\ & (2.0) \end{aligned}$ | $\begin{aligned} & 13.4 \\ & (1.6) \end{aligned}$ | $\begin{aligned} & 12.4 \\ & (1.9) \end{aligned}$ | $\begin{aligned} & 13.5 \\ & (1.3) \end{aligned}$ | $\begin{aligned} & 16.0 \\ & (2.8) \end{aligned}$ | $\begin{aligned} & 12.1 \\ & (2.0) \end{aligned}$ |
| Total species : Shrubs | 11 | 6 | 7 | 7 | 10 | 9 | 7 | 16 |
| Herbs | 20 | 7 | 14 | 24 | 24 | 13 | 17 | 49 |
| Bryo. \& lichens | 32 | 20 | 22 | 25 | 36 | 22 | 20 | 26 |
| Species diversity |  |  |  |  |  |  |  |  |
| $\text { Shrubs \& herbs : Exp }\left(H^{\prime}\right)^{2}$ | $5.3$ | $4.6$ | $5.5$ | $9.6$ | $5.4$ | $8.8$ | $13.3$ | $19.7$ |
| $1 / \lambda^{3}$ | $6.3$ | $3.8$ | $3.8$ | $6.4$ | $3.7$ | $6.2$ | $10.6$ | $11.7$ |
| $\text { Bryo. \& lichens : } \operatorname{Exp}_{1 / \lambda^{3}}\left(H^{\prime}\right)^{2}$ | $\begin{aligned} & 18.7 \\ & 14.6 \end{aligned}$ | 10.1 7.6 | 11.6 8.8 | 11.5 9.2 | 12.7 9.3 | $\begin{array}{r} 12.8 \\ 9.7 \end{array}$ | $\begin{array}{r} 12.1 \\ 9.4 \end{array}$ | $\begin{aligned} & 14.5 \\ & 11.4 \end{aligned}$ |

Table 26 : Abies group tree strata summary table (see table 15 for community type codes).

| Community types | A1 | A2 | A3 | A4 | A5 | A6 | A7 | AI | A2 | A3 | A4 | A5 | A6 | A7 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Number of plots | 3 | 2 | 4 | 4 | 12 | 2 | 5 |  |  |  |  |  |  |  |
| Tree (> 10 cm DBH) | Mean | rela | tive | imp | orta | nce | value (\%) |  | Constancy (\%) |  |  |  |  |  |
| Abies amabilis | 12 | 25 | 45 | 65 | 74 | 25 | 4 | 66 | 100 | 100 | 100 | 100 | 100 | 40 |
| Chamaecyparis nootkatensis | 2 |  |  |  |  |  |  | 66 |  |  |  |  |  |  |
| Picea sitchensis |  |  |  |  |  |  | 9 |  |  |  |  |  |  | 20 |
| Pseudotsuga menziesii | 25 | 7 |  |  |  |  | 17 | 100 | 50 |  |  |  |  | 60 |
| Thuja plicata | 8 | 22 |  |  | 3 | 11 | 13 | 60 | 100 |  |  | 16 | 50 | 40 |
| Tsuga heterophylla | 52 | 45 | 55 | 35 | 23 | 64 | 57 | 100 | 100 | 100 | 100 | 91 | 100 | 100 |
| Saplings (0-10 cm DBH) | Mean relative density (\%) |  |  |  |  |  |  |  |  |  |  |  |  |  |
| Abies amabilis | 44 | 39 | 27 | 80 | 46 | 1 | 1 | 100 | 100 | 100 | 100 | 100 | 50 | 20 |
| Thuja plicata | 3 | 2 |  |  | + |  | 1 | 33 | 50 |  |  | 8 |  | 20 |
| Tsuga heterophylla | 48 | 58 | 73 | 19 | 54 | 99 | 98 | 100 | 100 | 100 | 50 | 100 | 100 | 100 |
| Seedlings (below BH) | Mean relative density (\%) |  |  |  |  |  |  |  |  |  |  |  |  |  |
| Abies amabilis | 41 | 30 | 12 | 48 | 27 | 5 | 3 | 100 | 100 | 100 | 100 | 100 | 100 | . 40 |
| Thuja plicata | 3 | 8 | 1 | 3 | 6 |  | 2 | 66 | 50 | 50 | 50 | 33 |  | 40 |
| Tsuga heterophylla | 54 | 62 | 87 | 49 | 67 | 95 | 94 | 66 | 100 | 100 | 100 | 100 | 100 | 100 |

Table 26 (continued)

| Community types | A1 | A2 | A3 | A4 | A5 | A6 | A7 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Species richness ${ }^{1}$ | $\begin{gathered} 5.0 \\ (1.0)^{2} \end{gathered}$ | $\begin{gathered} 3.0 \\ (1.4) \end{gathered}$ | $\begin{gathered} 2.7 \\ (0.9) \end{gathered}$ | $\begin{gathered} 3.5 \\ (1.9) \end{gathered}$ | $\begin{gathered} 2.5 \\ (0.8) \end{gathered}$ | $\begin{gathered} 2.5 \\ (0.7) \end{gathered}$ | $\begin{gathered} 2.8 \\ (0.4) \end{gathered}$ |
| Total species ${ }^{1}$ | 6 | 4 | 4 | 4 | 5 | 3 | 5 |
| $\text { Species diversity }{ }^{3}: \operatorname{Exp}_{1 / \lambda^{6}}\left(H^{\prime}\right)^{5}$ | $\begin{aligned} & 3.4 \\ & 2.8 \end{aligned}$ | $\begin{aligned} & 3.4 \\ & 3.1 \end{aligned}$ | $\begin{aligned} & 2.0 \\ & 2.0 \end{aligned}$ | $\begin{aligned} & 2.0 \\ & 1.9 \end{aligned}$ | $\begin{aligned} & 1.9 \\ & 1.7 \end{aligned}$ | $\begin{aligned} & 2.4 \\ & 2.1 \end{aligned}$ | $\begin{aligned} & 3.4 \\ & 2.6 \end{aligned}$ |
| Mean basal area ( $\left.\mathrm{m}^{2} / \mathrm{ha}\right)^{4}$ | $\begin{aligned} & 121.0 \\ & (44.5) \end{aligned}$ | $\begin{aligned} & 146.5 \\ & (47.4) \end{aligned}$ | $\begin{gathered} 79.5 \\ (11.9) \end{gathered}$ | $\begin{aligned} & 121.9 \\ & (29.6) \end{aligned}$ | $\begin{aligned} & 107.7 \\ & (20.4) \end{aligned}$ | $\begin{aligned} & 116.5 \\ & (43.1) \end{aligned}$ | $\begin{aligned} & 185.2 \\ & (37.4) \end{aligned}$ |
| Mean density (trees/ha) ${ }^{3}$ | $\begin{gathered} 700 \\ (295) \end{gathered}$ | $\begin{aligned} & 280 \\ & (28) \end{aligned}$ | $\begin{gathered} 435 \\ (153) \end{gathered}$ | $\begin{gathered} 420 \\ (140) \end{gathered}$ | $\begin{gathered} 517 \\ (149) \end{gathered}$ | $\begin{aligned} & 420 \\ & (56) \end{aligned}$ | $\begin{gathered} 348 \\ (114) \end{gathered}$ |
| Mean max. height (m) | $\begin{gathered} 44 \\ (12) \end{gathered}$ | $\begin{aligned} & 50 \\ & (-) \end{aligned}$ | $\begin{aligned} & 53 \\ & (6) \end{aligned}$ | $\begin{aligned} & 53 \\ & (3) \end{aligned}$ | 54 <br> (6) | 46 <br> (9) | 64 <br> (4) |

1 Includes tree, sapling and seedling strata
2 (standard deviation)
3 Includes tree stratum only
4 Includes trees and saplings
5 Antilog of Shannon's Index
${ }^{6}$ Reciprocal of Simpson's Index

Table 27 : Abies group understory strata summary table (see table 15 for community type codes).

| Community types | A1 | A2 | A3 | A4 | A5 | A6 | A7 | A1 | A2. A3 | A4 | A5 | A6 | A7 |
| :--- | ---: | ---: | ---: | ---: | ---: | ---: | ---: | ---: | ---: | ---: | ---: | ---: | ---: | ---: |
| Number of plots | 3 | 2 | 4 | 4 | 12 | 2 | 5 |  |  |  |  |  |  |


| Shrubs | Mean coverage (\%) |  |  |  |  |  | + | Constancy (\%) |  |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Gaultheria shallon | 34 | 2 | $+$ |  | 1 | 43 |  | 100 | 50 | 50 |  | 41 | 100 | 40 |
| Menziesia ferruginea | + | + | + |  | + | 2 |  | 66 | 50 | 25 |  | 41 | 50 |  |
| Oplopanax horridus |  |  |  | 1 | + |  |  |  |  |  | 75 | 16 |  |  |
| Rubus spectabilis |  | $+$ | 2 | + | 1 |  | + |  | 50 | 100 | 25 | 91 |  | 80 |
| Sorbus sitchensis | $+$ |  |  |  |  |  |  | 66 |  |  |  |  |  |  |
| Vaccinium alaskaense | 41 | 43 | 2 | 31 | 25 | 6 | 5 | 100 | 100 | 100 | 100 | 100 | 100 | 100 |
| Vaccinium ovalifolium | 1 | 1 |  | + | 3 |  |  | 66 | 50 |  | 25 | 66 |  |  |
| Vaccinium parvifolium | 27 | 14 | 22 | 4 | 10 | 28 | 9 | 100 | 100 | 100 | 100 | 100 | 100 | 100 |

Herbs

| Achlys triphylla | $+$ | 4 | $+$ | 4 | 3 | + | $+$ | 33 | 50 | 25 | 100 | 50 | 50 | 40 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Adiantum pedatum |  |  |  | 1 | $+$ |  | + |  |  |  | 50 | 16 |  | 20 |
| Athyrium filix-femina |  | + |  | 3 | + | + | $+$ |  | 50 |  | 75 | 50 | 50 | 60 |
| Blechnum spicant | 1 | 16 | 11 | 7 | 17 | 15 | 25 | 33 | 100 | 75 | 100 | 100 | 100 | 100 |
| Clintonia uniflora |  |  |  | 1 | + |  |  |  |  |  | 50 | 16 |  |  |
| Cornus canadensis | 1 | $+$ |  | + | + |  |  | 33 | 50 |  | 50 | 16 |  |  |
| Dryopteris austriaca |  |  | + | 1 | 2 |  | + |  |  | 25 | 50 | 66 |  | 60 |
| Gymnocarpium dryopteris |  |  |  | 1 | + |  |  |  |  |  | 75 | 25 |  |  |
| Hypopitys monotropa | + |  | + | + | + |  |  | 33 |  | 50 | 25 | 25 |  |  |
| Listera caurina |  | $+$ | $+$ | + | + |  |  |  | 100 | 50 | 75 | 16 |  |  |
| Listera cordata | $+$ | + |  |  |  | $+$ | $+$ | 66 | 50 |  |  |  | 50 | 20 |
| Maianthemum dilatatum | + | $+$ | + | 1 | $+$ |  | + | 33 | 50 | 25 | 50 | 41 |  | 20 |
| Polystichum munitum | + | 4 | 2 | 2 | 3 | $+$ | 29 | 33 | 50 | 100 | 75 | 83 | 100 | 100 |
| Rubus pedatus |  | 1 |  | 31 | 3 |  |  |  | 50 |  | 75 | 50 |  |  |
| Streptopus amplexifolius |  | + | + |  | + |  | + |  | 50 | 75 |  | 41 |  | 60 |

Table 27 (continued)

| Community types | A1 | A2 | A3 | A4 | A5 | A6 | A7 | A1 | A2 | A3 | A4 | A5 | A6 | A7 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Herbs (continued) |  |  |  |  |  |  |  |  |  |  |  |  |  |  |
| Streptopus roseus | + | $+$ |  | 5 | + |  | + | 33 | 50 |  | 100 | 25 |  | 20 |
| Streptopus streptopoides |  |  |  | 8 | $+$ |  |  |  |  |  | 100 | 8 |  |  |
| Tiarella laciniata |  | $+$ | + | + | $+$ | $+$ | + |  | 50 | 50 | 75 | 50 | 50 | 60 |
| Tiarella trifoliata |  | 3 | $+$ | 3 | 4 | + | $+$ |  | 50 | 25 | 75 | 91 | 50 | 80 |
| Trautvetteria caroliniensis |  |  |  | 1 | + | + |  |  |  |  | 50 | 33 | 50 |  |
| Trillium ovatum | + | 1 | + | + | $+$ | $+$ | + | 33 | 50 | 75 | 100 | 58 | 50 | 60 |
| Bryophytes \& 1ichens |  |  |  |  |  |  |  |  |  |  |  |  |  |  |
| Cephalozia bicuspidata |  |  | 2 | 1 | 3 |  | 1 |  |  | 50 | 25 | 83 |  | 80 |
| Dicranum fuscescens | 1 | 1 | + | + | + |  | $+$ | 100 | 100 | 50 | 100 | 33 |  | 20 |
| Diplophyllum albicans |  |  | 2 |  | 1 |  |  |  |  | 50 |  | 16 |  |  |
| Eurhynchium pulchellum |  |  |  | + |  |  |  |  |  |  | 50 |  |  |  |
| Heterocladium macounii |  |  | 2 |  | + |  | + |  |  | 25 |  | 8 |  | 80 |
| Hookeria lucens |  | + | + | + | 1 | + | + |  | 50 | 75 | 50 | 75 | 50 | 80 |
| Hylocomium splendens | 3 | + | 1 | + | 3 | 7 | $+$ | 100 | 50 | 25 | 25 | 83 | 50 | 20 |
| Hypnum circinale | 2 | 3 | 2 | 3 | + | + | 1 | 66 | 100 | 75 | 100 | 25 | 100 | 80 |
| Isopterygium elegans |  | + | 2 | + | 1 | + | 1 |  | 50 | 100 | 25 | 66 | 50 | 40 |
| Isothecium stoloniferum | 2 | 1 | 2 | 1 | 2 | 2 | 4 | 66 | 100 | 75 | 100 | 83 | 100 | 100 |
| Lepidozia reptans | + | + | + |  | + | 3 |  | 33 | 50 | 75 |  | 8 | 50 |  |
| Plagiochila porelloides |  |  |  | + | $+$ | $+$ | $+$ |  |  |  | 50 | 58 | 100 | 20 |
| Plagiothecium undulatum | + | 4 | 5 | 8 | 8 | 11 | 4 | 33 | 100 | 100 | 100 | 100 | 100 | 80 |
| Rhizomnium glabrescens |  | 1 | 4 | 1 | 7 | 6 | 2 |  | 50 | 75 | 75 | 100 | 100 | 80 |
| Rhytidiadelphus loreus | 17 | 6 | 3 | 11 | 17 | 1 | 2 | 100 | 100 | 50 | 75 | 100 | 100 | 80 |
| Rhytidiopsis robusta | 3 | 3 |  | 1 |  |  |  | 66 | 100 |  | 100 |  |  |  |
| Scapania bolanderi | 3 | 4 | 6 | 2 | 2 | 7 | 4 | 100 | 100 | 75 | 100 | 91 | 100 | 100 |
| Stokesiella oregana | 5 | + | $+$ | + | 7 | 24 | 2 | 66 | 50 | 25 | 25 | 75 | 100 | 100 |
| Rock |  |  | 1 | 3 | $+$ |  | 5 |  |  | 50 | 25 | 33 |  | 60 |

Table 27 (continued)

| Community types | A1 | A2 | A3 | A4 | A5 | A6 | A7 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Strata mean coverage (\%) |  |  |  |  |  |  |  |
| Shrubs | $\begin{aligned} & 104 \\ & (34)^{1} \end{aligned}$ | $\begin{gathered} 61 \\ (33) \end{gathered}$ | $\begin{gathered} 25 \\ (25) \end{gathered}$ | $\begin{gathered} 36 \\ (30) \end{gathered}$ | $\begin{gathered} 40 \\ (21) \end{gathered}$ | $\begin{gathered} 80 \\ (29) \end{gathered}$ | $\begin{array}{r} 15 \\ (8) \end{array}$ |
| Herbs | $\begin{gathered} 3 \\ (1) \end{gathered}$ | $\begin{gathered} 31 \\ (42) \end{gathered}$ | $\begin{array}{r} 14 \\ (9) \end{array}$ | $\begin{gathered} 70 \\ (60) \end{gathered}$ | $\begin{gathered} 35 \\ (31) \end{gathered}$ | $\begin{gathered} 16 \\ (20) \end{gathered}$ | $\begin{gathered} 56 \\ (27) \end{gathered}$ |
| Bryo. \& lichens | $\begin{gathered} 39 \\ (16) \end{gathered}$ | $\begin{gathered} 25 \\ (12) \end{gathered}$ | $\begin{gathered} 33 \\ (18) \end{gathered}$ | $\begin{gathered} 30 \\ (20) \end{gathered}$ | $\begin{gathered} 54 \\ (17) \end{gathered}$ | $\begin{gathered} 61 \\ (11) \end{gathered}$ | $\begin{array}{r} 22 \\ (7) \end{array}$ |
| Total understory | $\begin{aligned} & 147 \\ & (44) \end{aligned}$ | $\begin{aligned} & 116 \\ & (88) \end{aligned}$ | $\begin{gathered} 72 \\ (18) \end{gathered}$ | $\begin{aligned} & 137 \\ & (73) \end{aligned}$ | $\begin{aligned} & 129 \\ & (48) \end{aligned}$ | $\begin{aligned} & 158 \\ & (20) \end{aligned}$ | $\begin{gathered} 93 \\ (22) \end{gathered}$ |
| Species richness (.05 ha) (20) (18) (20) |  |  |  |  |  |  |  |
| Shrubs | $\begin{gathered} 5.3 \\ (1.1) \end{gathered}$ | $\begin{gathered} 4.0 \\ (2.8) \end{gathered}$ | $\begin{gathered} 3.8 \\ (0.5) \end{gathered}$ | $\begin{gathered} 3.3 \\ (1.0) \end{gathered}$ | $\begin{gathered} 4.8 \\ (1.0) \end{gathered}$ | $\begin{gathered} 3.5 \\ (0.7) \end{gathered}$ | $\begin{gathered} 3.4 \\ (0.9) \end{gathered}$ |
| Herbs | $\begin{gathered} 5.0 \\ (2.6) \end{gathered}$ | $\begin{aligned} & 11.0 \\ & (2.7) \end{aligned}$ | $\begin{gathered} 7.0 \\ (5.4) \end{gathered}$ | $\begin{aligned} & 15.0 \\ & (5.1) \end{aligned}$ | $\begin{aligned} & 10.9 \\ & (7.1) \end{aligned}$ | $\begin{gathered} 6.0 \\ (5.6) \end{gathered}$ | $\begin{gathered} 7.8 \\ (2.5) \end{gathered}$ |
| Bryo. \& lichens | $\begin{gathered} 7.3 \\ (2.1) \end{gathered}$ | $\begin{aligned} & 10.5 \\ & (4.9) \end{aligned}$ | $\begin{aligned} & 11.3 \\ & (2.9) \end{aligned}$ | $\begin{aligned} & 11.0 \\ & (2.9) \end{aligned}$ | $\begin{aligned} & 12.7 \\ & (2.5) \end{aligned}$ | $\begin{array}{r} 11.0 \\ (-) \end{array}$ | $\begin{aligned} & 11.4 \\ & (3.6) \end{aligned}$ |
| Total species : Shrubs <br> Herbs <br> Bryo. \& lichens | $\begin{array}{r} 7 \\ 14 \\ 10 \end{array}$ | $\begin{array}{r} 6 \\ 20 \\ 14 \end{array}$ | $\begin{array}{r} 5 \\ 16 \\ 23 \end{array}$ | $\begin{array}{r} 5 \\ 25 \\ 19 \end{array}$ | $\begin{array}{r} 9 \\ 41 \\ 30 \end{array}$ | $\begin{array}{r} 4 \\ 10 \\ 14 \end{array}$ | $\begin{array}{r} 5 \\ 17 \\ 21 \end{array}$ |
| Species diversity |  |  |  |  |  |  |  |
| $\text { Shrubs \& herbs : } \operatorname{Exp}_{1 / \lambda^{3}}\left(H^{\prime}\right)^{2}$ | $\begin{aligned} & 4.9 \\ & 3.8 \end{aligned}$ | $\begin{aligned} & 6.3 \\ & 3.7 \end{aligned}$ | $\begin{aligned} & 4.8 \\ & 3.4 \end{aligned}$ | 9.8 8.0 | $\begin{aligned} & 9.8 \\ & 6.4 \end{aligned}$ | $\begin{aligned} & 4.5 \\ & 3.6 \end{aligned}$ | $\begin{aligned} & 5.5 \\ & 4.7 \end{aligned}$ |
| $\text { Bryo. \& lichens : } \operatorname{Exp}_{1 / \lambda^{3}}\left(H^{\prime}\right)^{2}$ | $\begin{aligned} & 8.5 \\ & 7.4 \end{aligned}$ | 8.1 6.6 | 13.0 10.4 | 9.7 7.7 | 12.2 8.8 | $\begin{aligned} & 7.6 \\ & 6.1 \end{aligned}$ | $\begin{array}{r} 11.9 \\ 9.4 \end{array}$ |

1 (standard deviation) 2 Antilog of Shannon's Index $\quad 3$ Reciprocal of Simpson's Index

Table 28 : Subalpine vegetation group tree strata summary table.

|  | Mean relative importance value (\%) | Constancy <br> (11 plots) (\%) |
| :---: | :---: | :---: |
| Trees ( $>10 \mathrm{~cm} \mathrm{DBH}$ ) |  |  |
| Abies amabilis | 14 | 82 |
| Chamaecyparis nootkatensis | 7 | 36 |
| Pseudotsuga menziesii | 10 | 64 |
| Thuja plicata | 8 | 36 |
| Tsuga heterophylla | 51 | 91 |
| Tsuga mertensiana | 10 | 45 |
| Saplings (0-10 cm DBH) | Mean rel. density (\%) |  |
| Abies amabilis | 78 | 100 |
| Chamaecyparis nootkatensis | 3 | 27 |
| Pseudotsuga menziesii | + | 9 |
| Thuja plicata | 3 | 45 |
| Tsuga heterophylla | 13 | 82 |
| Tsuga mertensiana | 3 | 45 |
| Seedlings (below BH) | Mean rel. density (\%) |  |
| Abies amabilis | 46 | 100 |
| Chamaecyparis nootkatensis | 3 | 45 |
| Pseudotsuga menziesii | 1 | 54 |
| Thuja plicata | 3 | 36 |
| Tsuga heterophylla | 42 | 91 |
| Tsuga mertensiana | 5 | 18 |

Mean basal area ( $\mathrm{m}^{2} / \mathrm{ha}$ ) : 113.2 (26.3)
Mean density (trees/ha) : 520 (243)

Table 29 : Classification of community type soils to the subgroup level (see table 15 for community type codes).

Soil subgroups (C.S.S.C., 1978)

| Community <br> types | ODB |
| :--- | :--- | :--- | :--- | :--- | :--- | :--- | :--- | :--- | :--- | :--- |
| EDB |  |$\quad$ GDB | OHFP |
| :--- |


| $\mathrm{SA}^{1}$ | 1 | 7 | 1 | 1 | 1 |  |
| :--- | :--- | :--- | :--- | :--- | :--- | :--- |
| D1 |  |  |  |  |  | 3 |

D2 1

F1 1
61
F2
$\begin{array}{lll}\text { P1 } & 3\end{array}$
$\begin{array}{lll}\mathrm{P} 2 & 1 & 3\end{array}$
P3 5
$\begin{array}{ll}\mathrm{P} 4 & 2\end{array}$
$\begin{array}{lllll}\text { P5 } & 2 & 12 & 1 & 2\end{array}$
$\begin{array}{lll}\text { P6 } & 1 & 10\end{array}$
$\begin{array}{lll}\text { P7 } & 1\end{array}$
T1 1
$\begin{array}{llll}\mathrm{T} 2 & 1 & 4\end{array}$
$\begin{array}{llllll}\text { T3 } & 1 & 1 & 1 & 2 & 2 \\ \text { T4 } & 3 & 2 & 3 & 4 & 7\end{array}$
T5
A1 3
A2 2
A3 4

| A4 |  |  | 1 | 1 | 1 | 1 | 1 |
| :--- | :--- | :--- | :--- | :--- | :--- | :--- | :--- |
| A5 | 1 | 1 | 6 | 1 | 2 |  |  |
| A6 |  |  |  | 2 |  |  |  |
| A7 |  |  | 3 | 1 | 1 |  |  |


| DB : Dystric Brunisol | SB : Sombric Brunisol | E : Eluviated |
| ---: | ---: | ---: |
| HFP: : Humo-Ferric Podzol | CR : Cumulic Regosol | G : Gleyed |
| FHP : Ferro-Humic Podzol | OR : Orthic Regosol | OT : Ortstein |
| HG : Humic Gleysol | 0 : Orthic | DU : Duric |

1 SA : Subalpine vegetation group
2 Typic Folisols

TTable 30 : Mean species richness of community types (for .05 ha ).

| Community types |  | Mean species richness |  |
| :---: | :---: | :---: | :---: |
|  |  | vascular | total ${ }^{2}$ |
|  | Dry Pinus-Pseudotsuga forests | 30.3 (8.6) $^{1}$ | 49.0 (6.0) |
|  | Coastal dry Pinus forests | 20.7 (1.2) | 41.2 (0.9) |
| F1 | Floodplain forests | 29.9 (7.8) | 42.0 (6.9) |
|  | Floodplain forests (Lysichitum variant) | 25.5 (3.5) | 41.5 (0.7) |
| P1 | Dry Pseudotsuga forests | 32.2 (3.9) | 46.0 (2.1) |
| P2 | Pseudotsuga-Thuja-Acer forests | 31.0 (4.2) | 39.7 (6.2) |
| P3 | Pseudotsuga-Linnaea forests | 28.0 (7.7) | 39.0 (8.5) |
| P4 | Pseudotsuga-Berberis forests | 24.4 (4.9) | 35.2 (6.7) |
| P5 | Tsuga-Pseudotsuga-Polystichum forests | 19.8 (7.6) | 30.8 (9.1) |
|  | Montane Tsuga forests | 18.1 (6.6) | 30.1 (7.2) |
| P7 | Montane Tsuga-Gaultheria forests | 14.3 (7.6) | 24.1 (8.2) |
|  | Coastal dry Thuja forests | 13.3 (1.5) | 26.3 (2.3) |
|  | Coastal Tsuga-B1echnum-Polystichum forests | 13.8 (3.8) | 25.5 (5.1) |
|  | Coastal montane Thuja forests | 20.4 (5.3) | 33.8 (5.0) |
|  | Coastal Thuja forests | 14.6 (4.4) | 27.0 (5.7) |
|  | Coastal wet Thuja forests | 19.0 (2.6) | 32.5 (2.4) |
|  | Montane Tsuga-Abies-Gaultheria forests | 15.3 (2.9) | 22.7 (2.1) |
|  | Montane Abies-Tsuga forests | 18.0 (17.0) | 28.5 (21.9) |
|  | Montane Tsuga-Abies forests | 13.5 (5.8) | 24.7 (8.3) |
|  | Montane Abies-Streptopus forests | 21.7 (5.4) | 32.7 (7.9) |
|  | Lowland Abies forests | 18.2 (7.6) | 30.8 (9.3) |
| A6 | Tsuga-Gaultheria-Blechnum forests | 12.0 (5.6) | 23.0 (5.6) |
|  | Tsuga-Blechnum-Polystichum forests | 14.0 (2.5) | 25.4 (9.3) |
|  | (standard deviation) |  |  |
|  | also includes bryophytes and lichens |  |  |

Table 31 : Homogeneity and richness of vegetation strata within community types compared with a fire disturbance index (see table 15 for community type codes).


