CARBOCYCLE CONSTRUCTION IN TERPENOID SYNTHESIS. THE TOTAL SYNTHESES OF (±)-SARCODONIN G AND (±)-1-EPI-9-NORPRESILPHIPERFOLAN-9-ONE.

by

MICHAEL W. GILBERT

B.Sc., University of Ottawa, 1995

A THESIS SUBMITTED IN PARTIAL FULFILMENT OF THE REQUIREMENTS FOR THE DEGREE OF DOCTOR OF PHILOSOPHY

in

THE FACULTY OF GRADUATE STUDIES

(Department of Chemistry)

We accept this thesis as conforming

ي د

to the required standard

THE UNIVERSITY OF BRITISH COLUMBIA

JUNE, 2002

© Michael W. Gilbert, 2002

In presenting this thesis in partial fulfilment of the requirements for an advanced degree at the University of British Columbia, I agree that the Library shall make it freely available for reference and study. I further agree that permission for extensive copying of this thesis for scholarly purposes may be granted by the head of my department or by his or her representatives. It is understood that copying or publication of this thesis for financial gain shall not be allowed without my written permission.

Chemis

Department of _____

The University of British Columbiav Vancouver, Canada

Date June 11/2002

ABSTRACT

Carbocycle construction represents a significant challenge in synthetic studies geared towards terpenoid natural products. A step-wise annulation approach to the construction of the diterpenoid framework of the cyathanes (54) was investigated, ultimately leading to the first reported total synthesis of (\pm)-sarcodonin G (36). The sesquiterpenoid carbon skeleton of the presilphiperfolanes (270) was assembled *via* a tandem cyclization strategy, affording the known ketone, (\pm)-1-epi-9-norpresilphiperfolan-9-one (306).

The construction of the cyathane 5-6-7 fused tricyclic framework commenced with 3-methyl-2-cyclohexen-1-one **38**, representing the B-ring. The annulation sequences for A and C-rings utilized the bifunctional reagents **39** and **44**, respectively. With the help of these reagents, the 5-6-6 fused tricycle **155** was quickly assembled from **38**. Intermediate **155** was further elaborated, including a key SmI₂-mediated ring expansion, to afford the 5-6-7 fused tricycle **159**, which displays the complete cyathane framework. The target structure, (\pm)-sarcodonin G (**36**), was obtained in several steps from **159**. In addition, an advanced intermediate in the synthesis of **36** was transformed into substance **269**, a likely synthetic precursor of the corresponding keto triol natural product, (\pm)-cyathin A₄ (**37**).

Assembly of the presilphiperfolane 5-5-6 fused tricyclic carbon skeleton 270 was approached *via* a tandem cyclization strategy involving free-radical reactions. The key intermediates of this synthetic study, xanthates 367, were readily prepared from enone 49 in several steps. Bu₃SnH-mediated free-radical reaction of 367 yielded a mixture of

cyclization products, alkenes **368**, whose frameworks include the carbon skeleton of the natural product presilphiperfolan-9-ol (**53**). Alkenes **368** were subjected to oxidative cleavage conditions, affording the known ketone (\pm) -1-epi-9-norpresilphiperfolan-9-one (**306**).

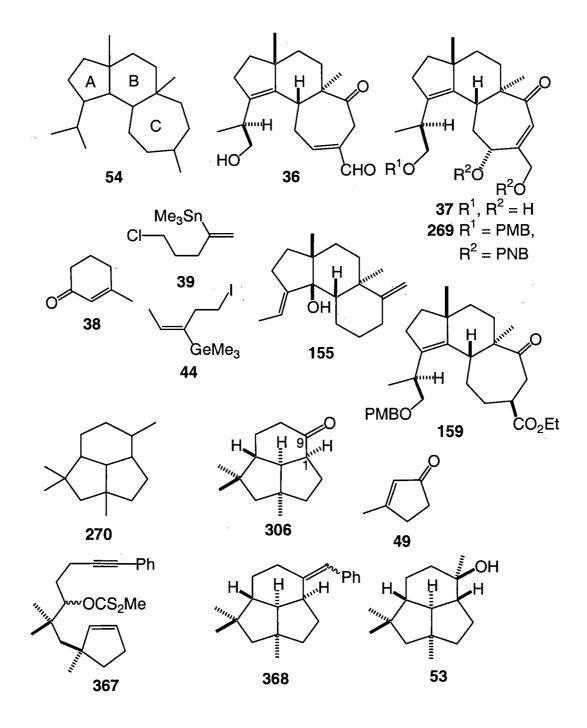


TABLE OF CONTENTS

ABSTRACT		ii
TABLE OF CONT	ENTS	iv
LIST OF TABLES.		vi
LIST OF FIGURES	5	ix
LIST OF ABBREV	IATIONS	x
ACKNOWLEDGE	MENTS	xiii
I. INTRODUC	TION	1
		1 10
-	AND DISCUSSION	10
2.1. Studies To	wards Cyathane Diterpenoid Synthesis	14
2.1.1. Backs	ground	14
2.1.1.1.	Cyathane History and Biogenesis	14
2.1.1.2.	Hydrocarbon Cyathanes	17
2.1.1.3.	Oxidized Cyathanes	17
2.1.1.4.	Biological Activity of the Cyathanes	. 24
2.1.2. Previ	ous Studies in Cyathane Synthesis	26
2.1.2.1.	Total Syntheses of Cyathanes	26
2.1.2.2.	Approaches to the Cyathanes	32
2.1.3. Total	Synthesis of (±)-Sarcodonin G	42
2.1.3.1.	Retrosynthetic Analysis	42
2.1.3.2.	Preparation of 5-6-6 Fused Tricycle 151	43
	2.1. Preparation of Bifunctional Reagent 44	44
2.1.3.	2.2. Preparation of <i>cis</i> -Fused Bicyclic	
	Dimethylhydrazone 102	55
2.1.3.	1	58
	2.4. Preparation of Alcohol 151	60
2.1.3.3.	Preparation of Ketone 112.	66
2.1.3.4.	Preparation of Iodides 160	76
2.1.3.5.	Preparation of Keto Ester 159.	83
2.1.3.6.	Preparation of Keto Aldehyde 240	88
2.1.3.7.	Preparation of (±)-Sarcodonin G (36)	98

--

	2.1.4.	Studi	ies Towards the Synthesis of (±)-Cyathin A ₄	103
	2.1	.4.1.	Retrosynthetic Analysis	103
2.1.4.2.		.4.2.	Preparation of Keto Alcohol 254	
	2.1	.4.3.	Attempted Preparation of Diol 231	110
2.2	. Stud	lies To	wards Presilphiperfolane	
			penoid Synthesis	122
	2.2.1.	Back	ground: Presilphiperfolane Isolation, Biogenesis	
			Biological Activity	122
	2.2.2.		ious Syntheses	126
	2.2.3.	Studi	ies Towards the Synthesis of	
		(±)-P	resilphiperfolan-9-ol	129
	2.2	2.3.1.	Retrosynthetic Analysis	130
	2.2	2.3.2.	Preparation of Ketone 315	132
	2.2	2.3.3.	Preparation of Aldehyde 314	135
	2.2	2.3.4.	Preparation of the Tandem Cyclization	
			Precursors, Xanthates 50	138
	2.2	2.3.5.	Tandem Free-radical Cyclization	140
III.	CON	CLUSI	IONS	155
IV.	EXPE	RIME	ENTAL	162
4.1	. Gen	eral		162
	4.1.1.	Data	Acquisition and Presentation	162
	4.1.2.	Solve	ents and Reagents	164
4.2			thesis of (±)-Sarcodonin G	167
			Studies Towards (±)-Cyathin A4	219
4.4	. Tota	al Synt	thesis of (±)-1-Epi-9-presilphiperfolan-9-one	232
V.	T) TATAT	זאקומי		040
V.	KEFE	KEIN	CES AND FOOTNOTES	249

v

LIST OF TABLES

Table 1.	Addition of Germylcopper(I) Reagents (167-170) to Ethyl 2-Butynoate (171)		
Table 2.	Isomerization of C–3 Substituted Cycloheptenones 251 and 252	96	
Table 3.	Comparison of IR and MS Spectral Data for Synthetic (±)-Sarcodonin G (36) with those Reported for Natural (–)-Sarcodonin G (36)	99	
Table 4.	Comparison of ¹ H NMR Data for Synthetic (±)-Sarcodonin G (36) (CDCl ₃ , 400 MHz)with those Reported for Natural (–)-Sarcodonin G (36) (CDCl ₃ , 250 MHz)	100	
Table 5.	Comparison of ¹³ C NMR Data for Synthetic (±)-Sarcodonin G (36) (CDCl ₃ , 100.6 MHz) with those Reported for Natural (–)-Sarcodonin G (36) (CDCl ₃ , 62.5 MHz)	· 101	
Table 6.	¹ H NMR Data for Synthetic (±)-Triol 268 (CDCl ₃ , 400 MHz)	116	
Table 7.	Comparison of the ¹ H NMR (CDCl ₃ , 400 MHz) Data for Ketone 306 with those Reported for (±)-1-Epi-9-norpresilphiperfolan-9-one (306)	151	
Table 8.	Comparison of the ¹³ C NMR (CDCl ₃ , 100.6 MHz) Data for Ketone 306 and those Reported for (±)-1-Epi-9-norpresilphiperfolan-9-one (306)	152	
Table 9.	¹ H NMR (CDCl ₃ , 400 MHz) Data for Ethyl (<i>Z</i>)-3-Tri- methylgermyl-2-pentenoate (165): NOED Experiment	169	
Table 10.	 ¹³C NMR (CDCl₃, 100.6 MHz) and ¹H NMR (CDCl₃, 400 MHz) Data for Ethyl (Z)-3-Trimethylgermyl-2- pentenoate (165): HMQC Experiment 	170	
Table 11.	¹ H NMR (CDCl ₃ , 400 MHz) Data for Ethyl (<i>E</i>)-3-Tri- methylgermyl-2-pentenoate (174): NOED Experiment	172	

Table 12.	 ¹³C NMR (CDCl₃, 100.6 MHz) and ¹H NMR (CDCl₃, 400 MHz) Data for Ethyl (Z)-3-Trimethylgermyl- 2-pentenoate (174): HMQC Experiment 	173
Table 13.	¹ H NMR (CDCl ₃ , 400 MHz) Data for Ethyl (<i>E</i>)-3-Tri- methylgermyl-3-pentenoate (164): NOED Experiment	175
Table 14.	¹ H NMR (CDCl ₃ , 400 MHz) Data for Keto Ester 159 : NOED Experiment	206
Table 15.	¹³ C NMR (CDCl ₃ , 100.6 MHz) and ¹ H NMR (CDCl ₃ , 400 MHz) Data for Keto Ester 159 : HMQC Experiment	207
Table 16.	¹³ C NMR (CDCl ₃ , 100.6 MHz) and ¹ H NMR (CDCl ₃ , 400 MHz) Data for Ketone 240 : HMQC Experiment	214
Table 17.	Comparison of ¹ H NMR Data for Synthetic $(1R^*, 6R^*, 9R^*)$ - 3-[(S^*)-2-Hydroxy-1-methylethyl]-6,9-dimethyl-10-oxo- tricyclo[7.5.0.0 ^{2,6}]tetradeca-2,12-diene-12-carboxaldehyde [(±)-Sarcodonin G] (36) (CDCl ₃ , 400 MHz) with those Reported for Natural (–)-Sarcodonin G (36) (CDCl ₃ , 250 MHz)	217
Table 18.	Comparison of ¹³ C NMR Data for Synthetic ($1R^*, 6R^*, 9R^*$)- 3-[(S^*)-2-Hydroxy-1-methylethyl]-6,9-dimethyl-10-oxo- tricyclo[7.5.0.0 ^{2,6}]tetradeca-2,12-diene-12-carboxaldehyde [(±)-Sarcodonin G] (36) (CDCl ₃ , 400 MHz) with those Reported for Natural (–)-Sarcodonin G (36) (CDCl ₃ , 62.5 MHz)	218
Table 19.	¹ H NMR (CDCl ₃ , 400 MHz) Data for Diester 269 : NOED Experiment	228
Table 20.	¹³ C NMR (CDCl ₃ , 100.6 MHz) and ¹ H NMR (CDCl ₃ , 400 MHz) Data for Diester 269 : HMQC Experiment	230
Table 21.	 ¹³C NMR (CDCl₃, 100.6 MHz) and ¹H NMR (CDCl₃, 400 MHz) Data for 1-(1-Methyl-cyclopent-2-enyl)- propan-2-one (315): HMQC Experiment 	234
Table 22.	¹³ C NMR (CDCl ₃ , 100.6 MHz) and ¹ H NMR (CDCl ₃ , 400 MHz) Data for the 1:1 Mixture of 2-Methyl-3- (1-methyl-cyclopent-2-enyl)-propionaldehydes (333): HMQC Experiment.	237

Table 23.	 ¹³C NMR (CDCl₃, 100.6 MHz) and ¹H NMR (CDCl₃, 400 MHz) Data for 2,2-Dimethyl-3-(1-methyl-cyclopent-2-enyl)-propionaldehyde (314): HMQC Experiment 	239
Table 24.	¹³ C NMR (CDCl ₃ , 100.6 MHz) and ¹ H NMR (CDCl ₃ , 400 MHz) Data for a 1:1 Mixture of 2,2-Dimethyl-1- (1-methyl-cyclopent-2-enyl)-7-phenyl-hept-6-yn-3-ol (366): HMQC Experiment.	242
Table 25.	Comparison of the ¹ H NMR (CDCl ₃ , 400 MHz) Data for (1R [*] ,4R [*] ,7S [*] ,8R [*])-4,6,6-Trimethyltricyclo- [5.3.1.0 ^{4,11}]undecan-9-one (306) with those Reported for (±)-1-Epi-9-norpresilphiperfolan-9-one (306)	247
Table 26.	Comparison of the ¹³ C NMR (CDCl ₃ , 100.6 MHz) Data for $(1R^*, 4R^*, 7S^*, 8R^*)$ -4,6,6-Trimethyltricyclo- [5.3.1.0 ^{4,11}]undecan-9-one (306) and those Reported for (±)-1-Epi-9-norpresilphiperfolan-9-one (306)	248

LIST OF FIGURES

FIGURE 1.	¹ H NMR Spectrum for Synthetic (±)-Sarcodonin G (36)		
	(CDCl ₃ , 400 MHz)	102	

LIST OF ABBREVIATIONS

α	-	below the plane of the ring or 1,2 relative position
Ac	-	acetyl
AIBN	-	azobisisobutyronitrile
anal.	-	analysis
APT	-	attached proton test
ax	-	axial
β	- ·	above the plane of a ring or 1,3-relative position
9-BBN	-	9-borabicyclo[3.3.0]nonane
Bn		benzyl
bp	-	boiling point
br	-	broad
Bu	-	butyl
Bz	-	benzoyl
calcd	-	calculated
cm	-	centimeter
COSY	-	(¹ H- ¹ H)-homonulcear correlation spectroscopy
18-cr-6	-	18-crown-6 (1,4,7,10,13,16-hexaoxacyclooctadecane)
CSA		camphorsulfonic acid
C–x	-	carbon number x
d	-	doublet
δ	-	chemical shift in parts per million from tetramethylsilane
Δ	-	heat
DBN	-	1,5-diazabicyclo[4.3.0]non-5-ene
DBU	-	1,8-diazabicylco[5.4.0]undec-7-ene
DDQ	-	2,3-dichloro-5,6-dicyano-1,4-benzoquinone
DEAD		diethyl azodicarboxylate
DIBALH	-	diisobutylaluminum hydride
DIEA	_ ·	diisopropylethylamine
DMAP		4-dimethylaminopyridine
DMSO	-	dimethyl sulfoxide
DNA	- _	deoxyribonucleic acid
E	_	entgegen (configuration)
ed.	_	edition
ED., Eds.	_	editor, editiors
ED_{50}	-	effective dose
epi	_	epimeric
eq	_	equatorial
equiv.	-	equivalent(s)
Et	-	ethyl
	-	
g alc	-	gram
glc h	-	gas-liquid chromatography
		hour(s) (¹ H- ¹³ C) heteronuclear multiple bond coherence
HMBC	-	
HMPA	-	hexamethylphosphoramide

HMQC	-	(¹ H- ¹³ C) heteronuclear multiple quantum coherence
HPLC	-	high performance liquid chromatography
HRMS	_	high resolution mass spectroscopy
H–x	-	hydrogen number x
Hz	-	hertz
i	-	iso
IC_{50}	_	inhibitory concentration (for 50% of a biological sample)
<i>i</i> -PrDMS	-	<i>iso</i> -propyldimethylsilyl
IR	-	infrared
J	_	coupling constant in hertz
KDA	-	potassium diisopropylamide
KHMDS	-	potassium 1,1,1,3,3,3,-hexamethyldisilazide
LAH	-	lithium aluminum hydride
LDA	-	lithium diisopropylamide
LiHMDS	-	lithium 1,1,1,3,3,3-hexamethyldisilazide
m	-	multiplet
m	-	meta
М	-	molar
<i>m</i> CPBA	-	meta-chloroperoxybenzoic acid
Me	_	methyl
mg	-	milligram(s)
μg	-	microgram(s)
MHz	-	megahertz
min	-	minute(s)
mL	-	milliliter(s)
μL	-	microliter(s)
mm	-	millimeter(s)
μm	. 	micrometer(s)
mmol	-	millimole(s)
mol.	-	molecular
mp	-	melting point
Ms	-	methanesulfonyl
n	-	normal
NBS	-	N-bromosuccinimide
NIS	-	<i>N</i> -iodosuccinimide
NMR	-	nuclear magnetic resonance
NOE	-	nuclear Overhauser enhancement
p	-	page
p	-	para
PCC	-	pyridinium chlorochromate
pH	_	-log ₁₀ [H ⁺]
Ph	-	phenyl
PMB	-	para-methoxybenzyl
PNB	-	para-nitrobenzoyl
PP	-	pyrophosphate
ppm	_	parts per million
rr		r F

PPTS	-	pyridinium <i>p</i> -toluenesulfonate
Pr	-	propyl
pyr	-	pyridine
q	-	quartet
R	-	rectus (configuration)
S	-	singlet
S	-	sinister (configuration)
sec	-	secondary
t	-	triplet
t	-	tertiary
TBAF	-	tetrabutylammonium fluoride
TBDPS	-	tert-butyldiphenylsilyl
TBS	-	tert-butlydimethylsilyl
tert	-	tertiary
Tf	-	trifluoromethanesulfonyl
TFA	-	trifluoroacetic acid
THF	-	tetrahydrofuran
tlc	-	thin layer chromatography
TMS	-	trimethylsilyl
TPAP	-	tertrapropylammonium perruthenate
Ts	-	para-toluenesulfonyl
uv	-	ultraviolet
-ve	-	negative
v/v	-	volume to volume ratio
w/v	-	weight to volume ratio
•	-	coordination or complex
±	. –	racemic

ACKNOWLEDGMENTS

Firstly, I would like to thank Dr. Edward Piers, my supervisor and mentor, for continued guidance throughout my stay at UBC. For giving me the opportunity to join his research group and for teaching me the science of organic synthesis, I am forever grateful.

Many thanks to the members of Piers research group, past and present, for creating an atmosphere of cooperative learning. I am especially appreciative of the advice, friendly competition and many heated debates provided by Robert Britton from Day 1 of our graduate studies. I would also like to thank Arturo Orellana for the countless hours spent proofreading this manuscript.

The assistance of the technical staff (NMR, MS and Microanalysis labs) in the Department of Chemistry at UBC is gratefully acknowledged. I would like to extend a special thank-you to Mrs. L. Darge and Mrs. M. Austria of the NMR lab for their ongoing help and advice.

I will always treasure the friends I made in Vancouver, especially the Brew-Club members, Rob B., Art, Rob G., Carl, Mark, Todd and Roger, who made my stay in B.C. an unforgettable one.

This thesis is dedicated to my parents, whose support and encouragement continuously fuel my ambitions.

Finally, I wish to thank Sara for being her wonderful self and making everything worthwhile.

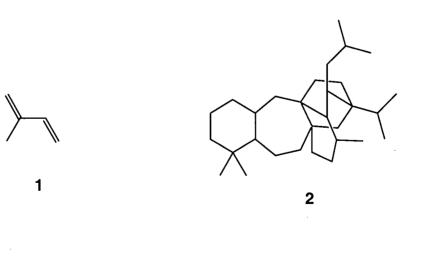
I. INTRODUCTION

1.1. General

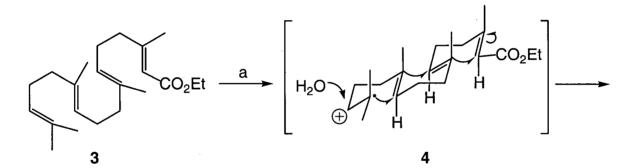
As we enter the 21st century, synthetic organic chemistry is at center stage of scientific and technological discovery. Since its conception nearly two centuries ago, this discipline of chemistry has played in increasingly important role in biological and medicinal research. As an example, the ability to synthesize organic compounds represents the foundation of pharmaceutical drug-discovery programs, which have lead to numerous biomedical breakthroughs. The evolution of synthetic chemistry has been possible, in large, because of technological advances in the fields of chemical separation and identification. With a growing library of proven synthetic methods and an upward number of commercially available reagents, synthetic chemistry allows for increasingly short and efficient preparations of target structures.

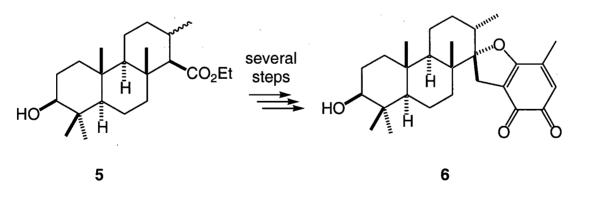
There are numerous motives that might entice a chemist in choosing to synthesize a target structure from commercially available materials, a study known as total synthesis. The target structures are often naturally occurring compounds or families of structurally related compounds that have limited availability from natural sources. In some cases, these natural compounds have demonstrated potentially important biological activity and total synthesis can provide additional quantities of material for further biological and chemical evaluation. Total synthesis also provides a testing ground for novel methods of bond construction. The limitations of a novel reaction developed in simple models systems are often revealed in complex total syntheses. To be sure, total synthesis represents an exciting yet challenging study through which one can learn the many facets of chemistry.