[^4]Table 32 : Tree seedling abundance on undecomposed wood and forest floor substrata within community types (see table 15 for community type codes).

| Community types |  | Mean seedling density/m² (S.D.) |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: |
| Substrata types |  | Tsuga heterophyl1a | Thuja plicata | Abies amabilis | Pseudotsuga menziesii |
| $\begin{gathered} \mathrm{P} 4 \\ (5)^{1} \end{gathered}$ | wood (33) ${ }^{2}$ | 2.82 (6.13) | 0.09 (0.38) |  | 0.06 (0.24) |
|  | floor (67) | 1.10 (4.93) | 0.10 (0.53) |  | 0.10 (0.47) |
|  | $z^{3}$ | 1.40 | 0.15 |  | 0.62 |
|  | sig. ${ }^{4}$ | n.s. | n.s. |  | n.s. |
| $\begin{gathered} \text { P5 } \\ (13) \end{gathered}$ | wood (111) | 3.22 (7.31) | 0.37 (1.57) |  |  |
|  | floor (149) | 0.70 (2.57) | 0.09 (0.51) |  |  |
|  | $z$ | 3.47 | 1.82 |  |  |
|  | sig. | *** | * |  |  |
| $\begin{gathered} \text { P6 } \\ (10) \end{gathered}$ | wood (86) | 17.74 (27.25) | 0.80 (1.99) |  | 0.13 (0.50) |
|  | floor (114) | 6.36 (11.60) | 0.59 (1.35) |  | 0.14 (0.55) |
|  | - | 3.63 | 0.85 |  | 0.17 |
|  | sig. | *** | n.s. |  | n.s. |
|  | wood (63) | 6.60 (9.94) | 0.71 (1.90) | 0.27 (1.01) |  |
|  | floor (57) | 1.88 (3.84) | 0.16 (0.53) | 0.21 (0.67) |  |
|  | $z$ | 3.50 | 2.23 | 0.37 |  |
|  | sig. | *** | ** | n.s. |  |
| $\begin{array}{r} \mathrm{T} 3 \\ (6) \end{array}$ | wood (66) | 9.05 (15.27) | 5.20 (10.47) | 0.82 (2.20) |  |
|  | floor (54) | 1.56 (4.82) | 0.30 (1.25) | 0.24 (0.97) |  |
|  | $z$ | 3.76 | 3.77 | 1.92 |  |
|  | sig. | *** | *** | * |  |
| $\begin{gathered} \text { T4 } \\ (8) \end{gathered}$ | wood (80) | 1.00 (1.92) | 0.73 (2.23) | 0.05 (0.22) |  |
|  | floor (80) | 0.18 (0.69) | 0.04 (0.19) | 0.04 (0.19) |  |
|  | $z$ | 3.62 | 2.74 | 0.36 |  |
|  | sig. | *** | ** | n.s. |  |
| $\begin{array}{r} \text { A5 } \\ (7) \end{array}$ | wood (88) | 5.60 (10.97) |  | 3.98 (6.94) |  |
|  | floor (52) | 1.79 (3.37) |  | 4.19 (7.28) |  |
|  | 2 | 3.03 |  | 0.27 |  |
|  | sig. | ** |  | n.s. |  |
| $\begin{array}{r} \text { F1 } \\ (7) \end{array}$ | wood (41) | 0.78 (1.85) |  |  |  |
|  | floor (99) | 0.07 (0.38) |  |  |  |
|  | z | 2.43 |  |  |  |
|  | sig. | ** |  |  |  |

[^5]Figure 1 : Study area and plot location map.


Figure 2 : Climate diagrams.

Abscissa in months, ordinate with one division to $10^{\circ} \mathrm{C}$ or 20 mm precipitation (except 100 mm at top of diagram),

A = elevation above sea level,
$B=$ distance drom the coast,
$C=$ length of record,
$\mathrm{D}=$ mean annual temperature $\left({ }^{\circ} \mathrm{C}\right)$,
$\mathrm{E}=$ mean annual precipitation (mm),
$\mathrm{F}=$ highest temperature on record,
$\mathrm{G}=$ mean daily maximum of the warmest month (July),
$\mathrm{H}=$ mean daily minimum of the coldest month (January),
I = lowest temperature on record,
$\mathrm{J}=$ mean monthly precipitation curve,
$\mathrm{K}=$ mean monthly temperature curve,
L (vertical shading, two scales) = relative humid season,

M (dotted shading) = relative period of drought,
$\mathrm{N} \quad$ (neutral shading) $=$ months with mean daily minimum below $0^{\circ} \mathrm{C}$,
$0 \quad$ (diagonal shading) $=$ months with lowest temperature on record below $0^{\circ} \mathrm{C}$,
$P=$ frost-free period.
[following Walter and Lieth (1967), data from
Atmospheric Environment Service (Anon. 1982) ].


Figure 3 : Watersheds sampled in the study area.

1. Sproat Lake;
2. Cous Creek;
3. Nahmint Lake;
4. Kennedy River;
5. Estevan;
6. Cypre River;
7. China Creek;
8. Museum Creek;
9. Coleman Creek;
10. Nitinat River;
11. Sarita River;
12. Pachena River;
13. K1anawa River.


Figure 4 : Microplot sampling designs.
a) systematic microplot placement within $500 \mathrm{~m}^{2}$ plot;
b) stratified random microplot placement within plot;
c) cover classes used in conjunction with microplots.


Figure 5 : Reciprocal averaging ordination of forest vegetation data from 172 plots.

Variance explained is $11.0 \%$ by the first axis and 8.8 \% by the second axis. Solid triangles ( $\mathbf{\Delta}$ ) indicate plots from the subalpine vegetation group, squares indicate plots from the floodplain vegetation group and circles indicate plots from the Pinus contorta vegetation group. Community types : floodplain forests (F1), $\boldsymbol{\square}$; floodplain forests (Lysichitum variant) (F2), $\square$; dry Pinus-Pseudotsuga forests (D1), ; coastal dry Pinus forests (D2), O ; dry Pseudotsuga forests (P1), $\triangle$. Species names are approximately located where they appear in the species ordination also produced by RA. Nonclassified plots are represented by small dots.


Figure 6 : Reciprocal averaging ordination of forest vegetation data from 140 plots.

Variance explained is 13.6 \% by the first axis and 9.8 \% by the second axis. Circles ( ${ }^{()}$ indicate plots from the Pseudotsuga group, squares ( $\square$ ) indicate plots from the Abies group, and triangles ( $\mathbf{\Delta}$ ) indicate plots from the Thuja group. Species names are approximately located where they appear in the species ordination also produced by RA. A single non-classified plot is represented by a small dot.


Figure 7 : Reciprocal averaging ordination (a) and direct ordination
(b) of 59 plots from the Pseudotsuga vegetation group.

Variance explained is 14.4 \% by the first axis and
9.6 \% by the second axis. Community types :

Pseudotsuga-Thuja-Acer forests (P2), ■ ;
Pseudotsuga-Linnaea forests (P3), O ;
Pseudotsuga-Berberis forests (P4), $\triangle$;
Tsuga-Pseudotsuga-Polystichum forests (P5), $\square$;
montane Tsuga forests (P6), $\mathbf{\Delta}$;
montane Tsuga-Gaultheria forests (P7), ; added to the direct ordination, dry Pinus-Pseudotsuga
forests (D1), . Species names are approximately
located where they appear in the species ordination also produced by RA. Non-classified plots are represented by small dots. The topographical gradient is modified from Whittaker (1960).



Figure 8 : Reciprocal averaging ordination (a) and direct ordination (b) of 40 plots from the Thuja vegetation group.

Variance explained is 23.4 \% by the first axis and 13.5 \% by the second axis. Community types : coastal dry Thuja forests (T1), ;
coastal Tsuga-Blechnum-Polystichum forests (T2), $\square$; coastal montane Thuja forests (T3), $\triangle$;
coastal Thuja forests (T4), $\boldsymbol{\Delta}$;
coastal wet Thuja forests (T5), O ;
added to the direct ordination, coastal dry Pinus forests (D2), $\diamond$. Species names are approximately located where they appear in the species ordination also produced by RA. A single non-classified plot is represented by a small dot. The topographical gradient is modified from Whittaker (1960).



Figure 9 : Reciprocal averaging ordination (a) and direct ordination (b) of 40 plots from the Abies vegetation group.

Variance explained is 16.7 \% by the first axis and 10.3 \% by the third axis. Community types : montane Tsuga-Abies-Gaultheria forests (A1), ; montane Abies-Tsuga forests (A2), $\triangle$; montane Tsuga-Abies forests (A3), $\boldsymbol{A}$; montane Abies-Streptopus forests (A4), ■ ; lowland Abies forests (A5), $\square$; Tsuga-Gaultheria-Blechnum forests (A6), O ; Tsuga-Blechnum-Polystichum forests (A7), . Species names are approximately located where they appear in the species ordination also produced by RA. Non-classified plots are represented by small dots. The topographical gradient is modified from Whittaker (1960).



Figure 10 : Reciprocal averaging ordination of 105 modal vegetation plots.

Variance explained is 15.0 \% by the first axis and 8.8 \% by the second axis. Circles are modal plots from the Pseudotsuga group, squares are modal plots from the Abies group, and triangles are modal plots from the Thuja group. Modal plots not classified into community types are represented by small dots. Community types : P2, © $\mathrm{P} 3, \boldsymbol{\theta}$; $\mathrm{P} 4, \mathrm{O}$; $\mathrm{P} 5, \mathrm{O}^{-} \mathrm{P} 6, \mathrm{O} ; \mathrm{P} 7, \ominus$; $\mathrm{T} 2, \boldsymbol{\Delta} ; \mathrm{T} 3, \triangle$; $\mathrm{T} 4, \boldsymbol{A} ; \mathrm{A} 3, \square$; $\mathrm{A} 6, \square$; A7, $\square$ (see table 15 for community type codes).


Figure 11 : Relationships between species basal areas, LFH thickness/ effective rooting depth ratios, and distance from the coast in 105 modal vegetation plots.

Polynomial regression equations for basal area ( $\mathrm{m}^{2} / \mathrm{ha}$ )
of species :
P. menziesii $=-4.202 \cdot D+0.261 \cdot D^{2}-0.003 \cdot D^{3}+16.629$ ( $\mathrm{P}=.0001, \mathrm{~F}=20.8$ );
T. plicata $=29.204 \cdot D-1.801 \cdot D^{2}+0.039 \cdot D^{3}-0.00029 \cdot D^{4}$

- $34.556(\mathrm{P} .=.0001, \mathrm{~F}=18.2)$;
A. amabilis $:=-0.419 \cdot \mathrm{D}+21.76(\mathrm{P}=.0001, \mathrm{~F}=29.9)$;
T. $\underline{\text { heterophylla }}=-9.538 \cdot \mathrm{D}+0.692 \cdot \mathrm{D}^{2}-0.0174 \cdot \mathrm{D}^{3}+$
$0.00014 \cdot D^{4}+67.189(P=.021, F=3.03)$;
LFH/E.R.D. ratio $=-6.185 \cdot D+0.552 \cdot D^{2}-0.0167 \cdot D^{3}$
$+0.0015 \cdot \mathrm{D}^{4}+79.906(\mathrm{P}=.0001, \mathrm{~F}=13.1)$.
$\mathrm{D}=$ distance from the coast in km.


Figure 12 : Isoline maps of vascular species richness, LFH thickness/ effective rooting depth ratio, and climate variables within the study area [c1imate maps adapted from Colidago (1980)] .


Figure 13 : Canonical analyses of vegetation groups, and community types within three groups, based on environmental data.

Shown are means of plot scores, and $90 \%$ confidence circles, on the first two canonical axes. Codes to vegetation groups and community types are listed in Table 15.


Thuja types


Figure 14 : Canonical analysis of twenty-two community types and the subalpine group based on environmental data.

Shown are means of plot scores, and $90 \%$ confidence circles, on the first two canonical axes. Codes to community types and group are listed in Table 15.


Figure 15 : Tree size-class structure : Pseudotsuga group community types and dry Pinus-Pseudotsuga forests (D1).

Hand-fitted and smoothed curves from the data of all plots within each community type. Codes to community types are listed in Table 15.

Ps = Pseudotsuga menziesii,
Ts = Tsuga heterophy11a,
Th $=$ Thuja plicata,
Pi $=$ Pinus contorta.


Figure 16 : Tree size-class structure : Thuja group community types and coastal dry Pinus forests (D2).

Hand-fitted and smoothed curves from the data of all plots within each community type. Codes to community types are listed in Table 15.

Th = Thuja plicata, Ts = Tsuga heterophy11a, $\mathrm{Ab}=\underline{\text { Abies }}$ amabilis, Pi $=$ Pinus contorta.


Figure 17 : Tree size-class structure : Abies group community types and coastal Tsuga-Blechnum-Polystichum forests (T2).

Hand-fitted and smoothed curves from the data of all plots within each community type. Codes to community types are listed in Table 15.
$\mathrm{Ab}=\underline{\text { Abies }}$ amabilis,
Ts = Tsuga heterophylla,
Ps $=$ Pseudotsuga menziesii.


Figure 18 : Community type photographs.
a) montane Tsuga-Gaultheria forests (P7, plot 32);
b) Pseudotsuga-Berberis forests ( $\mathrm{P} 4, \mathrm{plot} 123$ );
c) Tsuga-Pseudotsuga-Polystichum forests (P5, plot 124);
d) coastal Thuja forests (T4, plot 49);
e) Tsuga-Blechnum-Polystichum forests (A7, plot 91);
f) lowland Abies forests (A5, plot 162).


## Appendix 1 : List and constancy of species found in vegetation plots.

Life-form divisions used for shrubs and herbs (following Scoggan 1978-1979) :

Phanerophytes
(woody stems, perennating buds above 25 cm from ground)
Ms Mesophanerophytes, $8-30 \mathrm{~m}$ in height Mc Microphanerophytes, $2-8 \mathrm{~m}$ in height $\mathrm{N} \quad$ Nanophanerophytes, 25 cm to 2 m in height

Chamaephytes
Ch (woody stems, perennating buds within 25 cm of ground)

Hemicryptophytes
(perennating buds at ground surface)
Hp Protohemicryptophyte without runners
Hpr Protohemicryptophyte with runners
Hs Hemicryptophyte, semi-rosette, without runners
Hsr Hemicryptophyte, semi-rosette, with runners
Hr Hemicryptophyte, rosette, without runners
Hrr Hemicryptophyte, rosette, with runners

Cryptophytes
(perennating buds or structure under ground surface)
Gp Saprophytic or parasitic geophyte
Grh Rhizome geophyte, perennating bud terminating a deep rhizome
Gst Stem-tuber geophyte, perennating by tubers or corms
Grt Root-tuber geophyte, perennating by tuberous roots
$\mathrm{Gb} \quad$ Bulb geophyte, perennating by a bulb or bulbs
He1 Helophyte, perennating buds and lower part of plant submersed or in mud

Therophytes
(perennating as a seed)

T
Therophyte, plant annual

| Trees |  |  | Constan | y (\%) |  |
| :---: | :---: | :---: | :---: | :---: | :---: |
| Abbr. | Species | All plots $(172)^{1}$ | $\begin{aligned} & \text { Pse. }{ }^{2} \\ & (59) \end{aligned}$ | Thu. <br> (40) | Abies (40) |
| ABA | Abies amabilis (Doug1.) Forbes | 59.3 | 30.5 | 82.5 | 92.5 |
| ABG | Abies grandis (Doug1.) Forbes | 2.3 | 5.0 |  |  |
| ACM | Acer macrophyllum Pursh | 13.3 | 22.0 |  | 2.5 |
| ALR | Alnus rubra Bong. | 4.6 | 8.5 | 2.5 |  |
| ARB | Arbutus menziesii Pursh | 3.4 |  |  |  |
| CHN | Chamaecyparis nootkatensis (D.Don) Spach | 11.6 | 6.8 | 7.5 | 5.0 |
| COR | Cornus nuttallii Aud. | 9.3 | 22.0 |  |  |
| PIS | Picea sitchensis (Bong.) Carr. | 8.1 |  | 12.5 | 2.5 |
| PIC | Pinus contorta Doug1. | 7.5 |  | 10.0 |  |
| PIM | Pinus monticola Dougl. | 9.8 | 11.8 | 7.5 |  |
| PSE | Pseudotsuga menziesii (Mirbe1) Franco | 55.2 | 98.3 | 10.0 | 30.0 |
| TAX | Taxus brevifolia Nutt. | 22.6 | 11.8 | 60.0 | 7.5 |
| THU | Thuja plicata Donn. | 73.2 | 81.3 | 95.0 | 55.0 |
| TSH | Tsuga heterophyl1a (Raf.) Sarg. | 96.5 | 100.0 | 100.0 | 100.0 |
| TSM | Tsuga mertensiana (Bong.) Carr. | 5.8 | 3.3 |  |  |
|  | TOTAL TREE Species | 15 | 12 | 10 | 8 |

1 number of plots
2 vegetation groups

| Shrubs |  | Lifeforms | Constancy (\%) |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Abbr. | Species |  | All plots (172) | Pse. (59) | Thu . <br> (40) | Abies (40) |
| ACGL | Acer glabrum Torr. | Mc | 2.9 | 6.7 |  |  |
| AMAL | Amelanchier alnifolia Nutt. | N | 8.1 | 13.5 |  |  |
| ARCO | Arctostaphylos columbiana Piper | N | 2.3 |  |  |  |
| BENE | Berberis nervosa Pursh | N | 32.5 | 72.8 |  | 7.5 |
| CosT | Cornus stolonifera Michx. | N | 0.5 |  |  |  |
| GASH | Gau1theria shallon Pursh | N (Mc) | 68.6 | 69.4 | 97.5 | 50.0 |
| HODI | Holodiscus discolor (Pursh) Maxim. | N (Mc) | 4.6 | 6.7 |  |  |
| LOCI | Lonicera ciliosa (Pursh) DC. | N (Mc) | 1.1 | 1.6 |  |  |
| MEFE | Menziesia ferruginea Smith | N (Mc) | 34.3 | 5.0 | 85.0 | 35.0 |
| OPHO | Oplopanax horridus (Smith) Miq. | N (Mc) | 6.9 | 3.3 |  | 17.5 |
| PAMY | Pachistima myrsinites (Pursh) Raf. | N | 1.7 |  |  |  |
| PHYS | Physocarpus opulifolius (L.) Maxim. | Mc | 0.5 |  |  |  |
| PYUS | Pyrus fusca Raf. | Mc (Ms) | 4.6 |  | 15.0 |  |
| RHAM | Rhamnus purshiana DC. | Mc (Ms) | 2.9 | 1.6 | 10.0 |  |
| RHHA | Rhododendron albiflorum Hook. | N | 0.5 |  |  |  |
| RIBB | Ribes bracteosum Doug1. | N (Mc) | 4.0 | 1.6 |  | 2.5 |
| RIBL | Ribes lacustre (Pers.) Poir. | N | 2.3 | 5.0 |  |  |
| ROGY | Rosa gymnocarpa Nutt. | N | 17.4 | 32.2 | 5.0 | 2.5 |
| RUPA | Rubus parviflorus Nutt. | Hp | 4.6 | 5.0 |  | 2.5 |
| RUSP | Rubus spectabilis Pursh | Hpr | 43.6 | 16.9 | 72.5 | 65.0 |
| RUUR | Rubus ursinus Cham. \& Schlecht. | Hpr | 19.7 | 44.0 |  |  |
| SALI | Salix sp. | Mc | 1.7 |  |  |  |


| Shrubs |  |  | Constancy (\%) |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  |  | Lifeforms | $\begin{aligned} & \text { A11 plots } \\ & (172) \end{aligned}$ | Pse. (59) | Thu. (40) | Abies (40) |
| SAMR | Sambucus racemosa L. | Mc | 3.4 | 3.3 |  | 5.0 |
| SOSI | Sorbus sitchensis Roemer | Mc | 3.4 |  | 2.5 | 7.5 |
| SYAL | $\left[\begin{array}{l}\text { Symphoricarpos albus (L.) Blake } \\ \text { ( Symphoricarpos mollis Nutt. }\end{array}\right.$ | N <br> Ch | 5.2 | 11.8 |  |  |
| VAAL | Vaccinium alaskaense Howe 11 | N | 70.3 | 38.9 | 95.0 | 95.0 |
| VAME | Vaccinium membranaceum Doug1. | N | 4.0 | 3.3 |  | 2.5 |
| VAOL | Vaccinium ovalifolium Smith | N | 33.1 | 11.8 | 55.0 | 40.0 |
| VAOT | Vaccinium ovatum Pursh | $N(M c)$ | 21.5 | 13.5 | 47.5 |  |
| VAPA | Vaccinium parvifolium Smith | $N$ (Mc) | 99.4 | 100 | 100 | 100 |
|  | TOTAL SHRUB Species |  | 31 | 23 | 11 | 14 |


| Herbs |  | Lifeforms | Constancy (\%) |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Abbr. | Species |  | All plots $(172)$ | Pse. <br> (59) | Thu. $(40)$ | $\begin{aligned} & \text { Abies } \\ & (40) \end{aligned}$ |
| ACHI | Achillea millefolium L. | Hsr | 1.1 |  |  |  |
| ACTR | Achlys triphylla (Smith) DC. | Grh | 51.7 | 86.4 | 2.5 | 52.5 |
| ADBI | Adenocaulon bicolor Hook. | Hs | 10.4 | 13.5 |  | 12.5 |
| ADPE | Adiantum pedatum L. | Grh | 12.7 | 18.6 |  | 15.0 |
| AGAL | Agrostis sp. | Hs | 1.1 |  |  |  |
| AGSC | Agrostis hyemalis (Walt.) BSP. | Hs | 2.3 |  |  |  |
| ALVI | Allotropa virgata T. \& G. | Gp | 5.8 | 13.5 |  |  |
| ANLY | Anemone 1yallii Britt. | Grh | 2.3 | 5.0 |  |  |
| ANNE | Antennaria neglecta Greene | Ch | 0.5 |  |  |  |
| APAN | Apocynum androsaemifolium L. | Grh (Hp) | ) 2.9 | 1.6 |  |  |
| ARCE | Arceuthobium campy1opodum Engelm. | parasite | e | - | - | - |
| ARUV | Arctostaphylos uva-ursi (L.) Spreng. | Ch | 1.1 |  |  |  |
| AREN | Arenaria macrophylla Hook. | Hpr | 2.9 | 3.3 |  |  |
| ARUY | Aruncus sylvester Kostel | Hp | 3.4 | 1.6 |  |  |
| ASTR | Asplenium trichomanes L. | Hr | 0.5 | 1.6 |  |  |
| ATFI | Athyrium filix-femina (L.) Roth. | Hr | 22.0 | 11.8 | 10.0 | 40.0 |
| BLSP | Blechnum spicant (L.) Roth. | Hr | 63.9 | 33.8 | 100 | 87.5 |
| ВОНО | Boschniakia hookeri Walpers | Gp | 11.0 | 11.8 | 12.5 |  |
| BOMU | Botrychium multifidum (Gme1.) Trevis | Grh | 0.5 |  | 2.5 |  |
| BOVI | Botrychium virginianum (L.) Swartz | Grh | 0.5 | 1.6 |  |  |
| BOYE | Boykinia elata (Nutt.) Greene | Hs | 5.8 | 1.6 | 5.0 | 7.5 |
| BROV | Bromus vulgaris (Hook.) Shear | Hs | 4.6 | 6.7 |  |  |
| CALA | Calamagrostis nutkaensis (Presl) Steud. | Hsr | 1.7 |  | 7.5 |  |


| Herbs |  |  | Constancy (\%) |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  |  | Lifeforms | All plots (172) | Pse. (59) | Thu. (40) | Abies (40) |
| CALY | Calypso bulbosa (L.) Oakes | Gst | 4.6 | 11.8 |  |  |
| CAMP | Campanula scouleri Hook. | Hpr | 5.8 | 11.8 |  |  |
| CARD | Carex deweyana Schw. | Hs | 0.5 |  |  |  |
| CARH | Carex hendersonii Bailey | Hs | 1.7 | 1.6 |  |  |
| CARI | Carex laeviculmis Meinsh. | Hs | 0.5 |  | 2.5 |  |
| CARL | Carex leptalea Wahl. | Hsr | 0.5 |  | 2.5 |  |
| CARM | Carex mertensii Prescott | Grh | 0.5 |  |  | 2.5 |
| CARO | Carex obnupta Bailey | Grh | 5.2 |  | 15.0 |  |
| CARR | Carex sp. 1 |  | 0.5 |  | 2.5 |  |
| CARS | Carex sp. 2 |  | 0.5 |  | 2.5 |  |
| CHME | Chimaphila menziesii (R.Br.) Spreng. | Hpr | 22.0 | 40.6 |  | 12.5 |
| CHUM | Chimaphila umbellata (L.) Bart. | Hpr | 20.3 | 37.2 |  | 10.0 |
| CIRC | Circaea alpina L. | Grh | 0.5 |  |  |  |
| CLUN | Clintonia uniflora (Schult.) Kunth. | Grh | 6.3 | 1.6 |  | 10.0 |
| COAA | Collomia heterophy11a Hook. | T | 0.5 |  |  |  |
| COAS | Coptis asplenifolia Salisb. | Hrr | 6.9 |  | 15.0 | 12.5 |
| COMA | Corallorhiza maculata Raf. | Grh | 6.9 | 10.1 |  | 7.5 |
| COME | Corallorhiza mertensiana Bong. | Grh | 13.9 | 27.1 | 2.5 | 5.0 |
| CORN | Cornus canadensis L. | Hpr | 23.8 | 6.7 | 45.0 | 27.5 |
| CRCR | Cryptogramma crispa (L.) R. Br . | Hr | 1.7 |  |  |  |
| CYST | Cystopteris fragilis (L.) Bernh. | Hr | 0.5 |  |  | 2.5 |
| DASP | Danthonia spicata (L.) Beauv. | Hs | 4.6 |  |  |  |
| DESC | Deschampsia caespitosa (L.) Beauv. | Hs | 0.5 |  | 2.5 |  |


| Herbs |  | $\begin{aligned} & \text { Life- } \\ & \text { forms } \end{aligned}$ | Constancy (\%) |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  |  |  | All plots (172) | $\begin{aligned} & \text { Pse. } \\ & \text { (59) } \end{aligned}$ | Thu . $(40)$ | $\begin{aligned} & \text { Abies } \\ & (40) \end{aligned}$ |
| DICE | Dicentra formosa (Andr.) Walp. | Grh | 0.5 | 1.6 |  |  |
| DIHO | Disporum hookeri (Torr.) Nicholson | Grh | 8.1 | 15.2 |  | 5.0 |
| DISM | Disporum smithii (Hook.) Piper | Grh | 1.7 | 1.6 |  | 5.0 |
| DRAU | Dryopteris austriaca (Jacq.) Woynar | Hr (Grh) | ) 16.8 | 10.1 | 7.5 | 37.5 |
| EQTE | Equisetum telmateia Ehrh. | Grh | 1.1 |  | 5.0 |  |
| ERLA | Eriophyllum lanatum (Pursh) Forbes | Hp | 0.5 |  |  |  |
| FEOC | Festuca occidentalis Hook. | Hs | 7.5 | 11.8 |  |  |
| FEOV | Festuca ovina L. | Hs | 1.1 |  |  |  |
| FESA | Festuca subulata Trin. | Hs | 1.7 |  |  |  |
| FESU | Festuca subuliflora Scribn. | Hs | 17.4 | 42.3 | 2.5 | 2.5 |
| FRAG | Fragaria virginiana Duchesne | Hrr | 2.3 |  |  |  |
| FRIT | Fritillaria camschatcensis (L.) Ker-Gawl. | Gb | 1.1 |  |  |  |
| GALI | Galium triflorum Michx. | Hp | 13.3 | 11.8 | 5.0 | 15.0 |
| GAOV | Gaultheria ovatifolia Gray | Ch | 3.4 | 3.3 |  | 2.5 |
| GOOB | Goodyera oblongifolia Raf. | Hr r | 37.7 | 55.9 | 12.5 | 15.0 |
| GYDR | Gymnocarpium dryopteris (L.) Newm. | Grh | 7.5 |  |  | 20.0 |
| HADI | Habenaria dilatata (Pursh) Hook. | Grt | 0.5 |  |  | 2.5 |
| HAEL | Habenaria elegans (Lindl.) Boland. | Grt | 0.5 |  |  |  |
| HASA | Habenaria saccata Greene | Grt | 0.5 |  | 2.5 |  |
| HASP | Habenaria sp. | Grt | 0.5 |  |  |  |
| HECO | Hemitomes congestum Gray | Gp | 7.5 | 13.5 |  | 5.0 |
| HEMI | Heuchera micrantha Dougl. | Hr | 0.5 | 1.6 |  |  |
| HIAL | Hieracium albiflorum Hook. | Hs | 6.9 | 8.4 |  |  |
| HYPA | Hypochaeris radicata L . |  | 2.3 |  |  |  |

Herbs

| HYPO | Hypopitys monotropa Crantz | Gp | 19.1 | 23.7 |  | 25.0 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| LAMU | Lactuca muralis (L.) Fresen. |  | 12.7 | 27.1 |  | 5.0 |
| LANE | Lathyrus nevadensis Wats. | Grh | 2.3 | 6.7 |  |  |
| LIAO | Lilium columbianum Hanson | Gb | 2.9 | 1.6 |  |  |
| LIBO | Linnaea borealis L . | Ch | 35.4 | 50.8 | 30.0 | 7.5 |
| LICA | Listera caurina Piper | Grh | 23.8 | 15.2 | 20.0 | 40.0 |
| LICO | Listera cordata (L.) R. Br. | Grh | 39.5 | 49.1 | 45.0 | 22.5 |
| LUPI | Lupinus sp. |  | 0.5 |  |  |  |
| LUZC | Luzula campestris (L.) DC. | Hs | 1.1 |  |  |  |
| LUZP | Luzula parviflora (Ehrh.) Desv. | Hs | 8.1 | 1.6 | 2.5 | 10.0 |
| LYCL | Lycopodium clavatum L . | Ch | 4.0 | 3.3 | 5.0 | 2.5 |
| LYSE | Lycopodium selago L. | Ch | 2.9 | 1.6 | 2.5 | 2.5 |
| LYSI | Lysichitum americanum Hu1ten \& St.John | Grh | 6.3 |  | 17.5 | 2.5 |
| MAAD | Madia madioides (Nutt.) Greene | Hs | 1.1 |  |  |  |
| MADI | Maianthemum dilatatum (Wood) Nels. \& Macbr. | Grh | 31.3 | 8.4 | 55.0 | 35.0 |
| MECU | Melica subulata (Griseb.) Scribn. | Gst | 3.4 | 1.6 |  |  |
| MIOV | Mitella ovalis Greene | Hrr | 2.9 |  | 5.0 | 2.5 |
| MONE | Moneses uniflora (L.) Gray | Hr | 4.6 | 3.3 | 5.0 | 10.0 |
| MONO | Monotropa uniflora L . | Gp | 6.9 | 16.9 |  | 2.5 |
| MOPA | Montia parvifolia (Moc.) Greene | Hsr | 4.0 | 10.1 |  |  |
| MOSI | Montia sibirica (L.) Howell | T (Hs) | 3.4 | 5.0 |  | 2.5 |
| NEPH | $\frac{\text { Nephrophy11idium }}{\text { (Menzies) Gilg. }}$ | Grh (He | ) 0.5 |  | 2.5 |  |


|  | Constancy (\%) |  |  |  |
| :--- | :---: | :---: | :---: | :---: |
| Life- <br> forms | All plots <br> $(172)$ | Pse. <br> (59) | Thu <br> (40) | Abies <br> $(40)$ |


| OXOR | Oxalis oregana Nutt. |  | 1.1 |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| PAOC | Panicum occidentale Scribn. | Hs | 0.5 |  |  |  |
| PEDI | Pedicularis racemosa Dougl. | Hp | 1.1 | 1.6 |  |  |
| PENS | Penstemon davidsonii Greene | Ch | 0.5 |  |  |  |
| PENT | Penstemon serrulatus Menzies | Hp (Ch) | 0.5 |  |  | 2.5 |
| PHYL | Phyllodoce empetriformis (Sw.) D.Don | Ch | 1.1 |  |  |  |
| PLER | Pleuropogon refractus (Gray) Benth. | Hs | 0.5 |  |  |  |
| POAM | Poa marcida Hitchc. | Hs | 2.9 |  |  | 5.0 |
| POLY | Polypodium glycyrrhiza D.C. Eat. | Grh | 20.3 | 33.8 | 7.5 | 2.5 |
| POMU | Polystichum munitum (Kaulf.) Presl | Hr | 71.5 | 89.8 | 35.0 | 80.0 |
| PRAL | Prenanthes alata (Hook.) D. Dietr. | Hp | 1.7 |  | 2.5 | 5.0 |
| PRUN | Prunella vulgaris L. | Hsr | 1.1 |  | 2.5 |  |
| PTAQ | Pteridium aquilinum (L.) Kuhn | Grh | 16.2 | 33.8 |  | 5.0 |
| PTEA | Pterospora andromedea Nutt. | Gp | 2.3 | 6.7 |  |  |
| PYAP | Pyrola aphylla Smith | Hrr | 1.1 | 3.3 |  |  |
| PYAS | Pyrola asarifolia Michx. | Hrr | 2.3 | 1.6 |  | 2.5 |
| PYPI | Pyrola picta Smith | Hrr | 9.8 | 18.6 |  | 5.0 |
| PYSE | Pyrola secunda L. | Hrr | 8.7 | 3.3 |  | 15.0 |
| RUNI | Rubus nivalis Doug1. | Hpr | 0.5 | 1.6 |  |  |
| RUPE | Rubus pedatus J.E. Smith | Hpr | 16.8 | 1.6 | 10.0 | 37.5 |
| SAXF | Saxifraga ferruginea Graham | Hr | 2.9 |  |  |  |
| SEWA | Selaginella wallacei Hieron. | Ch | 4.6 | 3.3 |  |  |
| SMRA | Smilacina racemosa (L.) Desf. | Grh | 4.0 | 10.1 |  |  |

Herbs

|  | Constancy (\%) |  |  |  |
| :--- | :---: | :---: | :---: | :---: |
| Life- <br> forms | Al1 plots <br> $(172)$ | Pse. <br> $(59)$ | Thu. <br> $(40)$ | Abies <br> $(40)$ |
| Grh | 4.0 | 5.0 | 7.5 |  |
| Gst | 2.3 | 1.6 |  |  |
| Gb | 0.5 | 1.6 |  |  |
| Grh | 23.2 | 6.7 | 27.5 | 35.0 |
| Grh | 13.3 | 5.0 |  | 35.0 |
| Grh | 6.9 |  |  | 17.5 |
| Hsr | 40.1 | 35.5 | 37.5 | 52.5 |
| Hsr | 51.1 | 49.1 | 40.0 | 67.5 |
| Hsr | 1.1 |  |  | 2.5 |
| Hsr | 1.1 |  |  | 22.5 |


| TRIE | Trientalis arctica Fisch. | Hpr | 0.5 |  | 2.5 |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| TRLA | Trientalis latifolia Hook. | Gst | 20.3 | 45.7 |  |  |
| TROV | Trillium ovatum Pursh | Grh | 45.9 | 47.4 | 32.5 | 65.0 |
| TRMA | Trisetum canescens Buck1. | Hs | 0.5 |  |  |  |
| TRMC | Trisetum cernuum Trin. | Hs | 2.3 |  |  |  |
| VEVI | Veratrum viride Ait. | Grh | 10.4 |  | 15.0 | 7.5 |
| VIGL | Viola glabella Nutt. | Hsr | 6.3 | 1.6 | 5.0 | 5.0 |
| VIOR | Viola orbiculata Geyer | Hs | 1.7 |  |  | 2.5 |
| VISE | Viola sempervirens Greene | Hsr | 22.0 | 33.8 | 5.0 | 15.0 |
| ZYVE | Zigadenus venenosus Wats. | Gb | 0.5 |  |  |  |

Mosses

Abbr. Species

Constancy (\%)
All plots Pse. Thu. Abies
(172) (59) (40) (40)
1.1

ANDR Andreaea rupestris Hedw.
ANTI Antitrichia curtipendula (Hedw.) Brid.
1.1
1.7
2.5
2.5

BUXB Buxbaumia piperi Best
CAMY Campylopus atrovirens De Not.
1.1

CLOA Claopodium crispifolium (Hook.) Ren. \& Card. 4.6
59.3
77.9
40.0
52.5

DICM Dicranum majus Sm .
1.1
5.0

DICS Dicranum scoparium Hedw.
8.1
$3.3 \quad 7.5$
DITR Ditrichum sp.
0.5

EURP Eurhynchium pulchellum (Hedw.) Jenn. var. barnesii (Ren. \& Card.) Crum, Steere \& Anders.

HETE Heterocladium macounii Best
HETP $\frac{\text { Heterocladium procurrens (Mitt.) }}{\text { Rau \& Herv. }}$
HOLU Hookeria lucens (Hedw.) Sm.
43.0
78.4
$\begin{array}{llll}64.5 & 84.7 & 47.5 & 65.0\end{array}$

| HYPU | Hypnum circinale Hook. | 64.5 | 84.7 | 47.5 | 65.0 |
| :---: | :---: | :---: | :---: | :---: | :---: |
| HYPV | Hypnum dieckii Ren. \& Card. ex Roell. | 0.5 |  | 2.5 |  |
| HYPP | Hypopterygium fauriei Besch. | 1.1 | 1.6 |  | 2.5 |
| ISOP | Isopterygium elegans (Brid.) Lindb. | 43.0 | 44.0 | 32.5 | 52.5 |
| ISST | Isothecium stoloniferum Brid. | 72.0 | 74.5 | 80.0 | 87.5 |
| LEME | Leucolepis menziesii (Hook.) Steere | 16.2 | 20.3 | 5.0 | 12.5 | ex L. Koch

Constancy (\%)

| Al1 plots <br> $(172)$ | Pse. <br> $(59)$ | Thu. <br> $(40)$ | Abies <br> $(40)$ |
| :---: | :---: | :---: | :---: |

$\begin{array}{llll}\text { META } \\ \begin{array}{ll}\text { Metaneckera } \\ \text { menziesii }\end{array} \text { (Hook. ex Drumm.) } & 0.5 & 1.6 \\ \text { Steere }\end{array}$
MNIU Mnium spinulosum B.S.G.
$13.3 \quad 25.4$
0.5
8.18 .4
75.5
3.4
1.6

POGM Pogonatum alpinum (Hedw.) Roh1.
20.9
37.2
6.9
1.6
4.0

PONF Polytrichum formosum Hedw.
0.5
5.8
1.6

PONJ Polytrichum juniperinum Hedw.
2.9
1.6

PORO Porotrichum bigelovii (Sull.) Kindb.
0.5

RHAA Rhacomitrium aquaticum (Brid. ex Schrad.) 1.7 Brid.

RHAC Rhacomitrium canescens (Hedw.) Brid.
3.4

RHAH Rhacomitrium heterostichum (Hedw.) Brid. 6.9 2.3

RHGL Rhizomnium glabrescens (Kindb.) Kop. 67.4
RHLO Rhytidiadelphus loreus (Hedw.) Warnst. 88.9
$83.0 \quad 95.0$
85.0

RHTR Rhytidiadelphus triquetrus (Hedw.) Warnst. 9.8 20.3
RHYT Rhytidiopsis robusta (Hook.) Broth.
28.4
44.0
27.5

ROEL Roellia roellii (Broth.) Andr. ex Crum
0.5
2.5

| Constancy (\%) |  |  |
| :---: | :---: | :---: |
| A11 plots <br> $(172)$ | Pse. <br> $(59)$ | Thu. <br> $(40)$ |
| 0.5 |  | Abies <br> $(40)$ |
| 0.5 | 2.5 |  |
| 9.3 |  | 2.5 |

Liverworts

Abbr. Species

BAZZ Bazzania denudata (Torr. ex Gott.) Trev.
BLET Blepharostoma trichophyllum (L.) Dum.
CALF Calypogeia fissa (L.) Raddi
CALM Calypogeia muellerana (Schiffn.) K. Muell.
CEPH Cephalozia bicuspidata (L.) Dum.
COCO Conocephalum conicum (L.) Dum. ex Lindb.
DIPA Diplophyllum albicans (L.) Dum.
DIPP Diplophyllum plicatum Lindb.
HEBA Herbertus aduncus (Dicks.) S. Gray
JULE Jungermannia leiantha Grolle
KURZ Kurzia sp.
LEDO Lepidozia reptans (L.) Dum.
MARS Marsupella emarginata (Ehrh.) Dum. 0.5
METZ Metzgeria conjugata Lindb.
1.1
3.4
1.7
12.7
30.8 Lindenb.
12.7
10.4
0.5
8.7
31.3
3.4
11.6
2.9
1.7
9.3

MYTA Mylia taylorii (Hook.) S. Gray
NASC Nardia scalaris S. Gray
PELI Pellia neesiana (Gott.) Limpr.
PLAG Plagiochila porelloides (Torr. ex Nees)

Constancy (\%)

| All plots |
| :--- | :--- | :--- | :--- |
| $(172)$ | | Pse. |
| :--- |
| $(59)$ | | Thu. |
| :--- |
| $(40)$ | | Abies |
| :--- |
| $(40)$ |

$3.3 \quad 32.5$
15.0
$3.3 \quad 20.0$
15.0 2.5
$1.6 \quad 27.5$
12.5
12.5
6.31 .617 .5
1.15 .0
7.5
10.1
5.0
17.5
2.5
$\begin{array}{lllll}\text { PTIC Ptilidium californicum (Aust.) Underw. } & 1.1 & 1.6 & 2.5\end{array}$
PTIP Ptilidium pulcherrinum (G. Web.) Hampe
RICL Riccardia 1atifrons Lindb.
0.5
10.4
$30.0 \quad 10.0$

Liverworts

RICM Riccardia multifida (L.) S. Gray
SCAA Scapania americana K. Muell.
SCAB Scapania bolanderi Aust.
SCAP Scapania paludosa (K. Mue11.) K. Mue11.

TOTAL LIVERWORT Species

26
$11 \quad 22$
$11 \quad 22$
12
All plots Pse. Thu. Abies
(172) (59) (40) (40)
$0.5 \quad 2.5$
1.1
83.1
$89.8 \quad 77.5$
90.0
0.5
2.5

Lichens

Abbr. Species

CLAS Cladina impexa (Harm.) B. de Lesd.
CLAR Cladina rangiferina (L.) Harm.
CLEA Cladonia acuminata (Ach.) Norr1.
CLDB Cladonia bellidiflora (Ach.) Schaer.
CLDP Cladonia chlorophaea (Flk.) Spreng.
CLEC Cladonia furcata (Huds.) Schrad.
(172) (59) (40) (40)
$3.4 \quad 3.3$
5.2
1.6
0.5
0.5

CLDG Cladonia gracilis (L.) Willd.
4.0

CLDF Cladonia multiformis Merr. 4.0
CLEB Cladonia pyxidata (L.) Hoffm. 0.5
CLDS Cladonia squamosa (Scop.) Hoffm.
1.1

CLDU Cladonia uncialis (L.) Wigg.
2.9

LOBA Lobaria linita (Ach.) Rabh.
2.3
0.5
1.6

Lobaria oregana (Muill. Arg.) Hale
Peltigera aphtosa (L.) Willd.
1.7

PELO Peltigera leucophlebia (Ny1.) Gyeln.
Peltigera membranacea (Ach.) Ny1.
PELT Peltigera polydactyla (Neck.) Hoffm.
Peltigera praetextata (Somm.) Vain.
5.25 .0
$6.9 \quad 10.1$
2.5
5.0
11.6
20.3
0.5
2.5

STEO Stereocaulon subcoralloides Nyl.
2.9

STET Stereocaulon tomentosum Fr.

TOTAL LICHEN Species
$\qquad$

Appendix 2 : Environmental data descriptive statistics for vegetation groups and community types.

List of variables
(refer to Table 1 for definitions of classes for discrete variables*) :
1 - elevation (m)
2 - aspect ( $0-180$, NNE to SSW)
3-slope (\%)
4 - topographic position (1-6)*
5 - drainage (1-7)*
6 - effective rooting depth (cm)
7 - root restricting depth (cm)
8 - soil depth (cm)
9 - surficial material ( $0-4$ )*
10 - effective rooting depth/root restricting depth
11 - effective rooting depth/soil depth
12 - LFH thickness/effective rooting depth
13 - fire disturbance (0-1)*
14 - wind disturbance ( $0-1$ )*
15 - worms $(0-3)$ *
16 - LFH $\mathrm{pH}\left(\mathrm{H}_{2} \mathrm{O}\right)$
17 - LFH pH $\left(\mathrm{CaCl}_{2}\right)$
18 - LFH thickness (cm)
19 - A pH $\left(\mathrm{H}_{2} \mathrm{O}\right)$
$20-\mathrm{ApH}\left(\mathrm{CaCl}_{2}\right)$
21 - $\mathrm{B}_{1} \mathrm{pH}\left(\mathrm{H}_{2} \mathrm{O}\right)$
$22-\mathrm{B}_{1} \mathrm{pH}\left(\mathrm{CaCl}_{2}\right)$
$23-\mathrm{B}_{1} \%$ coarse fragments
$24-\mathrm{B}_{1}$ texture $(1-12)$ *
$25-\mathrm{B}_{1} \% \mathrm{~N}$
$26-\mathrm{B}_{1} \% \mathrm{C}$
27 - $\mathrm{B}_{1} \mathrm{C} / \mathrm{N}$ ratio
$28-\mathrm{B}_{2} \mathrm{pH}\left(\mathrm{H}_{2} \mathrm{O}\right)$
$29-\mathrm{B}_{2} \mathrm{pH}\left(\mathrm{CaCl}_{2}\right)$
30 - LFH \% C
31 - LFH \% N
32 - LFH C/N ratio

| Slope classes | (C.S.S.C., 1978) |
| :---: | :--- |
| $\%$ slope | terminology |
| $<2.5$ | level |
| $2-5$ | very gentle slopes |
| $6-9$ | gentle slopes |
| $10-15$ | moderate slopes |
| $16-30$ | strong slopes |
| $31-45$ | very strong slopes |
| $46-70$ | extreme slopes |
| $71-100$ | steep slopes |
| $>100$ | very steep slopes |

(Note : variable 15 cannot be interpreted from these data since it was not recorded in 1980 plots, and these are included in these statistics)

Appendix 2 : Subalpine vegetation group (SA)


Appendix 2 : Pinus contorta vegetation group (D)
descriptive measures

| VARIABLE | N | MINIMUM | MAXIMUM | MEAN | STD DEV |
| :---: | :---: | :---: | :---: | :---: | :---: |
| 1. ELEV | 7 | 45.000 | 322.00 | 156.71 | 117.01 |
| 2. ASPECT | 7 | 46.000 | 167.00 | 103.86 | 41.611 |
| 3. SLOPE | 7 | 10.000 | 65.000 | 37.143 | 19.334 |
| 4.PDSIT | 7 | 1.0000 | 1.0000 | 1.0000 |  |
| 5. DRAINAGE | 7 | 1.0000 | 1.0000 | 1.0000 |  |
| 6. EROOTDEP | 7 | 3.0000 | 23.000 | 12.857 | 8.4346 |
| 7.ROOTDEP | 7 | 3.0000 | 23.000 | 12.857 | 8.4346 |
| 8. SbILDEP | 7 | 3.0000 | 23.000 | 12.857 | 8.4346 |
| 9.MATERIAL | 7 | 0. | 1.0000 | . 28571 | . 48795 |
| 10.RATIO1 | 7 | 1.0000 | 1.0000 | 1.0000 |  |
| 11. RATIO2 | 7 | 1.0000 | 1.0000 | 1.0000 |  |
| 12.RATIO3 | 7 | . 13000 | 1.0000 | . 46857 | . 38133 |
| 13.FIRE | 7 | 0. | 1.0000 | . 42857 | . 53452 |
| 14.WIND | 7 | 0. | 1.0000 | . 42857 | . 53452 |
| 15. WORMS | 7 | 0. | 0. | 0. |  |
| 16. ORGPHWAT | 7 | 3.8000 | 4.4000 | 4.0429 | . 19881 |
| 17. ORGPHCAL | 7 | 3.3000 | 4.0000 | 3.6143 | . 25448 |
| 18.ORGTHICK | 7 | 1.0000 | 14.000 | 4.4286 | 4.4668 |
| 19. APHWAT | 1 | 3.8000 | 3.8000 | 3.8000 |  |
| 20.APHCAL | 1. | 3.2000 | 3.2000 | 3.2000 |  |
| 21.B1PHWAT | 5 | 4.2000 | 5.4000 | 4.7600 | . 43359 |
| 22.B1PHCAL | 5 | 3.7000 | 4.7000 | 4. 1800 | . 38987 |
| 23.CDARSE\% | 7 | 0. | 95.000 | 40.571 | 41.016 |
| 24.TEXTURE | 5 | 2.0000 | 6.0000 | 4.2000 | 1.7889 |
| 25. $\mathrm{B}^{1 \% \mathrm{~N}}$ | 5 | . 20000 | . 36000 | . 28000 | . 58310 |
| $26 . \mathrm{Bi} \mathrm{\% C}$ | 5 | 3. 1000 | 15.620 | 8.3240 | 4.7100 |
| 27. BICNRAT | 5 | 15.300 | 43.500 | 28.080 | 11.107 |
| 28.B2PHWAT | 0 |  |  |  |  |
| 29.82PHCAL | 0 |  |  |  |  |
| $30.0 \mathrm{RG} \mathrm{\%}$ | 7 | 36.150 | 49.760 | 44.830 | 4.7782 |
| 31. ORG\% | 7 | . 48000 | .88000 | . 63571 | . 13794 |
| 32. ORGCNRAT | 7 | 48.200 | 100. 10 | 73.871 | 19 |

Appendix 2 : Floodplain vegetation group (F)

## DESCRIPTIVE MEASURES

| VARIABLE | N | MINIMUM | MAXImum | MEAN | STD DEV |
| :---: | :---: | :---: | :---: | :---: | :---: |
| 1. ELEV | 10 | 15.000 | 95.000 | 47.900 | 37.563 |
| 2. ASPECT | 10 | 0. | 107.00 | 19.300 | 40.988 |
| 3. SLOPE | 10 | 0. | 5.0000 | 2. 1000 | 2.0790 |
| 4. POSIT | 10 | 5.0000 | 5.0000 | 5.0000 |  |
| 5. DRAINAGE | 10 | 4.0000 | 7.0000 | 4.9000 | 1.1972 |
| 6.EROOTDEP | 10 | 11.000 | 113.00 | 55.000 | 29.728 |
| 7. ROOTOEP | 10 | 11.000 | 130.00 | 72.500 | 42.009 |
| 8. SOILDEP | 10 | 62.000 | 130.00 | 96.100 | 18.947 |
| 9.MATERIAL | 10 | 3.0000 | 4.0000 | 3.5000 | . 52705 |
| 10.RATIO1 | 10 | . 61000 | 1.0000 | . 82600 | . 16304 |
| 11. RATIO2 | 10 | . 10000 | . 87000 | . 56000 | . 22730 |
| 12.RATIO3 | 10 | . $30000-1$ | . 19000 | . 77000 | . 61653 |
| 13.FIRE | 10 | 0. | 1.0000 | . 30000 | . 48305 |
| 14.WIND | 10 | 0. | 1.0000 | . 50000 | . 52705 |
| 15. WORMS | 10 | 0. | 3.0000 | 2.0000 | 1.0541 |
| 16. ORGPHWAT | 10 | 4.2000 | 5.6000 | 4.7900 | . 38427 |
| 17.ORGPHCAL | 10 | 3.9000 | 5.1000 | 4.4000 | . 35277 |
| 18. ORGTHICK | 10 | 1.0000 | 10.000 | 3.4000 | 2.6750 |
| 19. APHWAT | 8 | 4.0000 | 5.7000 | 4.8375 | . 63005 |
| 20. APHCAL | 8 | 3.6000 | 5.4000 | 4.4000 | . 62564 |
| 21.B4PHWAT | 10 | 4.3000 | 5.9000 | 5. 1600 | . 54467 |
| 22.B1PHCAL | 10 | 3.7000 | 5.4000 | 4.4800 | . 49621 |
| 23.COARSE\% | 10 | 0. | 10.000 | 1.0000 | 3.1623 |
| 24.TEXTURE | 10 | 3.0000 | 10.000 | 5.3000 | 2.8304 |
| $25 . \mathrm{B} 1 \% \mathrm{~N}$ | 10 | .60000-1 | . 30000 | . 20800 | . 79833 |
| $26 . \mathrm{B} 4 \% \mathrm{C}$ | 10 | 1.0200 | 7.2000 | 4.2130 | 2.0890 |
| 27. B ICNRAT | 10 | 16.700 | 28.200 | 19.860 | 4.1743 |
| 28.E2PHWAT | 10 | 4.5000 | 6.1000 | 5.2900 | . 45570 |
| 29. B2PHCAL | 10 | 3.9000 | 5.5000 | 4.6200 | . 48028 |
| 30.0kG\%C | 10 | 25.210 | 49.200 | 38.552 | 6.8562 |
|  | 10 | . 48000 | 1.0000 | . 71200 | . 16295 |
| 32. ORGCNRAT | 10 | 34.200 | 70.400 | 55.770 | 11.652 |

Appendix 2 : Pseudotsuga vegetation group ( P )

DESCRIPTIVE MEASURES

| VARIABLE | $N$ | MINIMUM |  | MAXIMUM | MEAN | STD DEV |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 1. ELEV | 59 | 15.000 |  | 805.00 | 344.71 | 195.15 |
| 2. ASPECT | 59 | 0. |  | 178.00 | 108.88 | 58.283 |
| 3. SLOPE | 59 | 4.0000 |  | 80.000 | 46.085 | 20.761 |
| 4.POSIT | 59 | 1.0000 |  | 5.0000 | 3. 1525 | . 73844 |
| 5. DRA INAGE | 59 | 1.0000 |  | 4.0000 | 2.4746 | . 79559 |
| 6. EROOTDEP | 59 | 8.0000 |  | 128.00 | 57.932 | 32.400 |
| 7. ROOTDEP | 59 | 15.000 |  | 128.00 | 78.644 | 26.835 |
| 3.50ILDEP | 59 | 15.000 |  | 164.00 | 86.508 | 31.111 |
| 9.MATERIAL | 59 | 0. |  | 3.0000 | 1.3051 | . 70109 |
| 10.RATIO1 | 59 | . 10000 |  | 1.0000 | . 74119 | . 27768 |
| 11. RATIO2 | 59 | . 60000 | -1 | 1.0000 | . 70153 | . 29389 |
| 12.RATIO3 | 59 | . 30000 | -1 | 1.0000 | . 22458 | . 22897 |
| 13.FIRE | 59 | 0. |  | 1.0000 | . 83051 | . 37841 |
| 14. WIND | 59 | 0. |  | 1.0000 | . 16949 | . 13019 |
| 15. WORMS | 59 | 0. |  | 3.0000 | . 94945 | 1.0073 |
| 16. DRGPHWAT | 58 | 3.2000 |  | 5.7000 | 4.1138 | . 59011 |
| 17. CRGPHCAL | 58 | 2.7000 |  | 5.4000 | 3.6534 | . 61991 |
| 18. ORGTHICK | 59 | 1.0000 |  | 23.000 | 8.4407 | 4.6024 |
| 13. LFHWAT | 7 | 3.8000 |  | 5.2000 | 4.3000 | .54772 |
| 20. APHCAL | 7 | 3.2000 |  | 4.8000 | 3.7286 | . 56484 |
| 21.3 PPHWAT | 56 | 3.5000 |  | 6.0000 | 4.9143 | . 43918 |
| 22.E1PHCAL | 56 | 3.0000 |  | 5.4000 | 4.3214 | . 40842 |
| 23. COARSE\% | 59 | 5.0000 |  | 100.00 | 60.169 | 23.717 |
| 24.texture | 56 | 1.0000 |  | 10.000 | 3.9286 | 1.9896 |
| 25.B1\%N | 56 | . 40000 | $-1$ | .35000 | . 14018 | . 70362 |
| 26. B \% C | 56 | 1.3000 |  | 14.440 | 4.6812 | 2.3227 |
| 27. EICNRAT | 56 | 15.300 |  | 94.300 | 36.668 | 17.234 |
| 28.E2PHWAT | 49 | 4.6000 |  | 6.6000 | 5. 1959 | . 39049 |
| 29. B2PHCAL | 49 | 4.1000 |  | 5.7000 | 4.6184 | . 38225 |
| 30.ORG\%C | 58 | 27.010 |  | 56.200 | 45.255 | 5.9177 |
| $31.0 \mathrm{OG} \% \mathrm{~N}$ | 58 | . 48000 |  | 4.5600 | . 86517 | . 21904 |
| 32. ORGCNRAT | 58 | 32.000 |  | 104.40 | 55.093 | 14.307 |

Appendix 2 : Thuja vegetation group (T)