Terpenoid natural products, which have attracted significant attention from synthetic chemists, display a seemingly unlimited complexity with regard to construction of carbon skeleta, the backbones of the structures. Biosynthetically evolving from 5-carbon building blocks (C_5) generally represented by isoprene (1), terpenoids are classified as either mono-(C_{10}), sesqui-(C_{15}), di-(C_{20}), sester-(C_{25}) or triterpenoids (C_{30}). Through isotope-labeling experiments and NMR studies on the resulting metabolites, chemists have uncovered many of nature's strategies for the transformation of simple chains of isoprene units into complex carbocyclic terpenoids. These biosynthetic pathways often involve a combination of enzyme-catalyzed cyclizations, rearrangements and alkyl shifts. The elaborate architecture displayed in structure **2**, representing the carbon skeleta of the recently discovered salvadione triterpenoids,¹ nicely illustrates the complexity of carbocycle construction in terpenoid chemistry. Several of the different tactics employed by synthetic chemists in terpenoid construction, including biomimetic, tandem and step-wise annulation approaches, shall be outlined.



A biomimetic synthetic strategy is one in which the key reactions are based on the proposed biogenesis of the target structure. For example, according to the Stork-Eschenmoser hypothesis,^{2,3} certain polyunsaturated molecules with *trans* C=C double bonds should cyclize in a stereospecific manner to furnish polycyclic systems with *trans,anti,trans* stereochemistry at the ring fusions. Recently, Demuth and coworkers reported⁴ a biomimetic cascade cyclization of a polyalkene, which led to a total synthesis of (±)-stypoldione (6) (Scheme 1). A photo-induced electron transfer (PET) reaction triggered cyclization of polyalkene 3, yielding a mixture of *trans-anti-trans* 6-6-6 fused tricyclic substances 5. The natural product, (±)-stypoldione (6), was ultimately obtained in 11 steps from 5.

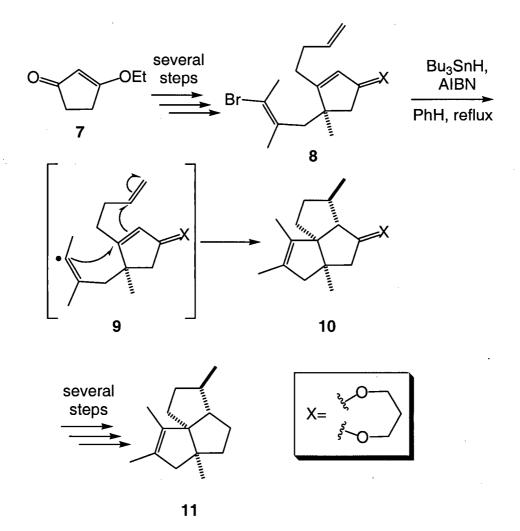




Reagents: hv, poly(oxy-1,2-ethanediyloxycarbonyl-1,4-phenylenecarbonyl), 1,4-dicyano-2,3,5,6-tetramethylbenzene, biphenyl, MeCN, H_2O .

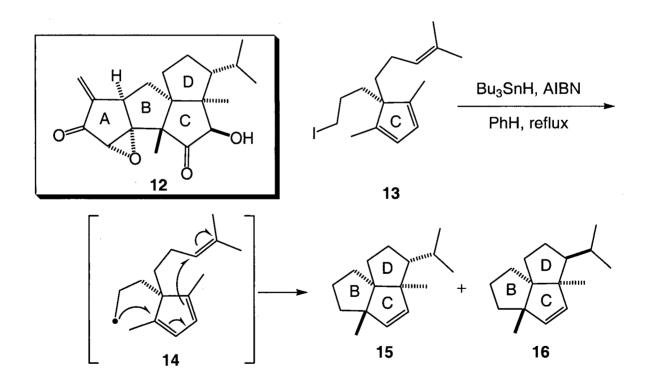
Scheme 1

Many types of reactions, including pericyclic, radical, cationic and anionic processes, have been incorporated in cascade or tandem sequences for the construction of carbocycles.⁵ For instance, a radical-based tandem cyclization was employed in the synthesis of triquinane silphiperfolene (11), as reported by Curran and coworkers (Scheme 2).⁶ The key intermediate of the study, bromide **8**, was prepared in several steps from enone **7**. Treatment of **8** with Bu₃SnH and AIBN gave tricycle **10** *via* the radical intermediate **9**. Transformation of **10** into the target structure **11** was accomplished in few steps.



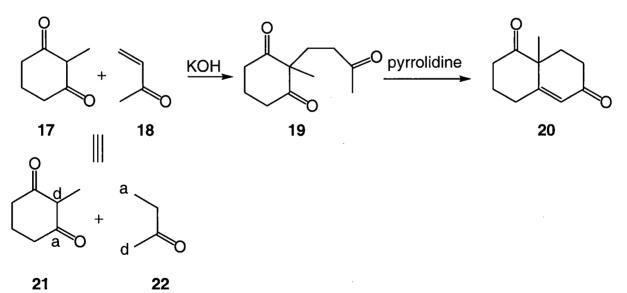


Although the combination of several bond-forming steps in one tandem sequence certainly appeals, the difficult task of controlling the selectivity in each step must also be considered. As an example, Curran and coworkers reported⁷ unexpected stereoselectivity difficulties in their investigation of a tandem cyclization approach to (\pm)-crinipellin A (12) (Scheme 3). In a model study directed towards the synthesis of the B-C-D ring system of 12, Bu₃SnH-mediated tandem free radical cyclization of iodide 13 afforded a 1:5 mixture of 15 and 16. Although this remarkable reaction showed the desired *cis* geometry at both ring fusions, the tandem approach was not carried through to the target structure 12, because the major isomer (16) displayed the isopropyl appendage in the unnatural β -configuration.



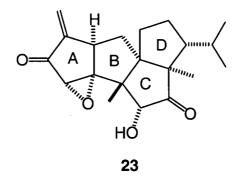


An attractive approach to the construction of carbocycles involves the use of bifunctional reagents in stepwise annulation sequences.⁸ An annulation sequence is a ring-forming process in which two molecular fragments are united with the formation of two new bonds.⁹ The reactive sites leading to the newly formed bonds can be described as either acceptor (a) or donor (d) sites.¹⁰ Upon analysis of the potential reactive sites of a substrate, one may envisage the annulation synthon that displays the desired corresponding acceptor or donor reactive sites. This relationship is illustrated in the reported¹¹ synthesis of the Wieland-Miescher ketone¹² **20** *via* the well-known Robinson annulation¹³ (Scheme 4). Under base-mediated conditions, diketone **17** undergoes a Michael addition with methyl vinyl ketone (**18**) to give adduct **19**. Treatment of **19** with pyrrolidine results in an intramolecular aldol condensation, yielding the Wieland-Miescher ketone (**20**). Therefore, methyl vinyl ketone (**18**), the synthetic equivalent of the 2-butanone a⁴,d¹-synthon **22**, represents an excellent annulation reagent for diketone **17**, corresponding to donor-acceptor synthon **21**.



Scheme 4

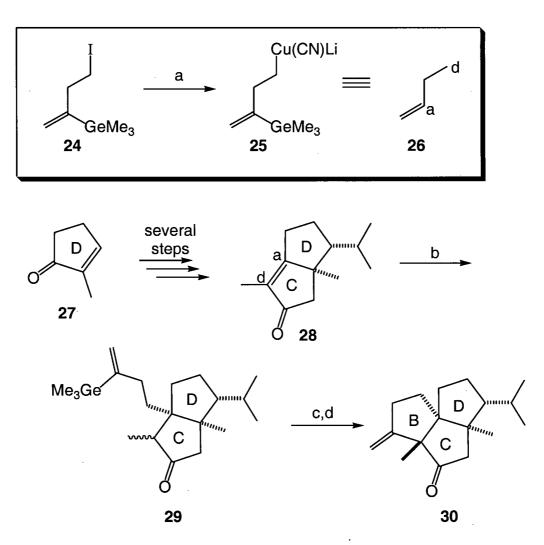
Recently, one of the areas of study in the Piers group has been the development of novel bifunctional reagents and their application in annulation sequences. For example, the first total synthesis of (\pm) -crinipellin B $(23)^{14,15}$ employed novel bifunctional reagents in the annulation sequences leading to the A- and B-rings, as outlined in Schemes 5 and 6.



The bifunctional reagent 24, which is readily transformed into cuprate 25, the synthetic equivalent of the 1-butene a^2 , d^4 -synthon 26, was employed in the preparation of the B-ring (Scheme 5). The synthesis commenced with 2-methyl-2-cyclopentenone (27), which was transformed in several steps into the 5,5-fused bicyclic compound 28, representing the C-D ring system. The annulation sequence began with a conjugate addition of a cuprate 25 to enone 28, affording adduct 29. Treatment of 29 with iodine provided the corresponding alkenyl iodide. Slow addition of *t*-BuOK/*t*-BuOH to solution of this alkenyl iodide and (Ph₃P)₄Pd in THF resulted in the triquinane product 30.

·. .

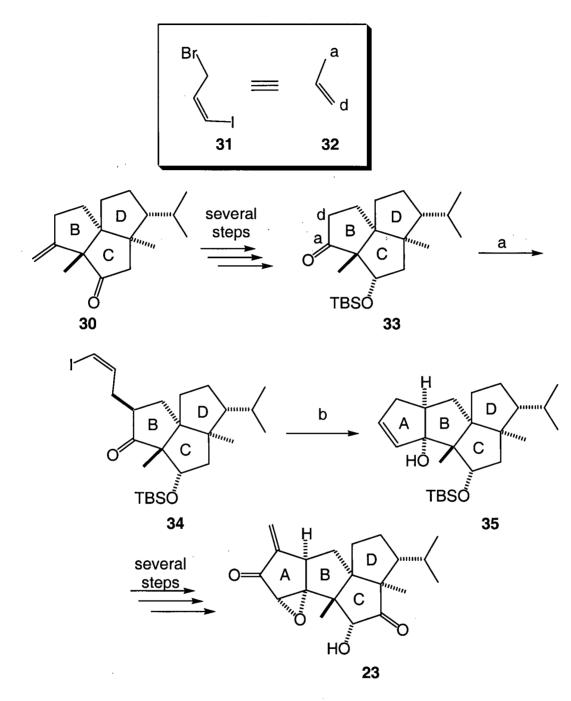




Reagents: a) *t*-BuLi, THF; CuCN; b) **25**, TMSBr, THF; NH₄Cl, NH₄OH, H₂O; c) I_2 , CH₂Cl₂; d) (Ph₃P)₄Pd, *t*-BuOK, *t*-BuOH-THF.

Scheme 5

The annulation sequence leading to the A-ring employed bifunctional reagent 31, the synthetic equivalent of the 1-propene a^3 , d^1 -synthon 32 (scheme 6). The conversion of triquinane 30 into ketone 33 was accomplished in a few steps. Alkylation of the enolate derived from 33 with reagent 31 yielded adduct 34, as the major product. Treatment of 34 with butyllithium resulted in a cyclization, affording the tetracyclic alcohol 35. The target structure, (±)-crinipellin B (23), was ultimately obtained from 35 in 10 steps.

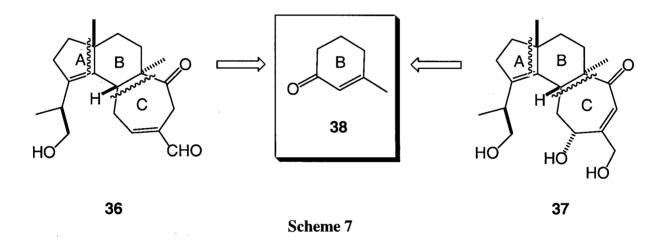


Reagents: a) LDA, THF; 31; b) BuLi, THF; NaHCO₃, H₂O.

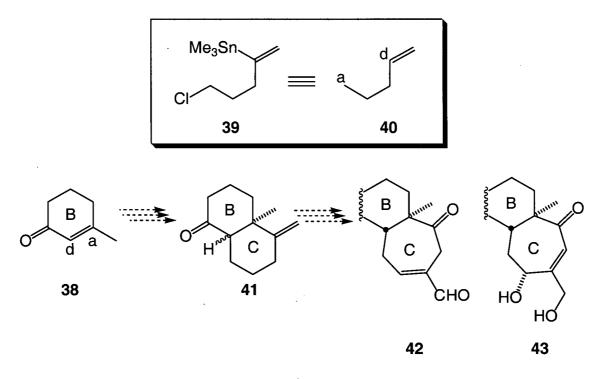
Scheme 6

1.2. Proposal

In the first part of the research program described in this thesis, it was planned to employ a sequential annulation strategy to the assembly of 5-6-7 fused ring systems. It was anticipated that the carbon framework of the structurally-related diterpenoids, sarcodonin G (**36**) and cyathin A_4 (**37**),^{16,17} would be accessible from commercially available 3-methyl-2-cyclohexen-1-one (**38**), which would ultimately serve as the B-ring of these two natural products (Scheme 7).

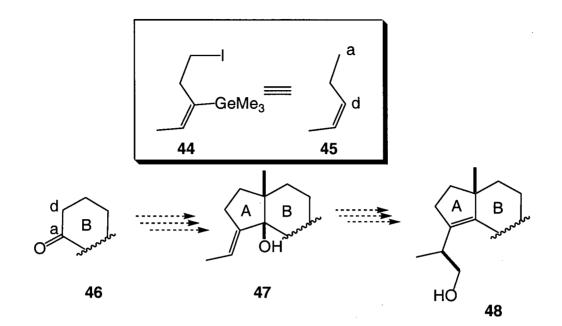


The annulation sequence leading to construction of the C-ring was anticipated to involve bifunctional reagent **39**, representing the 1-pentene a^5 , d^2 -synthon **40** (Scheme 8). The preparation of the 6-6 fused bicycles **41**, utilizing enone **38** and reagent **39**, was previously reported by our group.¹⁸ The 6-membered C-ring in **41** was thought to be a suitable precursor to the 7-membered C-rings illustrated in partial structures **42** and **43**, representing sarcodonin G (**36**) and cyathin A₄ (**37**), respectively.



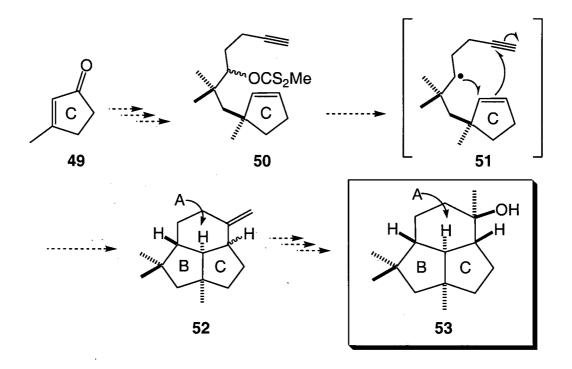
Scheme 8

The A-ring construction was anticipated to result from an annulation sequence employing the (Z)-2-pentene a^5 , d^3 -synthon 45, which translates to bifunctional reagent 44 (Scheme 9). A representation of the sequence, starting with partial structure 46, is illustrated. The annulation sequence, which includes a methylation step, was anticipated to give alcohol 47. Partial structure 48, representing the A- and B-rings of both target compounds 42 and 43, was expected to be available from 47 *via* a Still-Mitra rearrangement protocol.¹⁹



Scheme 9

The second part of the research study described herein revolves around the possible use of a proposed tandem cyclization strategy, envisaged for the construction of the 5-5-6 fused framework of (±)-presilphiperfolan-9-ol (53) (Scheme 10).²⁰ It was hoped that the key cyclization precursors, intermediates 50, would be available in a few steps from enone 49, which would become the C-ring of target 53. Tandem formation of the A-and B-rings was anticipated to involve a radical-mediated cyclization of 50 *via* the intermediate 51. The β -isomer of the proposed cyclization products 52, which display a 5-5-6 fused ring system, is a known²⁰ precursor of (±)-presilphiperfolan-9-ol (53).



Scheme 10

II. RESULTS AND DISCUSSION

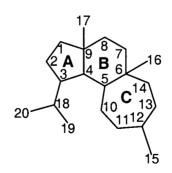
2.1. Studies Towards Cyathane Diterpenoid Synthesis

2.1.1. Background

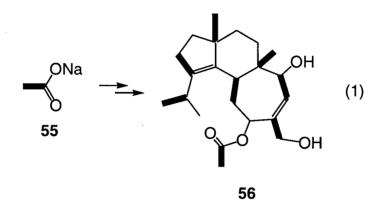
2.1.1.1. Cyathane History and Biogenesis

The history of the cyathane family of diterpenoids begins in 1966 with Brodie's discovery²¹ of the bird's nest fungus *Cyathus helenae* in the Canadian Rockies. In a chance observation, cultures of *C. helenae* were shown²² to inhibit the growth of bacteria. Starting in the early 1970s, Ayer and co-workers characterized²³ numerous biologically active cyathane components from several *Cyathus* fungi. To date, over forty different cyathanes have been isolated and characterized.

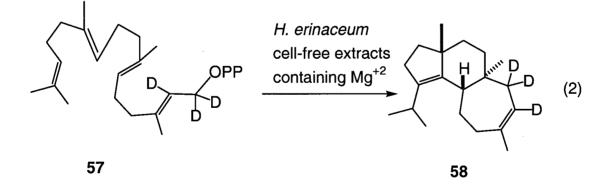
Using bioassay guided fractionation methods, Ayer and coworkers isolated several C_{20} compounds from the cyathin extracts. Each of these novel diterpenoids was named cyathin and was classified according to its molecular formula. The cyathin name includes a letter that denotes the degree of unsaturation (A for 30 hydrogens, B for 28 hydrogens and C for 26 hydrogens) and a subscript number that indicates the number of oxygen atoms. A trivial numbering scheme was assigned to the novel 5-6-7 fused tricyclic core (54) of the cyathanes.



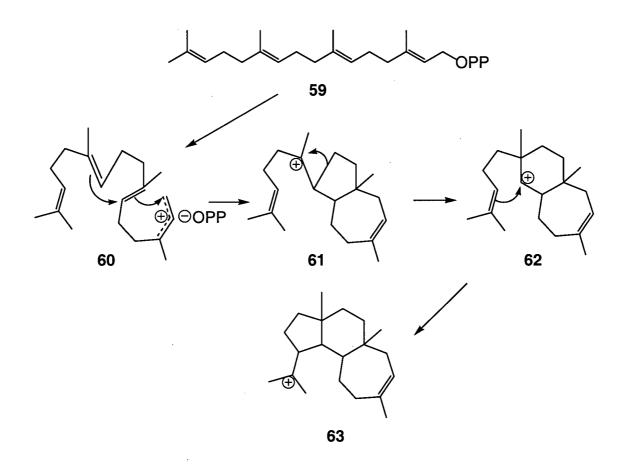
In 1979, Ayer and coworkers reported their investigations into cyathane biogenesis through labeling experiments.²⁴ Since the biosynthetic pathway was presumed to involve acetate incorporation into geranylgeranyl pyrophosphate, $[1,2^{-13}C_2]$ -labeled acetate (55) was fed to *Cyathus earlei* and the labeling pattern of the resulting 11-*O*-acetylcyathatriol product (56) was determined by ¹³C NMR analysis (eq 1).



Recently, Sassa and coworkers reported²⁵ a study of the fungal diterpene cyclase, cyathadiene synthase. Treatment of a cell-free extract of *Hericum erinaceum* with deuterated all-trans-geranylgeranyl diphosphate (57) for 2 h at 30 °C provided the deuterated cyatha-3,12-diene 58 in 30% yield (eq 2).



Both Ayer's and Sassa's results (*vide supra*) support the biosynthetic pathway postulated²⁴ by Ayer in 1979 (Scheme 11).

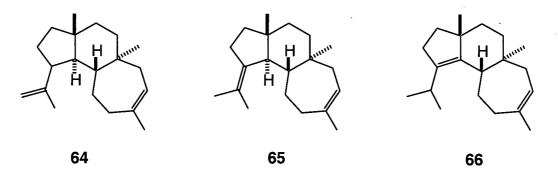




The first steps of the postulated biosynthetic pathway involve tandem cationic ring closures of all-trans-geranylgeranyl diphosphate (**59**) to form the hydroazulene-like cation (**61**), which undergoes a Wagner-Meerwein migration to provide a 6-7 fused bicyclic intermediate (**62**). Cyclization of the isopropylidene moiety onto the bicyclic core of **62** provides the 5-6-7 fused tricyclic cation **63**, which displays the cyathin skeletal core.

2.1.1.2. Hydrocarbon Cyathanes

Several cyathadiene hydrocarbons, postulated²⁵ progenitors of the oxygenated cyathanes, have recently been isolated. In 1993, Cassidy reported²⁶ the isolation of cyatha-12,18-diene (**64**) from a non-polar fraction of *Higginsia sp* extract. In 2001, Sassa and coworkers isolated²⁵ cyatha-3(18),12-diene (**65**) and cyatha-3,12-diene (**66**) from the erinacine-producing basodiomycete *Hericium erinaceum*.

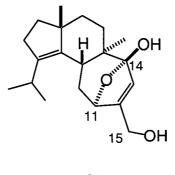


The isolation of hydrocarbons **64-66** suggests that intermediate **63** in Ayer's proposed pathway (*vide supra*) can follow different sequences of elimination and/or hydride shifts prior to oxidation.²⁵

2.1.1.3. Oxygenated Cyathanes

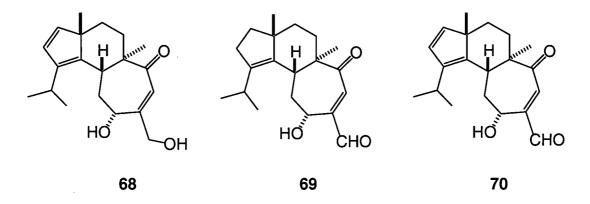
Nature's elaboration of the cyathane core includes, primarily, oxidation at allylic positions. Oxygenation on the three cyathin rings as well as on the methyl and isopropyl appendages has been noted. Certain oxygenation patterns introduce, from a synthetic viewpoint, considerable structural complexity to the cyathin structures. The diverse oxygenation patterns of several structurally elucidated cyathanes are discussed below.

The most common sites of oxygenation on the cyathin core are the carbons of the C-ring and its methyl appendage, C–15. Cyathin A₃ (67), the first cyathane to yield to structural elucidation,²⁷ exhibits oxygen functions at C–11, 14 and 15.

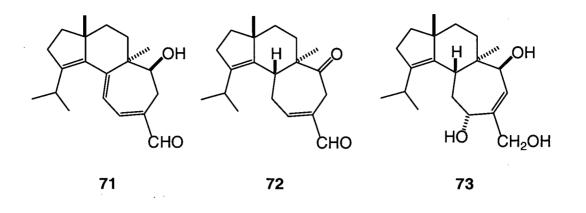


67

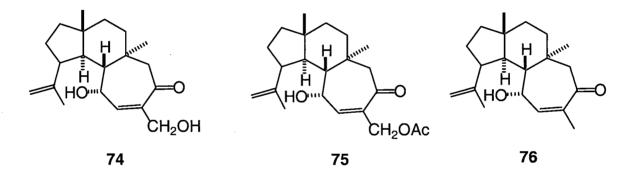
In their study of *Cyathus helenae* extracts, Ayer and coworkers reported several cyathanes that display an oxidation pattern similar to that of cyathin A_3 . The structures of allocyathin B_3 (68), cyathin B_3 (69), and cyathin C_3 (70) were elucidated through chemical correlations with cyathin A_3 (67).^{27,28}



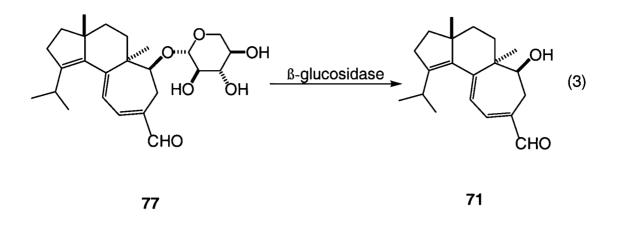
Extracts of the tropical species of bird's nest fungus *Cyathus earlei* yielded several acetylated cyathin-type compounds as well as three new cyathanes: allocyathin B_2 (71), cyathin B_2 (72) and cyathatriol (73).²⁹



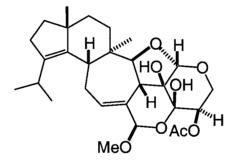
In 1985, Cassidy and co-workers isolated³⁰ cyathanes **74-76** from the extracts of the marine sponge *Higginsia sp*. The structure of cyathane **74**, which includes oxygen functions at C-10, 13 and 15, was verified by X-ray crystallographic analysis.



The erinacines, which are glycosylated cyathanes, were isolated from the mycelia of the fungus *Herimicium erinaceum*.³¹⁻³⁴ The xylose moiety of erinacine A (77) was cleaved³¹ using a β -glucosidase and the resulting aglycone product was identified as allocyathin B₂(71) (eq 3).²⁹

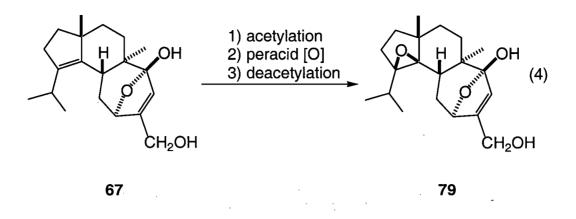


Hecht and coworkers reported three novel diterpenoids, named straitins, from the fungus *Cyathus striatus*.³⁵ The structure of straitin A (**78**) displays a cyathin core that is triple-linked to a pentose carbohydrate.

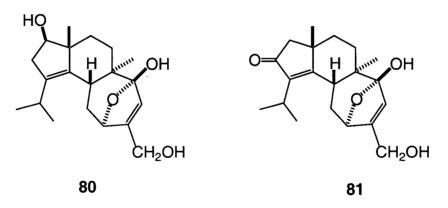


78

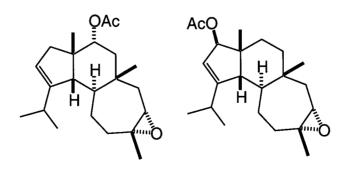
A number of cyathanes exhibit extensive oxygenation on the A-, B- and/or Crings. The structure¹⁷ of neoallocyathin A_4 (**79**), which displays an epoxide function in the A ring, was confirmed through chemical correlations with cyathin A_3 (**67**) (eq 4).



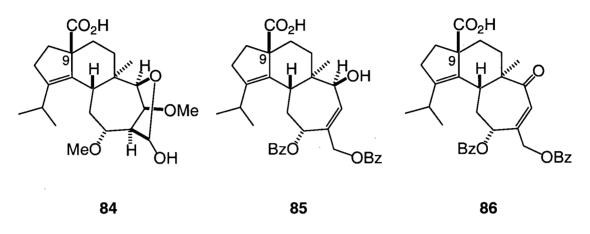
Oxygenation on both the A- and C-rings was noted in cyafrins A_4 (80) and B_4 (81), which were isolated³⁶ from the extracts of *Cyathus africanas*, a fungus collected in Tanzania.



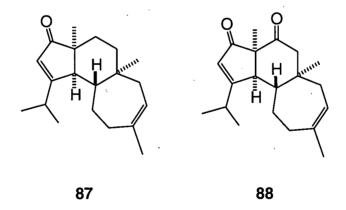
Cyathanes **82** and **83**, isolated³⁷ from the marine sponge *Myrekioderma styx*, have oxygenation patterns that include an epoxide on the C-ring.



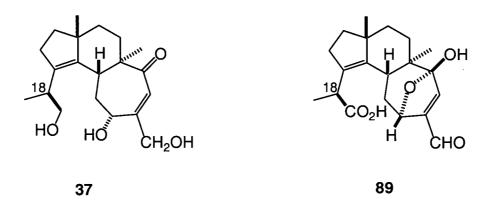
A novel group of cyathanes, named scabronines, were isolated³⁸⁻⁴⁰ from the fruiting bodies of the mushroom *Sarcodon scabrosus*. As in all scabronines, scabronines A (84), B (85) and C (86) display an acid function at the C–9 angular carbon.



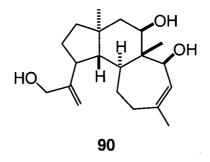
Few members of the cyathane family display an oxygenation pattern that excludes the C-ring. Cyanthiwigins A (87) and C (88), which display oxygenation on the A- and/or B-rings, were isolated by Green and coworkers from the extracts of *Apipolasis reiswigi*.⁴¹



Oxygenation of the isopropyl group attached to C-3 is a rare feature in the cyathane family. In 1978, Ayer and coworkers reported¹⁷ the isolation of cyathins A₄ (**37**) and C₅ (**89**), which display oxygenation of the C-3 isopropyl group.⁴²

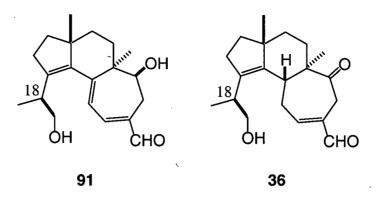


Onychiol B (90), isolated⁴³ from the rhizome of the fern *Onchium japonicum*, displays oxygenation on the B- and C-rings and on the isopropenyl group attached at A-3.



In 1989, Shibata and coworkers reported¹⁶ the isolation of eight novel cyathanetype diterpenoids from the fruiting bodies of *Sarcodon scabrosus*: sarcodonins A-H. Of these eight sarcodonins, only A (91) and G (36), which display oxygenation on the isopropyl moiety, yielded to structural elucidation. The absolute stereochemistry of sarcodonin G (36) was determined by X-ray crystallographic analysis of its *p*bromobenzoate derivative.

. .



2.1.1.4. Biological Activity of the Cyathanes

Members of the cyathane family have been shown to exhibit diverse biological activities. The cyathin complexes of the *Cyathus* fungi *C. helenae, C. earli and C. striatus* exhibit^{17,23,29} pronounced antibacterial and antifungal activities. The cyathanes isolated from *Myrmekioderma styx* exhibit³⁷ moderate cytotoxic activity against the P388 leukemia cell line and the A549 human lung tumor cell line, with IC₅₀'s as low as 5.6 and 4.0 μ g/mL, respectively. The cyanthiwigin cyathanes exhibit⁴¹ cytotoxic activity against P388 leukemia cells, with IC₅₀'s as low as 2.5 μ g/mL. The straitin cyathane-xylosides were shown to have leishmanicidal activity.⁴⁴

Saito and coworkers recently reported⁴⁵ that certain erinacines^{31-34,46} behave as potent kappa opioid receptor agonists. Activation of membrane-bound opioid receptors (μ , δ and κ), which are located throughout the nervous system,⁴⁷ can induce various physiological effects including antinociception and neurotransmitter release.⁴⁸ Morphine, an opioid drug used in the clinical management of pain, acts on the μ receptor and causes untoward side effects such as tolerance, dependence and respiratory depression. Stein and coworkers have reported⁴⁹ that activation of the κ receptor induces hyperalgesia in a rat model of inflammation. It is postulated that the application of erinacines as selective κ receptor agonists may produce the desired antinociceptive activity without the adverse side effects observed with μ receptor agonists.

Certain members of the erinacine³¹⁻³³ and scabronine^{38-40,50} cyathanes exhibit the ability to stimulate the biosynthesis of nerve growth factor (NGF) *in vitro*. NGFs play an important role in the early development of the embryonic central nervous system as well as in the phenotypic maintenance and cell regulation of neurons in adults. The role of NGFs in a novel mode of treatment for neurodegenerative disorders is currently under investigation.⁵¹ However, the inability⁵² of NGFs to penetrate the blood brain barrier (BBB) has limited their clinical application to the direct infusion into the cerebral spinal fluid.⁵³ The inconvenience of this medical implementation initiated a search for low-molecular-weight NGF inducers, such as the erinacines and scabronines, which can penetrate the BBB.

25

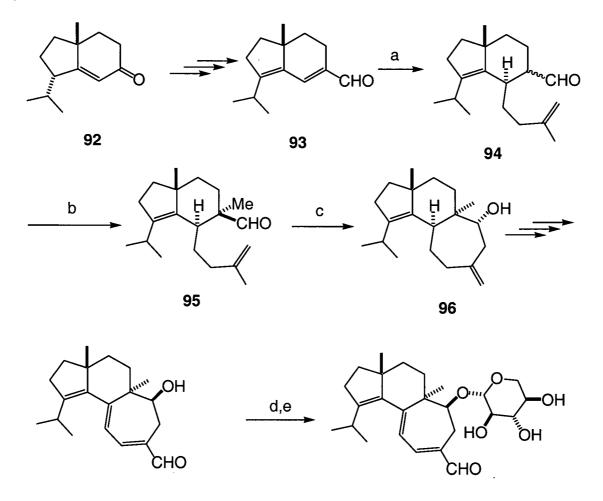
2.1.2. Previous Studies in Cyathane Synthesis

The diverse biological activities and structural novelty displayed by the cyathanes have made them attractive targets for total synthesis.⁵⁴ From a synthetic viewpoint, the diverse oxygenation patterns, the *anti* 1,4 angular methyls and the 5-6-7 fused tricyclic core represent significant challenges. As an illustration of the diverse approaches to the construction of the cyathane framework, the outlines of the studies described in this section focus primarily on key ring-forming steps.

2.1.2.1. Total Syntheses of Cyathanes

Although the first structural elucidation of a cyathane appeared in the literature in the early 1970s,²³ the first account of a cyathane total synthesis, reported by Snider and coworkers,^{55,56} was not published until 1996. Their synthetic sequence leading to (\pm) -allocyathin B₂ (**71**) and (+)-erinacine A (**77**) relied on an intramolecular carbonyl ene reaction as a key step (Scheme 12). The synthesis commenced with the known 5-6 fused bicyclic ketone **92**, which was converted to aldehyde **93** in four steps. TMS-accelerated copper(I)-catalyzed conjugate addition of isopentenylmagnesium bromide to **93** provided a mixture of epimeric aldehydes **94**. Methylation of **94** afforded aldehyde **95**. Treatment of **95** with Me₂AlCl resulted in an intramolecular carbonyl ene reaction, furnishing alcohol **96**, which displayed the complete cyathane skeleton. This material was transformed into (!)-allocyathin B₂ (**71**) in ten chemical operations. Glycosylation of racemic **71** with 2,3,4-tri-*O*-acetyl- α -D-xylopyranosyl bromide provided a 1:1 mixture of

(+)-erinacine A triacetate and its diastereomer. Deacetylation of the former with potassium carbonate in methanol afforded (+)-erinacine A (77).



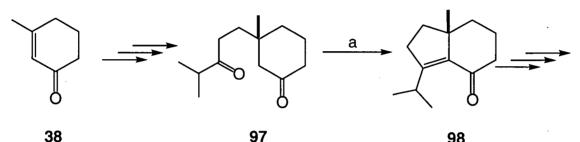
71

77

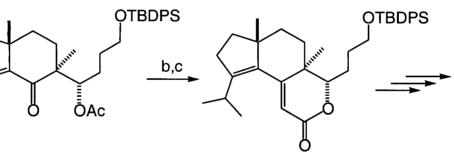
Reagents: a) $H_2C=C(CH_3)CH_2CH_2MgBr$, CuBr DMS, TMSCl, HMPA; b) *t*-BuOK, MeI; c) Me₂AlCl; d) 2,3,4-tri-*O*-acetyl- α -D-xylopyranosyl bromide, Hg(CN)₂, HgCl₂; e) K₂CO₃, MeOH.

Scheme 12

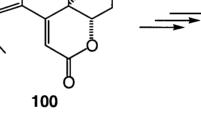
In 1998, Tori and coworkers reported⁵⁷ the second total synthesis of (!)allocyathin B_2 (71), which involved three key aldol condensation reactions (Scheme 13). Dione 97, precursor for the first-aldol condensation, was prepared in five steps from 3methyl-2-cyclohexen-1-one (38). Intramolecular condensation of 97 provided the fused hydrindenone system 98, which was transformed into compound 99 in four chemical operations. Base-mediated intramolecular cyclization of 99, followed by dehydration of the resultant tertiary alcohol using SOCl₂, furnished the α , β -unsaturated lactone 100. Conversion of 100 to dialdehyde 101 was accomplished in five steps. Aldol condensation of 101, with concomitant acetate hydrolysis, provided (\pm) -allocyathin B₂ (71).



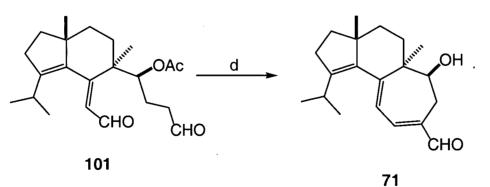








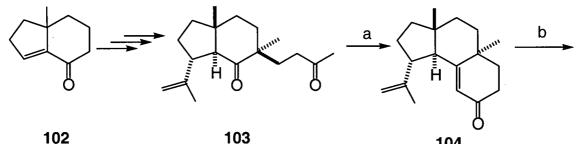
98



Reagents: a) 5% aq. KOH, MeOH, *****; b) LiHMDS; c) SOCl₂, pyr; d) KOH, MeOH.

Scheme 13

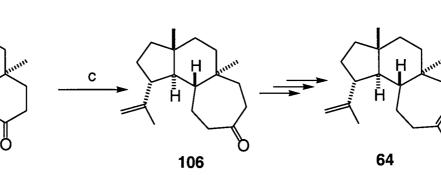
Taking advantage of an advanced intermediate in a synthetic study of verrucosane natural products,⁵⁸ Piers and Boulet developed a synthesis⁵⁹ of (!)-cyatha-12,18-diene (64) (Scheme 14). The sequence commenced with the known bicyclic enone 102, which was transformed into dione 103 in six steps. Base-mediated intramolecular aldol condensation of 103 afforded tricyclic enone 104. Metal-ammonia reduction of 104 provided ketone 105, which displayed the requisite *trans* stereochemistry at the ring fusion. The branching point in a divergent synthetic plan, compound 105 was also employed in the total syntheses of several verrucosane natural products.⁵⁸ Lewis-acid catalyzed homologation of 105, followed by decarboxylation of the resultant β -keto ester, provided the 5-6-7 fused tricyclic ketone 106. Final conversion of 106 to (!)-cyatha-12,18-diene (64) was accomplished in two steps.





Ĥ

105



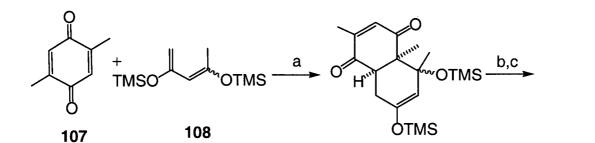
104

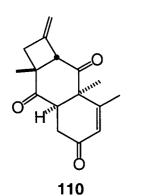
Reagents: a) NaOMe, •; b) Li, NH₃, *t*-BuOH; solid NH₄Cl; c) BF₃·OEt, N₂CHCO₂Et; DMSO, NaCl, **•**.

Scheme 14

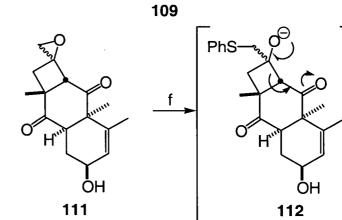
29

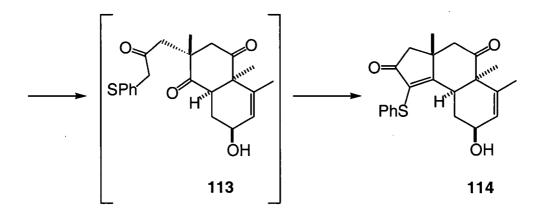
Recently, Ward and coworkers completed the total synthesis of allocyathin B_3 (68), a synthetic endeavor that began while Ward was a student in Ayer's group.⁶⁰⁻⁶² The first part of the synthesis, reported by Ayer and coworkers,⁶⁰ relied on two key cycloaddition reactions (Scheme 15). Diels-Alder reaction between benzoquinone 107 and dienes 108, gave the *cis*-fused bicycle 109. Photochemical [2+2] cycloaddition of 109 with allene, followed hydrolysis of the silyl enol ether function, afforded tricycle 110 as the major isomer. Epoxidation of the exocyclic double bond, followed by 1,2 reduction of the enone, furnished a mixture of epoxides 111. Treatment of 111 with benzenethiol and potassium hydroxide caused a sequence of reactions, commencing with a nucleophilic ring-opening of the epoxide to give intermediate 112. Fragmentation of 112, followed by intramolecular aldol condensation of the resultant triketone 113, ultimately provided tricycle 114.





d,e

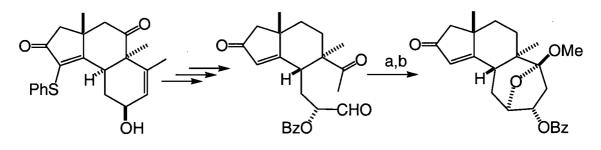




Reagents: a) xylene, \checkmark ; b) allene, hv; c) Rexyn 101 acidic ion exchange resin; d) *m*-CPBA; e) 9-BBN; f) PhSH, 5% aq. KOH, \checkmark .

Scheme 15

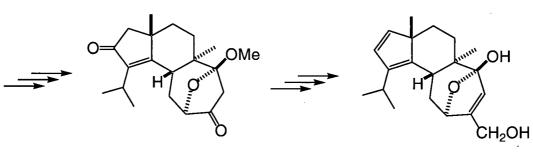
Transformation of Ayer's advanced intermediate **114** (*vide supra*) to (\pm) allocyathin B₃ (**68**) was completed by Ward and coworkers.^{61,62} Conversion of **114** to keto aldehyde **115** required ten chemical operations (Scheme 16). Acid-promoted aldol cyclization of **115** proceeded with a concomitant intramolecular benzoyl transfer. The resultant hemiketal was methylated, providing 5-6-7 fused tricycle **116**. The task of affixing an isopropyl moiety at C–3 (cyathane numbering) of **116** was accomplished in eight steps, affording intermediate **117**. (\pm) Allocyathin B₃ (**68**) was obtained from **117** in three steps.



114

115

116



117

68

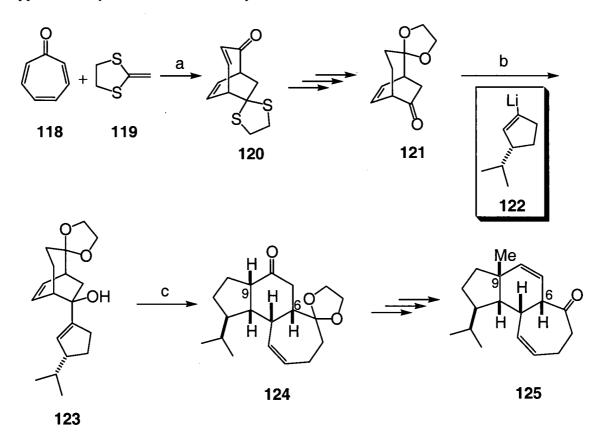
Reagents: a) p-TsOH; b) MeI, Ag₂O.