DESCRIPTIVE MEASURES
$\left.\begin{array}{llllll}\text { VARIABLE } & \text { N MINIMUM } & \text { MAXIMUM } & \text { MEAN } & \text { STD DEV } \\ \text { 1.ELEV } & 40 & 15.000 & 610.00 & 173.00 & 155.56 \\ \text { 2.ASPECT } & 40 & 0 . & 169.00 & 76.450 & 53.926 \\ \text { 3. SLOPE } & 40 & 0 . & 96.000 & 29.900 & 20.836 \\ \text { 4. POSIT } & 40 & 1.0000 & 6.0000 & 3.5000 & 1.1094 \\ \text { 5. ORAINAGE } & 40 & 1.0000 & 7.0000 & 4.5500 & 1.4667 \\ \text { 6. EROOTDEP } & 40 & 40 & 5.0000 & 110.00 & 31.025\end{array}\right) 22.564$

Appendix 2 : Abies vegetation group (A)
descriptive measures

| VARIABLE | $N$ | MINIMUM | MAXIMUM | MEAN | STD DEV |
| :---: | :---: | :---: | :---: | :---: | :---: |
| 1. ELEV | 40 | 25.000 | 915.00 | 355.72 | 235.33 |
| 2. ASPECT | 40 | 0. | 172.00 | 57.925 | 51.816 |
| 3.SLOPE | 40 | 3.0000 | 80.000 | 38.800 | 22.253 |
| 4.POSIT | 40 | 2.0000 | 5.0000 | 3.4250 | . 81296 |
| 5. DRA INAGE | 40 | 2.0000 | 6.0000 | 3.6500. | 1.0754 |
| 6. EROOTDEP | 40 | 5.0000 | 109.00 | 33.350 | 25.077 |
| 7. ROOTDEP | 40 | 11.000 | 134.00 | 61.400 | 29.846 |
| 8. SDILDEP | 40 | 17.000 | 196.00 | 96.400 | 32.762 |
| 9. MATERIAL | 40 | 1.0000 | 3.0000 | 1.7250 | .71567 |
| 10. RATIOI | 40 | .12000 | 1.0000 | .57850 | . 30590 |
| 11. RATIO2 | 40 | . $50000-1$ | 1.0000 | . 38250 | . 28372 |
| 12. RATIO3 | 40 | $.40000-1$ | 1.0000 | . 58375 | .34013 |
| 13. FIRE | 40 | 0. | 1.0000 | .30000 | . 46410 |
| 14.WIND | 40 | 0. | 1.0000 | .40000 | . 49614 |
| 15. WORMS | 40 | 0. | 3.0000 | 1.1750 | 1.1297 |
| 16. ORGPHWAT | 40 | 3.0000 | 6.2000 | 3.7500 | . 53060 |
| 17. ORGPHCAL | 40 | 2.4000 | 5.4000 | 3.1875 | . 53166 |
| 18. ORGTHICK | 40 | 3.0000 | 27.000 | 13.225 | 5.8287 |
| 19. APHWAT | 20 | 3.5000 | 6.2000 | 3.9750 | . 62228 |
| 20. APHCAL | 20 | 3.0000 | 5.4000 | 3.3800 | . 56345 |
| 21.81PHWAT | 40 | 3.8000 | 6.3000 | 4.6725 | . 46243 |
| 22.B1PHCAL | 40 | 3.5000 | 5.6000 | 4.1900 | .44133 |
| 23. COARSE\% | 40 | 5.0000 | 95.000 | 46.625 | 23.949 |
| 24.TEXTURE | 40 | 3.0000 | 9.0000 | 4.7500 | 1.7209 |
| $25 . \mathrm{B} 1 \% \mathrm{~N}$ | 40 | $.80000-1$ | .38000 | . 20700 | . $76902-1$ |
| 26. B 1\%C | 40 | 2.4000 | 12.670 | 5.9607 | 2.5819 |
| 27. B 1CNRAT | 40 | 12.500 | 51.100 | 29.462 | 8.8890 |
| 28. E2PHWAT | 36 | 4.3000 | 6.5000 | 5.0194 | .40905 |
| 29.82PHCAL | 36 | 3.8000 | 5.6000 | 4.5389 | . 40162 |
| 30. ORG\%C | 40 | 30.100 | 53.300 | 45.826 | 5.4391 |
| 31. ORG\%N | 40 | .66000 | 1.6800 | 1.0892 | .28331 |
| 32. ORGCNRAT | 40 | 23.500 | 74.500 | 44.655 | 11.825 |

Appendix 2 : Dry Pinus-Pseudotsuga forests (D1)

DESCRIPTIVE MEASURES

| VARIABLE | $N$ | MINIMUM | MAXIMUM | MEAN | STO DEV |
| :---: | :---: | :---: | :---: | :---: | :---: |
| 1. ELEV | 3 | 60.000 | 322.00 | 215.67 | 137.79 |
| 2. ASPECT | 3 | 117.00 | 167.00 | 135.00 | 27.785 |
| 3. SLOPE | 3 | 10.000 | 60.000 | 31.667 | 25.658 |
| 4. POSIT | 3 | 1.0000 | 1.0000 | 1.0000 |  |
| 5. DRAINAGE | 3 | 1.0000 | 1.0000 | 1.0000 |  |
| 6. EROOTDEP | 3 | 3.0000 | 23.000 | 15.333 | 10.786 |
| 7. ROOTDEP | 3 | 3.0000 | 23.000 | 15.333 | 10.786 |
| 8.SOILDEP | 3 | 3.0000 | 23.000 | 15.333 | 10.786 |
| 9.MATERIAL | 3 | 0. | 1.0000 | . 66667 | .57735 |
| 10. RATIO1 | 3 | 1.0000 | 1.0000 | 1.0000 |  |
| 11.RATIO2 | 3 | 1.0000 | 1.0000 | 1.0000 |  |
| 12.RATIO3 | 3 | . 13000 | 1.0000 | . 54333 | . 43662 |
| 13.FIRE | 3 | 1.0000 | 1.0000 | 1.0000 |  |
| 14.WIND | 3 | 0. | 1.0000 | . 66667 | . 57735 |
| 15. WORMS | 3 | 0. | 0. | 0. |  |
| 16. ORGPHWAT | 3 | 4.0000 | 4.4000 | 4.2000 | . 20000 |
| 17. ORGPHCAL | 3 | 3.3000 | 4.0000 | 3.7000 | . 36056 |
| 18.ORGTHICK | 3 | 3.0000 | 4.0000 | 3.3333 | . 57735 |
| 19. APHWAT | 0 |  |  |  |  |
| 20. APHCAL | 0 |  |  |  |  |
| 21.81FHWAT | 2 | 4.8000 | 4.8000 | 4.8000 |  |
| 22.BIPHCAL | 2 | 4.3000 | 4.3000 | 4.3000 |  |
| 23.COARSE\% | 3 | 0. | 95.000 | 58.000 | 50.863 |
| 24.TEXTURE | 2 | 6.0000 | 6.0000 | 6.0000 |  |
| $25.81 \% N$ | 2 | . 20000 | . 28000 | . 24000 | . $56569-1$ |
| 26.B1\%C | 2 | 3. 1000 | 9.7500 | 6.4250 | 4.7023 |
| 27. B1CNRAT | 2 | 15.300 | 34.900 | 25.100 | 13.859 |
| 28.E2PHWAT | 0 |  |  |  |  |
| 29.82PHCAL | 0 |  |  |  |  |
| 30.ORG\%C | 3 | 36.150 | 47.840 | 43.030 | 6.1137 |
| 31. ORG\%N | 3 | .61000 | . 88000 | . 74667 | . 13503 |
| 32.ORGCNRAT | 3 | 48.200 | 78.400 | 59.300 | 16.614 |

Appendix 2 : Coastal dry Pinus forests (D2)

DESCRIPTIVE MEASURES

| VARIABLE | N | MINIMUM | MAXIMUM | MEAN | Sto dev |
| :---: | :---: | :---: | :---: | :---: | :---: |
| 1. ELEV | 4 | 45.000 | 250.00 | 112.50 | 92.961 |
| 2.ASPECT | 4 | 46.000 | 129.00 | 80.500 | 35.369 |
| 3. SLOPE | 4 | 30.000 | 65.000 | 41.250 | 16.008 |
| 4.POSIT | 4 | 1.0000 | 1.0000 | 1.0000 |  |
| 5. DRAINAGE | 4 | 1.0000 | 1.0000 | 1.0000 |  |
| 6.EROOTDEP | 4 | 5.0000 | 20.000 | 11.000 | 7.3485 |
| 7. ROOTDEP | 4 | 5.0000 | 20.000 | 11.000 | 7.3485 |
| 8. SOILDEP | 4 | 5.0000 | 20.000 | 11.000 | 7.3485 |
| 9.MATERIAL | 4 | 0 . | 0. | 0. |  |
| 10.RATIO1 | 4 | 1.0000 | 1.0000 | 1.0000 |  |
| 11. RATIO2 | 4 | 1.0000 | 1.0000 | 1.0000 |  |
| 12.RATIO3 | 4 | . 20000 | 1.0000 | . 41250 | . 39238 |
| 13.FIRE | 4 | 0. | 0. | 0 . |  |
| 14.WIND | 4 | 0. | 1.0000 | . 25000 | . 50000 |
| 15. WORMS | 4 | 0. | 0. | 0. |  |
| 16. ORGPHWAT | 4 | 3.8000 | 4.0000 | 3.9250 | . $95743-1$ |
| 17. ORGPHCAL | 4 | 3.4000 | 3.7000 | 3.5500 | . 17321 |
| 18. ORGTHICK | 4 | 1.0000 | 14.000 | 5.2500 | 6.1305 |
| 19. APHWAT | 1 | 3.8000 | 3.8000 | 3.8000 |  |
| 20. APHCAL | 1 | 3.2000 | 3.2000 | 3.2000 |  |
| 21. BIPHWAT | 3 | 4.2000 | 5.4000 | 4.7333 | . 61101 |
| 22.B4PHCAL | 3 | 3.7000 | 4.7000 | 4. 1000 | . 52915 |
| 23. COARSE\% | 4 | 0. | 75.000 | 27.500 | 33.292 |
| 24.texture | 3 | 2.0000 | 4.0000 | 3.0000 | 1.0000 |
| 25.E1\%N | 3 | . 26000 | . 36000 | . 30667 | . $50332-1$ |
| 26. B 1\%C | 3 | 6.4000 | 15.620 | 9.5900 | 5.2251 |
| 27.EICNRAT | 3 | 22.600 | 43.500 | 30.067 | 11.658 |
| 28.B2PHWAT | 0 |  |  |  |  |
| 29.B2PHCAL | 0 |  |  |  |  |
| 30.0RG\%C | 4 | 40.800 | 49.760 | 46.180 | 3.8826 |
| 31.0RG\% ${ }^{\text {N }}$ | 4 | . 48000 | . 63000 | . 55250 | . $66018-1$ |
| 32. ORGCNRAT | 4 | 70.300 | 100.10 | 84.800 | 15.227 |

Appendix 2 : Floodplain forests (FI)

DESCRIPTIVE MEASURES


## Appendix 2 : Floodplain forests (Lysichitum variant) (F2)



## Appendix 2 : Dry Pseudotsuga forests (P1)

DESCRIPTIVE MEASURES


Appendix 2 : Pseudotsuga-Thuja-Acer forests (P2)

DESCRIPTIVE MEASURES


DESCRIPTIVE MEASURES

| VARIABLE | $N$ | Minimum | MAXIMUM | MEAN | STD DEV |
| :---: | :---: | :---: | :---: | :---: | :---: |
| 1.ELEV | 5 | 200.00 | 590.00 | 324.00 | 157.26 |
| 2. ASPECT | 5 | 13.000 | 157.00 | 80.400 | 67.859 |
| 3. SLOPE | 5 | 15.000 | 60.000 | 37.800 | 17.880 |
| 4.POSIT | 5 | 2.0000 | 3.0000 | 2.8000 | . 44721 |
| 5. DRAINAGE | 5 | 2.0000 | 4.0000 | 2.6000 | . 89443 |
| $6 . E R O O T D E P$ | 5 | 15.000 | 99.000 | 55.400 | 40.955 |
| 7. ROOTDEP | 5 | 15.000 | 99.000 | 59.600 | 40.698 |
| 8.SOILDEP | 5 | 15.000 | 125.00 | 64.800 | 47.997 |
| 9.MATERIAL | 5 | 1.0000 | 3.0000 | 1.4000 | . 89443 |
| 10.RATIO! | 5 | .68000 | 1.0000 | . 93600 | . 14311 |
| 11.RATIO2 | 5 | . 68000 | 1.0000 | . 89400 | . 15027 |
| 12.ratio3 | 5 | .50000.-1 | 33000 | . 19000 | . 11979 |
| 13.FIRE | 5 | 1.0000 | 1.0000 | 1.0000 |  |
| 14.WIND | 5 | 0. | 0. | 0 . |  |
| 15. WORMS | 5 | 0. | 2.0000 | . 60000 | . 89443 |
| 16. ORGPHWAT | 5 | 4.0000 | 5.7000 | 4.9000 | . 73824 |
| 17. ORGPHCAL | 5 | 3.5000 | 5.4000 | 4.5400 | . 78294 |
| 18.ORGTHICK | 5 | 5.0000 | 11.000 | 6.8000 | 2.6833 |
| 19. APHWAT | 0 |  |  |  |  |
| 20. APHCAL | 0 |  |  |  |  |
| 21.81PHWAT | 5 | 4.6000 | 6.0000 | 5.2000 | . 68920 |
| 22.81PHCAL | 5 | 4.1000 | 5.3000 | 4.6000 | . 51962 |
| 23. COARSE\% | 5 | 40.000 | 77.000 | 56.400 | 13.686 |
| 24. TEXTURE | 5 | 3.0000 | 4.0000 | 3.6000 | . 54772 |
| $25 . \mathrm{B} 1 \% \mathrm{~N}$ | 5 | . $70000-1$ | . 20000 | . 15000 | . 54314 |
| 26.81\%C | 5 | 4.6000 | 14.440 | 8.0220 | 3.8233 |
| 27.B1CNRAT | 5 | 30.400 | 84.400 | 56.440 | 21.380 |
| 28.B2PHWAT | 3 | 4.9000 | 6.6000 | 5.8667 | . 87369 |
| 29.B2PHCAL | 3 | 4.3000 | 5.6000 | 5.1333 | . 72342 |
| 30.0RG\%C | 5 | 35.630 | 49.880 | 43.402 | 6.4640 |
| $31.0 R G \% N$ | 5 | . 77000 | 1.2900 | . 95000 | . 20087 |
| 32. ORGCNRAT | 5 | 36.300 | 64.800 | 47.260 | 12.608 |

## Appendix 2 : Pseudotsuga-Berberis forests (P4)

## descriptive measures

| VARIABLE | $N$ | MINIMUM | MAXIMUM | MEAN | Sto dev |
| :---: | :---: | :---: | :---: | :---: | :---: |
| 1. ELEV | 11 | 150.00 | 465.00 | 252.00 | 107.99 |
| 2. ASPECT | 11 | 37.000 | 173.00 | 108.27 | 51.219 |
| 3. SLOPE | 11 | 5.0000 | 60.000 | 40.909 | 18.987 |
| 4.POSIT | 11 | 3.0000 | 4.0000 | 3.1818 | . 40452 |
| 5. DRAINAGE | 11 | 1.0000 | 3.0000 | 2.2727 | . 64667 |
| 6. EROOTDEP | 11 | 26.000 | 117.00 | 67.455 | 29.784 |
| 7.ROOTDEP | 11 | 29.000 | 117.00 | 78.364 | 27.332 |
| 8. SOILDEP | 11 | 29.000 | 117.00 | 79.273 | 26.710 |
| 9. MATERIAL | 11 | 1.0000 | 3.0000 | 1.5455 | . 82020 |
| 10.RATIO1 | 11 | . 31000 | 1.0000 | . 86909 | . 21925 |
| 11. RATIO2 | 11 | . 31000 | 1.0000 | . 85545 | . 21491 |
| 12.R4TIO3 | 11 | . $30000-1$ | . 40000 | . 15818 | . 11957 |
| 13. FIRE | 11 | 0. | 1.0000 | . 90909 | . 30151 |
| 14.WIND | 11 | 0. | 0. | 0. |  |
| 15.WORMS | 11 | 0. | 2.0000 | . 63636 | . 92442 |
| 16. ORGPHWAT | 11 | 3.6000 | 5.6000 | 4. 1545 | . 62508 |
| 17. ORGPHCAL | 11 | 3.3000 | 5.3000 | 3.7364 | . 63918 |
| 18.ORGTHICK | 11 | 3.0000 | 23.000 | 8.6364 | 5.4639 |
| 19. APHWAT | 2 | 3.9000 | 4.8000 | 4.3500 | . 63640 |
| 20. APHCAL | 2 | 3.2000 | 4.0000 | 3.6000 | . 56569 |
| 21.B1PHWAT | 11 | 4.6000 | 5.4000 | 4.9727 | . 26492 |
| 22.B1PHCAL | 11 | 4.0000 | 4.8000 | 4.2909 | . 25082 |
| 23. COARSE\% | 11 | 50.000 | 84.000 | 71.636 | 10.828 |
| 24.texture | 11 | 2.0000 | 9.0000 | 3.8182 | 1.9400 |
| $25.81 \% \mathrm{~N}$ | 11 | . $40000-1$ | . 25000 | . 11636 | . 56617 -1 |
| 26.81\%C | 11 | 1.5600 | 6.7000 | 3.6164 | 1.7445 |
| 27.81CNRAT | 11 | 18.000 | 77.300 | 33.145 | 15.870 |
| 28.82PHWAT | 9 | 4.8000 | 5.8000 | 5. 1667 | . 36056 |
| 29. 22 PHCAL | 9 | 4.2000 | 5.0000 | 4.4778 | . 29059 |
| 30.0RG\%C | 11 | 27.010 | 51.800 | 44.104 | 7.5492 |
| 31.ORG\%N | 11 | . 54000 | 1.3200 | . 87273 | . 22055 |
| 32. ORGCNRAT | 11 | 37.500 | 82.300 | 52.473 | 12.859 |

Appendix 2 : Tsuga-Pseudotsuga-Polystichum forests (P5)

DESCRIPTIVE MEASURES


## Appendix 2 : Montane Tsuga forests (P6)

DESCRIPTIVE MEASURES

| VARIABLE | N | MINIMUM | MAXIMUM | MEAN | STD DEV |
| :---: | :---: | :---: | :---: | :---: | :---: |
| 1. ELEV | 12 | 102.00 | 795.00 | 486.00 | 195.36 |
| 2. ASPECT | 12 | 27.000 | 177.00 | 116.67 | 50.869 |
| 3. SLOPE | 12 | 14.000 | 75.000 | 52.083 | 19.374 |
| 4.POSIT | 12 | 2.0000 | 4.0000 | 3.0000 | . 42640 |
| 5. DRAINAGE | 12 | 1.0000 | 3.0000 | 2.4167 | .66856 |
| 6. EROOTDEP | 12 | 10.000 | 95.000 | 51.250 | 28.661 |
| 7. ROOTDEP | 12 | 43.000 | 111.00 | 75.667 | 20.169 |
| 8. SOILDEP | 12 | 43.000 | 120.00 | 85.583 | 26.919 |
| 9.MATERIAL | 12 | 1.0000 | 2.0000 | 1. 1667 | . 38925 |
| 10.RATIO1 | 12 | .15000 | 1.0000 | . 68083 | . 32520 |
| 11. RATIO2 | 12 | . 11000 | 1.0000 | . 64833 | . 35022 |
| 12.RATIO3 | 12 | $.50000-1$ | 1.0000 | . 31000 | .34351 |
| 13.FIRE | 12 | 0. | 1.0000 | . 91667 | . 28868 |
| 14.WIND | 12 | 0. | 0. | 0. |  |
| 15. WORMS | 12 | 0. | 3.0000 | 1.2500 | 1. 1382 |
| 16. ORGPHWAT | 12 | 3.5000 | 4.7000 | 3.8250 | . 37447 |
| 17.ORGPHCAL | 12 | 2.9000 | 4. 1000 | 3.3333 | . 39158 |
| 18.ORGTHICK | 12 | 4.0000 | 18.000 | 8.5000 | 4. 1010 |
| 19. APHWAT | 0 |  |  |  |  |
| 20. APHCAL | 0 |  |  |  |  |
| 21.B1PHWAT | 11 | 4.5000 | 5.1000 | 4.8182 | . 19400 |
| 22.B1PHCAL | 11 | 4.0000 | 4.5000 | 4.2727 | . 17373 |
| 23.COARSE\% | 12 | 20.000 | 90.000 | 53.750 | 21.440 |
| 24. TEXTURE | 11 | 2.0000 | 6.0000 | 3.5455 | 1.2933 |
| 25.B1\%N | 11 | . $50000-1$ | . 20000 | .11818 | . $44004-1$ |
| 26.E1\%C | 11 | 2.2400 | 7.1700 | 4.9145 | 1.6014 |
| 27. B1CNRAT | 11 | 27.000 | 94.300 | 45.882 | 22. 196 |
| 28. B2PHWAT | 10 | 5.0000 | 5.4000 | 5. 1500 | . 12693 |
| 29. B2PHCAL | 10 | 4.3000 | 5.2000 | 4.6100 | . 25582 |
| $30.0 R G \% C$ | 12 | 39.200 | 50.110 | 45.880 | 3.7412 |
| 31.0RG\%N | 12 | .48000 | 1.1800 | . 77667 | .19690 |
| 32. ORGCNRAT | 12 | 37.500 | 104.40 | 63.025 | 18.709 |

Appendix 2 : Montane Tsuga-Gaultheria forests (P7)
descriptive measures


## Appendix 2 : Coastal dry Thuja forests (Tl)

DESCRIPTIVE MEASURES

| VARIABLE | $N$ | MINIMUM | MAXIMUM | MEAN | STD DEV |
| :---: | :---: | :---: | :---: | :---: | :---: |
| 1.ELEV | 3 | 60.000 | 90.000 | 76.667 | 15.275 |
| 2. ASPECT | 3 | 59:000 | 137.00 | 107.33 | 42.218 |
| 3. SLOPE | 3 | 40.000 | 54.000 | 46.333 | 7.0946 |
| 4. POSIT | 3 | 1.0000 | 3.0000 | 1.6667 | 1. 1547 |
| 5. ORAINAGE | 3 | 1.0000 | 3.0000 | 1.6667 | 1.1547 |
| 6. EROOTDEP | 3 | 10.000 | 18.000 | 13.333 | 4.1633 |
| 7. ROOTDEP | 3 | .10 .000 | 27.000 | 16.333 | 9.2916 |
| 8. SOILDEP | 3 | 10.000 | 27.000 | 16.333 | 9.2916 |
| 9. MATERIAL | 3 | 0. | 2.0000 | . 66667 | 1. 1547 |
| 10. RAT 101 | 3 | .67000 | 1.0000 | . 89000 | . 19053 |
| 11. RATIO2 | 3 | .67000 | 1.0000 | . 89000 | . 19053 |
| 12. RATIO3 | 3 | . 17000 | 1.0000 | . 72333 | . 47920 |
| 13.FIRE | 3 | 0. | 0. | 0. |  |
| 14.WIND | 3 | 1.0000 | 1.0000 | 1.0000 |  |
| 15.WORMS | 3 | 0. | 2.0000 | 1.0000 | 1.0000 |
| 16. ORGPHWAT | 3 | 4.1000 | 4.5000 | 4.3000 | .20000 |
| 17. ORGPHCAL | 3 | 3.8000 | 4.1000 | 3.9333 | . 15275 |
| 18.ORGTHICK | 3 | 3.0000 | 12.000 | 8.3333 | 4.7258 |
| 19: APHWAT | 1 | 4.1000 | 4. 1000 | 4. 1000 |  |
| 20. APHCAL | 1 | 3.5000 | 3.5000 | 3.5000 |  |
| 21.EIPHWAT | 1 | 5.1000 | 5. 1000 | 5.1000 |  |
| 22.B1PHCAL | 1 | 4.6000 | 4.6000 | 4. 6000 |  |
| 23. COARSE\% | 3 | 0. | 10.000 | 3.3333 | 5.7735 |
| 24. TEXTURE | 0 |  |  |  |  |
| $25 . \mathrm{B} 1 \% \mathrm{~N}$ | 1 | . 13000 | . 13000 | 13000 |  |
| 26. $\mathrm{B}^{1 \%} \mathrm{C}$ | 1 | 4.3800 | 4.3800 | 4.3800 |  |
| 27.E1CNRAT | 1 | 32.400 | 32.400 | 32.400 |  |
| 28.82PHWAT | 0 |  |  |  |  |
| 29.B2PHCAL | 0 |  |  | : |  |
| 30. ORG\%C | 3 | 40.020 | 49.400 | 46.153 | 5.3147 |
| 31. ORG\%N | 3 | . 54000 | . 86000 | . 70667 | . 16042 |
| 32. DRGCNRAT | 3 | 57.000 | 74. 100 | 66.567 | 8.7295 |

Appendix 2 : Coastal Tsuga-Blechnum-Polystichum forests (T2)

DESCRIPTIVE MEASURES

| VARIABLE | N | MINIMUM | MAXIMUM | MEAN | STD DEV |
| :---: | :---: | :---: | :---: | :---: | :---: |
| 1. ELEV | 6 | 120.00 | 358.00 | 238.00 | 100.65 |
| 2. ASPECT | 6 | 29.000 | 157.00 | 80.500 | 50.425 |
| 3. SLOPE | 6 | 34.000 | 60.000 | 46.667 | 11.343 |
| 4.POSIT | 6 | 3.0000 | 3.0000 | 3.0000 |  |
| 5. DṘAINAGE | 6 | 3.0000 | 4.0000 | 3.5000 | . 54772 |
| 6. EROOTDEP | 6 | 6.0000 | 70.000 | 33.833 | 23.241 |
| 7. ROOTDEP | 6 | 26.000 | 118.00 | 59.833 | 32.474 |
| 8.SOILDEP | 6 | 26.000 | 158.00 | 92.333 | 53.377 |
| 9.MATERIAL | 6 | 1.0000 | 3.0000 | 1.5000 | . 83666 |
| 10.RATIO 1 | 6 | . 23000 | 1.0000 | . 61667 | . 41994 |
| 11. RATIO2 | 6 | . 18000 | 1.0000 | . 41167 | . 31301 |
| 12.RATIO3 | 6 | . 21000 | 1.0000 | . 46500 | . 28836 |
| 13. FIRE | 6 | 0. | 0. | 0 . |  |
| 14.WIND | $\epsilon$ | 0. | 1.0000 | . 66667 | . 51640 |
| 15. WORMS | 6 | 0. | 3.0000 | 1.6667 | 1.0328 |
| 16. ORGPHWAT | 6 | 3.5000 | 4.2000 | 3.8500 | . 23452 |
| 17. ORGPHCAL | 6 | 2.9000 | 3.8000 | 3.3167 | . 29269 |
| 18. ORGTHICK | 6 | 3.0000 | 40.000 | 14.667 | 13.501 |
| 19. APHWAT | 0 |  |  |  |  |
| 20. APHCAL | 0 |  |  |  |  |
| 21.BIPHWAT | 6 | 4.3000 | 4.9000 | 4.5333 | . 22509 |
| 22.B1PHCAL | 6 | 3.8000 | 4.3000 | 4.0333 | . 21602 |
| 23. COARSE\% | 6 | 10.000 | 60.000 | 43.333 | 20.656 |
| 24. TEXTURE | 6 | 4.0000 | 6.0000 | 5.0000 | 1.0954 |
| $25.81 \% N$ | 6 | . 17000 | . 86000 | . 37333 | . 24476 |
| 26.E1\%C | 6 | 5.6500 | 24.410 | 10.502 | 7:0714 |
| 27.B1CNRAT | 6 | 20.300 | 32.600 | 28.217 | 4.8139 |
| 28.E2PHWAT | 5 | 4.4000 | 5.0000 | 4.6600 | . 27019 |
| 29.82PHCAL | 5 | 4.0000 | 4.7000 | 4. 1800 | . 29496 |
| 30. ORG\%C | 6 | 41.200 | 51.950 | 47.207 | 4.2463 |
| $31.0 R G \% \mathrm{~N}$ | 6 | . 69000 | 1.4800 | 1.0650 | . 25665 |
| 32. ORGCNRAT | 6 | 27.800 | 74.900 | 47.400 | 16.017 |

Appendix 2 : Coastal montane Thuja forests (T3)

DESCRIPTIVE MEASURES


Appendix 2 : Coastal Thuja forests (T4)
descripitive measures

| VARIABLE | $N$ | MINIMUM | MAXIMUM | MEAN | STD DEV |
| :---: | :---: | :---: | :---: | :---: | :---: |
| 1. Elev | 19 | 15.000 | 380.00 | 115.11 | 90.941 |
| 2. ASPECT | 19 | 0 . | 169.00 | 81.526 | 53.127 |
| 3. SLOPE | 19 | 0. | 50.000 | 22.632 | 14.415 |
| 4.POSIT. | 19 | 2.0000 | 6.0000 | 3:6842 | 1. 1082 |
| 5. DRAINAGE | 19 | 3.0000 | 7.0000 | 4.8947 | 1. 2425 |
| 6.ERDOTDEP | 19 | 5.0000 | 60.000 | 30.105 | 15.051 |
| 7. RODTDEP | 19 | 20.000 | 100.00 | 55.211 | 24.503 |
| 8.SOILOEP - | 19 | 41.000 | 138.00 | 92.632 | 26.790 |
| 9.material | 19 | 1.0000 | 4.0000 | 2.3158 | . 74927 |
| 10.RATIO1 | 19 | .60000-1 | 1.0000 | . 62789 | . 29293 |
| 11.RATIO2 | 19 | . $40000-1$ | . 77000 | . 34737 | . 18113 |
| 12.RAT103 | 19 | . 31000 | 1.0000 | . 73421 | . 29615 |
| 13.FIRE | 19 | 0 . | 1.0000 | . 10526 | . 31530 |
| 14.WIND | 19 | 0. | 1.0000 | . 89474 | . 31530 |
| 15. WORMS | $\therefore 19$ | 0. | 3.0000 | 1.3158 | 1.0569 |
| 16. ORGPHWAT | 19 | 3.7000 | 4.8000 | 4. 1263 | . 37095 |
| 17. ORGPHCAL | 19 | 3. 1000 | 4.4000 | 3.6105 | . 37401 |
| 18.ORGTHICK | 19 | 3.0000 | 43.000 | 19.842 | 9.4473 |
| 19. APHWAT | 9 | 3.8000 | 5.4000 | 4.2222 | . 48419 |
| 20. APHCAL | 9 | 3.1000 | 4.6000 | 3.6556 | . 40961 |
| 21. B1PHWAT | 19 | 4.2000 | 6.0000 | 4.7105 | . 40537 |
| 22.B1PHCAL | 19 | 3.6000 | 5.4000 | 4. 1632 | . 41394 |
| 23. COARSE\% | 19 | 0. | 95.000 | 42.053 | 28.415 |
| 24.texture | 19 | 2.0000 | 12.000 | 4.5789 | 2. 1938 |
| $25 . \mathrm{B} \% \mathrm{~N}$ | 19 | . $20000-1$ | . 32000 | . 20316 | . 81789 |
| $26 . \mathrm{B} 1 \% \mathrm{C}$ | 19 | . 18000 | 13.400 | 6.4284 | 3.1632 |
| 27.81 CNRAT | 19 | 10.000 | 44.200 | 30.700 | 8.0325 |
| $28.82 P H W A T$ | . 17 | 4.5000 | 5.3000 | 4.9176 | . 24808 |
| 29. ${ }^{\text {2 PPHCAL }}$ | 17 | 3.8000 | 5.2000 | 4.4471 | . 41400 |
| 30.ORG\%C | 19 | 32.300 | 49.100 | 43.278 | 5.1703 |
| 31.ORG\% | 19 | . 67000 | 1.3000 | 1.0347 | . 17302 |
| $32.0 R G C N R A T$ | 19 | 33.900 | 56.300 | 42.642 | 6.9708 |

Appendix 2 : Coastal wet Thuja forests (T5)

DESCRIPTIVE MEASURES


Appendix 2 : Montane Tsuga-Abies-Gaultheria forests (Al)

DESCRIPTIVE MEASURES

| VARIABLE | N MINIMUM | MAXIMUM | MEAN | STD DEV |  |
| :--- | :---: | :---: | :---: | :---: | :---: |
| 1.ELEV | 3 | 238.00 | 670.00 | 457.00 | 216.06 |
| 2.ASPECT | 3 | 125.00 | 172.00 | 149.00 | 23.516 |
| 3.SLOPE | 3 | 15.000 | 65.000 | 44.000 | 25.942 |
| 4.POSIT | 3 | 3 | 2.0000 | 3.0000 | 2.6667 |

## Appendix 2 : Montane Abies-Tsuga forests (A2)

DESCRIPTIVE MEASURES

| VARIABLE | N | MINIMUM | MAXIMUM | MEAN | STO DEV |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 1.ELEV | 2 | 497.00 | 650.00 | 573.50 | 108. 19 |  |
| 2.ASPECT | 2 | 42.000 | 147.00 | 94.500 | 74.246 | 4 |
| 3. SLOPE | 2 | 60.000 | 65.000 | 62.500 | 3.5355 | \% |
| 4.POSIT | 2 | 3.0000 | 3.0000 | 3.0000 |  | \% |
| 5. DRAINAGE | 2 | 4.0000 | 5.0000 | 4.5000 | . 70711 | \% |
| 6. EROOTDEP | 2 | 16.000 | 17.000 | 16.500 | . 70711 |  |
| 7. ROOTDEP | 2 | 19.000 | 55.000 | 37.000 | 25.456 |  |
| 8. SOILDEP | 2 | $106.00$ | 125.00 | 115.50 | 13.435 |  |
| 9.MATERIAL | 2 | 1.0000 | 2.0000 | 1.5000 | . 70711 |  |
| 10.RATIO1 | 2 | . 31000 | . 84000 | . 57500 | . 37477 |  |
| 11. RATIO2 | 2 | . 14000 | . 15000 | . 14500 | . 70711 -2 |  |
| 12.RATIO3 | 2 | . 71000 | 1.0000 | . 85500 | . 20506 |  |
| 13.FIRE | 2 | 0. | 1.0000 | . 50000 | . 70711 | , |
| 14.WIND | 2 | 0. | 1.0000 | . 50000 | . 70711 |  |
| 15. WORMS | 2 | 0. | 3.0000 | 1. 5000 | 2. 1213 |  |
| 16. ORGPHWAT | 2 | 3.4000 | 3.7000 | 3.5500 | . 21213 | \% |
| 17. ORGPHCAL | 2 | 2.9000 | 3. 1000 | 3.0000 | . 14142 |  |
| 18.ORGTHICK | 2 | 9.0000 | 12.000 | 10.500 | 2. 1213 |  |
| 19. APHWAT | 2 | 3.7000 | 3.8000 | 3.7500 | . 70711 - 1 |  |
| 20. APMCAL | 2 | 3.1000 | 3.3000 | 3.2000 | . 14142 |  |
| 21.E!PHWAT | 2 | 4.4000 | 4. 5000 | 4.4500 | .70711-1 |  |
| 22.B1PHCAL | 2 | 4.0000 | 4.1000 | 4.0500 | . 70711 -1 |  |
| 23.COARSE\% | 2 | 50.000 | 75.000 | 62.500 | 17.678 | ; |
| 24.TEXTURE | 2 | 3.0000 | 6.0000 | 4.5000 | 2.1213 |  |
| $25 . \mathrm{B} 1 \% \mathrm{~N}$ | 2 | . 22000 | . 23000 | . 22500 | . 70711 -2 |  |
| 26.B1\%C | 2 | 8.5300 | 8.5400 | 8.5350 | . 70711 -2 |  |
| 27. BicNRAT $^{\text {I }}$ | 2 | 37.800 | 39.300 | 38.550 | 1.0607 |  |
| 28.B2PHWAT | 2 | 4.8000 | 5.2000 | 5.0000 | . 28284 |  |
| 29.92PHCAL | 2 | 4.2000 | 5.0000 | 4.6000 | . 56569 |  |
| 3). ORG\%C | 2 | 48.480 | 51.510 | 49.995 | 2. 1425 |  |
| 31.ORG\%N | 2 | . 86000 | 1. 1600 | 1.0100 | . 21213 |  |
| 32. ORGCNRAT | 2 | 41.800 | 59.900 | 50.850 | 12.799 |  |

Appendix 2 : Montane Tsuga-Abies forests (A3)

DESCRIPTIVE MEASURES


Appendix 2 : Montane Abies-Streptopus forests (A4)

| VVARIABLE | $N$ | MINIMUM | MAXIMUM | MEAN | STD DEV |
| :---: | :---: | :---: | :---: | :---: | :---: |
| 1.ELEV | 4 | 580.00 | 710.00 | 672.50 | 61.847 |
| 2.ASPECT | 4 | 12.000 | 67.000 | 43.250 | 24.281 |
| 3.SLOPE | 4 | 40.000 | 45.000 | 42.250 | 2.6300 |
| 4.POSIT | 4 | 2.0000 | 3.0000 | 2.5000 | . 57735 |
| 5. DRA INAGE | 4 | 3.0000 | 5.0000 | 4.5000 | 1.0000 |
| 6. EROOTDEP | 4 | 16.000 | 103.00 | 41.750 | 41.040 |
| 7. ROOTDEP | 4 | 33.000 | 103.00 | 67.250 | 33.130 |
| 8. SOILDEP | 4 | 69.000 | 127.00 | 93.000 | 27.653 |
| 9.MATERIAL | 4 | 1.0000 | 1.0000 | 1.0000 |  |
| 10.RATIO9 | 4 | . 30000 | 1.0000 | . 58000 | . 32445 |
| 11. RATIO2 | 4 | . 22000 | . 81000 | . 40000 | . 27653 |
| 12.RATIO3 | 4 | . 26000 | 1.0000 | . 74750 | . 34903 |
| 13.FIRE | 4 | 0. | 0. | 0. |  |
| 14.WIND | 4 | 0. | 1.0000 | . 50000 | . 57735 |
| 15. WORMS | 4 | 0. | 2.0000 | . 50000 | 1.0000 |
| 16. ORGPHWAT | 4 | 3.0000 | 3.6000 | 3.2500 | . 26458 |
| 17. ORGPHCAL | 4 | 2.4000 | 3.0000 | 2.7250 | . 27538 |
| 18.ORGTHICK | 4 | 16.000 | 27.000 | 21.000 | 4.6904 |
| 19. APHWAT | 2 | 3.6000 | 3.7000 | 3.6500 | .70719-1 |
| 20. $\triangle P H C A L$ | 2 | 3.0000 | 3.0000 | 3.0000 |  |
| 21.B1PHWAT | 4 | 4.0000 | 4.8000 | 4.3500 | .. 36968 |
| 22.BIPHCAL | 4 | 3.6000 | 4.5000 | 3.9250 | . 42720 |
| 23. COARSE\% | 4 | 5.0000 | 50.000 | 26.250 | 22.127 |
| 24.TEXTURE | 4 | 3.0000 | 6.0000 | 4.5000 | 1.7321 |
| $25.81 \% \mathrm{~N}$ | 4 | . $80000-9$ | . 29000 | . 16750 | . $88459-1$ |
| 26. B 1\% C | 4 | 2.8000 | 12.670 | 5.3350 | 4.8905 |
| 27. BICNRAT | 4 | 17.700 | 43.700 | 29.200 | 12.417 |
| 28.B2PHWAT | 4 | 4.4000 | 5.4000 | 4.7750 | . 47871 |
| 29.B2PHCAL | 4 | 3.8000 | 5.1000 | 4.3000 | . 55976 |
| 30.0RG\%C | 4 | 49.240 | 50.600 | 49.995 | . 59248 |
| 31.ORG\% | 4 | . 82000 | 1.6800 | 1. 1800 | . 41817 |
| 32. ORGCNRAT | 4 | 30.100 | 60.000 | 46.350 | 15.205 |

Appendix 2 : Lowland Abies forests (A5)
descriptive measures

| VARIABLE | $N$ | MINIMUM | MAXIMUM | MEAN | STD DEV |
| :---: | :---: | :---: | :---: | :---: | :---: |
| 1. ELEV | 12 | 55.000 | 415.00 | 194.67 | 116.39 |
| 2. ASPECT | 12 | 0. | 157.00 | 53.417 | 53.241 |
| 3. SLOPE | $\therefore 12$ | 3.0000 | 50.000 | 19.833 | 15.379 |
| 4.POSIT | 12 | 4.0000 | 5.0000 | 4.2500 | . 45227 |
| 5. DRAINAGE | 12 | 2.0000 | 5.0000 | 3. 6667 | . 88763 |
| 6. EROOTDEP | 12 | 10.000 | 56.000 | 32.000 | 16.586 |
| 7. ROOTOEP | 12 | 26.000 | 100.00 | 60.417 | 21.082 |
| 8. SOILDEP | 12 | 51.000 | 141.00 | 94.917 | 22.952 |
| 9. MATERIAL. | 12 | 1.0000 | 3.0000 | 2.3333 | . 77850 |
| 10.RATIO1 | 12 | . 24000 | 1.0000 | .54750 | . 27367 |
| 11. RATIO2 | 12 | $.90000-1$ | 1.0000 | . 36417 | . 25239 |
| 12. RATIO3 | 12 | .26000 | 1.0000 | . 58250 | . 31606 |
| 13. FIRE | 12 | 0 . | 1.0000 | . 16667 | . 38925 |
| 14.WIND | 12 | 0. | 1.0000 | . 41667 | . 51493 |
| 15. WORMS | 12 | 0. | 2.0000 | 1.0833 | . 99620 |
| 16. ORGPHWAT | 12 | 3.4000 | 4.9000 | 3.7917 | . 50355 |
| 17. ORGPHCAL | 12 | 2.8000 | 4.5000 | 3. 2083 | . 52477 |
| 18.ORGTHICK | 12 | 8.0000 | 25.000 | 14.750 | 4.8077 |
| 19. APHWAT | 8 | 3.5000 | 5.1000 | 3.9750 | .49497 |
| 20. APHCAL | 8 | 3.0000 | 4.3000 | 3.3500 | . 41057 |
| 21.EIPHWAT | 12 | 3.8000 | 5. 1000 | 4.5833 | . 36139 |
| 22.B1PHCAL | 12 | 3.6000 | 4.7000 | 4. 1333 | . 30251 |
| 23.COARSE\% | 12 | 10.000 | 95.000 | 42.500 | 27.593 |
| 24. TEXTURE | 12 | 3.0000 | 6.0000 | 4.5833 | 1.5050 |
| 25. $\mathrm{E} 1 \% \mathrm{~N}$ | 12 | $.90000-1$ | . 28000 | .20333 | . 65273 |
| 26.E1\%C | 12 | 2.4000 | 8.5800 | 5.4100 | 1.9536 |
| 27.8 ICNRAT | 12 | 18.700 | 37.200 | 26.733 | 5.7299 |
| 28.E2PHWAT | 11 | 4.7000 | 5.5000 | 5.0455 | . 23817 |
| 29, E2PHCAL | 11 | 4.1000 | 5.2000 | 4.5455 | . 30778 |
| $30.0 R G \% C$ | 12 | 36.300 | 53.300 | 46.277 | 6.3396 |
| 31. ORG\%N | 12 | .75000 | 1.6300 | 1.1725 | . 29815 |
| 32.ORGCNRAT | 12 | 23.500 | 53.000 | 41.283 | 8.4671 |

## Appendix 2 : Tsuga-Gaultheria-Blechnum forests (A6)

DESCRIPTIVE MEASURES

| VARIABLE | N | MINIMUM | MAXIMUM | MEAN | STD DEV |
| :---: | :---: | :---: | :---: | :---: | :---: |
| 1.ELEV | 2 | 125.00 | 200.00 | 162.50 | 53.033 |
| 2.ASPECT | 2 | 12.000 | 103.00 | 57.500 | 64.347 |
| 3. SLOPE | 2 | 27.000 | 30.000 | 28.500 | 2. 1213 |
| 4. POSIT | 2 | 3.0000 | 4.0000 | 3.5000 | . 70711 |
| 5. Dralnage | 2 | 4.0000 | 4.0000 | 4.0000 |  |
| 6. EROOTDEP | 2 | 31.000 | 47.000 | 39.000 | 11.314 |
| 7 7. ROOTDEP | 2 | 69.000 | 72.000 | 70.500 | 2. 1213 |
| 8. SOILDEP | 2 | 89.000 | 117.00 | 103.00 | 19.799 |
| 9. Material | 2 | 2.0000 | 2.0000 | 2.0000 |  |
| 10.RATIO1 | 2 | . 45000 | . 65000 | . 55000 | . 14142 |
| 11.RATIO2 | 2 | . 35000 | . 40000 | . 37500 | . $35355-1$ |
| 12. RATIO3 | 2 | . 25000 | . 35000 | . 30000 | . 70711 -1 |
| 13.FIRE | 2 | 0 . | 0 . | 0 . |  |
| 14.WIND | 2 | 0. | 1.0000 | . 50000 | . 70711 |
| 15. WORMS | 2 | 0. | 2.0000 | 1.0000 | 1.4142 |
| 16. ORGPHWAT | 2 | 3.5000 | 3.6000 | 3.5500 | .70711-1 |
| 17. ORGPHCAL | 2 | 2.9000 | 3.0000 | 2.9500 | . $70711-1$ |
| 18. ORGTHICK | 2 | 11.000 | 12.000 | 11.500 | .70711 |
| 19. APHWAT | 1 | 3.7000 | 3.7000 | 3.7000 |  |
| 20. APHCAL | 1 | 3.2000 | 3.2000 | 3.2000 |  |
| 21.E1PHWAT | 2 | 4.3000 | 5.1000 | 4.7000 | . 56569 |
| 22.B1PHCAL | 2 | 4. 1000 | 4.7000 | 4.4000 | . 42426 |
| 23.COARSE\% | 2 | 30.000 | 40.000 | 35.000 | 7.0711 |
| 24. TEXTURE | 2 | 3.0000 | 3.0000 | 3.0000 |  |
| 25. $\mathrm{B}^{1 \% \mathrm{~N}}$ | 2 | . 16000 | . 19000 | .17500 | . $21213-1$ |
| 26.B1\%C | 2 | 3.8500 | 6.1900 | 5.0200 | 1.6546 |
| 27.EICNRAT | 2 | 23.800 | 32.700 | 28.250 | 6.2933 |
| 28.B2PHWAT | 2 | 4.6000 | 4.8000 | 4.7000 | $.14142$ |
| 29.B2PHCAL | 2 | 4.2000 | 4.5000 | 4.3500 | . 21213 |
| 30.0RG\%C | 2 | 38.740 | 44.290 | 41.515 | 3.9244 |
| 31.0RG\%N | 2 | 1.0500 | 1.3400 | 1. 1950 | . 20506 |
| 32. ORGCNRAT | 2 | 33.000 | 36.900 | 34.950 | 2.7577 |

Appendix 2 : Tsuga-Blechnum-Polystichum forests (A7)

| VARIABLE | $N$ | MINIMUM | MAXIMUM | MEAN | STD DEV |
| :---: | :---: | :---: | :---: | :---: | :---: |
| 1.ELEV | 5 | 25.000 | 350.00 | 183.00 | 123.98 |
| 2. ASPECT | 5 | 15.000 | 72.000 | 44.800 | 26.376 |
| 3. Slope | 5 | 20.000 | 75.000 | 40.600 | 20.477 |
| 4.POSIT | 5 | 3.0000 | 4.0000 | 3.6000 | . 54772 |
| 5. DRAINAGE | 5 | 2.0000 | 4.0000 | 3.2000 | . 83666 |
| 6. EROOTDEP | 5 | 5.0000 | 45.000 | 22.800 | 17.922 |
| 7. RODTAEP | 5 | 25.000 | 100.00 | 51.000 | 32.581 |
| 8. SOILDEP | 5 | 82.000 | 113.00 | 100.40 | 12.857 |
| 9.MATERIAL | 5 | 1.0000 | 2.0000 | 1.6000 | . 54772 |
| 10.RAT101 | 5 | . 17000 | . 65000 | 42200 | . 17880 |
| 11. RATIO2 | 5 | . 50000 | -1.40000 | . 22200 | . 16208 |
| 12.RATIO3 | 5 | . 41000 | 1.0000 | . 76600 | . 32044 |
| 13.FIRE | 5 | 0. | 1.0000 | 40000 | . 54772 |
| 14.WIND | 5 | 0. | 1.0000 | 20000 | . 4472 |
| 15. WORMS | 5 | 1.0000 | 3.0000 | 2.4000 | . 89443 |
| 16. ORGPHWAT | 5 | 3.6000 | 4.0000 | 3.7600 | . 15166 |
| 17. ORGPHCAL | 5 | 2.9000 | 3.6000 | 3.1800 | . 25884 |
| 18. ORGTHICK | 5 | 5.0000 | 19.000 | 13.000 | 5.2440 |
| 19. APHWAT | 0 |  |  |  |  |
| 20.APHCAL | 0 |  |  |  |  |
| 21.BIPHWAT | 5 | 4.5000 | 5.2000 | 4.8600 | . 35071 |
| 22.B1PHCAL | 5 | 3.8000 | 5.0000 | 4.3400 | . 49800 |
| 23. COARSE\% | 5 | 30.000 | 75.000 | 59.000 | 18.84 |
| 24.texture | 5 | 3.0000 | 6.0000 | 4.6000 | 1.3416 |
| $25 . \mathrm{B} 1 \% \mathrm{~N}$ | 5 | . 17000 | . 36000 | 26000 | . 82158 |
| $26.81 \% \mathrm{C}$ | 5 | 2.8000 | 10.230 | 7.1420 | 2.9069 |
| 27. BICNRAT | 5 | 16.000 | 51.100 | 27.960 | 13.504 |
| 28.B2PHWAT | 5 | 4.8000 | 5.3000 | 5.1800 | . 21679 |
| 29. B2PHCAL | 5 | 4.3000 | 5.3000 | 4.7400 | . 37815 |
| 30.0 G\% $\%$ | 5 | 38.900 | 49.480 | 43.300 | 4.1590 |
| 31. ORG\%N | 5 | . 86000 | 1.5200 | 1.0680 | . 26621 |
| 32. ORGCNRAT | 5 | 25.600 | 57.500 | 42.680 | 11.539 |

Appendix 3 : Community types complete understory vegetation tables.

```
cons. = constancy (%)
m. cov. = mean coverage (%)
max. cov. = maximum coverage (%)
m. fre. = mean frequency (%)
m.i.v. = mean importance value (%)
        (i.v. = [relative cov.+ relative fre.]/2)
t. spp = total number of species
m. spp = mean number of species
```

(Refer to Appendix 1 for abbreviations of species names)

Appendix 3 : Dry Pinus-Pseudotsuga forests (DI) plots: 9110144

|  | cons. | m.cov | $\max . \operatorname{cov}$ | m.fre. | m.1.v |
| :---: | :---: | :---: | :---: | :---: | :---: |
| shrubs |  |  |  |  |  |
| amal | 66 | 0.007 | 0.01 | 0.67 | 0.033 |
| arco | 100 | 7.047 | 10.63 | 15.33 | 3.013 |
| bene | 100 | 4.733 | 8.52 | 20.00 | 2.540 |
| gash | 100 | 12.850 | 17.90 | 33.33 | 5.760 |
| hodi | 33 | 0.003 | 0.01 | 0.33 | 0.017 |
| pamy | 33 | 0.750 | 2.25 | 5.00 | 0.457 |
| rogy | 100 | 0.307 | 0.90 | 4.00 | 0.273 |
| ruur | 100 | 1.210 | 1.35 | 21.67 | 1.443 |
| sali | 33 | 0.003 | 0.01 | 0.33 | 0.017 |
| vame | 33 | 0.003 | 0.01 | 0.33 | 0.017 |
| vaot | 66 | 17.800 | 37.00 | 30.00 | 7.573 |
| vapa | 100 | 0.010 | 0.01 | 1.00 | 0.050 |
| neros |  |  |  |  |  |
| achi | 66 | 0.013 | 0.03 | 2.00 | 0.100 |
| actr | 33 | 0.003 | 0.01 | 0.33 | 0.017 |
| agal | 33 | 0.003 | 0.01 | 0.33 | 0.017 |
| agsc | 33 | 0.060 | 0.18 | 3.33 | 0.183 |
| anne | 33 | 0.050 | 0.15 | 1.67 | 0.090 |
| apan | 66 | 0.527 | 0.90 | 15.00 | 0.887 |
| aren | 33 | 0.003 | 0.01 | 0.33 | 0.017 |
| aruv | 33 | 10.627 | 31.88 | 23.33 | 4.360 |
| chum | 33 | 0.050 | 0.15 | 1.67 | 0.090 |
| crer | 66 | 0.020 | 0.05 | 3.67 | 0.170 |
| dasp | 100 | 4.073 | 11.93 | 38.67 | 3.030 |
| erla | 33 | 0.003 | 0.01 | 0.33 | 0.017 |
| feoc | 66 | 0.260 | 0.75 | 3.33 | 0.247 |
| feov | 66 | 1.000 | 2.97 | 11.67 | 0.933 |
| frag | 66 | 0.383 | 0.93 | 11.67 | 0.667 |
| goob | 100 | 0.160 | 0.18 | 6.67 | 0.380 |
| hial | 100 | 0.333 | 0.77 | 16.67 | 0.873 |
| nypa | 66 | 0.007 | 0.01 | 0.67 | 0.033 |
| 1 iao | 33 | 0.017 | 0.05 | 3.33 | 0.170 |
| 1 ibo | 33 | 1.000 | 3.00 | 6.67 | 0.607 |
| lica | 33 | 0.017 | 0.05 | 3.33 | 0.170 |
| 1 ico | 66 | 0.220 | 0.63 | 10.00 | 0.517 |
| luzc | 33 | 0.050 | 0.15 | 1.67 | 0.100 |
| poly | 66 | 0.013 | 0.03 | 2.00 | 0.103 |
| pomu | 66 | 0.007 | 0.01 | 0.67 | 0.033 |
| prun | 33 | 0.003 | 0.01 | 0.33 | 0.017 |
| saxf | 66 | 0.060 | 0.15 | 3.33 | 0.177 |
| sewa | 100 | 4.727 | 7.40 | 38.33 | 3.433 |
| trla | 66 | 0.210 | 0.45 | 8.33 | 0.457 |
| trma | 33 | 0.500 | 1.50 | 10.00 | 0.597 |
| vise | 33 | 0.003 | 0.01 | 0.33 | 0.017 |
| zyve | 33 | 0.010 | 0.03 | 1.67 | 0.077 |
| bryophytes and 1ichens |  |  |  |  |  |
| clar | 100 | 5.527 | 15.48 | 30.00 | 3. 113 |
| clas | 66 | 1.253 | 3.75 | 3.67 | 0.600 |
| cldb | 100 | 0.710 | 1.08 | 50.00 | 2.610 |
| cldf | 100 | 2.793 | 7.25 | 33.33 | 2.583 |
| cldg | 66 | 1.170 | 2.38 | 36.67 | 2.130 |
| cldp | 33 | 0.100 | 0.30 | 20.00 | 1.053 |


| cldu | 33 | 0.003 | 0.01 | 0.33 | 0.017 |
| :---: | :---: | :---: | :---: | :---: | :---: |
| cleb | 33 | 0.127 | 0.38 | 8.33 | 0.450 |
| dicf | 33 | 0.227 | 0.68 | 11.67 | 0.647 |
| dies | 66 | 10.260 | 20.63 | 60.00 | 6.267 |
| hylo | 100 | 2.117 | 3.55 | 15.00 | 1. 393 |
| isop | 33 | 0.060 | 0. 18 | 3.33 | 0.190 |
| isst | 33 | 1. 760 | 5.28 | 20.00 | 1.620 |
| pelo | 100 | 1.143 | 1.75 | 40.00 | 2.300 |
| pelt | 33 | 0.067 | 0.20 | 5.00 | 0.277 |
| plun | 33 | 0.050 | 0.15 | 1.67 | 0.103 |
| plzs | 33 | 3.100 | 9.30 | 10.00 | 1.413 |
| ponc | 100 | 0.900 | 1.65 | 10.00 | 0.783 |
| ponj | 100 | 3.810 | 5.13 | 78.33 | 5.003 |
| ponp | 33 | 0.450 | 1.35 | 23.33 | 1.297 |
| rhac | 100 | 3.217 | 6.55 | 25.00 | 2.250 |
| rhan | 33 | 0.710 | 2.13 | 11.67 | 0.837 |
| rhal | 33 | 7.900 | 23.70 | 23.33 | 3.797 |
| rhlo | 66 | 0.110 | 0.18 | 5.00 | 0.280 |
| rhtr | 66 | 0.677 | 1.88 | 3.33 | 0.393 |
| rhyt | 33 | 0.050 | 0.15 | 1.67 | 0.100 |
| scaa | 66 | 0.390 | 0.95 | 20.00 | 1. 140 |
| steo | 33 | 0.033 | 0.10 | 6.67 | 0.350 |
| stet | 66 | 0.727 | 2.00 | 23.33 | 1.317 |
| stor | 100 | 4.153 | 5.93 | 23.33 | 2.567 |
| tame | 33 | 0.777 | 2.33 | 6.67 | 0.537 |
| bare rock |  |  |  |  |  |
| rock | 100 | 27.937 | 38.50 | 71.67 | 0.000 |
| totals: |  |  |  |  |  |
|  |  | m.cov max.cov |  | t.spp m.spp |  |
| shrubs |  | 44.73 | 50.6 | 12 | 8.7 |
| herbs |  | 24.40 | 55.0 | 32 | 17.3 |
| bryo. |  | 54.37 | 59.0 | 31 | 18.7 |
| all spp. |  | 123.43 | 147.7 | 75 | 44.7 |

```
Note : Pelo = Peltigera leucophlebia (9, 100)
                and \(P\). aphtosa (144)
    Pelt \(=\underline{p}\). membranacea
```