Scheme 16

2.1.2.2. Approaches to the Cyathanes

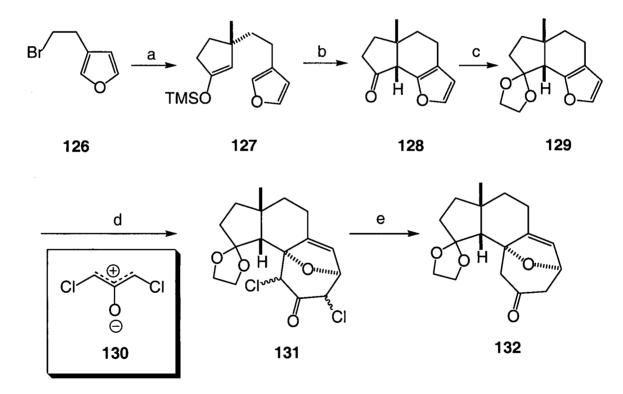
In addition to the total synthesis publications, there have been numerous reports that describe novel methods of constructing the cyathane skeletal core. In 1994, Dahnke and Paquette reported⁶³ a synthetic approach to the cyathane core *via* an anionic oxy-Cope rearrangement. An inverse demand Diels-Alder reaction between tropone (**118**) and

2-methylidene-1,3-dithiolane (119) provided bicyclo[3.2.2]nonane 120 (Scheme 17). Several steps, which included a resolution of enantiomers, afforded the levorotatory enantiomer of ketone 121 from intermediate 120. Treatment of (–)-121 with the enantiopure organolithium reagent 122 provided the rearrangement precursor 123. Oxyanion accelerated Cope rearrangement of 123 furnished the 5-6-7 fused tricycle 124. Compound 124 was transformed into ketone 125 in six steps, which included a methylation at C–9 from the β -face. Unfortunately, the requisite installation of a methyl group at C–6, which must occur from the α -face, proved difficult and this synthetic approach to cyathanes was ultimately abandoned.



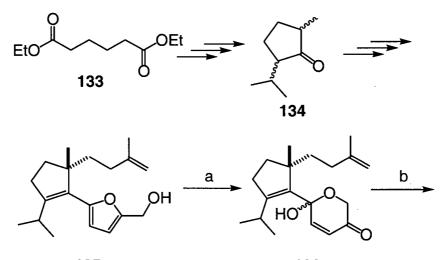
Reagents: a) Et₃N, **→**; b) **122**; c) KH, 18-cr-6, **→**.

In 1999, Wright and coworkers disclosed a novel approach to the cyathane core that relied on sequential oxidative coupling and [4+3] cycloaddition steps.⁶⁴ The sequence began with a TMSCl-accelerated copper(I)-catalyzed addition of the Grignard reagent derived from bromide **126** to 3-methyl-2-cyclopenten-1-one (Scheme 18). Exposure of the resultant adduct **127** to a carbon anode promoted an oxidative coupling reaction, yielding the tricyclic product **128**. Ketone **128** was converted to its cyclic ketal derivative **129**. Treatment of **129** with oxyallyl cation **130**, which was generated *in-situ* from trichloroacetone, resulted in a [4+3] cycloaddition. The cycloaddition products **131** were dechlorinated with a zinc–copper couple to afford compound **132**, which displayed a 5-6-7 fused carbon skeleton present in cyathanes.



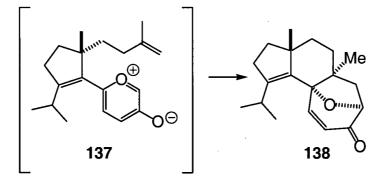
Reagents: a) Mg; CuI, TMSCl; 3-methyl-2-cyclopenten-1-one; b) carbon anode, LiClO₄, 2,6-lutidine; c) p-TsOH, HO(CH₂)₂OH, CH(OEt)₃; d) 1,1,3-trichloroacetone, NaOCH₂CF₃; e) Zn-Cu couple, NH₄Cl.

Magnus and Shen reported⁶⁵ a synthetic approach to the cyathane core that relied on a key [5+2] annulation reaction (Scheme 19). The sequence commenced with diethyl adipate (133), which was converted to the mixture of ketones 134. Transformation of 134 to alcohol 135 was accomplished in four steps. Irradiation of 135 in the presence of O_2 resulted in an oxidative rearrangement, affording the unstable pyranone 136. Treatment of 136 with TFA generated the corresponding pyrylium ylide 137, which underwent an intramolecular [5+2] cycloaddition to give the 5-6-7 fused tricycle 138.



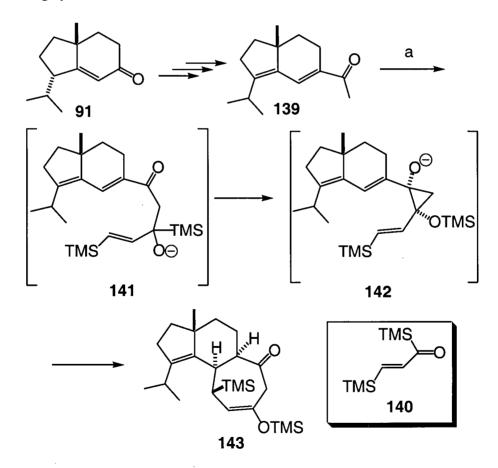


136



Reagents: a) O₂, rose bengal, hv; DMS; b) TFA.

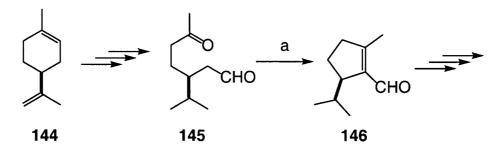
Takeda and coworkers reported a rapid assembly of the cyathane tricyclic core *via* a Brook rearrangement-mediated [3+4] annulation (Scheme 20).⁶⁶ Dienone **139**, precursor of the key annulation, was obtained in two chemical operations from the known enone **91**, which was also employed in Snider's cyathane studies (see Scheme 12, *vide supra*). Addition of the enolate of **139** to a solution of the acryloylsilane **140** resulted in a [3+4] annulation *via* intermediates **141** and **142**. 1,2-Adduct **141** underwent a Brook rearrangement to afford the divinylcyclopropane intermediate **142**. A homo-Cope rearrangement of **142** afforded compound **143**, which displayed the 5-6-7 fused tricyclic cyathane ring system.

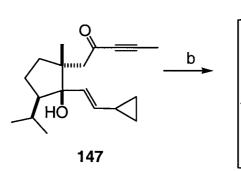


Reagents: a) LDA, 140.

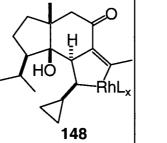
Scheme 20

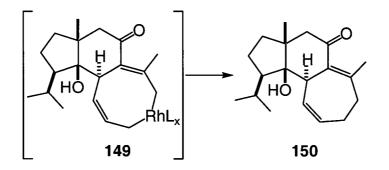
In 2001, Wender and coworkers reported an asymmetric approach to the cyathane core *via* a transition metal-catalyzed [5+2] cycloaddition.⁶⁷ (*S*)-(–)-Limonene (144) was converted to keto aldehyde 145 in two steps (Scheme 21). An intramolecular aldol condensation of 145 provided enal 146. Transformation of 146 to 147 required ten chemical operations. Treatment of 147 with $[Rh(CO)_2Cl]_2$ caused the key intramolecular [5+2] cycloaddition. The proposed pathway of this cycloaddition involved an initial complexation of the alkyne and vinylcyclopropane moieties of 147 with the Rh-catalyst. Upon oxidative addition, this complex provided intermediate 148. Strain-driven cleavage of the cyclopropane ring in 148 led to intermediate 149. Reductive elimination of 149 provided the cycloaddition product 150, which displays the cyathane 5-6-7 fused tricyclic ring-system.





÷





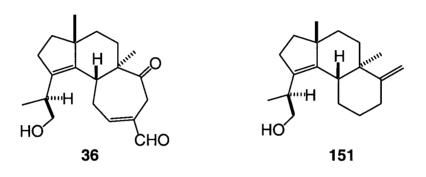
Reagents: a) piperidine, AcOH, •; b) 5 mol % [Rh(CO)₂Cl]₂, •.

Scheme 21

· · ·

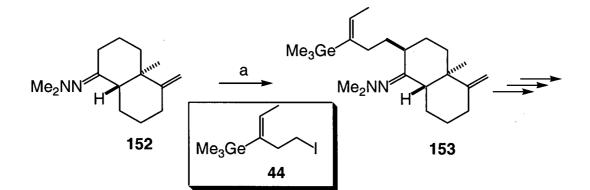
,

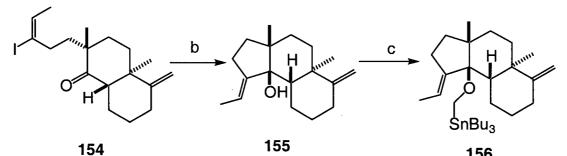
In a synthetic study directed towards (\pm)-sarcodonin G (**36**), Piers and Cook prepared the 5-6-6 fused tricycle **151** *via* sequential ring closure and Still-Mitra [2,3] sigmatropic rearrangement reactions.^{68,69}



ð

The synthesis of **151** commenced with an alkylation of the anion derived from *trans*-fused bicyclic dimethylhydrazone **152** with iodide **44**, affording adduct **153** (Scheme 22). Conversion of **153** to keto alkenyl iodide **154** was accomplished in four steps. A stereoselective BuLi-mediated anionic cyclization of **154** yielded the 5-6-6 fused tricyclic carbinol **155**. Conversion of **155** to the corresponding (tributylstannyl)methyl ether derivative **156**, followed by BuLi-mediated transmetallation, provided carbanion intermediate **157**. A [2,3]-sigmatropic rearrangement of **157** afforded alcohol **151**, which displayed the desired C–3,4 double bond and possessed the correct relative configuration of the stereogenic center at C–18 (cyathane numbering).

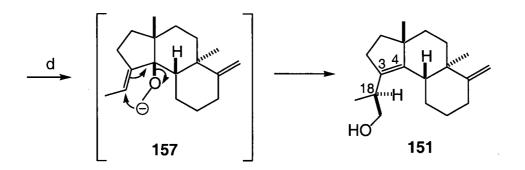




154

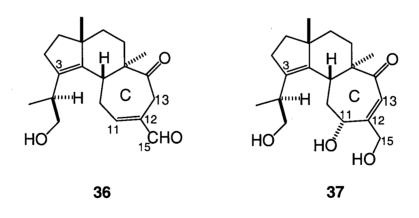


156



Reagents: a) KDA, HMPA; 44; b) BuLi; c) KH, 18-cr-6; Bu₃SnCH₂I d) BuLi.

The work described in this thesis involves synthetic studies directed towards the diterpenoids sarcodonin G (**36**) and cyathin A_4 (**37**). Both compounds display the cyathane 5-6-7 fused ring system with identical relative configurations at the ring-fusion centers. In addition, both compounds are among the few cyathanes that display a hydroxyl function on the isopropyl side chain at C-3 (see Section 2.1.1.3., *vide supra*). The structural differences between sarcodonin G (**36**) and cyathin A_4 (**37**) are restricted to the C-ring and its one-carbon appendage. Although both compounds display a ketone function at C-10, sarcodonin G (**36**) displays an aldehyde function at C-15 whereas cyathin A_4 (**37**) displays hydroxyl functions at C-11 and C-15. In addition, sarcodonin G (**36**) displays a C-11,12 double bond as opposed to the C-12,13 double bond exhibited by cyathin A_4 (**37**). These similarities and differences of structures **36** and **37** lend themselves to a centralized and divergent synthetic approach that addresses the oxygenation and unsaturation pattern of the C-ring in the later stages of the syntheses.*

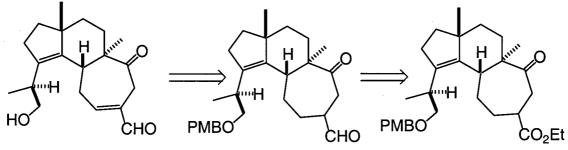


^{*} In the text of this thesis, Ayer's cyathane numbering system is used in most descriptions of cyathane-type natural products and a systematic IUPAC-based numbering system is used to describe the synthetic intermediates and the final products. The experimental section of this thesis contains IUPAC names for the synthetic intermediates and the final products.

2.1.3. Total Synthesis of (!)-Sarcodonin G

2.1.3.1. Retrosynthetic Analysis

Our proposed synthetic plan for the construction of the cyathane natural product sarcodonin G (36) is outlined in Scheme 23. The target compound 36 can presumably be derived from keto aldehyde 158 via a selenenylation/oxidation/elimination sequence to introduce a double bond in the seven-membered ring, followed by a cleavage of the PMB ether to generate the alcohol function. Reduction of keto ester 159, followed by Dess-Martin oxidation of the resultant diol, should provide the corresponding keto aldehyde **158.** In theory, the 5-6-7 fused tricyclic keto ester **159** can be obtained from the 5-6-6 fused tricycle 160 via a radical-based ring-expansion reaction. A sequence of ethoxy carbonylation and iodomethylation steps should provide the ring-expansion precursor 160 from ketone 161. Ketone 161 should be available from alcohol 160 through a protection of the hydroxyl function as a PMB ether and an oxidative cleavage of the exocyclic double bond. On the basis of previous work in our laboratories (see Section 2.1.2.2, vide supra),^{68,69} alcohol 160 was expected to be accessible from the bicyclic dimethylhydrazone 161 and the bifunctional reagent 44 via keto alkenyl iodide 154. Conversion of 154 to 151 would involve sequential BuLi-mediated cyclization and Still-Mitra [2,3]-sigmatropic rearrangement reactions.

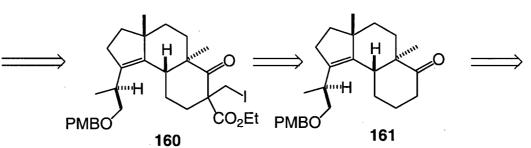


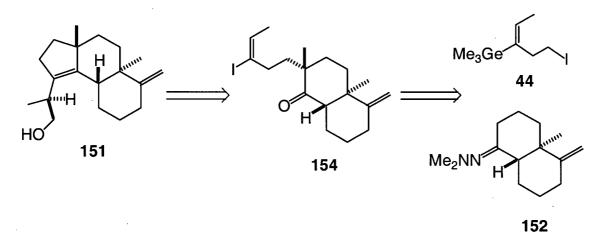
158





159



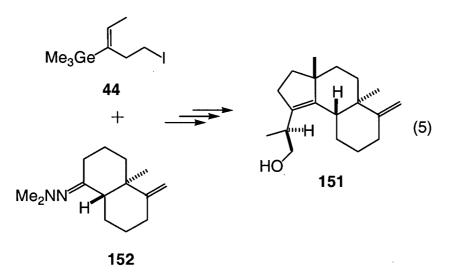




2.1.3.2. Preparation of 5-6-6 Fused Tricycle 101

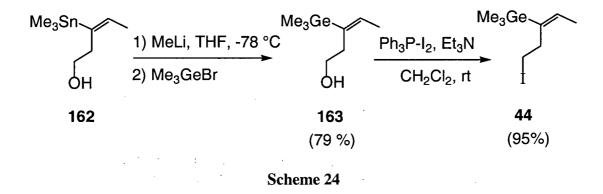
As described in Section 2.1.2.2 (Scheme 22, *vide supra*), previous work in our laboratories by K. L. Cook provided a synthetic route to the 5-6-6 fused tricyclic alcohol **151** from the bifunctional reagent **44** and *trans*-fused bicyclic dimethylhydrazone **152** (eq 5).⁶⁹ Since our study involved transformation of the advanced intermediate **151** into the

target compound sarcodonin G (*vide supra*), we embarked on a large-scale preparation of **151** and investigated several possible improvements to the previous synthesis.

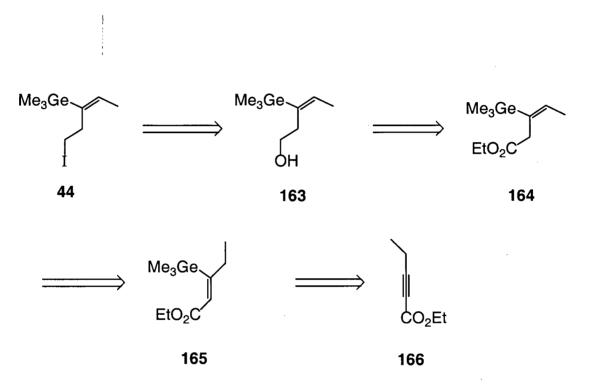


2.1.3.2.1. Preparation of Bifunctional Reagent 44

In Cook's study, the synthesis of iodide 44 (Scheme 24) commenced with the known alcohol 162, which was readily obtained in three steps from commercially available materials.⁷⁰ Transmetallation of 162 with MeLi, followed by addition of Me₃GeBr, provided the corresponding germane 163. Treatment of 163 with Ph₃P–I₂ and Et₃N furnished 44 in high yield.

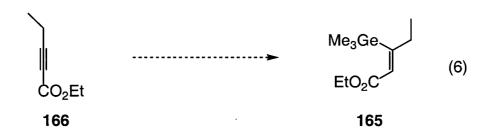


An alternative synthesis of 44, which would circumvent the tin-germanium exchange step (*vide supra*), is outlined in Scheme 25. Presumably, alcohol 163, a known precursor to iodide 44, could be obtained from a reduction of ester 164. On the basis of previous studies by our group, it was thought that a stereospecific deconjugation of the ethyl (Z)-2-pentenoate 165 would provide ethyl (E)-3-pentenoate 164. Compound 165 should to be available from ethyl 2-pentynoate (166) *via* a germylcuprate conjugate addition reaction also developed in our laboratories.



Scheme 25

The first step of our proposed synthesis of **44** involved the preparation of ethyl (Z)-3-trimethylgermyl-2-pentenoate (**165**) from the commercially available ethyl 2-pentynoate (**166**) (eq 6).



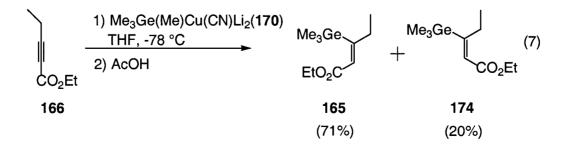
The preparation of ethyl 3-trimethylgermyl-2-butenoates *via* conjugate addition of (trimethylgermyl)copper(I) reagents **167-170** to ethyl 2-butynoates has recently been reported⁷¹ by Piers and Lemieux.

Treatment of alkynic ester **171** with germylcopper(I) reagents **167-170** was shown to afford the *E*- and *Z*-alkenylgermane isomers, **172** and **173** (**Table 1**). Conjugate addition of reagents **167** and **168** to alkynic ester **171** afforded the *E*-isomer **172** in high yields (entries 1-2, **Table 1**). A mixture of **172** and its geometric isomer **173** were obtained employing cuprate reagent **169** (entry 3, **Table 1**). Of particular interest to our study, conjugate addition of **170** favored formation of the *Z*-isomer **173** over the *E*-isomer **172** in a 3.9:1 ratio (entry 4, **Table 1**).

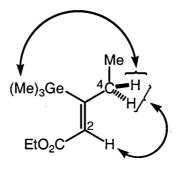
–) reagent, THF, -78 °C ♪ 2) AcOH; aq NH₄CI-NH₃	GeMe ₃ + EtO ₂ C	GeMe ₃ CO ₂ Et 173
Entry	Reagent	172:173 Ratio	Yield (%)
1	Me ₃ GeCuMe ₂ S 167	>99:1	81
2	Me ₃ GeCu(CN)Li 168	>99:1	90
3	(Me ₃ Ge) ₂ CuLi 169	1.7:1	90
4	Me ₃ Ge(Me)Cu(CN)Li ₂ 170	1:3.9	86

 Table 1. Addition of Germylcopper(I) Reagents (167-170) to Ethyl 2-Butynoate (171).

On the basis of the stereoselectivity displayed by the reaction summarized in entry 4 (**Table 1**), the conjugate addition of germylcuprate reagent Me₃Ge(Me)Cu(CN)Li₂ (**170**) was applied to the preparation of ethyl (*Z*)-3-trimethylgermyl-2-pentenoate (**165**). Ethyl 2-pentynoate (**166**) was added to a cold (-78 °C) solution of **170**, and the resultant mixture was treated with AcOH (eq 7). Upon aqueous work-up and purification of the resulting material by flash chromatography, ethyl (*Z*)-3-trimethylgermyl-2-pentenoate (**165**) and the corresponding *E*-isomer, **174**, were obtained in 71% and 20% yields, respectively.



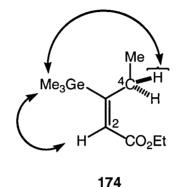
The structure of **165** was confirmed by spectrometric analysis. The IR spectrum of **165** exhibited an ester carbonyl stretch at 1718 cm⁻¹. The ¹H NMR spectrum of **165** displayed singlets at δ 0.29 (9H) and 6.21 (1H) attributed to the Me₃Ge– moiety and the olefinic proton, respectively. The ¹³C NMR resonances for the olefin carbons and the ester carbonyl carbon were observed at δ 126.6 and 166.8, and at δ 171.4, respectively.



¹H NMR NOED experiments were performed to establish the geometry of the alkene function in **165**. Thus, irradiation at δ 2.30, a quartet attributed to the allylic methylene protons (H–4), caused enhancement of the signals at δ 6.21 and at δ 0.29, assigned to the olefinic proton (H–2) and the Me₃Ge– protons, respectively. When the signal at δ 0.29 (Me₃Ge–) was irradiated, enhancement of the quartet at δ 2.30 (H–4) was observed. Irradiation of the signal at δ 6.21 (H–2) also caused the quartet at δ 2.30 (H–4) to be enhanced. These experiments, along with the spectral data presented above are consistent with the structural assignment for ethyl (*Z*)-3-trimethylgermyl-2-pentenoate (**165**).

Spectrometric analysis of the *E*-isomer 174 supported its assigned structure. The ester carbonyl stretch of 174 was observed at 1718 cm⁻¹ in its IR spectrum. The ¹H NMR

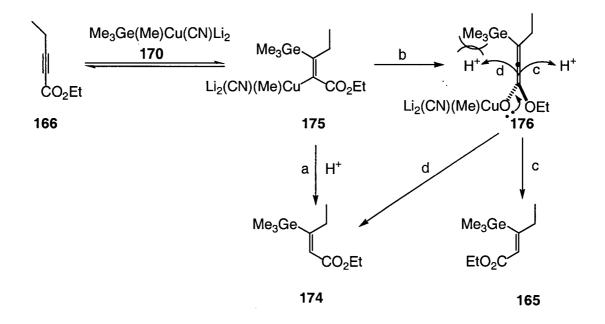
signals for the Me₃Ge– moiety and olefinic proton in **174** were displayed at δ 0.23 and 5.89, respectively. The ¹³C NMR spectrum of **174** displayed resonances at δ 124.5 and 164.9, and at δ 171.3, representing the olefin and carbonyl carbons, respectively.



The geometry of the alkene function in **174** was established by ¹H NMR NOED experiments. Irradiation at δ 2.73, a quartet attributed to the allylic methylene protons (H–4), caused enhancement of the singlet at δ 0.23, assigned to the protons of the Me₃Ge– group. Irradiation of the olefinic proton (H–2) signal at δ 5.89 caused enhancement of the singlet at δ 0.23 (Me₃Ge–). When the signal at δ 0.29 (Me₃Ge–) was irradiated, the signals at δ 2.73 (H–4) and 5.89 (H–2) were enhanced. These experiments confirm the *E* configuration of the alkene function in **174**.

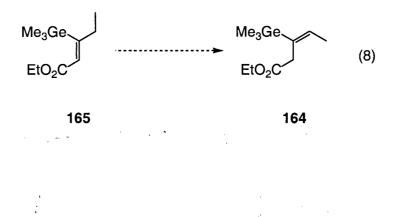
The formation of 165 and 174 can be rationalized by the possible pathway⁷¹ depicted in Scheme 26. A reversible *cis* addition of cuprate 170 to ethyl 2-pentynoate (166) provides the alkenyl copper immediate 175. Although direct protonation of 175 would produce the *E*-isomer 174 (path a), it appears that the majority of 175 isomerizes to the allenoate species 176 (path b). Protonation of allenoate 176 would occur predominantly from the side opposite to of the bulky Me₃Ge– moiety (path c), providing

primarily the Z-isomer 165. Protonation of allenoate 176 from the same side as the $Me_3Ge-moiety$ (path d) would provide E-isomer 174.



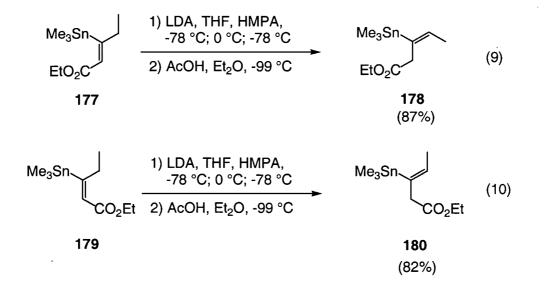
Scheme 26

With the ethyl (Z)-2-pentenoate 165 in hand, the next step involved a stereospecific deconjugation to provide ethyl (E)-3-pentenoate 164 (eq 8).

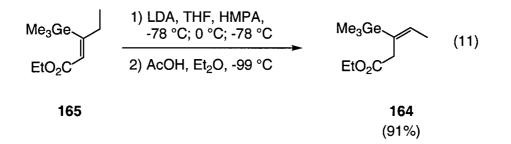


50

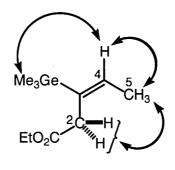
In 1990, Piers and Gavai reported⁷⁰ a study on the stereospecific deconjugations of ethyl 3-trimethylstannyl-2-pentenoates. For example, addition of (Z)-2-pentenoate **177** to a solution of LDA, followed by an inverse addition of the resultant enolate to a solution of AcOH, provided (*E*)-3-alkenoate **178** (eq 9). In a similar fashion, (*E*)-2alkenoate **179** was transformed, completely stereoselectively, into the corresponding (*Z*)-3-alkenoate **180** (eq 10).



It was expected that the application of this deconjugation reaction (*vide supra*) to ethyl (*Z*)-3-trimethylgermyl-2-pentenoate **165** would provide the corresponding ethyl (*E*)-3-pentenoate **164**. Thus, a solution of **165** in THF was added to a solution of LDA and HMPA in THF (eq 11). Subsequent inverse addition of the resultant enolate solution to a cold (-98 °C) solution of AcOH in Et₂O provided, upon work-up, a 91% yield of the desired ethyl (*E*)-3-pentenoate **164**.



Standard spectroscopic methods were used to confirm the structure of trimethylgermyl alkenoate **164**. A carbonyl absorption at 1737 cm⁻¹ in the IR spectrum of **164** was characteristic of an aliphatic ester. The ¹H NMR spectrum of **164** displayed a singlet (2H) at δ 3.16, attributed to the allylic methylene α to the ester function. The signals corresponding to the alkenyl methyl and proton moieties were observed at δ 1.69 (doublet, J = 6.7 Hz) and δ 5.87 (quartet, J = 6.7 Hz), respectively. The ¹³C NMR spectrum displayed resonances at δ 135.5 and 135.7, assigned to the olefinic carbons. The ¹³C signal for the ester carbonyl carbon was observed at δ 171.8.

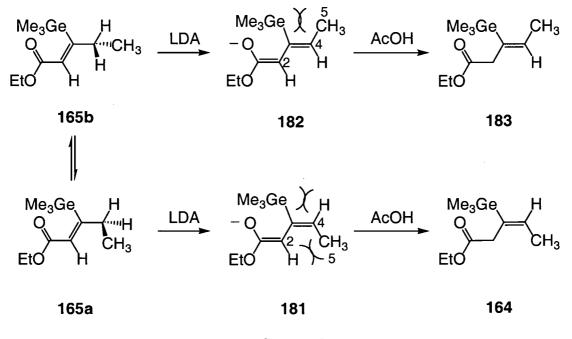


164

The geometry of the alkene function in **164** was confirmed by ¹H NMR NOED experiments. Irradiation at δ 1.69, attributed to the methyl protons (H–5), caused enhancement of resonances at δ 5.87 and 3.16, assigned to the olefinic proton (H–4) and

methylene protons (H–2), respectively. When the olefinic proton (H–4) signal at δ 5.87 was irradiated, the signals at δ 1.15 (Me₃Ge–) and 1.69 (H–5) were enhanced. These experiments, along with the spectral data presented above are consistent with the structural assignment for ethyl (*E*)-3-trimethylgermyl-3-pentenoate (**164**).

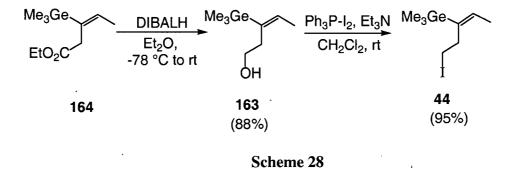
The stereoselective formation of 164 from 165 may be qualitatively rationalized⁷⁰ as shown in Scheme 27. The relative transition states for the kinetically controlled deprotonation of two ground state conformations, 165a and 165b, leading to the respective extended enolates 181 and 182, were considered. The transition state leading to the extended enolate 182 would be highly destabilized by the developing $A^{1,2}$ strain (between CH₃-5 and the Me₃Ge-group). On the other hand, the transition state leading to the extended enolate 181 would be destabilized by the relatively smaller steric repulsions due to A^{1,2} strain (H-4 and Me₃Ge-) and A^{1,3} strain (CH₃-5 and H-2). In addition, if the transition state leading to the extended enolate 181 contains some allylic anion-type character it may experience stabilization from the fact that the 181 contains a cis alkyl group.⁷² Strong evidence provided by experimental and theoretical studies suggests that allylic anions systems containing *cis* alkyl groups are more stable than those possessing *trans* alkyl moieties.⁷³ Hence, for this reason and for steric reasons, the deprotonation step would be expected to favor the formation of the allylic anion 181, which affords the *E*-alkenoate 164, upon protonation.



54

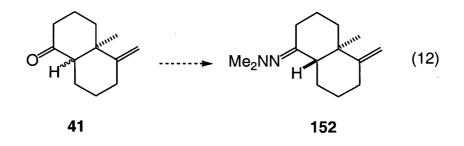


The final steps in the synthesis of bifunctional reagent 44 from ester 164 were straightforward (Scheme 28). DIBALH reduction of ester 164 provided alcohol 163 in 88% yield. As demonstrated by Cook (see Scheme 24 earlier), addition of 163 and Et₃N to a solution of Ph₃P–I₂ in CH₂Cl₂ provided the corresponding iodide 44 in high yield. All spectral data derived from compounds 163 and 44 were identical to those reported⁶⁹ by Cook.

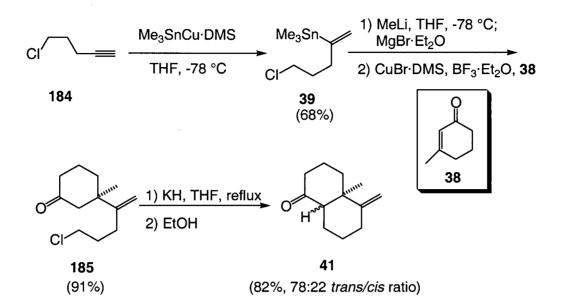


2.1.3.2.2. Preparation of cis-Fused Bicyclic Dimethylhydrazone 102

With the desired electrophile 44 in hand, preparation of the *trans*-fused bicyclic dimethylhydrazone 152 from the ketones 41 was investigated (eq 12).

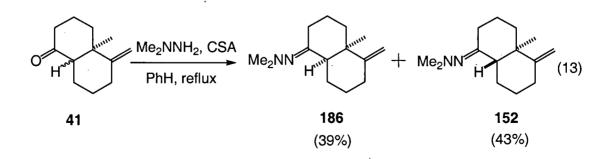


The known bicyclic ketones **41** were prepared following a 3-step procedure developed in our laboratories.¹⁸ Commercially available 5-chloro-1-pentyne (**184**) was added to a solution of Me₃SnCu·DMS complex, providing, upon work-up, the bifunctional reagent **39** (Scheme 29). A copper-catalyzed conjugate addition of the Grignard reagent derived from **39** to 3-methyl-2-cyclohexen-1-one (**38**) afforded the cyclic keto chloride **185**. KH-mediated cyclization of the keto chloride **185**, followed by an ethoxide-mediated epimerization reaction, yielded a 78:22 mixture of the *trans*- and *cis*- fused bicyclic ketones **41**.

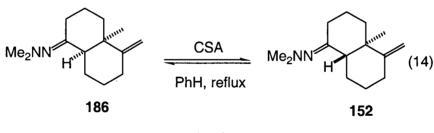


Scheme 29

As described in Cook's study,⁶⁹ the conversion of ketones **41** into the corresponding hydrazone derivatives was accomplished by treatment of a benzene solution of the former with dimethylhydrazine and a catalytic amount of CSA (eq 13). The resultant mixture was refluxed for 72 h with azeotropic removal of water. Upon solvent removal and purification of the resultant material by flash chromatography, the *cis*- and *trans*-fused bicyclic dimethylhydrazones, **186** and **152**, were obtained in 39% and 43% yields, respectively.



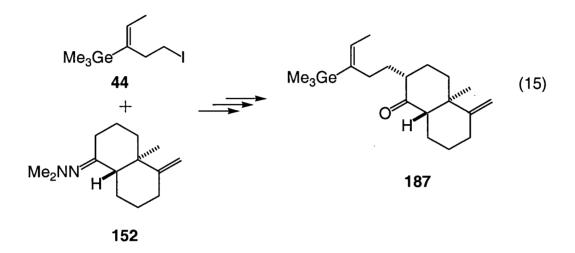
In the previous study, the *trans*-fused isomer **152** was carried through to the next step but the undesired *cis*-fused isomer **186** was abandoned. For our large-scale synthesis, a method of recycling the large quantities of the *cis*-fused **186** was sought. Although **186** did not appear to isomerize under standard base-mediated conditions (NaOMe/MeOH or *t*-BuOK/*t*-BuOH), acid-mediated isomerization conditions (CSA, benzene) yielded significant amounts of the desired *trans*-fused isomer **152** (eq 14). Thus, a catalytic amount of CSA was added to a benzene solution of the **186** and the resultant mixture was refluxed for 48 h. Upon work-up and purification of the resulting material by flash chromatography, **186** and **152** were obtained in nearly a 1:1 ratio. The *cis*-fused isomer **186** was recycled twice through the epimerization step, yielding additional **152** for a total overall yield of 71% from the mixture of ketones **41**.



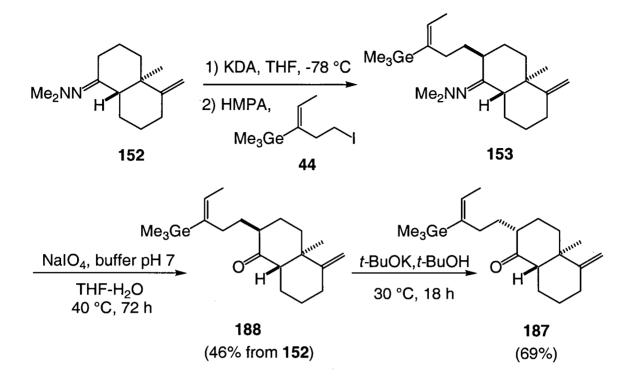
1:1 ratio of 186:152

2.1.3.2.3. Preparation of Ketone 187

With building blocks 152 and 44 in hand, the synthetic sequence leading to ketone 187 was investigated (eq 15).

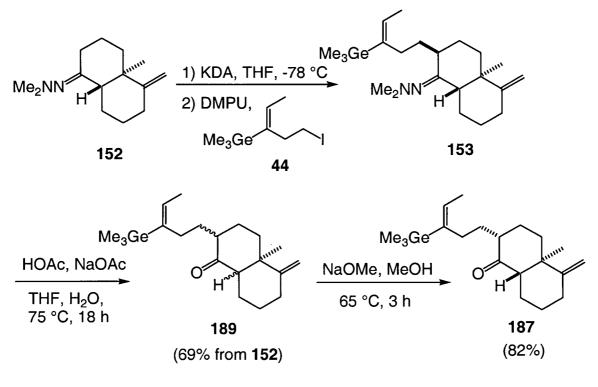


Cook's preparation⁶⁹ of **187** from **152** and **44** is outlined in Scheme 30. Deprotonation of the *trans*-fused hydrazone **152** with KDA, followed by addition of HMPA and iodide **44** to the resulting enolate solution, afforded adduct **153** after work-up. The crude **153** was taken up in a buffered aqueous THF mixture and the latter was treated with sodium periodate. The resultant mixture was heated to 40 °C and stirred for 72 h, affording ketone **188** in 46% yield. Isomerization of **188** was accomplished by treatment of this material with *t*-BuOK/*t*-BuOH, thus providing a 69% yield of isomer **187**. The overall yield from **152** to **187** was 32% and the reaction times for the last two steps totaled 90 h.



Scheme 30

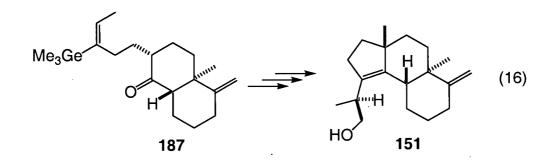
The optimized experimental procedures employed for our large-scale conversion of **152** to **187** are outlined in Scheme 31. Alkylation of the potassium enolate derived from **152** with iodide **44** was accomplished in the presence of DMPU, a less-toxic alternative to carcinogenic HMPA. The crude hydrazone **153** was subjected to hydrolysis with NaOAc/AcOH in aqueous THF, affording a mixture of isomeric ketones **189** in 69% yield from **152**. A 3 h isomerization of **189** in NaOMe/MeOH afforded a 66% yield of ketone **187** and 22% of three other diastereomers. The mixture of undesired ketone isomers was resubmitted to the same isomerization reaction to yield an additional quantity of **187** for a total overall yield of 82% from **189**. In comparison with the previous synthesis (*vide supra*), the total overall yield from **152** to **187** had been increased from 32% to 57% and the total reaction times of the latter two steps had decreased from 90 h to 24 h. All spectral data derived from **187** were identical with those of the corresponding data reported⁶⁹ by Cook.



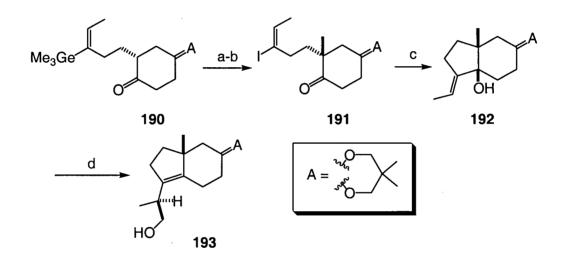
Scheme 31

2.1.3.2.4. Preparation of Alcohol 151

With ketone **187** in hand, the sequence of reactions leading from this substance to alcohol **151** was investigated (eq 16).



In 1996, Piers and Cook reported⁶⁸ an annulation method that relied on sequential BuLi-mediated ring closure and Still–Mitra [2,3]-sigmatropic rearrangement¹⁹ reactions. For example, methylation of **190**, followed by iododegermylation, gave keto alkenyl iodide **191** (Scheme 32). Treatment of **191** with BuLi resulted in an anionic ring closure, affording the *cis*-fused alcohol **192**. Conversion of **192** to its tributylstannylmethyl ether derivative, followed by a Still–Mitra [2,3]-sigmatropic rearrangement, provided alcohol **193**.

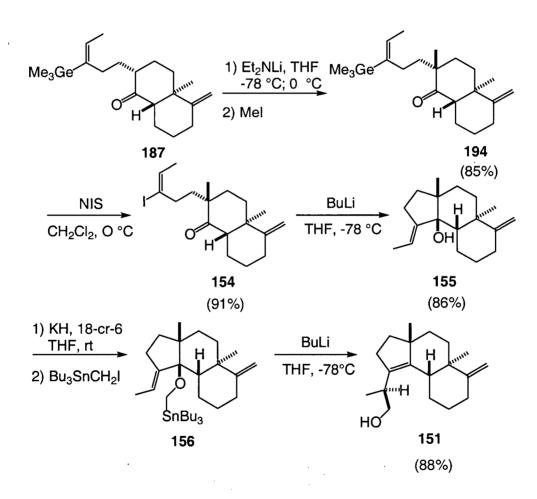


Reagents: a) *t*-BuOK, THF, HMPA; MeI; b) I₂, CH₂Cl₂; c) BuLi, THF; d) KH, 18-cr-6; Bu₃SnCH₂I; BuLi.

Scheme 32

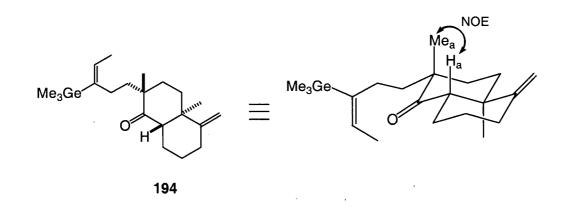
This novel annulation sequence (*vide supra*) was successfully applied to the conversion of **187** to **151**, as reported in Cook's previous study. The experimental procedures developed⁶⁹ by Cook were also employed in our large-scale preparation of **151** (Scheme 33). Methylation of **187** furnished **194** in 85% yield. The reaction involving iododegermylation of **194** with NIS in CH_2Cl_2 was stopped after 1 h, affording the corresponding alkenyl iodide **154** in 91% yield. In Cook's study, the NIS-mediated

iododegermylation process was stopped after 15 min, affording **154** in only 69% yield. A BuLi-mediated anionic ring-closure of keto alkenyl iodide **154** provided alcohol **155** in 86% yield. Treatment of **156**, the tributylstannylmethyl ether derivative of alcohol **155**, with BuLi resulted in the required [2,3]-sigmatropic rearrangement, affording alcohol **151** in 88% yield from **155**. All spectral data derived from the **194**, **154**, **155** and **151** were identical with those reported⁶⁹ by Cook.



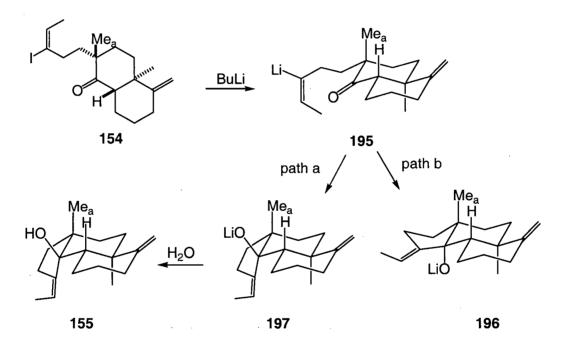
Scheme 33

Many stereochemical aspects of the sequence of transformations leading from 187 to 151 have been discussed elsewhere.^{68,69} However, several key features noted⁶⁹ by Cook that support the assigned relative configurations of intermediates 194, 155 and 151, are certainly worth outlining. The successful stereoselective axial methylation of 187 was confirmed by ¹H NMR NOED experiments performed on compound 194. In the ¹H NMR spectrum of 194, the newly introduced methyl group (Me_a) and the angular proton (H_a) gave rise to signals at δ 1.17 and 2.44, respectively. In NOED experiments, irradiations at δ 1.17 and 2.24 resulted in a mutual enhancement of these two resonances, thus establishing the *cis* relationship between the newly introduced methyl moiety (Me_a) and the angular proton (H_a).