| stac | 37 | 0.709 | 5.63 | 4.50 | 0.393 |
| :---: | :---: | :---: | :---: | :---: | :---: |
| stra | 75 | 0.294 | 1.88 | 2.75 | 0.209 |
| strr | 37 | 0.563 | 31.95 | 6.25 | $\therefore 0.465$ |
| tila | 62 | 0.550 | 3.17 | 11.88 | 0.630 |
| titr | 100 | 8.799 | 27.65 | 55.63 | 5 5.039 |
| tome | 25 | 1.369 | 10.80 | 5.63 | . 0.456 |
| trau | 100 | 9.890 | 36.13 | 42.50 | - 5.069 |
| trla | 25 | 0.005 | 0.03 | 0.75 | -0.038 |
| trmc | 25 | 0.005 | 50.03 | 0.75 | $\because 0.043$ |
| trov | 75 | 1.501 | 15.47 | 20.88 | 1.440 |
| vevi | 37 | 0.300 | 0.90 | 2.50 | 0.240 |
| vigl | 50 | 1.070 | - 7.45 | 10.75 | 0.625 |
| vise | 12 | 0.004 | 40.03 | 0.63 | 0.054 |
| oryoph | S | 11 chen | ens |  |  |
| bazz | 12 | 0.329 | 2.63 | 1.25 | 0.209 |
| ceph | 12 | 0.038 | 80.30 | 1.25 | 0.065 |
| coco | 37 | 0.647 | 4.9 .97 | 10.00 | 0.496 |
| dicf | 12 | 0.040 | 0.32 | 1.88 | 0.171 |
| dics | 12 | 0.019 | 0.15 | 0.63 | 0.026 |
| dipa | 12 | 0.023 | O. 0.18 | 1.25 | 0.112 |
| nete | 25 | 0.131 | 10.90 | 1.88 | 0.153 |
| nolu | 62 | 0.445 | 2.55 | 15.63 | 0.980 |
| hylo | 87 | 1.985 | 8.88 | 13.75 | 1.274 |
| hypu | 37 | 0.135 | 0.75 | 2.50 | 0.191 |
| 1sop | 50 | 0.629 | 3.05 | 10.00 | 0.730 |
| isst | 75 | 1.414 | 4.53 | 8.13 | 0.845 |
| ledo | 12 | 0.075 | 5 0.60 | 2.50 | 0.160 |
| leme | 75 | 5.810 | - 33.22 | 28.13 | 2.656 |
| peli | 50 | 0.266 | -1.58 | 10.00 | 0.581 |
| plag | 62 | 0.316 | -1.08 | 16.88 | 0.914 |
| plin | 75 | 2.169 | - 9.27 | 30.00 | 1.910 |
| plun | 62 | 2.006 | -8.75 | 31.88 | 2.191 |
| poge | 12 | 0. 112 | 20.90 | 1.25 | 0.095 |
| pogm | 12 | 0.188 | 1.50 | 1.25 | 0.165 |
| rigil | 100 | 2.114 | 4.80 | 36.25 | 2.365 |
| rhlo | 100 | 4.720 | 15.88 | 35.00 | 2.996 |
| ricl | 12 | 0.038 | -0.30 | 1.25 | 0.051 |
| scab | 87 | 0.498 | 80.93 | 8.13 | 0.553 |
| stor | 62 | 8.920 | 28.58 | 41.88 | 3.935 |
| stpr | 50 | 6.046 | -23.20 | 36.88 | 4. 169 |
| bare rock |  |  |  |  |  |
| rock | 12 | 1.016 | 8. 13 | 1.25 | 0.000 |
| totals: |  |  |  |  |  |
|  |  | m.cov max.cov |  | t.spp | m.spp |
| shrubs |  | 46.20 | 131.9 | 16 | 5.9 |
| herbs |  | 118.80 | 168.6 | 492 | 20.8 |
| bryo. |  | 39.11 | 86.2 | 26 | 12.1 |
| all |  | 204.10 | 278.1 | 91 | 38.1 |

Appendix 3 : Floodplain forests (Lysichitum variant) (F2)
plots: 4651

|  | cons. | m.cov | max.cov | m.fre. | m.i.v |
| :--- | ---: | ---: | ---: | ---: | ---: |
| shrubs |  |  |  |  |  |
| gash | 100 | 33.335 | 50.65 | 65.00 | 9.535 |
| mefe | 100 | 5.015 | 6.63 | 20.00 | 1.860 |
| rusp | 100 | 20.895 | 21.77 | 70.00 | 7.235 |
| vaal | 100 | 7.315 | 7.75 | 17.50 | 2.225 |
| vaol | 100 | 4.030 | 4.78 | 15.00 | 1.440 |
| vaot | 100 | 2.095 | 4.18 | 10.50 | 0.885 |
| vapa | 100 | 13.110 | 14.15 | 40.00 | 4.340 |


| herbs |  |  |  |  |  |
| :---: | ---: | ---: | ---: | ---: | ---: |
| adbi | 50 | 1.175 | 2.35 | 12.50 | 0.785 |
| atfi | 100 | 1.500 | 2.25 | 10.00 | 0.710 |
| blsp | 100 | 28.825 | 35.90 | 72.50 | 8.895 |
| boye | 100 | 0.010 | 0.01 | 1.00 | 0.045 |
| caro | 100 | 2.880 | 5.75 | 8.00 | 0.935 |
| fesa | 50 | 0.075 | 0.15 | 2.50 | 0.125 |
| gali | 50 | 0.100 | 0.20 | 7.50 | 0.345 |
| luzp | 50 | 0.015 | 0.03 | 2.50 | 0.110 |
| lysi | 100 | 26.125 | 46.03 | 60.00 | 7.860 |
| madi | 100 | 1.310 | 1.67 | 40.00 | 1.915 |
| pomu | 100 | 23.325 | 25.90 | 37.50 | 6.380 |
| stra | 100 | 0.525 | 0.90 | 7.50 | 0.410 |
| tila | 100 | 2.510 | 3.35 | 60.00 | 2.980 |
| titr | 100 | 5.260 | 5.57 | 65.00 | 3.760 |
| trmc | 100 | 0.230 | 0.45 | 8.00 | 0.390 |
| vevi | 50 | 0.005 | 0.01 | 0.50 | 0.025 |
| vigi | 100 | 0.240 | 0.47 | 10.50 | 0.505 |


| bryophytes and | 1 ichens |  |  |  |  |
| :--- | ---: | ---: | ---: | ---: | ---: |
| blet | 50 | 0.050 | 0.10 | 10.00 | 0.395 |
| calm | 50 | 0.700 | 1.40 | 17.50 | 0.825 |
| ceph | 50 | 0.050 | 0.10 | 10.00 | 0.395 |
| holu | 100 | 0.590 | 0.73 | 42.50 | 1.870 |
| hylo | 100 | 1.765 | 1.88 | 20.00 | 1.150 |
| isop | 50 | 0.075 | 0.15 | 2.50 | 0.125 |
| isst | 50 | 0.825 | 1.65 | 7.50 | 0.495 |
| leme | 100 | 9.760 | 19.20 | 30.00 | 3.270 |
| peli | 100 | 2.915 | 4.78 | 32.50 | 1.900 |
| plag | 100 | 3.295 | 5.32 | 62.50 | 3.185 |
| plin | 100 | 0.455 | 0.88 | 27.50 | 1.275 |
| plun | 100 | 3.660 | 4.22 | 65.00 | 3.390 |
| pogm | 50 | 0.005 | 0.01 | 0.50 | 0.020 |
| rhgl | 100 | 14.775 | 18.20 | 82.50 | 6.420 |
| rhlo | 100 | 2.690 | 4.15 | 35.00 | 1.975 |
| ricl | 50 | 1.200 | 2.40 | 10.00 | 0.635 |
| scab | 100 | 1.805 | 2.53 | 27.50 | 1.520 |
| sphh | 100 | 0.100 | 0.15 | 10.00 | 0.430 |
| stor | 50 | 0.005 | 0.01 | 0.50 | 0.020 |
| stpr | 100 | 17.135 | 18.75 | 85.00 | 7.025 |

totals:

|  | $m$. cov max.cov | $t$ spp | m.spp |  |
| :--- | ---: | ---: | ---: | ---: |
|  |  |  |  |  |
| shrubs | 85.80 | 103.7 | 7 | 7.0 |
| herbs | 94.15 | 113.9 | 17 | 14.5 |
| bryo. | 61.85 | 61.9 | 20 | 16.0 |
| all spp. | 241.75 | 243.6 | 44 | 37.5 |



| cldg | 25 | 0.055 | 0.22 | 5.00 | 0.210 |
| :---: | :---: | :---: | :---: | :---: | :---: |
| clec | 25 | 0.225 | 0.90 | 13.75 | 0.600 |
| dicf | 100 | 2.233 | 2.85 | 25.00 | 1.810 |
| dics | 50 | 1.688 | 5.25 | 12.50 | 1.008 |
| nylo | 100 | 21.203 | 46.78 | 45.00 | 7.565 |
| hypu | 50 | 0.075 | 0.15 | 2.50 | 0.148 |
| 1sst | 75 | 2.958 | 8.15 | 18.75 | 1.642 |
| pelo | 75 | 0.490 | 1.75 | 17.50 | 0.870 |
| pelt | 75 | 0.682 | 1.80 | 8.75 | 0.583 |
| piun | 25 | 0.038 | 0.15 | 1.25 | 0.075 |
| plzs | 25 | 0.038 | 0.15 | 1.25 | 0.075 |
| ponc | 25 | 0.008 | 0.03 | 1.25 | 0.065 |
| ponj | 100 | 1.683 | 5.93 | 30.00 | 1.810 |
| ponp | 25 | 0.275 | 1.10 | 6.25 | 0.325 |
| rhac | 75 | 2.128 | 5.88 | 13.75 | 1.168 |
| rhlo | 100 | 0.533 | 1.20 | 8.75 | 0.600 |
| rhtr | 75 | 0.438 | 0.93 | 8.75 | 0.548 |
| scie | 25 | 0.508 | 2.03 | 2.50 | 0.243 |
| stor | 100 | 27.688 | 50.10 | 73.75 | 10.563 |
| tame | 100 | 0.890 | - 2.50 | 11.50 | 0.847 |
| bare rock |  |  |  |  |  |
| rock | 50 | 1.813 | 5.75 | 6.25 | 0.000 |
| totals: |  |  |  |  |  |
|  |  | m.cov | max.cov | t.Spp m | m. SPP |
| shru |  | 98.35 | 132.7 | 14 | 7.8 |
| herb |  | 34.55 | 43.5 | 33 | 20.3 |
| bryo |  | 64.07 | 84.9 | 23 | 13.8 |
| all | $p$. | 197.00 | 213.1 | 70 | 41.8 |

Note : Pelt = Peltigera membranacea

Appendix 3 : Pseudotsuga-Thuja-Acer forests (P2)
plots: $\begin{array}{llll}13 & 14 & 16 & 131\end{array}$
cons. m.cov max.cov m.fre. m.i.v

| shrubs |  |  |  |  |  |
| :--- | ---: | ---: | ---: | ---: | ---: |
| acgi | 25 | 0.003 | 0.01 | 0.25 | 0.013 |
| amal | 25 | 0.003 | 0.01 | 0.25 | 0.013 |
| bene | 100 | 12.722 | 24.83 | 49.00 | 8.028 |
| gash | 75 | 13.282 | 26.58 | 40.00 | 5.948 |
| hodi | 25 | 0.003 | 0.01 | 0.25 | 0.013 |
| ribl | 25 | 0.038 | 0.15 | 1.25 | 0.070 |
| rogy | 100 | 1.757 | 4.07 | 12.75 | 1.128 |
| ruur | 100 | 8.212 | 31.90 | 33.75 | 3.830 |
| samr | 25 | 0.003 | 0.01 | 0.25 | 0.013 |
| syal | 50 | 0.232 | 0.90 | 3.75 | 0.253 |
| vaot | 25 | 0.003 | 0.01 | 0.25 | 0.025 |
| vapa | 100 | 2.290 | 6.45 | 17.75 | 1.765 |

nerbs

| actr | 100 | 10.590 | 22.52 | 63.75 | 6.440 |
| :--- | ---: | ---: | ---: | ---: | ---: |
| adbi | 50 | 0.265 | 1.05 | 4.00 | 0.275 |
| adpe | 25 | 0.600 | 2.40 | 5.00 | 0.395 |


| anly | 25 | 0.080 | 0.32 | 3.75 | 0.21 |
| :--- | :--- | :--- | :--- | :--- | :--- |


| brov | 50 | 0.075 | 0.15 |
| :--- | :--- | :--- | :--- |
| andy | 50 | 0.020 | 0.05 |

caly
camp
chme
chum
coma
feoc
fesu
gali heco
25
hypo
lamu

| lane | 25 |  |
| :--- | ---: | ---: |
| 1 ibo | 100 | 10 |

1 ico $\quad 75 \quad 10.100$

| mono | 25 | 0.045 | 0.10 |
| :--- | :--- | :--- | :--- |
| mopa | 50 | 0.048 | 0.15 |
| mos | 0.040 | 0.15 |  |


| mosi | 25 | 0.813 | 3.25 | 8.75 | 0.635 |
| :--- | ---: | ---: | ---: | ---: | ---: |
| poly | 100 | 0.710 | 2.60 | 13.75 | 0.908 |
| pomu | 100 | 18.248 | 41.38 | 41.25 | 7.873 |
| ptaq | 75 | 0.078 | 0.15 | 2.75 | 0.170 |
| ptea | 25 | 0.003 | 0.01 | 0.25 | 0.025 |
| pypi | 25 | 0.080 | 0.32 | 3.75 | 0.205 |
| smra | 25 | 0.003 | 0.01 | 0.25 | 0.013 |
| tila | 25 | 0.003 | 0.01 | 0.25 | 0.013 |
| titr | 50 | 0.577 | 2.30 | 11.50 | 0.710 |
| trla | 100 | 4.175 | 9.07 | 48.75 | 3.742 |
| trov | 25 | 0.003 | 0.01 | 0.25 | 0.043 |
| vise | 75 | 1.070 | 2.95 | 25.00 | 1.648 |
|  |  |  |  |  |  |
| bryophytes and | $11 c h e n s$ |  |  |  |  |
| dicf | 100 | 1.163 | 2.22 | 20.00 | 1.348 |
| dics | 25 | 0.038 | 0.15 | 1.25 | 0.070 |


| nylo | 100 | 22.005 | 53.20 | 60.00 | 9.455 |
| :---: | :---: | :---: | :---: | :---: | :---: |
| hypu | 50 | 0.450 | -1.50 | 5.00 | 0.375 |
| isst | 75 | 3.013 | 3.53 | 16.25 | 1.930 |
| leme | 75 | 2.683 | 3 7.30 | 25.00 | 2.435 |
| meta | 25 | 0.225 | - 0.90 | 2.50 | 0.190 |
| mniu | 25 | 0.038 | $8 \quad 0.15$ | 1.25 | 0.073 |
| plin | 25 | 0.287 | $7 \quad 1.15$ | 13.75 | 0.750 |
| plun | 50 | 0.020 | 0.05 | 3.75 | 0.245 |
| pogm | 50 | 1.295 | 5 5.03 | 3.75 | 0.568 |
| rhgl | 25 | 0.050 | 0.20 | 3.75 | - 0.197 |
| rhlo | 75 | 2.405 | - 4.82 | 17.50 | 1.598 |
| rhtr | 75 | 6.220 | - 16.88 | 32.50 | - 3.547 |
| scab | 50 | 0.118 | $8 \quad 0.32$ | 5.00 | 0.278 |
| stor | 100 | 24.340 | -48.78 | 75.00 | 13.655 |
| $t \mathrm{~mm}$ | 25 | 0.038 | $8 \quad 0.15$ | 1.25 | 0.070 |
| bare rock |  |  |  |  |  |
| rock | 50 | 9.070 | 032.00 | 17.50 | 0.000 |
| totals: |  |  |  |  |  |
|  |  | m.cov max.cov |  | t.spp | m.spp |
|  |  | 38.55 | 55.5 | 12 | 6.8 |
|  |  | 53.88 | 77.7 | 36 | 18.5 |
| herbs bryo. |  | 64.40 | 77.1 | 17 | 9.5 |
| all spp. |  | 156.80 | 483.5 | 65 | 34.8 |

Appendix 3 : Pseudotsuga-Linnaea forests (P3)
plots: 71112109138

|  | cons. | m.cov | max.cov | m.fre. | m.i.v |
| :--- | ---: | ---: | ---: | ---: | ---: |
| shrubs |  |  |  |  |  |
| amal | 40 | 0.032 | 0.15 | 1.20 | 0.074 |
| bene | 100 | 10.392 | 22.42 | 38.20 | 4.208 |
| gash | 100 | 43.694 | 93.13 | 65.00 | 15.446 |
| hodi | 40 | 1.906 | 9.52 | 3.20 | 0.504 |
| loci | 20 | 0.030 | 0.15 | 1.00 | 0.054 |
| rogy | 80 | 2.832 | 9.98 | 16.00 | 1.356 |
| ruur | 60 | 1.518 | 6.00 | 21.20 | 1.368 |
| syal | 40 | 0.566 | 2.65 | 5.00 | 0.370 |
| vaal | 20 | 0.150 | 0.75 | 1.00 | 0.100 |
| vame | 20 | 0.002 | 0.01 | 0.20 | 0.010 |
| vaot | 20 | 0.706 | 3.53 | 4.00 | 0.558 |
| vapa | 100 | 10.334 | 19.90 | 43.00 | 5.586 |

nerbs

| actr | 100 | 12.360 | 28.65 | 54.00 | 5.428 |
| :--- | ---: | ---: | ---: | ---: | ---: |
| adbi | 20 | 0.002 | 0.01 | 0.20 | 0.010 |
| adpe | 20 | 0.002 | 0.01 | 0.20 | 0.010 |
| alvi | 40 | 0.004 | 0.01 | 0.40 | 0.022 |
| anly | 20 | 0.030 | 0.15 | 1.00 | 0.054 |
| aren | 20 | 0.030 | 0.15 | 1.00 | 0.054 |
| boho | 60 | 0.022 | 0.05 | 4.20 | 0.280 |

$\begin{array}{lllll}\text { boho } & 60 & 0.022 & 0.05 & 4.20 \\ & 20 & 0.426 & 2.13 & 7.00 \\ 0.280\end{array}$

| brov | 20 | 0.426 | 2.13 | 7.00 | 0.404 |
| :--- | :--- | :--- | :--- | :--- | :--- |
| caly | 40 | 0.042 | 0.18 | 3.00 | 0.154 |


| caly | 40 | 0.042 | 0.18 | 3.00 | 0.154 |
| :--- | :--- | :--- | ---: | ---: | ---: |
| camp | 40 | 0.252 | 1.08 | 6.00 | 0.348 |
| chme | 80 | 0.200 | 0.47 | 15.00 | 0.804 |


| chme | 80 | 0.200 | 0.47 | 15.00 | 0.804 |
| :--- | ---: | ---: | ---: | ---: | ---: |
| chum | 100 | 2.480 | 4.22 | 31.20 | 2.498 |


| coma | 60 | 0.044 | 0.18 | 3.20 | 0.168 |
| :--- | :--- | :--- | :--- | :--- | :--- |


| come | 60 | 0.042 | 0.15 | 3.20 | 0.182 |
| :--- | ---: | ---: | ---: | ---: | ---: |

feoc $40 \quad 1.176 \quad 5.13 \quad 17.00 \quad 1.130$

| fesu | 100 | 1.698 | 6.38 | 17.20 | 1.380 |
| :--- | :--- | :--- | :--- | :--- | :--- |


| goob | 100 | 0.246 | 0.80 | 19.20 | 0.976 |
| ---: | ---: | ---: | ---: | ---: | ---: |
| $h e c o$ | 20 | 0.002 | 0.01 | 0.20 | 0.010 |


| heco | 20 | 0.002 | 0.01 | 0.20 | 0.010 |
| :--- | :--- | :--- | :--- | :--- | :--- |
| nial | 20 | 0.304 | 1.52 | 7.00 | 0.412 |


| hial | 20 | 0.304 | 1.52 | 7.00 | 0.412 |
| :--- | :--- | :--- | :--- | :--- | :--- |
| lamu | 20 | 0.002 | 0.01 | 0.20 | 0.010 |


| lane | 60 | 1.488 | 5.40 | 7.20 | 0.632 |
| :--- | :--- | :--- | :--- | :--- | :--- |


| 1 iao | 20 | 0.002 | 0.01 | 0.20 | 0.010 |
| ---: | ---: | ---: | ---: | ---: | ---: |


| 1 lbo | 100 | 19.622 | 38.40 | 79.00 | 9.046 |
| ---: | ---: | ---: | ---: | ---: | ---: |
| 1 ico | 80 | 0.150 | 0.30 | 15.00 | 0.882 |


| pedi | 20 | 1.556 | 7.78 | 11.00 | 0.996 |
| :--- | :--- | :--- | :--- | :--- | :--- |


| poly | 40 | 0.046 | 0.22 | 4.20 | 0.210 |
| :--- | ---: | ---: | ---: | ---: | ---: |


| pomu 100 | 3.764 | 12.40 | 11.20 | 1.590 |
| :--- | :--- | :--- | :--- | :--- |


| ptaq | 60 | 0.190 | 0.93 | 3.40 | 0.218 |
| :--- | :--- | :--- | :--- | :--- | :--- |
| pypi | 40 | 0.070 | 0.32 | 4.00 | 0.198 |


| pypi | 40 | 0.070 | 0.32 | 4.00 | 0.198 |
| :--- | :--- | :--- | :--- | :--- | :--- |


| sewa | 20 | 0.030 | 0.15 | 1.00 | 0.054 |
| :--- | :--- | :--- | :--- | :--- | :--- |
| stre | 20 | 0.002 | 0.01 | 0.20 | 0.012 |


| tila | 20 | 0.030 | 0.15 | 1.00 | 0.064 |
| :--- | :--- | :--- | :--- | ---: | ---: |
| titr | 20 | 0.006 | 0.03 | 1.00 | 0.048 |
| trla | 80 | 1.584 | 3.97 | 31.00 | 1.966 |


| trla | 80 | 1.584 | 3.97 | 31.00 | 1.966 |
| :--- | :--- | :--- | :--- | :--- | :--- |
| vise | 80 | 1.612 | 3.60 | 38.00 | 2.262 |


| dicf |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: |
| dicf | 100 | 0.726 | 1.13 | 18.00 | 1.286 |
| dics | 20 | 0.094 | 0.47 | 4.00 | 0.352 |


| hylo | 100 | 52.232 | 79.50 | 87.00 | 17.726 |
| :--- | :--- | :--- | :--- | :--- | :--- |


| nypu | 40 | 0.420 | 1.50 | 6.00 | 0.546 |
| :--- | ---: | ---: | ---: | ---: | ---: |
| isop | 20 | 0.240 | 1.20 | 4.00 | 0.292 |
| isst | 20 | 0.526 | 2.63 | 2.00 | 0.260 |
| leme | 60 | 0.300 | 1.20 | 6.00 | 0.376 |
| mniu | 40 | 0.036 | 0.15 | 2.00 | 0.110 |
| pelt | 20 | 0.180 | 0.90 | 6.00 | 0.326 |
| plun | 60 | 1.266 | 5.65 | 23.00 | 1.886 |
| pogm | 100 | 2.654 | 11.90 | 10.00 | 1.106 |
| ponj | 20 | 0.064 | 0.32 | 3.00 | 0.156 |
| ponp | 20 | 0.030 | 0.15 | 1.00 | 0.054 |
| rhgl | 20 | 0.036 | 0.18 | 2.00 | 0.172 |
| rhlo | 100 | 4.372 | 10.05 | 43.00 | 3.548 |
| rhtr | 80 | 1.466 | 3.83 | 28.00 | 1.838 |
| rhyt | 40 | 0.714 | 3.42 | 10.00 | 0.714 |
| scab | 100 | 0.616 | 2.00 | 11.00 | 0.778 |
| stor | 100 | 5.864 | 10.90 | 53.00 | 4.702 |
| tame | 40 | 1.490 | 7.30 | 18.00 | 1.286 |
|  |  |  |  |  |  |
| bare rock |  | 0.750 | 3.75 | 2.00 | 0.000 |
| rock | 20 | 0.750 |  |  |  |
|  |  |  |  |  |  |
| totals: |  | $m . c o v$ | max.cov | $t . s p p$ | $m . s p p$ |
|  |  |  |  |  |  |
| shrubs | 72.16 | 117.3 | 12 | 6.4 |  |
| herbs | 49.50 | 79.6 | 35 | 17.4 |  |
| bryo. | 73.34 | 104.8 | 20 | 11.0 |  |
| all spp. | 195.00 | 268.9 | 67 | 34.8 |  |