The BuLi-mediated cyclization of iodide 154 was completely stereoselective, affording the *cis*-fused tricycle 155, exclusively (Scheme 34). The *cis* configuration of the ring-fusion of 155 was not unambiguously determined. However, on the basis of previously reported studies⁶⁸ and molecular modeling, the presence of the angular methyl Me_a in 195, the intermediate derived from 154, was expected to result in the exclusive

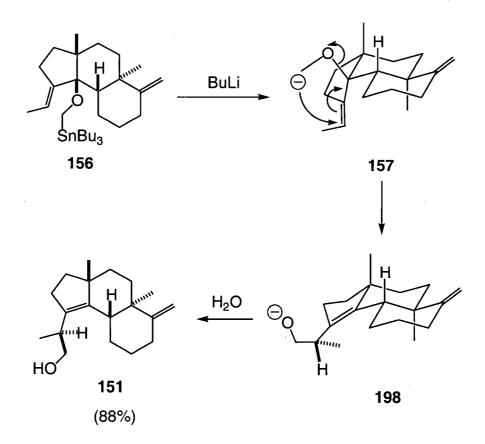
formation of a *cis*-fused cyclization product **197**. The molecular conformation of the transition state necessary for the cyclization of the alkenyllithium intermediate **195** leading to *cis*-fused alcohol **197** (path a) would appear to be considerably less strained than that leading to the corresponding *trans*-fused alcohol **196** (path b). The molecular conformation of the transition state leading to the *trans*-fused alcohol **196** would likely experience significant angle strain in the forming five membered ring in addition to the steric strain involving gauche interaction between the angular methyl Me_a and the incoming alkenyllithium moiety.



Scheme 34

The stereospecific nature of the Still–Mitra [2,3]-rearrangement, which provided alcohol **151** from tributylstannylmethyl ether **156**, has been well established (Scheme 35).^{19,74} Transmetallation of tributylstannylmethyl ether **156** with BuLi afforded the

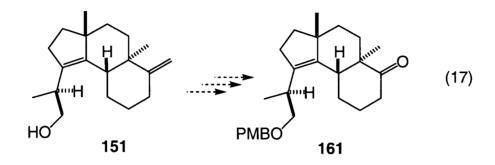
carbanion intermediate **157**. The rearrangement process would take place in a suprafacial manner, affording alkoxide **198**. Thus, the relative configuration of alcohol **151**, obtained upon aqueous work-up of **198** could be assigned with confidence.



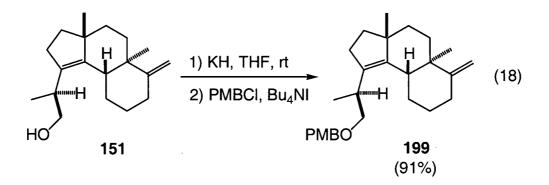
Scheme 35

2.1.3.3. Preparation of Ketone 112

Having completed a large-scale preparation of alcohol **151**, which represents the most advanced intermediate in Cook's study,⁶⁹ the preparation of ketone **161** was investigated (eq 17).

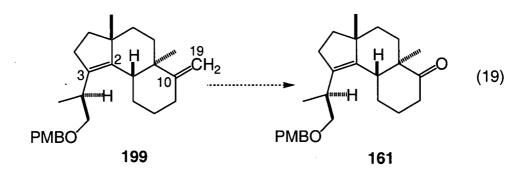


The first step was envisaged to be the protection of the hydroxyl moiety in **151** by converting this material into the corresponding *p*-methoxybenzyl ether **199** (eq 18). The *p*-methoxybenzyl (PMB) protecting group was chosen for its resistance to acidic and basic conditions. When required, the PMB group can be readily removed under mild neutral conditions with DDQ at room temperature.⁷⁵ Thus, the alkoxide derivative of alcohol **151**, which was generated *via* a deprotonation of **151** by KH in THF, was treated with *p*-methoxybenzyl chloride and a catalytic amount of Bu₄N⁺T. The Bu₄N⁺T caused a Finkelstein-type *in situ* conversion of the PMBCl into the more reactive electrophile, *p*-methoxybenzyl iodide. Standard work-up and purification of the resultant material by flash chromatography provided the *p*-methoxybenzyl ether **199** in 91% yield.

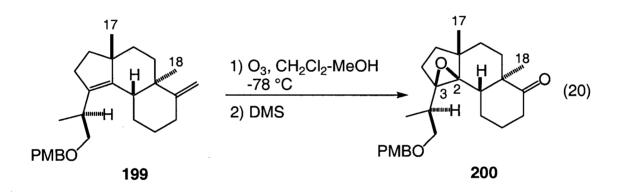


The successful formation of ether **199** was supported by ¹H NMR spectroscopic analysis of the product. The ¹H NMR spectrum of **199** displayed a 3-proton singlet at δ 3.78, representing the methoxy moiety of the PMB protecting group. In addition, two doublets were observed at δ 6.86 (2H, J = 8.9 Hz) and δ 7.23 (2H, J = 8.9 Hz), representing the aromatic protons of the *para*-substituted benzyl group.

With the ether diene **199** in hand, the next task was the chemoselective oxidative cleavage of the C–10(19) exocyclic double bond to form ketone **161** (eq 19). However, it was found (*vide infra*) that this chemoselective oxidation was difficult to achieve owing to the small difference in reactivity between the C–10(19) exocyclic double bond and the C–2 endocyclic double bond.

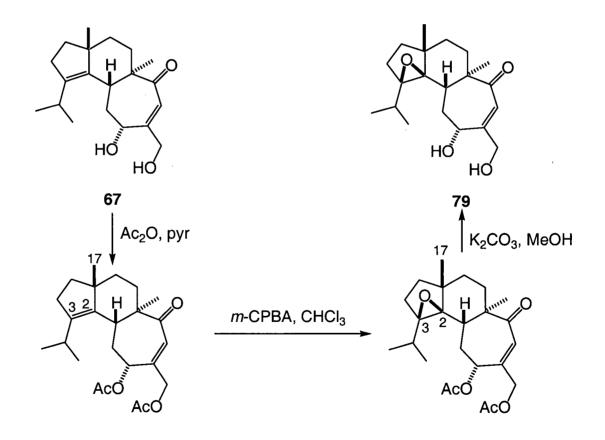


Ozonolysis was the first method investigated to achieve the required chemoselective oxidative cleavage. Following a standard ozonolysis procedure, a gaseous flow of ozone in O_2 was passed through a cold solution of diene **199** in MeOH/CH₂Cl₂ for 10 min, at which point the solution turned light blue, indicating the presence of excess ozone in the mixture (eq 20). Reductive work-up with dimethyl sulfide was followed by removal of the volatiles under reduced pressure. Purification of the resultant material by flash chromatography provided a major product, which was tentatively assigned structure **200**.



The assigned structure of compound **200** was supported by spectrometric analysis. A molecular mass peak at 412 in the mass spectrum of **200** confirmed its $C_{26}H_{36}O_4$ chemical formula. The ¹H NMR spectrum of **200** displayed 3-proton singlets at δ 1.24 and 0.86, attributed to the H–17 and H–18 angular methyl protons, respectively. Previously, the ¹H NMR signal for the corresponding H–17 and H–18 angular methyl protons in diene **200** had been noted at δ 1.01 and 0.92, respectively. The apparent deshielding of the H–17 angular methyl protons in **200** relative to those of **199** was attributed to the newly introduced β-face epoxide function at C–2,3 in **200**. This rationale for the proposed structure of **200** was based on an analogous argument by Ayer and coworkers (*vide infra*).

A similar β -face selective epoxidation was noted by Ayer and coworkers in their reported structural elucidation studies on neoallocyathin A₄ (**79**) involving spectroscopic correlations with a derivative of cyathin A₃ (**67**) (see Section 2.1.1.3., *vide supra*).¹⁷ Compound **201**, a diacetyl derivative of cyathin A₃ (**67**), was treated with *m*-CPBA to afford the β -epoxide **202** (Scheme 36). Treatment of a methanol solution of **202** with potassium carbonate gave neoallocyathin A₄ (**79**), which was spectroscopically identical with the corresponding natural product.



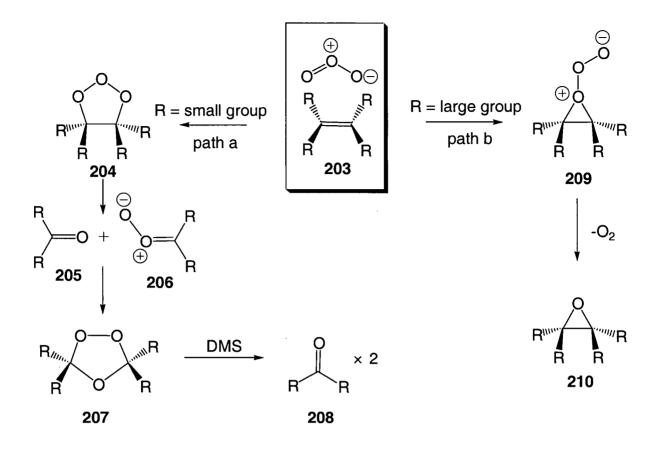




202

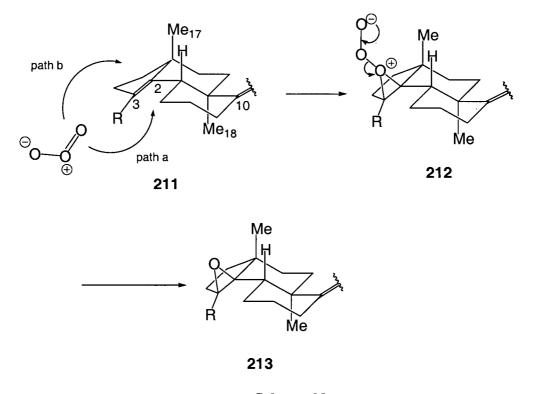
The rationale used to assign the structure of **200** in our study (*vide supra*) was based on the following evidence presented by Ayer and coworkers. The ¹H NMR signal attributed to the H–17 angular methyl protons of **202** was noted at δ 1.28. The ¹H NMR signal for the corresponding H–17 methyl protons in **201** had previously been noted at δ 1.07. The downfield shift of the H–17 signal for **202** relative to that of the H–17 signal for **201** was attributed to significant deshielding by the newly introduced C–2,3 β epoxide in **202**.

Although the epoxidation of 201 upon treatment with *m*-CPBA, as reported by Ayer and coworkers, was certainly expected, the epoxidation of 200 by ozonolysis, as noted in our study (*vide supra*), was not anticipated. A brief review of the literature provided some insight into this undesired ozonolysis side-reaction. The proposed mechanism for the ozonolysis reaction of non-hindered olefins (path a) involves three steps (Scheme 37).⁷⁶ The first step is an electrophilic cycloaddition of ozone to the alkene (see 203 \rightarrow 204, Scheme 37). Cleavage of the resultant ozonide 204 provides carbonyl 205 and carbonyl oxide zwitterion 206, which recombine to afford the cyclic ozonide 207. Reductive work-up of 207 with dimethyl sulfide provides two carbonyl compounds, 208. On the other hand, it has been shown that some hindered alkenes appear to be too sterically encumbered to allow for an electrophilic cycloaddition with ozone.⁷⁷ Alternatively, upon treatment with ozone, a hindered alkene (203) may form the peroxyepoxide intermediate 209 (path b). Loss of one molecule of dioxygen from 209 provides epoxide 210.⁷⁸



Scheme 37

In our study, the observed formation of a β -face epoxide upon ozonolysis of **199**, represented by partial structure **211**, may be rationalized with the help of Scheme 38. Only partial structures are illustrated because the state of the alkene at C–10, which is ultimately cleaved in this step, is not known at the time of the epoxidation. On the basis of molecular models, an approach by ozone to the α -face of C–2 double-bond in **211** (path a) would experience more steric interference, including 1,3-diaxial interaction with Me–18, than would the corresponding β -face approach (path b). Loss of dioxygen by the resulting peroxyepoxide **212** would afford intermediate **213**, displaying a β -configuration of the epoxide moiety.

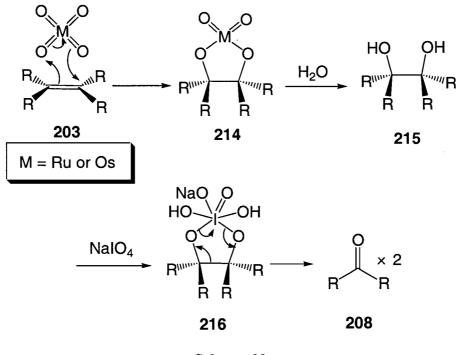


Scheme 38

In an attempt to prevent epoxide formation, the ozonolysis was repeated in the presence of Sudan Red 7B,⁷⁹ a dye that allows detection of low ozone concentrations. Despite careful observation and short reaction times, the epoxidation could not be prevented and the desired reaction could not be accomplished. Therefore, the ozonolysis approach to preparing **161** from **199** was abandoned and alternative methods for the chemoselective oxidative cleavage were investigated.

Certain transition metal-based oxidants have been shown to effect the cleavage of C-C double bonds *via* the pathway illustrated in Scheme 39.⁸⁰ Treatment of an alkene (203) with ruthenium tetroxide or osmium tetroxide generates intermediate 214 *via* a stereospecific *syn*-addition. Vicinal diol 215, derived from 214, can be oxidatively

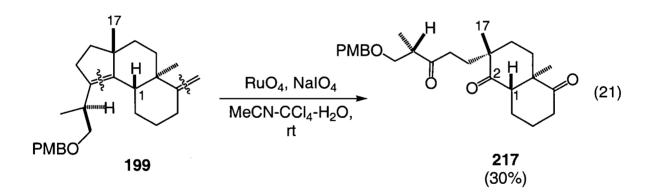
cleaved by treatment with a periodate reagent. The proposed mechanism of this transformation involves cleavage of the cyclic adduct **216** to afford carbonyl compounds **208**.



Scheme 39

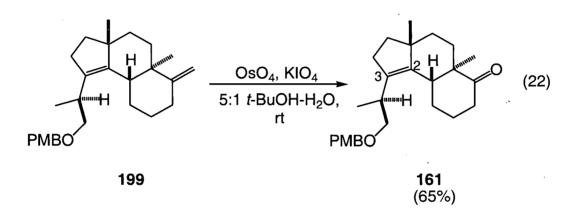
Ruthenium tetroxide is a toxic, volatile oxidant that is generally prepared by *in situ* oxidation of RuO_2 by a stoichiometric amount of periodate. The periodate reagent therefore serves the dual purpose of re-oxidizing the catalyst and cleaving the diol intermediate. Thus, in the presence of two equivalents of periodate and a catalytic amount of ruthenium dioxide, an olefin can undergo complete oxidative cleavage. Using modified conditions described by Sharpless and coworkers,⁸¹ a solution of the diene **199** in a 1:1:1.5 mixture of acetonitrile, carbon tetrachloride and water was treated with a catalytic amount of ruthenium dioxide and two equivalents of sodium periodate (eq 21).

Work-up and purification of the resultant material by flash chromatography provided one major product and 50% of recovered starting material (**199**). On the basis of spectroscopic analysis, the major product was tentatively assigned structure **217**, derived from the oxidative cleavage of both double bonds in diene **199**. A molecular mass peak at 428 in the mass spectrum of **217** confirmed its $C_{26}H_{36}O_5$ chemical formula. The ¹H NMR spectrum of **217** displayed signals at δ 1.15 and 2.65, attributed to H–17 and H–1, respectively. The corresponding ¹H NMR signals for the H–17 and H–1 in diene **199** were previously noted at δ 1.01 and 1.99, respectively. The downfield shifts noted for the signals of the H–17 and H–1 of **217** relative to those of diene **199** may be partially attributed to deshielding by the newly introduced carbonyl at C–2 in **217**.



Considering the lack of chemoselectivity displayed by ruthenium tetroxide, the less reactive osmium tetroxide oxidant was investigated. Like ruthenium tetroxide, osmium tetroxide is a toxic, volatile oxidant and is generally used in catalytic amounts with *in-situ* re-oxidation. Gratifyingly, treatment of diene **199** with a catalytic amount of osmium tetroxide and several equivalents of sodium periodate afforded the desired ketone **161** as the main product. However, the reaction was quite sluggish and, despite long reaction times (12-24 h) and increased reaction temperatures (from 20 °C to 50 °C),

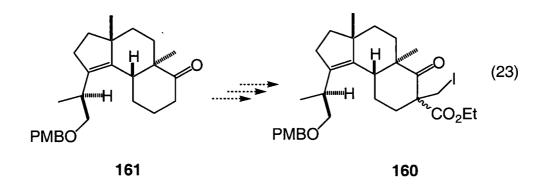
the yields were disappointingly low (30-40%). Addition of sodium bicarbonate and replacement of the sodium periodate co-oxidant with potassium periodate slightly improved the observed yields.⁵⁶ The optimal reaction conditions involved the addition of potassium periodate and sodium bicarbonate to a solution of diene **199** in 5:1 *t*-BuOH– H_2O (eq 22). A catalytic amount of osmium tetroxide was added and the resultant brown mixture was stirred for 72 h. Standard work-up and purification of the resultant material by flash chromatography provided the ketone **161** in 65% yield.



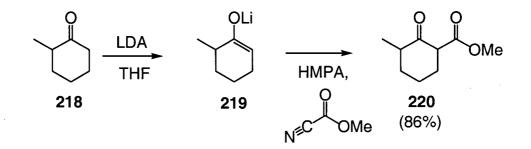
Spectroscopic analysis of product **161** confirmed that the required chemoselective oxidative cleavage of the exocyclic double bond in **199** had been achieved. The IR spectrum of the ketone **161** showed a sharp absorption band at 1703 cm⁻¹, characteristic of an aliphatic ketone function. The ¹³C NMR spectrum of ketone **161** displayed a carbonyl carbon signal at δ 216.3. Olefinic ¹³C NMR resonances were observed at δ 137.7 and 138.2, confirming the integrity of the C–2 double bond in **161**.

2.1.3.4. Preparation of Iodides 160

With ketone **161** in hand, the preparation of iodides **160** *via* sequential ethoxy carbonylation and halomethylation steps was investigated (eq 23).

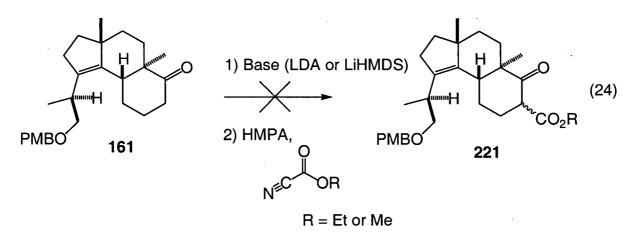


In 1983, Mander and coworkers reported an efficient method for effecting the alkoxycarbonylation of ketones with high *C*-selectivity (Scheme 40).⁸² Their method was based on the reaction of lithium enolates with alkyl cyanoformate reagents. For example, deprotonation of 2-methylcyclohexanone **218** with LDA generated the corresponding lithium enolate **219**, which, when treated with methyl cyanoformate in the presence of HMPA, afforded the β -keto ester **220**.

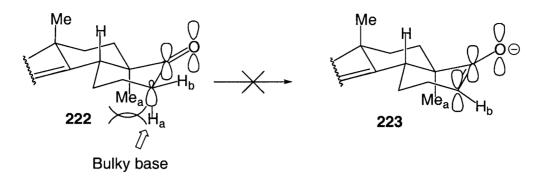


Scheme 40

The alkoxycarbonylation of ketone **161** following Mander's procedure was attempted several times with either LDA or LiHMDS as the base and either methyl or ethyl cyanoformate as the acylating reagent (eq 24). To our disappointment, the desired keto ester **221** was not produced and nearly quantitative recovery of starting material was noted in most attempts.

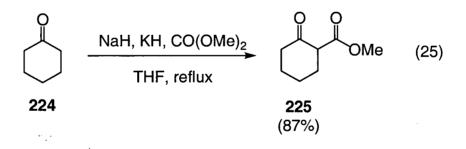


To determine whether or not the generation of the enolate of **161** had occurred, the ketone was again treated with base (LDA or LiHMDS) and the resulting solutions were treated with TMSCI. However, the corresponding enol silyl ether was not detected and, in each case, starting material was completely recovered. A rationale for the apparent difficulty in generating the requisite ketone enolate of **161** was based on steric and stereoelectronic effects (Scheme 41). In accordance with Corey's proposal,⁸³ the axial proton H_a in **222**, a partial structure representing ketone **161**, should be more readily abstracted than H_b, the equatorial proton, for stereoelectronic reasons. However, on the basis of possible steric interaction between the incoming base and the angular methyl group (Me_a), the transition state energy for the deprotonation of **222** with a bulky base (LDA or LiHMDS) leading to enolate **223** was expected to be relatively high.

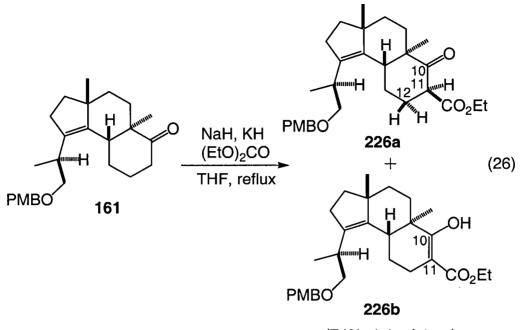


Scheme 41

In 1990, Weiler and coworkers reported an efficient method of generating β -keto esters, which employed the relatively small bases NaH and KH.⁸⁴ For example, a solution of ketone **224** and dimethyl carbonate in THF was refluxed in the presence of several equivalents of sodium hydride and 0.1 equivalent of potassium hydride (eq 25). Upon work-up, the desired β -keto ester **225** was obtained in 87% yield.



A diethyl carbonate version of Weiler's procedure⁸⁴ was applied to our synthesis. A mixture of ketone **161** and diethyl carbonate in THF was refluxed with sodium hydride and a catalytic amount of potassium hydride (eq 26). Upon work-up and purification of the resultant material by flash chromatography, product **226** was obtained in 74 % yield as a 1:1 tautomeric mixture of enol ester **226a** and keto ester **226b**.

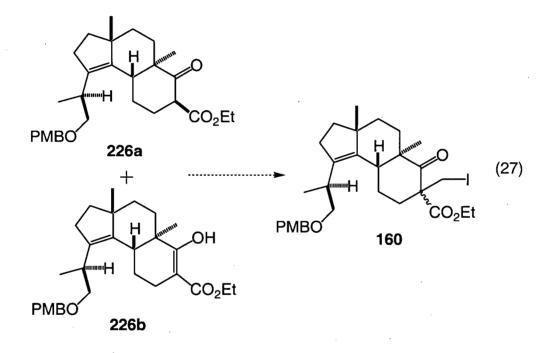


(74%, 1:1 mixture)

The structures of **226a** and **226b** were confirmed by spectroscopic methods. The IR spectrum of the mixture of tautomers **226** displayed an absorption at 1649 cm⁻¹, attributed to the ester carbonyl stretch of the enol ester **226b**. In addition, absorptions at 1709 and 1743 cm⁻¹, derived from the ketone and ester carbonyls of the keto ester **226a**, were also observed. The ¹H NMR spectrum of the mixture of tautomers **226** displayed overlapping triplets (3H) at δ 1.29 and overlapping quartets (2H) at δ 4.19, attributed to the methyl and methylene protons of the ethoxyl moieties. A doublet of doublets (1H, *J* = 5.5, 13.5 Hz) at δ 3.68 was assigned to the H–11 proton of the α -keto ester tautomer **226a**. The larger of the two coupling constants, 13.5 Hz, was attributed to the trans diaxial relationship between H–11 and the β -face H–12 proton. A singlet (1H) at δ 12.4 was attributed to the hydroxyl proton of the enol ester tautomer **226a** and **226b** was supported by an observed 1:1 integral ratio for the signals at δ 3.68 and 12.4, attributed to H–11 in **226a** and the hydroxyl proton in **226b**,

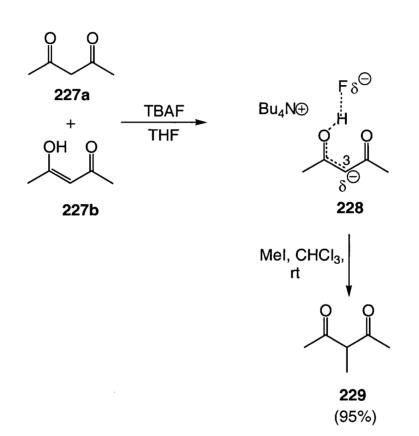
respectively. The ¹³C NMR resonances of the alkenic enol carbons in **226b** appeared at δ 170.6 (C–10) and 94.8 (C–11), respectively. The ester carbonyl carbons in **226a** and **226b** were displayed at δ 178.2and 173.5, respectively.

The next transformation, an iodomethylation reaction, was to be carried out on the mixture of tautomers, **226a** and **226b**, to give iodides **160** (eq 27).



A literature search revealed a protocol by Clark and Miller for the selective *C*-alkylation of β -dicarbonyl compounds through the intermediacy of tetrabutylammonium fluoride salt complexes (Scheme 42).⁸⁵ Treatment of a mixture of **227a** and **227b** with tetrabutylammonium fluoride (TBAF) in THF afforded the monosolvate **228**. On the basis of several NMR studies on **228**, the authors ruled out a proton transfer to the fluoride anion. They proposed that hydrogen-bonding between the fluoride anion and the

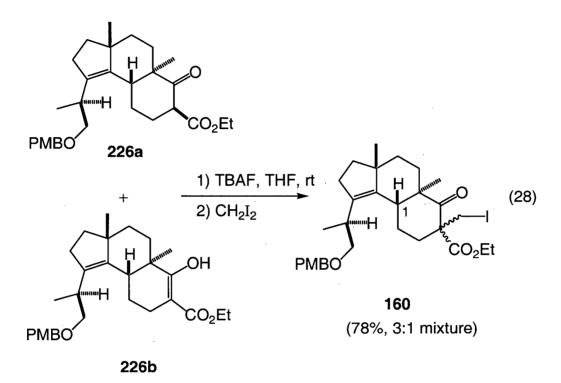
enolic hydroxyl proton results in a delocalization of the charge on the fluoride ion into the enol π -system. This delocalization appears to reduce the amount of enolic double bond character and increase the carbanion character at C-3. Treatment of **228** with iodomethane resulted in a highly *C*-selective alkylation, affording **229** in nearly quantitative yield.





It was hoped that the high degree of C-selectivity noted in the aforementioned alkylation of a TBAF monosolvate with iodomethane could be reproduced in our study with diiodomethane as the electrophile. Thus, a solution of a 1:1 mixture of enol ester and keto ester tautomers **226** in THF was treated with a solution TBAF in THF. The

resultant solution of the corresponding monosolvate was treated with ten equivalents of diiodomethane, providing a 3:1 mixture of the epimeric iodides **160** after work-up (eq 28).

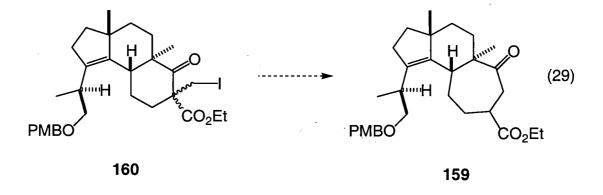


The structures of the epimeric iodides **160** were confirmed by spectroscopic methods. The IR spectrum of the mixture of iodides **160** exhibited the ketone and ester carbonyl stretching bands at 1708 and 1735 cm⁻¹, respectively. The ¹H NMR spectrum of the mixture of iodides **160** displayed overlapping multiplets (2H) between δ 4.36-4.45 attributed to the protons of the iodomethyl moieties. The assigned 3:1 ratio of the **160** isomers was supported by a 3:1 integral ratio of two ¹H NMR signals observed at δ 2.67 and δ 2.77, representing the angular ring-fusion protons (H–1) of the major and minor isomers, respectively. Since the mixture of isomers **160** was to be employed in the ensuing ring-expansion reaction, the respective relative configurations of the major and

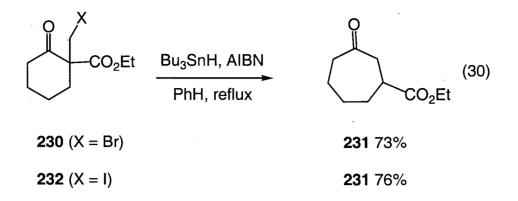
minor isomers were not determined. The ¹³C NMR spectrum of **160** displayed resonances at δ 169.3 and 210, assigned to the major isomer's ester and ketone carbonyl carbons, respectively. The ¹³C NMR resonances for the ester and ketone carbonyl carbons of the minor isomer were observed at δ 168.4 and 208.6, respectively.

2.1.3.5. Preparation of Keto Ester 159

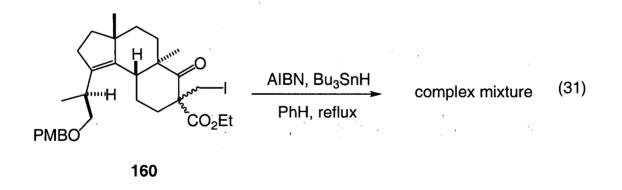
The next step in the synthesis of sarcodonin G involved a ring expansion of the α -halomethyl β -keto esters **160** to form the homologated γ -keto ester **159** (eq 29).



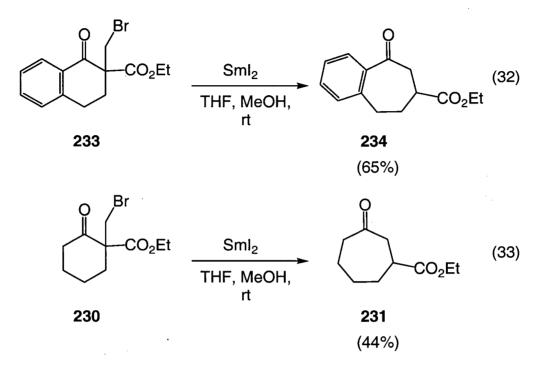
Concurrent studies by the research group of Dowd and that of Beckwith resulted in the discovery of a novel radical-mediated ring expansion of α -halomethyl β -keto esters. Dowd and coworkers reported studies on the Bu₃SnH-mediated ring expansion of α -bromomethyl β -keto esters to the corresponding γ -ketoester.^{86,87} As an example, treatment of bromide **230** with Bu₃SnH and a catalytic amount of AIBN in refluxing benzene afforded γ -keto ester **231** in a good yield (eq 30). Beckwith and coworkers reported an analogous ring expansion of iodide **232**, affording γ -keto ester **231**, also in good yield (eq 30).⁸⁸



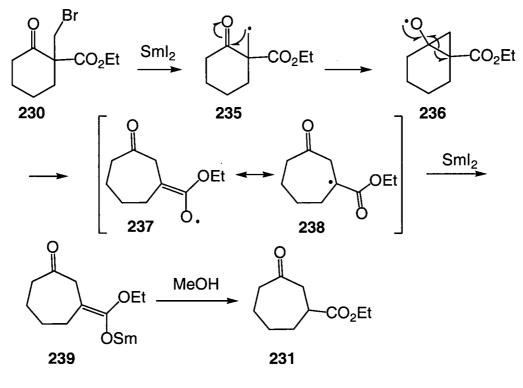
To our disappointment, attempts to effect Bu_3SnH -mediated ring expansion reactions of iodides **160**, following Dowd's procedure,⁸⁷ resulted in complex mixtures of unidentified products. (eq 31). Despite variations in the concentration of the iodides and addition time of the reagents, the desired ring-expansion product was not detected.



Recently, Hasegawa and coworkers reported an analogous SmI₂-mediated ringexpansion reaction to produce homologated γ -keto esters.⁸⁹ For example, treatment of aromatic α -bromomethyl β -keto ester 233 with 2 equiv of SmI₂ provided the ringexpansion product 234 (eq 32). However, addition of SmI₂ to a solution of the aliphatic α-bromomethyl β-keto ester **230**, a substrate used in Dowd's study (*vide supra*),⁸⁷ provided the ring-expansion product **231** in relatively modest yield (eq 33).

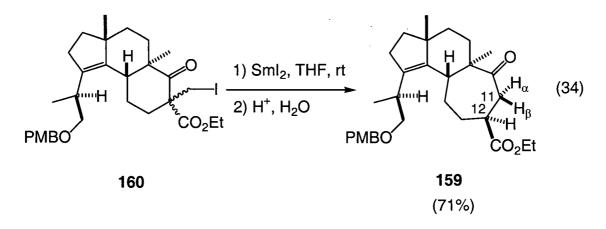


The proposed mechanism of the SmI₂-mediated ring expansion of α -bromomethyl β -keto ester 230, which is similar to that proposed for the analogous Bu₃SnH-mediated reactions, is depicted in Scheme 43.⁸⁹ The first step involves single electron transfer from SmI₂ to 230, providing the carbon radical 235. Attack of the resultant primary radical on to the ketone carbonyl function yields a cyclopropyloxy radical intermediate 236. Fragmentation of 236, which involves cleavage of the ring fusion bond, provides the seven-membered carbocycle 237. The resonance-stabilized radical 237 undergoes a one-electron transfer from a second equivalent of SmI₂, to yield the samarium enolate 239. In the presence of methanol or water, the samarium enolate is protonated to generate γ -keto ester 231.

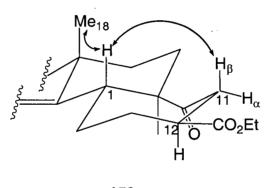


Scheme 43

Hasegawa's SmI₂-mediated ring-expansion protocol⁸⁹ was applied to the mixture of iodides **160**. A solution of the 3:1 mixture of epimeric iodides **160** in THF was treated with SmI₂ (eq 34). Aqueous acidic work-up and purification of the resultant material by flash chromatography afforded the ring-expansion product, γ -keto ester **159**, as a single isomer in 71% yield.



Evidence for the successful ring expansion of **160** was obtained by analysis of the ¹H and ¹³C NMR spectra derived of the γ -keto ester **159**. The ¹H NMR spectrum of **159** displayed a doublet of doublets (1H, J = 9.5 and 14 Hz) at δ 3.05 attributed to H–11_β. The ¹H signals for H–11_α and H–12 were attributed to overlapping signals observed between δ 2.77-2.84. The ¹³C NMR spectrum of **159** exhibited resonances at δ 39.3 and 38.7, assigned to C–11 and C–12, respectively. HMQC correlations were noted between the H–11 and H–12 signals and the corresponding C–11 and C–12 signals. The ¹³C NMR spectrum of **159** also displayed resonances at δ 174.5 and 215.0, representing the ester and ketone carbonyl carbons, respectively.



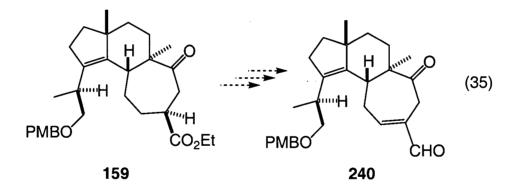
159

The proposed β -orientation of the ethoxycarbonyl moiety in **159** was supported by ¹H NMR NOED experiments on **159**. Irradiation at δ 2.65, a multiplet (1H) attributed the angular proton H–1, caused enhancement of a singlet at δ 1.06 assigned to H–18 and the doublet of doublets at δ 3.05 (J = 9.5, 14.4 Hz) assigned to H–11 $_{\beta}$. The larger (14.5 Hz) of the coupling constants for the doublet of doublets at δ 3.05 was attributed to the geminal coupling between H–11 $_{\beta}$ and H–11 $_{\alpha}$ whereas the smaller coupling (9.5 Hz) was

attributed to the coupling between H–11_{β} and H–12. The magnitude of the latter coupling suggests that H–11_{β} and H–12 have a trans diaxial-type relationship, which supports the proposed equatorial orientation of the ethoxycarbonyl group in **159**. Although the configuration at C–12 in **159** was clearly established, the reasons for the stereoselective α -face protonation, exclusively affording intermediate **159**, were not evident to us.

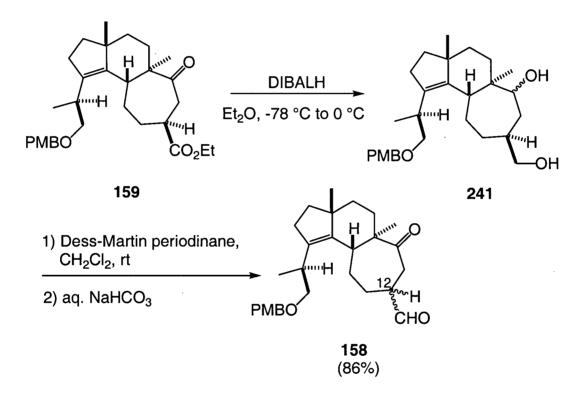
2.1.3.6. Preparation of Keto Aldehyde 240

With the ring-expansion step completed, the transformation of keto ester **159** into keto aldehyde **240** was investigated (eq 35).



The first step of this transformation was envisioned to be conversion of keto ester **159** to the corresponding keto aldehyde **158** (Scheme 44). Following a standard procedure, a solution of keto ester **159** in Et_2O was treated with several equivalents of DIBALH. A suitable work-up procedure provided the mixture of diols **241**. The crude

diols 241 were taken up in CH_2Cl_2 and treated with Dess-Martin's periodinane reagent.⁹⁰ After standard work-up and purification of the resultant material by flash chromatography, a 1:1 mixture of epimeric keto aldehydes 158 was obtained in 86% yield. Since the ratio of epimers varied with the duration of the work-up procedure, the C–12 epimerization of the highly enolizable aldehydes 158 presumably occurred during the aqueous NaHCO₃ wash.

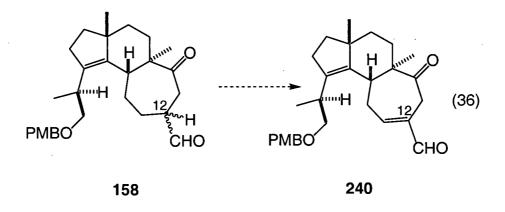


Scheme 44

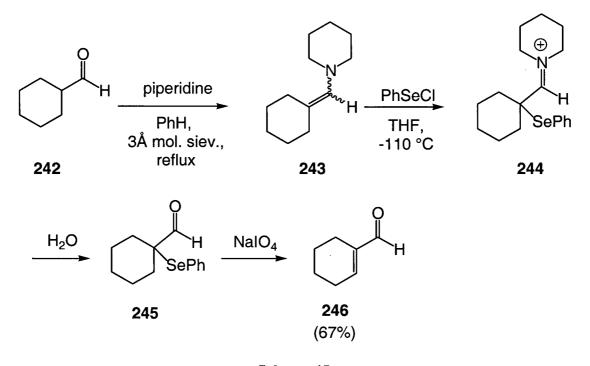
Evidence for the structure, and a measure of the ratio of the epimeric keto aldehydes **158**, was obtained by standard spectroscopic methods. The IR spectrum of the mixture of keto aldehydes **158** exhibited ketone and aldehyde carbonyl stretching bands at 1726 and 1697 cm⁻¹, respectively. The ¹H NMR spectrum of **158** displayed a pair of doublets at δ 9.60 and 9.63, attributed to the aldehydic proton signals. A 1:1 integral

ratio was observed for these aldehydic proton signals, indicating a 1:1 mixture of aldehydes. The ¹³C NMR spectrum displayed resonances at δ 201.0 and 201.5, representing the aldehydic carbonyl carbons. The ¹³C NMR resonances of the ketonic carbonyl carbons appeared at δ 214.9 and 215.8.

The next step in the synthesis of sarcodonin G involved the introduction of a double bond in the seven-membered ring at C-12 in 158 to give 240 (eq 36).



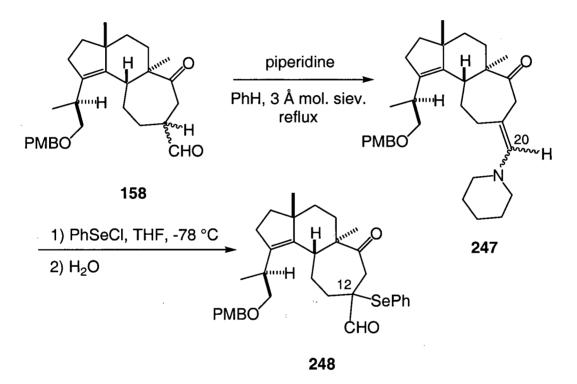
Williams and coworkers have reported a mild method of preparing α,β unsaturated aldehydes from the corresponding aldehydes *via* enamine derivatives.⁹¹ For example, conversion of aldehyde 242 to the corresponding enamine 243 was accomplished by heating a mixture of 242 and piperidine in benzene with removal of water with 3Å molecular sieves (Scheme 45). Treatment of the enamine derivative 243 with phenylselenenyl chloride provided the intermediate 244, which afforded the corresponding α -phenylselenoaldehyde 245 after aqueous work-up. A periodate oxidation of 245 generated the corresponding selenoxide intermediate, which underwent a *syn*-elimination reaction to give the desired α,β -unsaturated aldehyde 246.



Scheme 45

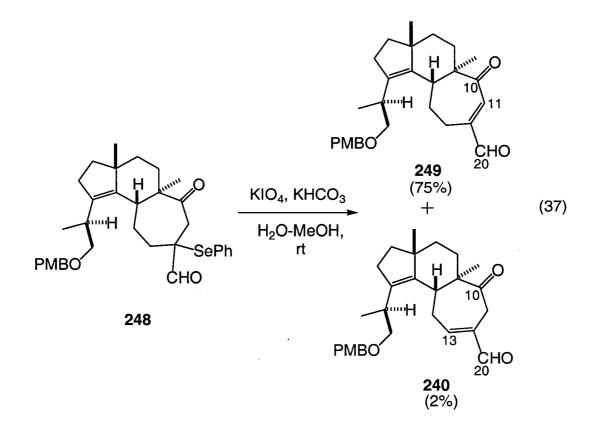
It was hoped that William's mild oxidation protocol (*vide supra*) could be successfully applied to the mixture of aldehydes **158**. On the basis of previous studies by Clinton and coworkers,⁹² the enamine formation step was expected to be chemoselective for the aldehyde function over that involving the ketone function in **158**. Thus, aldehydes **158** were converted into the corresponding piperidine enamines **247** by treatment of the former with 5 equivalents of piperidine in refluxing benzene over 3Å molecular sieves (Scheme 46). After removal of the solvent and excess piperidine, the resultant crude material was analyzed by ¹H NMR spectroscopy, which confirmed the successful formation of enamines **247**. The ¹H NMR spectrum of **247** displayed two singlets at δ 5.32 and 5.43, representing the olefinic enamine protons (H–20). Overlapping multiplets displayed between δ 1.4–1.6 and between δ 2.5-2.7 were attributed to the methylene protons of the piperidine rings. The crude enamines **247** were taken up in

THF and the solution was treated with freshly recrystallized phenylselenenyl chloride. After aqueous work-up, the resultant solution was concentrated and the α -phenylselenoaldehyde **248** was analyzed by ¹H NMR spectroscopy. The ¹H NMR spectrum of crude **248** displayed an aldehydic proton signal at δ 9.19. Several multiplets between δ 7.2 and 7.7 were also observed, attributed to the aromatic protons of a phenylselenide moiety. The relative configuration at C–12 was not determined.