Appendix 3 : Pseudotsuga-Berberis forests (P4)

|  | cons. | m.cov | max.cov | m.fre. | m.i.v |
| :---: | :---: | :---: | :---: | :---: | :---: |
| shrubs acgl | 18 | 0.523 | 5.00 | 0.91 | 0.205 |
| amal | 18 | 0.004 | 0.03 | 0.54 | 0.028 |
| bene | 100 | 11.493 | 26.40 | 41.00 | 7.648 |
| gash | 90 | 10.355 | 28.65 | 25.91 | 5.564 |
| hodi | 9 | 0.001 | 0.01 | 0.09 | 0.007 |
| mefe | 9 | 0.082 | 0.90 | 0.91 | 0.071 |
| opho | 9 | 0.001 | 0.01 | 0.09 | 0.005 |
| rogy | 54 | 0.457 | 4.20 | 5.27 | 0.365 |
| rusp | 9 | 0.001 | 0.01 | 0.09 | 0.007 |
| ruur | 72 | 0.551 | 4.15 | 9.64 | 0.773 |
| syal | 18 | 0.021 | 0.20 | 1.82 | 0.083 |
| vaal | 36 | 3.253 | 14.07 | 14.54 | 1.895 |
| vaol | 9 | 0.548 | 6.03 | 1.82 | 0.268 |
| vaot | 18 | 0.455 | 5.00 | 0.54 | 0.249 |
| vapa | 100 | 11.869 | 37.50 | 47.73 | 7.167 |
| nerbs |  |  |  |  |  |
| actr | 100 | 4.115 | 17.13 | 46.36 | 4.500 |
| adbi | 18 | 0.007 | 0.05 | 1.36 | 0.079 |
| adpe | 9 | 0.171 | 1.88 | 0.46 | 0.056 |
| alvi | 27 | 0.003 | 0.01 | 0.27 | 0.027 |
| aren | 9 | 0.014 | 0.15 | 0.46 | 0.026 |
| blsp | 9 | 0.001 | 0.01 | 0.09 | 0.006 |
| boho | 9 | 0.003 | 0.03 | 0.46 | 0.019 |
| boye | 9 | 0.001 | 0.01 | 0.09 | 0.005 |
| caly | 27 | 0.006 | 0.03 | 1.00 | 0.067 |
| camp | 9 | 0.001 | 0.01 | 0.09 | 0.008 |
| chme | 63 | 0.093 | 0.45 | 11.46 | 0.914 |
| chum | 54 | 0.025 | 0.18 | 2.18 | 0.158 |
| come | 27 | 0.018 | 0. 18 | 1.09 | 0.096 |
| corn | 9 | 0.136 | 1.50 | 0.91 | 0.104 |
| dino | 18 | 0.281 | 3.08 | 3.27 | 0.251 |
| feoc | 9 | 0.027 | 0.30 | 0.91 | 0.094 |
| fesu | 36 | 0.339 | 1.50 | 7.73 | 0.723 |
| goob | 81 | 0.165 | 0.43 | 16.46 | 1.073 |
| heco | 27 | 0.005 | 0.03 | 0.64 | 0.068 |
| hypo | 45 | 0.123 | 1.27 | 4.27 | 0.473 |
| lamu | 18 | 0.004 | 0.03 | 0.54 | 0.032 |
| 1 1bo | 81 | 8.530 | 24.10 | 43.18 | 5.202 |
| lica | 27 | 0.013 | 0. 10 | 2.36 | 0. 155 |
| lico | 54 | 0.096 | 0.38 | 10.00 | 0.692 |
| madi | 9 | 0.001 | 0.01 | 0.09 | 0.007 |
| mono | 36 | 0.005 | 0.03 | 0.73 | 0.056 |
| mopa | 9 | 0.014 | 0.15 | 0.46 | 0.026 |
| poly | 36 | 0.034 | 0.20 | 2.00 | 0.099 |
| pomu | 100 | 7.750 | 28.50 | 28.82 | 4.792 |
| ptaq | 63 | 0.488 | 2.92 | 4.36 | 0.586 |
| ptea | 9 | 0.001 | 0.01 | 0.09 | 0.006 |
| pyap | 9 | 0.003 | 0.03 | 0.46 | 0.052 |
| pyas | 9 | 0.003 | 0.03 | 0.46 | 0.029 |
| pypi | 27 | 0.010 | 0.05 | 1.82 | 0. 164 |
| smra | 18 | 0.148 | 1.58 | 3.18 | 0.207 |
| smst | 9 | 0.014 | 0.15 | 0.46 | 0.041 |
| tila | 54 | 0.065 | 0.30 | 3.64 | 0.256 |


| titr | 63 | 1. 110 | 7.45 | 13.64 | 1.026 |
| :---: | :---: | :---: | :---: | :---: | :---: |
| tria | 81 | 0.292 | 2.13 | 10.54 | 0.763 |
| trov | 54 | 0.074 | 0.32 | 5.64 | 0.325 |
| vise | 54 | 1.561 | 16.85 | 11.00 | 0.945 |
| bryopnytes and |  | d 1 ichen |  |  |  |
| blet | 9 | 0.003 | 0.03 | 0.46 | 0.029 |
| calm | 9 | 0.016 | 0.18 | 0.91 | 0.062 |
| ceph | 9 | 0.082 | 0.90 | 0.91 | 0.085 |
| cloa | 9 | 0.070 | 0.77 | 0.91 | 0.166 |
| dicf | 90 | 0.453 | 1.88 | 23.64 | 1.914 |
| nete | 27 | 0.753 | 4.53 | 10.91 | 0.780 |
| nylo | 100 | 22.647 | 58.03 | 56.82 | 13.516 |
| hypu | 81 | 1.515 | 5.50 | 20.46 | 2.594 |
| isop | 27 | 0.319 | 2.13 | 10.00 | 1.048 |
| isst | 81 | 2.484 | 7.78 | 29.54 | 2.980 |
| ledo | 9 | 0.068 | 0.75 | 0.46 | 0.045 |
| leme | 18 | 0.069 | 0.75 | 0.54 | 0.070 |
| mniu | 36 | 0.070 | 0.30 | 5.00 | 0.406 |
| pelt | 36 | 0.103 | 0.75 | 2.73 | 0.206 |
| plag | 9 | 0.003 | 0.03 | 0.46 | 0.025 |
| plin | 9 | 0.003 | 0.03 | 0.46 | 0.023 |
| plun | 45 | 0.142 | 0.63 | 7.73 | 0.564 |
| pogm | 9 | 0.014 | 0.15 | 0.46 | 0.022 |
| ptic | 9 | 0.014 | 0.15 | 0.46 | 0.043 |
| ringl | 54 | 0.362 | 2.30 | 8. 18 | 0.683 |
| rhlo | 90 | 6.130 | 16.63 | 40.91 | 4.910 |
| rhtr | 18 | 0.092 | 0.98 | 2.73 | 0.296 |
| rhyt | 63 | 2.123 | 14.25 | 20.46 | 2.128 |
| scab | 81 | 1.076 | 2.72 | 26.36 | 2.286 |
| stor | 100 | 20.375 | 77.28 | 77.73 | 14.259 |
| tame | 45 | 0.467 | 3.30 | 3.73 | 0.405 |
| bare rock |  |  |  |  |  |
| rock | 54 | 1.985 | 7.13 | 9.54 | 0.000 |
| totals: |  |  |  |  |  |
|  |  | m.cov max.cov |  | t.spp | m.spp |
| shrubs |  | 39.62 | 73.8 | 15 | 5.7 |
| heros |  | 25.71 | 84.0 | 41 | 14.3 |
| bryo. |  | 59.44 | 95.4 | 26 | 10.8 |
| all spp. |  | 124.80 | 226.6 | 82 | 30.8 |

[^6]Appendix 3 : Tsuga-Pseudotsuga-Polystichum forests (P5)


| pomu | 100 | 28.688 | 59.03 | 65.59 | 20.375 |
| :---: | :---: | :---: | :---: | :---: | :---: |
| ptaq | 23 | 0.411 | 3.70 | 2.41 | 0.446 |
| pyap | 5 | 0.001 | 0.01 | 0.06 | 0.009 |
| pyse | 5 | 0.001 | 0.01 | 0.06 | 0.004 |
| runi | 5 | 0.316 | 5.38 | 1.18 | 0.180 |
| sewa | 5 | 0.001 | 0.01 | 0.06 | 0.004 |
| smra | 11 | 0.092 | 1.55 | 1.23 | 0.093 |
| smst | 11 | 0.055 | 0.90 | 0.88 | 0.061 |
| stac | 5 | 0.111 | 1.88 | 0.29 | 0.115 |
| sten | 5 | 0.001 | 0.01 | 0.06 | 0.004 |
| stra | 5 | 0.128 | 2.17 | 0.88 | 0.081 |
| strr | 5 | 0.001 | 0.01 | 0.06 | 0.003 |
| tila | 41 | 0.384 | 6.13 | 5.77 | 0.501 |
| titr | 88 | 1.789 | 17.60 | 21.94 | 2.056 |
| trau | 5 | 0.001 | 0.01 | 0.06 | 0.004 |
| trla | 17 | 0.054 | 0.75 | 0.65 | 0.092 |
| trov | 76 | 0.365 | 1.70 | 11.76 | 0.986 |
| $v i g l$ | 5 | 0.004 | 0.01 | 0.06 | 0.003 |
| vise | 17 | 0.126 | 1.98 | 2.12 | 0.171 |
| bryoph | es | 1 ichen |  |  |  |
| bazz | 5 | 0.044 | 0.75 | 0.29 | 0.031 |
| blet | 5 | 0.006 | 0.10 | 1. 18 | 0.091 |
| ceph | 17 | 0.143 | 1.23 | 2.65 | 0.278 |
| cloa | 11 | 0.156 | 2.03 | 2.06 | 0.286 |
| dicf | 58 | 0.193 | 1.05 | 13.82 | 1.250 |
| hete | 52 | 1.214 | 4.97 | 17.35 | 1.748 |
| nolu | 23 | 0.033 | 0.45 | 2.06 | 0.161 |
| nylo | 76 | 2.524 | 16.75 | 14.88 | 2.248 |
| hypu | 94 | 2.611 | 5.97 | 31.47 | 4.439 |
| Isop | 76 | 0.468 | 1.13 | 12.65 | 1.381 |
| isst | 94 | 5.178 | 13.50 | 47.94 | 7.014 |
| ledo | 17 | 0.125 | 0.93 | 3.23 | 0.258 |
| leme | 11 | 0.019 | 0.18 | 0.88 | 0.052 |
| mniu | 29 | 0.242 | 1.90 | 6.47 | 0.522 |
| pelt | 23 | 0.065 | 0.75 | 1.47 | 0.154 |
| plag | 17 | 0.037 | 0.55 | 2.94 | 0.173 |
| plin | 17 | 0. 168 | 2.65 | 1.77 | 0.144 |
| plun | 70 | 2. 125 | 12.45 | 23.23 | 3.249 |
| pogm | 29 | 0.038 | 0.30 | 1.53 | 0.132 |
| rhah | 5 | 0.012 | 0.20 | 0.88 | 0.083 |
| ringl | 64 | 0.541 | 2.00 | 17.35 | 1.714 |
| rhlo | 76 | 3.804 | 31.77 | 22.94 | 3.218 |
| rhtr | 5 | 0.002 | 0.03 | 0.29 | 0.016 |
| rhyt | 17 | 0.145 | 1.50 | 2.06 | 0.162 |
| scab | 94 | 2.409 | 8.70 | 42.65 | 4.738 |
| stor | 94 | 12.915 | 50.15 | 62.35 | 12.459 |
| tame | 5 | 0.001 | 0.01 | 0.06 | 0.005 |
| bare rock |  |  |  |  |  |
| rock | 88 | 9.604 | 41.50 | 29.41 | 0.000 |

totals:

|  | m.cov max.cov | t.spp | m.spp |  |
| :--- | ---: | ---: | ---: | ---: |
|  |  |  |  |  |
| snrubs | 12.02 | 34.7 | 17 | 4.5 |
| herbs | 44.07 | 113.9 | 54 | 11.7 |
| bryo. | 35.21 | 80.2 | 27 | 11.0 |
| all spp. | 91.31 | 177.5 | 98 | 27.2 |

Note : Pelt = Peltigera polydactyla (98, 124, 142) and P. membranacea (133)

Appendix 3 : Montane Tsuga forests (P6)

nerbs
actr
adpe
alvi
anly
apan
atfi
blsp
camp
chme
chum.
coma
come 50
corn
diho
feoc
fesu
goob
heco
nial
nypo
1 amu
libo
lica 25
1100 50
lyci
madi
mone
mono
poly pomu ptaq ptea pypi 16
pyse
smra
stra 16
stre tila
titr
trla 33
4.
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0.
.138

| 13.48 | 27.08 | 3.402 |
| ---: | ---: | ---: |
| 0.01 | 0.08 | 0.005 |
| 0.01 | 0.17 | 0.011 |
| 0.01 | 0.08 | 0.006 |
| 0.01 | 0.08 | 0.006 |
| 0.01 | 0.08 | 0.005 |
| 21.17 | 7.58 | 1.323 |
| 0.15 | 0.50 | 0.035 |
| 1.33 | 9.42 | 0.646 |
| 0.45 | 3.92 | 0.308 |
| 0.03 | 0.42 | 0.024 |
| 0.15 | 1.92 | 0.144 |
| 0.01 | 0.08 | 0.006 |
| 0.01 | 0.08 | 0.007 |
| 0.01 | 0.08 | 0.005 |
| 0.01 | 0.17 | 0.011 |
| 0.60 | 7.50 | 0.507 |
| 0.03 | 0.50 | 0.033 |
| 0.03 | 0.50 | 0.030 |
| 0.03 | 1.00 | 0.082 |
| 0.01 | 0.25 | 0.019 |
| 5.43 | 6.67 | 0.941 |
| 0.08 | 1.75 | 0.111 |
| 0.20 | 5.42 | 0.427 |
| 4.50 | 1.67 | 0.283 |
| 0.01 | 0.08 | 0.005 |
| 1.27 | 3.75 | 0.375 |
| 0.01 | 0.08 | 0.005 |
| 0.01 | 0.25 | 0.020 |
| 10.75 | 10.92 | 1.847 |
| 0.01 | 0.08 | 0.006 |
| 0.15 | 0.42 | 0.035 |
| 0.01 | 0.17 | 0.013 |
| 0.01 | 0.08 | 0.008 |
| 0.03 | 0.42 | 0.027 |
| 0.01 | 0.17 | 0.010 |
| 0.01 | 0.08 | 0.005 |
| 2.47 | 4.75 | 0.398 |
| 6.15 | 8.33 | 0.813 |
| 0.15 | 1.00 | 0.075 |
| 0.0 |  |  |



Note : Pelo = Peltigera aphtosa (33)

```
Pelt = P. polydactyla (33, 64, 66, 103, 130, 139)
            and P. membranacea (96, 128)
Loba = Lobaria oregana (21)
```

Appendix 3 : Montane Tsuga-Gaultheria forests (P7) plots: 323840626567145

|  | cons | m.cov | $\max . \operatorname{cov}$ | m.fre. | m.1.v |
| :---: | :---: | :---: | :---: | :---: | :---: |
| shrubs |  |  |  |  |  |
| bene | 57 | 3.966 | 22.55 | 13.57 | 2.031 |
| gash | 100 | 74.924 | 95.63 | 98.57 | 30.344 |
| mefe | 14 | 0.021 | 0.15 | 0.71 | 0.040 |
| rogy | 14 | 1.181 | 8.27 | 2.86 | 0.427 |
| ruur | 28 | 0.954 | 4.88 | 6.43 | 0.946 |
| vaal | 14 | 8.429 | 59.00 | 14.29 | 2.357 |
| vaol | 14 | 1.929 | 13.50 | 8.57 | 0.810 |
| vaot | 14 | 3.607 | 25.25 | 6.43 | 1.410 |
| vapa | 100 | 6.781 | 15.88 | 38.57 | 4.953 |
| nerbs |  |  |  |  |  |
| actr | 42 | 0.013 | 0.05 | 2.29 | 0.221 |
| alvi | 14 | 0.001 | 0.01 | 0.14 | 0.009 |
| blsp | 28 | 0.003 | 0.01 | 0.29 | 0.016 |
| boho | 42 | 0.034 | 0.20 | 3.00 | 0.219 |
| chum | 42 | 0.544 | 2.28 | 8.00 | 0.559 |
| coma | 14 | 0.001 | 0.01 | 0.14 | 0.007 |
| come | 28 | 0.006 | 0.03 | 0.86 | 0.091 |
| corn | 14 | 0.197 | 1.38 | 7.14 | 0.401 |
| feoc | 14 | 0.257 | 1.80 | 2.86 | 0.220 |
| fesu | 28 | 0.740 | 5.03 | 5.00 | 0.464 |
| gaov | 14 | 0.393 | 2.75 | 5.71 | 0.367 |
| goob | 42 | 0.010 | 0.03 | 1.57 | 0.090 |
| hial | 14 | 0.007 | 0.05 | 1.43 | 0.083 |
| hypo | 14 | 0.001 | 0.01 | 0.14 | 0.009 |
| $l a m u$ | 14 | 0.021 | 0.15 | 0.71 | 0.046 |
| libo | 28 | 3.080 | 17.98 | 25.00 | 2.394 |
| 1 ica | 28 | 0.006 | 0.03 | 0.86 | 0.049 |
| lico | 85 | 0.316 | 0.80 | 34.29 | 2.703 |
| poly | 14 | 0.004 | 0.03 | 0.71 | 0.057 |
| pomu | 57 | 0.489 | 2.65 | 3.14 | 0.313 |
| ptaq | 14 | 0.107 | 0.75 | 0.71 | 0.130 |
| ptea | 14 | 0.001 | 0.01 | 0.14 | 0.009 |
| pypi | 28 | 0.026 | 0.15 | 1.43 | 0.106 |
| rupe | 14 | 0.447 | 3. 13 | 0.71 | 0.123 |
| trla | 28 | 0.270 | 1.88 | 0.86 | 0.111 |


| bryophytes and lichens |  |  |  |  |  |
| :--- | ---: | :--- | ---: | ---: | ---: |
| bazz | 14 | 0.021 | 0.15 | 0.71 | 0.059 |
| clar | 14 | 0.021 | 0.15 | 0.71 | 0.040 |
| cldo | 14 | 0.001 | 0.01 | 0.14 | 0.007 |
| clds | 14 | 0.004 | 0.03 | 0.71 | 0.041 |
| dicf | 100 | 1.944 | 3.70 | 50.00 | 4.539 |
| hylo | 100 | 34.777 | 69.03 | 74.43 | 13.331 |
| hypu | 85 | 1.333 | 2.85 | 25.00 | 2.663 |
| isst | 42 | 0.179 | 0.77 | 4.29 | 0.347 |
| ledo | 14 | 0.021 | 0.15 | 0.71 | 0.059 |
| pelo | 28 | 0.026 | 0.15 | 1.43 | 0.083 |
| pelt | 28 | 0.153 | 0.77 | 2.86 | 0.274 |
| plun | 57 | 0.379 | 1.67 | 6.43 | 0.626 |
| plzs | 14 | 0.983 | 6.88 | 2.14 | 0.299 |
| pogm | 28 | 0.171 | 0.75 | 2.86 | 0.219 |
| rhgl | 14 | 0.021 | 0.15 | 0.71 | 0.059 |
| rhlo | 85 | 14.584 | 30.55 | 55.71 | 7.477 |



Note : Pelt $=\frac{\text { Peltigera }}{\text { and Pembranacea } \text { polydactyla (62) }}$ (65)

Appendix 3 : Coastal dry Thuja forests (T1)
plots: 94149159

|  | cons. | m.cov | max.cov | m.fre. | m.i.v |
| :---: | ---: | ---: | ---: | ---: | ---: |
| shrubs |  |  |  |  |  |
| gash | 100 | 64.543 | 70.25 | 100.00 | 20.507 |
| mefe | 100 | 2.537 | 4.82 | 15.33 | 1.507 |
| rusp | 33 | 0.010 | 0.03 | 1.67 | 0.093 |
| vaal | 66 | 0.253 | 0.75 | 2.00 | 0.173 |
| vaot | 100 | 50.200 | 78.75 | 76.67 | 15.730 |
| vapa | 100 | 7.160 | 11.35 | 55.00 | 4.843 |
|  |  |  |  |  |  |
| herbs |  |  |  |  |  |
| blsp | 100 | 32.527 | 63.42 | 52.00 | 10.500 |
| boho | 66 | 0.007 | 0.01 | 0.67 | 0.043 |
| corn | 33 | 0.010 | 0.03 | 1.67 | 0.093 |
| libo | 66 | 0.927 | 2.63 | 5.00 | 0.473 |
| lico | 33 | 0.003 | 0.01 | 0.33 | 0.023 |
| lyse | 33 | 0.010 | 0.03 | 1.67 | 0.093 |
| madi | 33 | 0.483 | 1.45 | 15.00 | 0.923 |


| bryoph | S | 1 |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: |
| blet | 33 | 0.033 | 0.10 | 6.67 | 0.377 |
| calm | 33 | 0.150 | 0.45 | 5.00 | 0.307 |
| ceph | 33 | 0.760 | 2.28 | 13.33 | 0.927 |
| dicf | 100 | 0.350 | 0.38 | 20.00 | 1.237 |
| dipa | 33 | 1.073 | 3.22 | 20.00 | 1.563 |
| dipp | 33 | 1.250 | 3.75 | 20.00 | 1.417 |
| hete | 66 | 0.200 | 0.45 | 6.67 | 0.427 |
| nolu | 100 | 0.077 | 0.15 | 6.67 | 0.403 |
| hylo | 100 | 13.793 | 22.23 | 88.33 | 8.193 |
| hypu | 66 | 0.300 | 0.75 | 3.33 | 0.270 |
| isop | 33 | 0.010 | 0.03 | 1.67 | 0.113 |
| isst | 100 | 5.443 | 7.78 | 31.67 | 3.040 |
| kurz | 33 | 0.033 | 0.10 | 6.67 | 0.377 |
| plag | 33 | 0.010 | 0.03 | 1.67 | 0.093 |
| plun | 100 | 2.523 | 3.42 | 71.67 | 4.863 |
| rhgl | 100 | 0.353 | 0.70 | 20.00 | 1.223 |
| rhio | 100 | 19.797 | 30.17 | 81.67 | 9.187 |
| scab | 33 | 1.657 | 4.97 | 23.33 | 1.630 |
| sphg | 66 | 1.717 | 4.40 | 5.00 | 0.663 |
| stor | 100 | 12.870 | 20.40 | 93.33 | 8.333 |
| bare rock |  |  |  |  |  |
| rock | 33 | 0.877 | 2.63 | 3.33 | 0.000 |

totals:
m.cov max.cov t.spp m.spp

| shrubs | 124.70 | 160.8 | 6 | 5.0 |
| :--- | ---: | ---: | ---: | ---: |
| herbs | 33.97 | 63.6 | 7 | 3.7 |
| bryo. | 62.40 | 85.6 | 20 | 13.0 |
| a11 spp. | 221.07 | 230.7 | 33 | 21.7 |



Appendix 3 : Coastal montane Thuja forests (T3)

plots: 60.70 147152153154172

|  | cons. | m.cov | max.cov | m.fre. | m.i.v |
| :--- | ---: | ---: | ---: | ---: | ---: |
| shrubs |  |  |  |  |  |
| gash | 100 | 45.101 | 74.38 | 80.00 | 15.684 |
| mefe | 85 | 4.109 | 13.77 | 19.43 | 1.930 |
| rusp | 57 | 0.011 | 0.03 | 1.71 | 0.083 |
| sosi | 14 | 0.001 | 0.01 | 0.14 | 0.006 |
| vaal | 100 | 14.906 | 35.38 | 39.29 | 5.390 |
| vaol | 71 | 2.479 | 14.40 | 7.43 | 0.993 |
| vapa | 100 | 13.599 | 20.63 | 64.29 | 7.244 |
| herbs |  |  |  |  |  |
| blsp | 100 | 47.724 | 67.90 | 83.57 | 17.346 |
| boye | 28 | 0.003 | 0.01 | 0.29 | 0.016 |
| coas | 57 | 2.987 | 8.35 | 24.43 | 2.091 |
| come | 14 | 0.001 | 0.01 | 0.14 | 0.009 |
| corn | 71 | 2.086 | 7.50 | 23.57 | 1.699 |
| fesu | 14 | 0.004 | 0.03 | 0.71 | 0.044 |
| goob | 28 | 0.164 | 1.00 | 5.00 | 0.259 |
| libo | 42 | 1.307 | 7.63 | 7.86 | 0.813 |
| lica | 71 | 0.034 | 0.10 | 6.43 | 0.379 |
| lico | 85 | 0.063 | 0.22 | 12.29 | 0.686 |
| lycl | 14 | 0.107 | 0.75 | 0.71 | 0.057 |
| lysi | 42 | 1.197 | 6.32 | 6.57 | 0.759 |
| madi | 71 | 1.804 | 5.60 | 29.43 | 1.873 |
| mone | 14 | 0.001 | 0.01 | 0.14 | 0.009 |
| pomu | 42 | 0.717 | 5.00 | 1.71 | 0.337 |
| pral | 14 | 0.107 | 0.75 | 0.71 | 0.071 |
| rupe | 57 | 2.747 | 11.95 | 21.57 | 1.634 |
| stra | 85 | 0.347 | 1.50 | 3.29 | 0.253 |
| tila | 85 | 1.143 | 7.30 | 15.29 | 1.333 |
| titr | 85 | 0.764 | 3.80 | 13.14 | 0.997 |
| trov | 42 | 0.024 | 0.15 | 1.00 | 0.053 |
| vevi | 28 | 0.066 | 0.45 | 2.29 | 0.113 |
| vigl | 14 | 0.001 | 0.01 | 0.14 | 0.009 |
| vise | 14 | 2.411 | 16.88 | 9.29 | 1.531 |

bryophytes and lichens

| bazz | 28 | 0.376 | 1.88 | 1.43 | 0.153 |
| :--- | ---: | ---: | ---: | ---: | ---: |
| ceph | 71 | 2.351 | 6.38 | 33.57 | 2.193 |
| dicf | 42 | 0.133 | 0.75 | 2.14 | 0.147 |
| dipa | 14 | 0.129 | 0.90 | 1.43 | 0.119 |
| holu | 100 | 0.543 | 0.85 | 33.57 | 1.797 |
| hylo | 85 | 3.686 | 14.52 | 34.29 | 2.469 |
| hypu | 85 | 0.326 | 1.05 | 5.71 | 0.404 |
| hypv | 14 | 0.021 | 0.15 | 0.71 | 0.040 |
| isop | 57 | 0.171 | 0.75 | 5.71 | 0.379 |
| isst | 100 | 2.646 | 9.13 | 21.43 | 2.004 |
| jule | 14 | 0.190 | 1.33 | 9.29 | 0.414 |
| metz | 14 | 0.004 | 0.03 | 0.71 | 0.044 |
| myta | 14 | 0.150 | 1.05 | 2.14 | 0.139 |
| nasc | 14 | 0.026 | 0.18 | 1.43 | 0.063 |
| peli | 57 | 0.181 | 0.93 | 4.43 | 0.259 |
| plag | 42 | 0.057 | 0.20 | 4.29 | 0.247 |
| plun | 100 | 4.159 | 7.20 | 83.57 | 5.509 |
| pogc | 14 | 0.043 | 0.30 | 1.43 | 0.067 |
| rhaa | 14 | 0.004 | 0.03 | 0.71 | 0.043 |




| isst | 89 | 1.412 | 4.28 | 15.26 | 1.403 |
| :---: | :---: | :---: | :---: | :---: | :---: |
| jule | 5 | 0.026 | 0.50 | 1.32 | 0.070 |
| kurz | 10 | 0.205 | 3.30 | 2.63 | 0.285 |
| 1 edo | 5 | 0.168 | 3.20 | 3.95 | 0.237 |
| leme | 10 | 0.048 | 0.77 | 0.79 | 0.047 |
| myta | 5 | 0.002 | 0.03 | 0.25 | 0.014 |
| nasc | 5 | 0.001 | 0.01 | 0.05 | 0.003 |
| peli | 15 | 0.445 | 8.05 | 3.16 | 0.235 |
| plag | 63 | 0.415 | 3.65 | 11.84 | 0.812 |
| plun | 100 | 4.122 | 9.63 | 78.68 | 6.202 |
| pogm | 5 | 0.001 | 0.01 | 0.05 | 0.004 |
| pore | 5 | 0.002 | 0.03 | 0.26 | 0.013 |
| ringl | 94 | 5.348 | 32.00 | 51.58 | 4.340 |
| rhio | 94 | 5.935 | 25.90 | 42.37 | 3.887 |
| ricl | 42 | 0.834 | 8.57 | 10.05 | 0.677 |
| scab | 78 | 3. 184 | 19.00 | 41.05 | 3.797 |
| sphf | 5 | 0.288 | 5.47 | 1.58 | 0.126 |
| sphg | 10 | 0.040 | 0.75 | 0.32 | 0.027 |
| sphn | 5 | 0.016 | 0.30 | 0.53 | 0.027 |
| stor | 100 | 12.654 | 26.58 | 80.79 | 8.307 |
| totals: |  |  |  |  |  |
|  |  | m.cov | $x . c o v$ | t.spp | m.spp |
| shrubs |  | 87.09 | 124.6 | 10 | 6.1 |
| nerbs |  | 65.19 | 91.4 | 24 | 4.8 |
| bryo. |  | 47.07 | 92.9 | 36 | 12.4 |
| all spp. |  | 199.33 | 269.3 | 70 | 23.3 |


| Appendix 3 | : Coas | al w | $t$ Thuja | forests | s (T5) |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | plots: | 4748 | 77. 148 |  |  |  |
|  |  | cons. | m.cov | max.cov | m.fre. | . m.i.v |
|  | shrubs gash | 100 | 75.290 | 80.63 | 98.75 | 17.402 |
|  | mefe | 100 | 5.165 | 7.28 | 23.75 | 1.960 |
|  | pyus | 100 | 7.660 | 13.13 | 14.00 | 1.938 |
|  | rham | 25 | 0.783 | 3.13 | 1.25 | 0.182 |
|  | rusp | 75 | 1.615 | 5.43 | 22.50 | 1.295 |
|  | vaal | 100 | 5.663 | 8.52 | 17.50 | 1.745 |
|  | vaol | 25 | 0.225 | 0.90 | 2.50 | 0.165 |
|  | vaot | 100 | 33.693 | 50.67 | 65.00 | 8.590 |
|  | vapa | 100 | 13.230 | 19.33 | 70.00 | 5.410 |
|  | herbs |  |  |  |  |  |
|  | blsp | 100 | 64.910 | 78.75 | 96.25 | 15.515 |
|  | boho | 50 | 0.015 | 0.05 | 2.75 | 0.118 |
|  | caia | 50 | 0.970 | 3. 13 | 2.50 | 0.270 |
|  | caro | 75 | 5.255 | 14.38 | 11.50 | 1.432 |
|  | carr | 25 | 0.658 | 2.63 | 2.50 | 0.217 |
|  | corn | 100 | 9.080 | 10.95 | 51.25 | 3.840 |
|  | eqte | 25 | 0.038 | 0. 15 | 1.25 | 0.067 |
|  | 1 ibo | 100 | 5.430 | 12.18 | 46.25 | 2.900 |
|  | lico | 25 | 0.008 | 0.03 | 1.25 | 0.052 |
|  | lys | 25 | 0.188 | 0.75 | 1.25 | 0.095 |
|  | madi | 100 | 1.948 | 3.13 | 47.50 | 2.367 |
|  | stra | 25 | 0.038 | 0.15 | 1.25 | 0.067 |
|  | vevi | 50 | 0.005 | 0.01 | 0.50 | 0.023 |
|  | kryoph | tes a | and lichen |  |  |  |
|  | bazz | 25 | 0.013 | 0.05 | 2.50 | 0.125 |
|  | blet | 50 | 0.920 | 3.63 | 18.75 | 0.962 |
|  | ceph | 75 | 2.045 | 5.63 | 28.75 | 1.608 |
|  | dicf | 25 | 0.055 | 0.22 | 5.00 | 0.220 |
|  | dics | 75 | 0.158 | 0.30 | 6.25 | 0.308 |
|  | dipa | 75 | 0.933 | 2.05 | 31.25 | 1.480 |
|  | d\$pp | 50 | 0.645 | 2.13 | 12.50 | 0.702 |
|  | heba | 75 | 0.863 | 3.00 | 8.75 | 0.515 |
|  | holu | 100 | 0.425 | 0.90 | 17.50 | 0.848 |
|  | nylo | 100 | 7.750 | 11.35 | 81.25 | 4.883 |
|  | hypu | 25 | 0.038 | 0.15 | 2.50 | 0.115 |
|  | myta | 50 | 0.195 | 0.63 | 7.50 | 0.347 |
|  | plag | 75 | 0.445 | 1.55 | 15.00 | 0.723 |
|  | plun | 100 | 3.635 | 6.57 | 87.50 | 4.475 |
|  | rhgl | 100 | 1.410 | 4.57 | 32.50 | 1.645 |
|  | rhio | 100 | 11.880 | 14.43 | 80.00 | 5.530 |
|  | ricm | 25 | 0.563 | 2.25 | 3.75 | 0.287 |
|  | scab | 25 | 2. 138 | 8.55 | 21.25 | 1.247 |
|  | sphg | 75 | 3.272 | 12.88 | 11.50 | 1.035 |
|  | sphh | 25 | 0.003 | 0.01 | 0.25 | 0.013 |
|  | stor | 75 | 17.143 | 36.25 | 62.50 | 5.540 |
|  | stpr | 25 | 3.095 | 12.38 | 23.75 | 1.735 |
|  | totals |  |  |  |  |  |
|  |  |  | m.cov max | ax.cov t | $t . \operatorname{spp} \mathrm{m}$ | m.spp |
|  | shrub |  | 143.30 | 152.5 | 9 | 7.3 |
|  | herbs |  | 88.55 | 102.9 | 13 | 7.5 |
|  | bryo |  | 57.63 | 81.5 | $22$ | 13.5 |
|  | all | pp. | 289.42 | 300.3 | 442 | 28.3 |



Appendix 3 : Montane Abies-Tsuga forests (A2)


Appendix 3 : Montane Tsuga-Abies forests (A3)

|  | cons. | m.cov | max.cov | m.fre. | m.i.v |
| :---: | :---: | :---: | :---: | :---: | :---: |
| shrubs |  |  |  |  |  |
| gasn | 50 | 0.005 | 0.01 | 0.50 | 0.048 |
| mefe | 25 | 0.003 | 0.01 | 0.25 | 0.033 |
| rusp | 100 | 1.645 | 6.25 | 12.50 | 2.668 |
| vaal | 100 | 2.123 | 5.55 | 12.75 | 2.405 |
| vapa | 100 | 21.640 | 55.42 | 63.75 | 20.133 |
| nerbs |  |  |  |  |  |
| actr | 25 | 0.475 | 1.90 | 10.00 | 1.283 |
| blsp | 75 | 11.115 | 20.88 | 33.75 | 11.478 |
| chum | 25 | 0.003 | 0.01 | 0.25 | 0.025 |
| dino | 25 | 0.380 | 1.52 | 3.75 | 0.650 |
| drau | 25 | 0.003 | 0.01 | 0.25 | 0.025 |
| goob | 25 | 0.003 | 0.01 | 0.25 | 0.025 |
| hypo | 50 | 0.010 | 0.03 | 1.50 | 0.130 |
| 1 amu | 25 | 0.188 | 0.75 | 1.25 | 0. 268 |
| 1tca | 50 | 0.015 | 0.05 | 2.75 | 0.255 |
| mad 1 | 25 | 0.003 | 0.01 | 0.25 | 0.025 |
| poly | 25 | 0.008 | 0.03 | 1.25 | 0.115 |
| pomu | 100 | 1.863 | 4.63 | 9.00 | 2.583 |
| stra | 75 | 0.008 | 0.01 | 0.75 | 0.083 |
| tila | 50 | 0.005 | 0.01 | 0.50 | 0.058 |
| titr | 25 | 0.008 | 0.03 | 1.25 | 0.115 |
| trov | 75 | 0.080 | 0.30 | 3.00 | 0.343 |


| bryophytes and | lichens |  |  |  |  |
| :--- | ---: | :--- | ---: | ---: | ---: |
| anti | 25 | 0.237 | 0.95 | 10.00 | 1.085 |
| blet | 25 | 0.050 | 0.20 | 3.75 | 0.332 |
| ceph | 50 | 2.445 | 8.40 | 18.75 | 3.005 |
| dicf | 50 | 0.180 | 0.47 | 23.75 | 2.100 |
| dipa | 50 | 1.592 | 3.97 | 26.25 | 4.238 |
| hete | 25 | 1.875 | 7.50 | 2.50 | 1.780 |
| holu | 75 | 0.048 | 0.15 | 2.75 | 0.325 |
| hylo | 25 | 1.133 | 4.53 | 13.75 | 1.885 |
| hypu | 75 | 2.375 | 7.85 | 31.25 | 4.813 |
| isop | 100 | 1.953 | 4.28 | 28.75 | 4.355 |
| isst | 75 | 2.390 | 4.13 | 31.25 | 4.302 |
| ledo | 75 | 0.487 | 1.05 | 6.25 | 0.955 |
| loba | 25 | 0.038 | 0.15 | 1.25 | 0.143 |
| mniu | 25 | 0.038 | 0.15 | 1.25 | 0.143 |
| pelt | 25 | 0.008 | 0.03 | 1.25 | 0.115 |
| plun | 100 | 5.130 | 6.78 | 66.25 | 9.823 |
| pogc | 25 | 0.013 | 0.05 | 2.50 | 0.313 |
| pogm | 25 | 0.038 | 0.15 | 1.25 | 0.135 |
| rhaa | 25 | 0.038 | 0.15 | 1.25 | 0.143 |
| rhgl | 75 | 4.260 | 13.13 | 32.50 | 5.755 |
| rhlo | 50 | 2.640 | 10.55 | 10.25 | 2.418 |
| scab | 75 | 5.895 | 9.77 | 53.75 | 8.255 |
| stor | 25 | 0.003 | 0.01 | 0.25 | 0.025 |
| bare rock |  |  |  |  | $\ldots$ |

totals:
m.cov max.cov t.spp m.spp

| shrubs | 25.40 | 61.1 | 5 | 3.8 |
| :--- | ---: | ---: | ---: | ---: |
| herbs | 14.15 | 21.3 | 16 | 7.0 |
| bryo. | 32.83 | 56.3 | 23 | 11.3 |
| all spp. | 72.43 | 89.4 | 44 | 22.0 |



Note : Pelt = Peltigera membranacea (25)

Appendix 3 : Lowland Abies forests (A5)

|  | cons. | m.cov | max.cov | m.fre. | m.i.v |
| :---: | :---: | :---: | :---: | :---: | :---: |
| shrubs |  |  |  |  |  |
| gash | 41 | 1.302 | 10.38 | 6.42 | 0.930 |
| mefe | 41 | 0.160 | 1.88 | 0.75 | 0.094 |
| opho | 16 | 0.002 | 0.01 | 0.17 | 0.012 |
| ribb | 8 | 0.001 | 0.01 | 0.08 | 0.004 |
| rusp | 91 | 1.373 | 4.38 | 14.58 | 1.339 |
| samr | 8 | 0.001 | 0.01 | 0.08 | 0.004 |
| vaal | 100 | 24.761 | 76.50 | 57.08 | 13.152 |
| vaol | 66 | 2.798 | 13.27 | 9.17 | 1.428 |
| vapa | 100 | 10.107 | 22.10 | 56.67 | 8.363 |

herbs actr adb 1 adpe atfi blsp boye clun coas coma corn dino drau gali goob gyar heco hypo itca luzp madi miov mone poam pomu pral pyse rupe smst stra strr strs tila titr trau trov vigl
vise 50
25
16
50
100
16
8
16
16
8
16
8
66
25
8
25
8
25

| 25 | 0 |
| ---: | ---: |
| 8 | 0 |
| 16 | 0 |


| bryophytes and |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: |
| bazz | 16 | 0.307 | 2.63 | 2.08 | 0.239 |
| blet | 25 | 0.096 | 0.75 | 8.75 | 0.682 |
| calm | 16 | 0.336 | 3.85 | 5.42 | 0.503 |
| ceph | 83 | 2.693 | 7.13 | 32.08 | 3.453 |
| coco | 8 | 0.001 | 0.01 | 0.08 | 0.005 |
| dicf | 33 | 0.099 | 0.47 | 7.50 | 0.542 |
| dipa | 16 | 0.672 | 7.13 | 7.92 | 0.933 |
| hete | 8 | 0.003 | 0.03 | 0.42 | 0.035 |
| nolu | 75 | 0.707 | 3.45 | 15.83 | 1.214 |
| hylo | 83 | 2.868 | 9.27 | 22.17 | 2.663 |
| nypp | 8 | 0.015 | 0.18 | 0.83 | 0.042 |
| hypu | 25 | 0.382 | 2.25 | 5.42 | 0.548 |
| isop | 66 | 0.582 | 1.95 | 8.75 | 0.779 |
| isst | 83 | 1.842 | 6.38 | 22.92 | 2.707 |
| ledo | 8 | 0.025 | 0.30 | 0.83 | 0.074 |
| leme | 25 | 0.014 | 0.15 | 0.58 | 0.035 |
| peli | 25 | 0.353 | 3.90 | 2.50 | 0.246 |
| pelt | 16 | 0.038 | 0.30 | 1.25 | 0.080 |
| plag | 58 | 0.117 | 0.93 | 6.67 | 0.483 |
| plin | 8 | 0.013 | 0.15 | 0.42 | 0.022 |
| piun | 100 | 7.685 | 19.30 | 76.67 | 8.423 |
| poge | 33 | 0.090 | 0.77 | 1.75 | 0.137 |
| pogm | 8 | 0.158 | 1.90 | 0.83 | 0.076 |
| ptic | 8 | 0.063 | 0.75 | 0.42 | 0.067 |
| rhgi | 100 | 6.700 | 15.10 | 60.00 | 6.698 |
| rhlo | 100 | 16.952 | 44.20 | 64.58 | 10.271 |
| ricl | 25 | 0.763 | 8.10 | 5.83 | 0.664 |
| scab | 91 | 1.651 | 5.70 | 36.33 | 2.916 |
| sphg | 33 | 1.662 | 19.88 | 5.92 | 0.780 |
| stor | 75 | 7.140 | 20.65 | 40.42 | 6.358 |
| bare rock |  |  |  |  |  |
| rock | 33 | 0.475 | 3.90 | 4.17 | 0.000 |
| totals: |  |  |  |  |  |
|  |  | m.cov max.cov |  | t.spp | m.spp |
| shrubs |  | 40.51 | 84.1 | 9 | 4.8 |
| herbs |  | 34.90 | 103.7 | 41 | 10.9 |
| bryo. |  | 54.01 | 76.5 | 30 | 12.7 |
| all spp. |  | 129.40 | 214.1 | 80 | 28.3 |

Note : Pelt = Peltigera membranacea (29)
and P . polydactyla (45)

Appendix 3 : Tsuga-Gaultheria-Blechnum forests (A6)

|  | cons. | m.cov | max.cov | m.fre. | . m.i.v |
| :---: | :---: | :---: | :---: | :---: | :---: |
| shrubs |  |  |  |  |  |
| gash | 100 | 43.525 | 55.25 | 77.50 | 19.610 |
| mefe | 50 | 2.190 | 4.38 | 2.50 | 0.810 |
| vaal | 100 | 6.500 | 7.25 | 20.00 | 3.730 |
| vapa | 100 | 28.260 | 35.42 | 82.50 | 15.355 |
| nerbs |  |  |  |  |  |
| actr | 50 | 0.375 | 0.75 | 2.50 | 0.350 |
| atfi | 50 | 0.005 | 0.01 | 0.50 | 0.045 |
| blsp | 100 | 15.285 | 28.17 | 45.00 | 9.040 |
| dism | 50 | 0.005 | 0.01 | 0.50 | 0.045 |
| 1 ico | 50 | 0.015 | 0.03 | 2.50 | 0.225 |
| pomu | 100 | 0.080 | 0.15 | 3.00 | 0.280 |
| tila | 50 | 0.075 | 0.15 | 2.50 | 0.245 |
| titr | 50 | 0.005 | 0.01 | 0.50 | 0.045 |
| trau | 50 | 0.375 | 0.75 | 2.50 | 0.350 |
| trov | 50 | 0.150 | 0.30 | 5.00 | 0.490 |
| bryophytes and |  | 1 ichens |  |  |  |
| bazz | 50 | 0.375 | 0.75 | 2.50 | 0.350 |
| holu | 50 | 0.015. | 0.03 | 2.50 | 0.180 |
| hylo | 50 | 6.840 | 13.68 | 30.00 | 5.020 |
| hypu | 100 | 0.165 | 0.18 | 7.50 | 0.670 |
| isop | 50 | 0.375 | 0.75 | 2.50 | 0.285 |
| isst | 100 | 1.930 | 2.78 | 17.50 | 2.015 |
| $l$ edo | 50 | 3.000 | 6.00 | 30.00 | 3.010 |
| plag | 100 | 0.150 | 0.15 | 5.00 | 0.445 |
| plun | 100 | 10.725 | 11.85 | 82.50 | 9.815 |
| ponf | 50 | 0.075 | 0.15 | 2.50 | 0.245 |
| ring 1 | 100 | 5.965 | 11.63 | 52.50 | 5.565 |
| rhlo | 100 | 1.200 | 1.50 | 10.00 | 1. 185 |
| scab | 100 | 6.680 | 10.48 | 65.00 | 6.990 |
| stor | 100 | 23.825 | 26.27 | 77.50 | 13.595 |
| totals: |  |  |  |  |  |
|  |  | m.cov max.cov |  | t.spp m | m. spp |
| shrubs |  | 80.50 | 100.8 | 4 | 3.5 |
| nerbs |  | 16.35 | 30.3 | 10 | 6.0 |
| bryo. |  | 61.30 | 69.3 | 14 | 11.0 |
| all spp. |  | 158.15 | 172.5 | 28 | 20.5 |

Appendix 3 : Tsuga-Blechnum-Polystichum forests (A7) plots: 56879199107

|  | cons. | m.cov | max.cov | m.fre. | m.i.v |
| :---: | :---: | :---: | :---: | :---: | :---: |
| shrubs |  |  |  |  |  |
| gash | 40 | 0.490 | 2.25 | 6.00 | 0.834 |
| rusp | 80 | 0.066 | 0.22 | 13.00 | 1. 110 |
| samr | 20 | 0.002 | 0.01 | 0.20 | 0.016 |
| vaal | 100 | 5.302 | 18.30 | 18.20 | 4.684 |
| vapa | 100 | 8.828 | 18.35 | 39.00 | 7.988 |
| nerbs |  |  |  |  |  |
| actr | 40 | 0.192 | 0.95 | 4.20 | 0.452 |
| adpe | 20 | 0.002 | 0.01 | 0.20 | 0.024 |
| atfi | 60 | 0.408 | 1.88 | 2.20 | 0.354 |
| blsp | 100 | 25.002 | 47.08 | 63.00 | 17.902 |
| dism | 20 | 0.030 | 0.15 | 1.00 | 0.100 |
| drau | 60 | 0.154 | 0.75 | 1.40 | 0.182 |
| 1 l | 20 | 0.002 | 0.01 | 0.20 | 0.024 |
| luzp | 20 | 0.006 | 0.03 | 1.00 | 0.096 |
| madi | 20 | 0.016 | 0.08 | 3.00 | 0.250 |
| mone | 20 | 0.006 | 0.03 | 1.00 | 0.112 |
| pomu | 100 | 28.820 | 50.38 | 52.00 | 19.048 |
| ptaq | 20 | 0.510 | 2.55 | 5.00 | 0.748 |
| stra | 60 | 0.182 | 0.75 | 2.20 | 0.270 |
| strr | 20 | 0.002 | 0.01 | 0.20 | 0.018 |
| tila | 60 | 0.048 | 0.20 | 4.20 | 0.358 |
| titr | 80 | 0.162 | 0.35 | 7.20 | 0.712 |
| trov | 60 | 0.416 | 1.92 | 10.20 | 1.080 |
| bryophytes and lichens |  |  |  |  |  |
| bazz | 20 | 0.030 | 0.15 | 1.00 | 0.090 |
| calm | 40 | 0.258 | 0.77 | 12.00 | 1.076 |
| ceph | 80 | 1.326 | 2.88 | 25.00 | 2.864 |
| cloa | 20 | 0.030 | 0.15 | 1.00 | 0.030 |
| dicf | 20 | 0.076 | 0.38 | 5.00 | 0.510 |
| hete | 80 | 0.500 | 1.80 | 12.00 | 1.364 |
| holu | 80 | 0.226 | 0.75 | 11.00 | 1.070 |
| hylo | 20 | 0.040 | 0.20 | 3.00 | 0.268 |
| hypu | 80 | 1.300 | 4.20 | 17.00 | 2.322 |
| isop | 40 | 0.616 | 2.03 | 5.00 | 0.858 |
| isst | 100 | 3.666 | 8.05 | 38.00 | 5.328 |
| leme | 20 | 0.030 | 0.15 | 1.00 | 0.090 |
| plag | 20 | 0.006 | 0.03 | 1.00 | 0.092 |
| plun | 80 | 4.106 | 13.25 | 49.00 | 6.632 |
| pogm | 40 | 0.072 | 0. 18 | 4.00 | 0.404 |
| poro | 20 | 0.180 | 0.90 | 2.00 | 0.236 |
| rhgl | 80 | 1.734 | 3.05 | 35.00 | 3.804 |
| rhio | 80 | 1.760 | 6.20 | 13.00 | 1.934 |
| ricl | 20 | 0.076 | 0.38 | 5.00 | 0.410 |
| scab | 100 | 3.796 | 6.25 | 53.00 | 6.796 |
| stor | 100 | 2.262 | 5.50 | 27.00 | 3.648 |
| bare rock |  |  |  |  |  |
| rock | 60 | 4.776 | 17.50 | 13.00 | 0.000 |

totals:
m.Cov max.cov t.spp m.spp

| shrubs | 14.68 | 20.6 | 5 | 3.4 |
| :--- | ---: | ---: | ---: | ---: |
| herbs | 55.96 | 102.4 | 17 | 7.8 |
| bryo. | 22.08 | 28.6 | 21 | 11.4 |
| all spp. | 92.70 | 127.1 | 43 | 22.6 |

Appendix 4 : Discriminant analysis results.

Table 1 : Forward stepwise discriminant analysis of vegetation groups based on environmental data.

Step forward separation:

| Variables $\quad$ F | F-statistic | Signif. |
| :---: | :---: | :---: |
| 1 - Drainage | 32.80 | . 000 |
| 2 - Elevation | 18.58 | . 000 |
| 3 - Topographic position | 14.03 | . 000 |
| 4 - LFH thick./effect. root. depth | h 10.14 | . 000 |
| 5 - Wind | 9.54 | . 000 |
| 6 - Slope (\%) | 6.28 | . 000 |
| 7 - B1 \% coarse fragments | 5.84 | . 000 |
| 8 - LFH $\mathrm{pH}\left(\mathrm{CaCl}_{2}\right)$ | 6.15 | . 000 |
| $9-\mathrm{LFH} \mathrm{C} / \mathrm{N}$ | 5.88 | . 000 |
| $10-\mathrm{B}_{1} \mathrm{C} / \mathrm{N}$ | 4.29 | . 001 |
| 11 - Aspect | 4.30 | . 001 |
| 12 - Root restricting depth | 3.40 | . 002 |
| 13 - Worms | 4.08 | . 002 |
| 14 - Fire | 3.46 | . 006 |

Table 2 : Classification of vegetation plots into groups using discriminant functions (plots $=167$; non-classified plots $=10$; total classified plots $=157$ ).

Group $\underline{n} \quad$ Classification success (\%)

| Subalpine | 10 | 90.0 \% |
| :---: | :---: | :---: |
| Floodplain | 10 | 100.0 \% |
| Pinus | 5 | 100.0 \% |
| Pseudotsuga | 55 | 90.9 \% |
| Thuja | 37 | 86.5 \% |
| Abies | 40 | 60.0\% |
| Total | 157 | 82.8 \% |

## Appendix 4 (continued)

Table 3 : Forward stepwise discriminant analysis of the community types of the Pseudotsuga group based on environmental data.

Step forward separation :

| Variables | F-statistic | Signif. |
| :--- | :---: | :---: |
| 1 - Topographic position | 8.08 | .000 |
| 2 - Elevation | 6.01 | .000 |
| 3 - B1 \% C | 5.12 | .000 |
| $4-$ LFH pH $\left(\mathrm{CaCl}_{2}\right)$ | 3.91 | .003 |

Table 4 : Classification of Pseudotsuga group plots into types using discriminant functions (plots $=60$; non-classified plots $=4$; total classified plots $=56$ ).

| Type | $\underline{n}$ | Classification success (\%) |
| :--- | ---: | :--- |
|  |  |  |
| P1 | 4 | $75.0 \%$ |
| P2 | 4 | $100.0 \%$ |
| P3 | 5 | $40.0 \%$ |
| P4 | 11 | $54.5 \%$ |
| P5 | 14 | $71.4 \%$ |
| P6 | 11 | $63.6 \%$ |
| P7 | 7 | $42.8 \%$ |
| Total | 56 | $62.5 \%$ |

Table 5 : Forward stepwise discriminant analysis of the community types of the Thuja group based on environmental data.

Step forward separation :

| Variables |  |  |  | F-statistic | Signif. |
| :--- | :---: | :---: | :---: | :---: | :---: |
|  |  |  |  |  |  |
| $2-$ Elevation | 19.17 | .000 |  |  |  |
| $2-\mathrm{B}_{1} \% \mathrm{~N}$ | 5.81 | .003 |  |  |  |

## Appendix 4 (continued)

Table 6 : Classification of Thuja group plots into types using discriminant functions (plots $=39$; non-classified plots $=3$; classified plots $=36$ ).

| Type | $\underline{n}$ | Classification success (\%) |
| :---: | ---: | :---: |
|  |  |  |
| T2 | 6 | $33.3 \%$ |
| T3 | 7 | $85.7 \%$ |
| T4 | 19 | $68.4 \%$ |
| T5 | 4 | $100.0 \%$ |
| Tota1 | 36 | $69.4 \%$ |

Table 7 : Forward stepwise discriminant analysis of the community types of the Abies group based on environmental data.

Step forward separation :

| Variables | F-statistic | Signif. |
| :--- | :---: | :---: |
|  |  |  |
| 1 - Elevation | 10.87 | .000 |
| 2 - Slope (\%) | 3.86 | .008 |
| 3 - LFH thickness | 3.38 | .015 |
| 4 - Texture | 2.68 | .042 |

Table 8 : Classification of Abies group p1ots into types using discriminant functions (plots $=32$ ).

| Type | $\underline{n}$ | Classification success (\%) |
| :--- | ---: | :--- |
|  |  |  |
| A1 | 3 | $33.3 \%$ |
| A2 | 2 | $100.0 \%$ |
| A3 | 4 | $50.0 \%$ |
| A4 | 4 | $100.0 \%$ |
| A5 | 12 | $66.6 \%$ |
| A6 | 2 | $100.0 \%$ |
| A7 | 5 | $40.0 \%$ |
| Total | 32 | $65.6 \%$ |

## Appendix 4 (continued)

Table 9 : Forward stepwise discriminant analysis of community types based on environmental data.

Step forward separation :

| Variables F - | F-statistic | Signif. |
| :---: | :---: | :---: |
| 1 - Elevation | 16.38 | . 000 |
| 2 - Drainage | 12.27 | . 000 |
| 3 - Topographic position | 7.35 | . 000 |
| $4-\mathrm{LFH} \mathrm{pH}\left(\mathrm{CaCl}_{2}\right)$ | 5.49 | . 000 |
| 5 - LFH C/N | 4.43 | . 000 |
| 6 - Slope (\%) | 4.28 | . 000 |
| 7 - Fire | 4.06 | . 000 |
| $8-\mathrm{B}_{1} \mathrm{C} / \mathrm{N}$ | 3.35 | . 000 |
| $9-\mathrm{B}_{1} \% \mathrm{~N}$ | 2.57 | . 001 |
| $10-\mathrm{B}_{1} \%$ coarse fragments | 2.12 | . 006 |
| 11 - LFH thick./effect. root. depth | 1.91 | . 015 |
| 12 - Wind | 1.91 | . 015 |
| 13 - Aspect | 2.03 | . 008 |

## Appendix 4 (continued)

## Table 10 : Classification of plots into types using discriminant functions (plots = 159; non-classified plots = 10; total classified plots $=149$ ).

| Type | $\underline{n}$ | Classification success (\%) |
| :--- | ---: | :---: |
|  | $\underline{n}$ |  |
| Sub-alpine | 10 | $90.0 \%$ |
| F1 | 8 | $100.0 \%$ |
| F2 | 2 | $100.0 \%$ |
| D1 | 2 | $100.0 \%$ |
| D2 | 3 | $100.0 \%$ |
| P1 | 4 | $50.0 \%$ |
| P2 | 4 | $75.0 \%$ |
| P3 | 5 | $80.0 \%$ |
| P4 | 11 | $54.5 \%$ |
| P5 | 14 | $78.6 \%$ |
| P6 | 11 | $54.5 \%$ |
| P7 | 7 | $28.6 \%$ |
| T2 | 6 | $50.0 \%$ |
| T3 | 7 | $85.7 \%$ |
| T4 | 19 | $73.7 \%$ |
| T5 | 4 | $75.0 \%$ |
| A1 | 3 | $66.6 \%$ |
| A2 | 2 | $100.0 \%$ |
| A3 | 4 | $100.0 \%$ |
| A4 | 4 | $50.0 \%$ |
| A5 | 12 | $100.0 \%$ |
| A6 | 2 | $80.0 \%$ |
| A7 | 5 |  |
|  |  | $72.5 \%$ |
| Tota1 | 149 |  |


[^0]:    1 Modal community types are also equivalent to ecosystem associations (Klinka et al., 1979).

[^1]:    1 Beta diversity $=(\log a-\log z) / \log 2$, where $a=$ replicate plots similarity, and $z=$ extreme plots similarity. Plot similarities were measured using the cosine function which reflects quantitative changes in species representation.

[^2]:    1 Includes tree, sapling and seedling strata
    2 (standard deviation)
    3 Includes tree stratum on 1 y
    4 Includes trees and saplings
    5 Antilog of Shannon's Index
    6 Reciprocal of Simpson's Index

[^3]:    1 (standard deviation) $\quad 2$ Antilog of Shannon's Index $\quad 3$ Reciprocal of Simpson's Index

[^4]:    ${ }^{1}$ Homogeneity coefficient
    2 Species richness, average for .05 ha
    ${ }^{3}$ Fire : $0=$ absence, $1=$ presence (S.D.)

[^5]:    ${ }^{1}$ Number of plots; ${ }^{2}$ Number of microplots; 3 z-value of two sample z-test $4 * * *=p \leqslant .001 ; * *=.001<p \leqslant .01 ; *=.01<p \leqslant .05 ;$ n.s. $=p>.05$

[^6]:    Note : Pelt = Peltigera membranacea (19, 39) and P . polydactyla (123, 166)