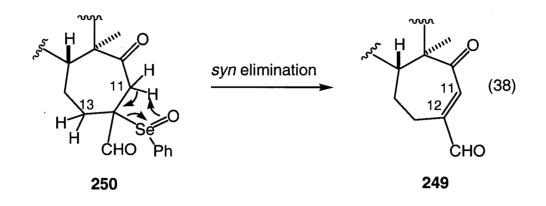


The crude α -phenylselenoaldehyde **248** was taken up MeOH and the solution was treated with solid KIO₄ and aqueous KHCO₃ (eq 37). Aqueous work-up and purification of the resultant material by flash chromatography afforded two isomeric α , β -unsaturated aldehydes, **249** and **240**, in 75% and 2 % yields, respectively

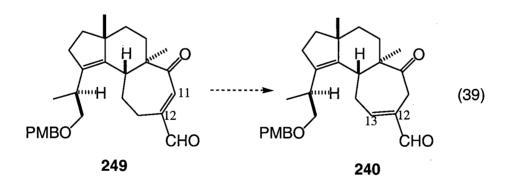


The structure of **249** was confirmed by spectrometric analysis. The IR spectrum of **249** exhibited a broad band at 1697 cm⁻¹, representing the carbonyl stretches of the conjugated ketone and aldehyde functions. The ¹H NMR spectrum of **249** displayed a 1proton singlet at δ 9.56 (1H), attributed to the aldehydic H–20 proton. The ¹H NMR signal for the olefinic C–11 proton in **249** was observed a singlet at δ 6.63 (1H). The lack of H-H coupling displayed by this olefinic signal supported the proposed C–11 position of the double bond in **249**. The ¹³C NMR resonances of the two alkene functions in **249** appeared at δ 136.9, 139.6, 144.2 and 146.1. The ¹³C NMR resonances of the aldehydic (C–20) and ketonic (C–10) carbonyl carbons appeared at δ 194.7 and 209.6, respectively The structure of the minor product, aldehyde **240**, was also confirmed by spectrometric analysis. The ketone and aldehyde functions in **240** were represented by carbonyl stretches in IR spectrum at 1704 and 1688 cm⁻¹, respectively. The ketone stretch at 1704 cm⁻¹ is characteristic of an aliphatic ketone, supporting the C–12 position of the double bond in **240**. The ¹H NMR signals for H–13 and H–20 for **240** were observed as a multiplet at δ 6.63–6.68 and a singlet at δ 9.33, respectively. The multiplicity of the olefinic H–13 signal, attributed to coupling with the adjacent H–14 protons, supports the C–12 positioning of the double bond. The ¹H NMR signals for the geminal H–11 protons were observed as a pair of doublets (J = 14 Hz) at δ 3.46 and 3.72. The ¹³C NMR resonances at δ 192.3 and 210.6 were attributed to the aldehydic and ketonic carbonyl carbons, respectively.

The ratio of the two products **249** and **240** derived from the selenoxide elimination reaction can be rationalized by comparison of the relative acidities of the H–11 and H–13 protons of selenoxide **250** (eq 38). The H–11 protons in **250**, which are α to a carbonyl group, are much more acidic than the H–13 protons since the former experience favorable overlap of the C–H sigma bonds with the π -bonds of the adjacent carbonyl function. Thus, the irreversible *syn*-elimination of the selenoxide moiety should favor the removal of the more acidic proton, H–11, affording primarily α , β -unsaturated ketone **249**.



Since the C-ring alkene function in the target structure, sarcodonin G, is positioned at C-12, the next task at hand involved an isomerization of **249**, which displays a C-11 double bond, to afford **240**, which displays the requisite C-12 double bond (eq 39).



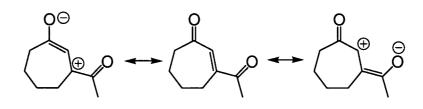
In 1984, Mease and Hirsch reported a study on the syntheses and base-catalyzed isomerizations of medium-ring cycloalkenones.⁹³ The effect of C–3 substituents on the base-catalyzed equilibration ratio of carbocycles **251** and **252** was investigated (**Table 2**). In the unsubstituted case, the product composition of the thermodynamically controlled isomerization of **251** or **252** significantly favored the 2-cycloheptenone isomer **251** (entry

1, **Table 2**). On the other hand, the isomerization of compounds with electronwithdrawing substituents at C-3 favored the 3-cycloheptenone isomer **252** (entries 2-4, **Table 2**). The authors' explanation for the substituent effect was based on the premise that the 3-cycloheptenone isomers **252** were more effectively conjugated with less conformational demands on the ring system than the corresponding 2-cycloheptenone isomers **251**.

$\begin{array}{c} \begin{array}{c} & \\ \end{array} \\ \end{array} \\ \end{array} \\ \begin{array}{c} \\ \end{array} \\ \begin{array}{c} \\ \end{array} \\ \end{array} \\ \begin{array}{c} \\ \end{array} \\ \begin{array}{c} \\ \end{array} \\ \end{array} \\ \begin{array}{c} \\ \end{array} \\ \begin{array}{c} \\ \end{array} \\ \end{array} \\ \begin{array}{c} \\ \end{array} \\ \begin{array}{c} \\ \end{array} \\ \end{array} \\ \begin{array}{c} \\ \end{array} \\ \begin{array}{c} \\ \end{array} \\ \end{array} \\ \begin{array}{c} \\ \end{array} \\ \end{array} \\ \begin{array}{c} \\ \end{array} \\ \begin{array}{c} \\ \end{array} \\ \end{array} \\ \begin{array}{c} \\ \end{array} \\ \begin{array}{c} \\ \end{array} \\ \end{array} \\ \end{array} \\ \begin{array}{c} \\ \end{array} \\ \end{array} \\ \end{array} \\ \begin{array}{c} \\ \end{array} \\ \end{array} \\ \end{array} \\ \end{array} \\ \end{array} \\ \end{array} \\ \begin{array}{c} \\ \end{array} \\ $				
Entry	Substituent (X)	Composition % of 251	Composition % of 252	
1	Н	76.8	23.2	
.2	CO ₂ CH ₃	16.8	83.2	
3	CN	15.1	84.9	
4	COCH ₃	20.7	79.3	

 Table 2. Isomerization of C-3 Substituted Cycloheptenones 251 and 252.

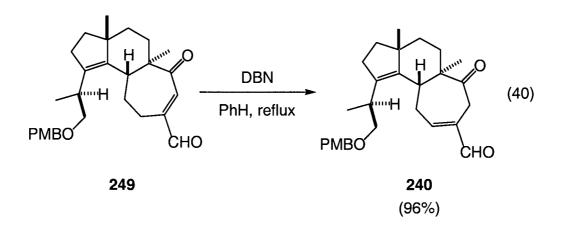
The authors also suggested that the double bonds in the 2-cycloheptenone isomers **251** with electron-withdrawing substituents experience destabilization due to similar electronic interactions at both ends of the double bond (entries 2-4, **Table 2**). As an example, the contributing resonances structures available to 2-cycloheptenone **251** with an electron-withdrawing acyl substituent (entry 4, **Table 2**) place positives charges at opposite ends of the double bond (Scheme 47).



251 (entry 4, Table 2)

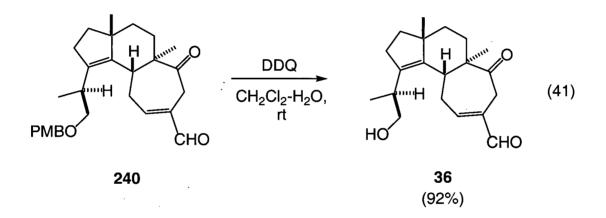
Scheme 47

On the basis of findings by Mease and Hirsch (*vide supra*), it was expected that **249** could be isomerized under thermodynamically controlled equilibrating conditions to **240**. Thus, a benzene solution of **249** containing 2 equivalent of DBN was refluxed for 12 h (eq 40). After removal of solvent and purification of the resultant material by flash chromatography, the desired isomer **240** was isolated in nearly quantitative yield (96%).



2.1.3.7. Preparation of (±)-Sarcodonin G (36)

The preparation of (\pm)-sarcodonin G (**36**) from **240** involved removal of the PMB protecting group (eq 41). Following optimized conditions reported by Oikawa,⁷⁵ a solution of keto aldehyde **240** in aqueous CH₂Cl₂ was treated with DDQ. After a standard work-up procedure and purification of the resultant material by flash chromatography, (\pm)-sarcodonin G (**36**) was isolated in 92% yield.⁹⁴



The synthetic (\pm)-sarcodonin G (**36**) exhibited spectral data in full accordance with those reported¹⁶ for the isolated natural product, (–)-sarcodonin G.⁹⁵ The IR, MS, ¹H NMR and ¹³C NMR spectra of the synthetic material and isolated material are compared in **Tables 3**, **4** and **5**. The ¹H NMR spectrum derived from synthetic (\pm)-sarcodonin G (**36**) is displayed in Fig. 1.

					(±)-Sarcodonin	G (36)
with those	e Reported ¹⁶	for Natural	1 (-)-Sarcodor	nin G (36)		

Data	Synthetic (±)-Sarcodonin G (36)	Natural (–)-Sarcodonin G (36)	
	3445	3400	
	1703	1705	
$\operatorname{IR}^{\mathrm{a}}$ (cm ⁻¹)	1640	1640	
()	1450	1450	
	1376	1380	
MS ^b (m/z)	316.2038 (DCI+)	316 (EIMS)	

.

^a The IR spectra were recorded using KBr pellets. ^b Exact mass calcd for C₂₀H₂₈O: 316.2039.

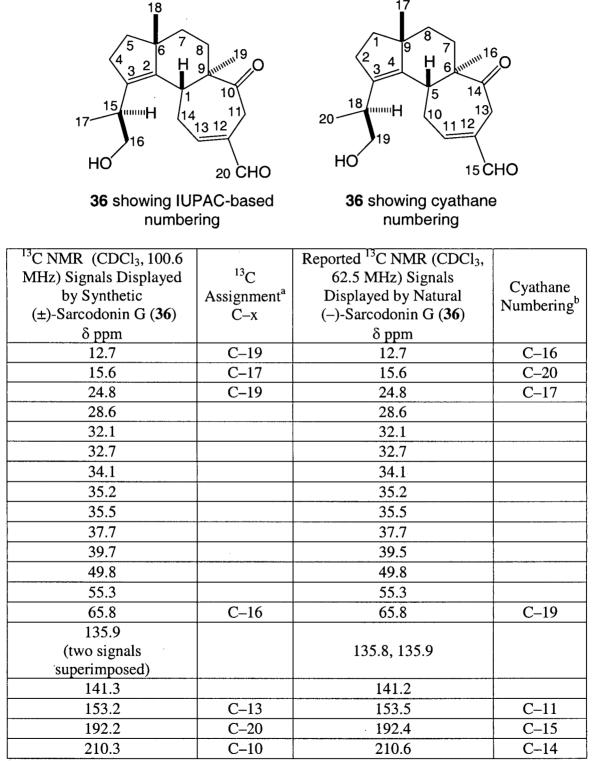
.

-	101 Matural ()-	5arcouolilli ((50) (CDC13, 25	U IVITIZJ.
	8 9 10 11 13 12	$\begin{array}{c} 1 \\ 1 \\ 2 \\ 3 \\ 4 \\ 4 \\ 1 \\ 1 \\ 1 \\ 1 \\ 1 \\ 1 \\ 1 \\ 1$)
ne	20 ĈHO	150	СНО
36 showing IUPAC-based numbering		36 showing cyathane numbering	
¹ H NMR (CDCl ₃ , 400 MHz) Signals Displayed ^a by Synthetic (±)-Sarcodonin G (36) δ ppm (Multiplicity, J (Hz))	¹ H Assignment ^b H–x	Reported ¹ H NMR (CDCl ₃ , 250 MHz) Signals Displayed by Natural (–)-Sarcodonin G (36) δ ppm (Multiplicity, J (Hz))	Cyathane Numbering ^c
0.97 (d, 6.9)	H–17	0.96 (d, 7.3)	H–20
1.03 (s)	H–19	1.01 (s)	H–16
1.14 (s)	H–18	1.12 (s)	H–17
1.24–1.30 (m)		1.24 (dt, 3.9, 13.2)	
1.46 (s)		part of m at 1.59	-
1.49–1.55 (m)		part of m at 1.59	
1.58–1.70 (m)		part of m at 1.59	
1.95 (dt, 5, 13.4)		1.94 (dt, 5.3, 13.3)	
2.21–2.40 (m)		2.31 (m)	
2.69–2.80 (m)		2.74 (m)	
2.98–3.08 (m)	H–15	3.02 (sextet, 7.3)	H–18
3.10–3.20 (m)		3.16 (dd, 13.4, 6.4)	
3.36 (br d, 12.6)		3.34 (d, 12)	
3.41–3.51 (m)		3.42 (d, 13.9), 3.45 (d, 7.3)	
3.71–3.75 (m)		3.72 (br d, 13.6)	
6.70–6.73 (m)	H–13	6.71 (m)	H–11
9.35 (s)	H–20	9.31(s)	H–15
^a The difference between o	beerved and re	ported & is likely due to d	ifferent CDCl.

Table 4. Comparison of ¹H NMR Data for Synthetic (\pm)-Sarcodonin G (**36**) (CDCl₃, 400 MHz) with those Reported¹⁶ for Natural (–)-Sarcodonin G (**36**) (CDCl₃, 250 MHz).

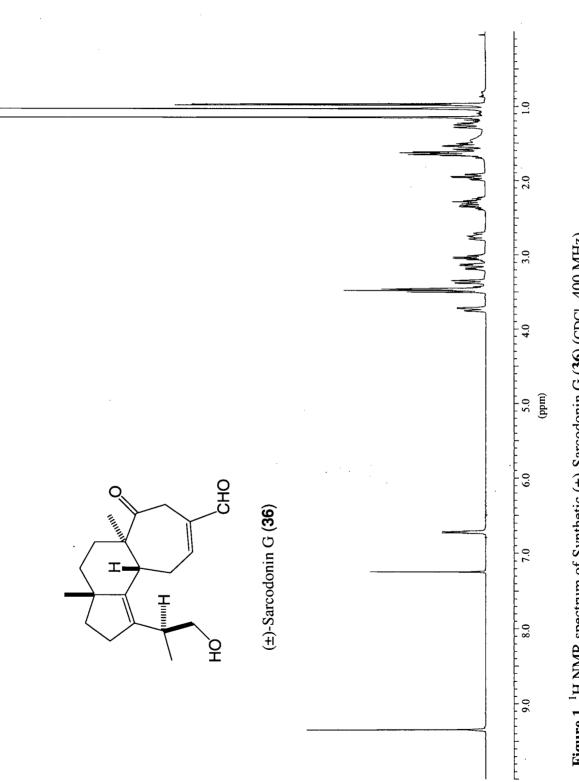
^a The difference between observed and reported δ is likely due to different CDCl₃ reference assignments. ^b Systematic IUPAC-based numbering system. ^c Ayer's cyathane numbering system.²³

Table 5. Comparison of ¹³C NMR Data for Synthetic (±)-Sarcodonin G (**36**) (CDCl₃, 100.6 MHz) with those Reported¹⁶ for Natural (–)-Sarcodonin G (**36**) (CDCl₃, 62.5 MHz).



^a Systematic IUPAC-based numbering system.

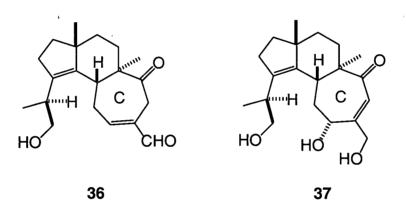
^b Ayer's cyathane numbering system.²³





2.1.4. Studies Towards the Synthesis of (±)-Cyathin A4

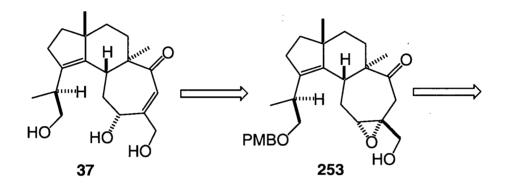
After the completion of total synthesis of (\pm) -sarcodonin G (**36**), the synthesis of the structurally related cyathane diterpenoid (\pm) -cyathin A₄ (**37**)¹⁷ was attempted. Our synthetic approach to sarcodonin G (**36**), which addressed the construction of oxygenation and unsaturation pattern of the C-ring in the later stages of the route, was expected to provide an advanced branching point from which cyathin A₄ (**37**) could be derived.

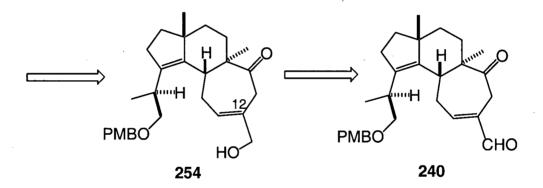


2.1.4.1. Retrosynthetic Analysis

Our proposed synthetic plan for the construction of cyathin A₄ (**37**) is outlined in Scheme 48. The target structure **37** should be available from compound **253** via ring opening of the epoxide function to give the corresponding allylic alcohol and cleavage of the PMB ether to generate the hydroxyl function. A chemo- and stereoselective epoxidation of C–12 double bond in **254** should provide epoxide **253**. Finally, compound

should be available *via* a chemoselective reduction of aldehyde **240**, an advanced intermediate available from our previously described total synthesis of sarcodonin G (see Section 2.1.3. *vide supra*).

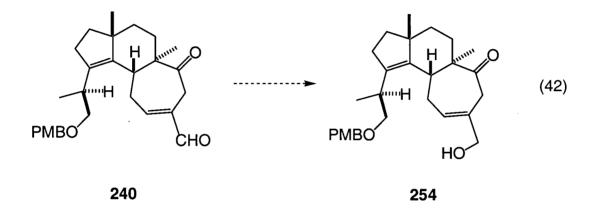




Scheme 48

2.1.4.2. Preparation of Keto Alcohol 254

The first step in our proposed synthesis of cyathin A_4 involved a chemoselective reductive transformation of keto aldehyde **240** to give keto alcohol **254** (eq 42). This chemoselective reduction proved difficult to achieve owing to a small difference in reactivity between the aldehyde and ketone carbonyl functions (*vide infra*).



Both steric and stereoelectronic effects are important factors that may contribute to the relative reactivity of saturated and unsaturated carbonyl functions towards a hydride source (reducing agent).⁹⁶ For steric reasons, carbonyls with one alkyl substituent (aldehydes) should be more readily reduced than those with two alkyl substituents (ketones). The steric bulk of a ketone's two alkyl substituents significantly restricts the approach of the nucleophile to the carbonyl and increases the energy of the transition state leading from a trigonal intermediate to the tetrahedral product. For stereoelectronic reasons, the carbonyl function of an α,β -unsaturated aldehyde or ketone function is significantly less electrophilic than that of the corresponding substance lacking the alkene function. The decreased electrophilicity of the former may be partially attributed to a decrease in carbonyl double-bond character as illustrated by contributing resonance structures **255** (Scheme 49).

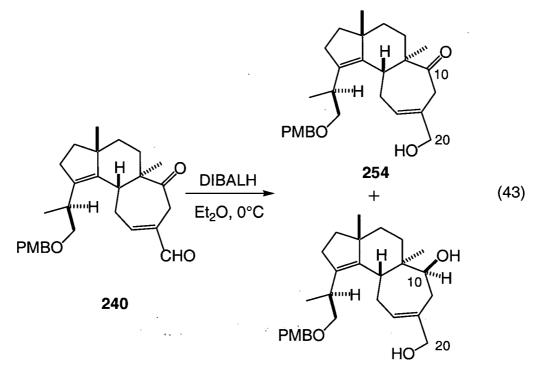


255

Scheme 49

On balance, the relative transition state energies involved in the reduction of the ketone function and unsaturated aldehyde function of compound **240** might be expected to be energetically similar, since the former is disfavored for steric reasons whereas the later is disfavored for stereoelectronic reasons.

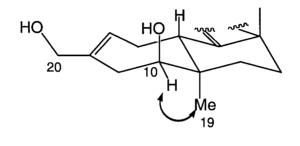
Diisobutylaluminum hydride (DIBALH) was the first reducing agent investigated for the chemoselective reduction of **240**. Treatment of a cold (-78 °C) Et₂O solution of keto aldehyde **240** with one equivalent of DIBALH simply resulted in the complete recovery of starting material. Consequently, the DIBALH reduction of **240** was repeated in Et₂O at 0 °C, which provided a 46% yield of keto alcohol **254** and diol **256** in a 1:4 ratio, along with 40% recovered starting material (eq 43). The three compounds were easily separated by silica gel flash chromatography.





(46%, 1:4 ratio of **254**:**256**)

Evidence for the successful chemoselective reduction of keto aldehyde **240** to provide keto alcohol **254** was obtained by spectroscopic methods. The IR spectrum of **254** exhibited a broad signal at 3435 cm⁻¹, attributed to the OH stretching vibration. An absorption band observed at 1701 cm⁻¹ in the IR spectrum of **254** was characteristic of a saturated ketone. The ¹H NMR spectrum of **254** displayed a multiplet (2H) between δ 3.95–4.08 assigned to the H–20 methylene protons. The ¹³C NMR spectrum of **254** displayed resonances at δ 68.8 and 211.3, attributed to the C–20 methylene and the C–10 ketone carbons, respectively. The structure of diol product **256** was also confirmed by spectroscopic analysis. The IR spectrum of **256** displayed a broad absorption at 3368 cm⁻¹, representing the OH stretch. The ¹H NMR signal for H–10 proton in **256** was observed at δ 3.42. A multiplet between δ 3.92–3.99 in the ¹H NMR spectrum was assigned to the H–20 protons. The ¹³C NMR resonance for the C–10 carbon in **256** was displayed at δ 75.3.

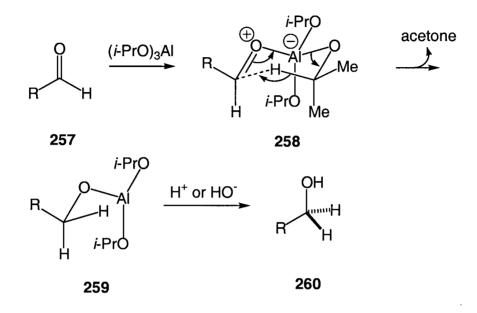


256

The relative configuration of the C-10 hydroxyl moiety was confirmed by ¹H NMR NOED experiments on **256**. Irradiation of the H-10 signal, displayed at δ 3.42 in the ¹H NMR of **256**, caused enhancement of a singlet at δ 0.89, attributed to the H-19 methyl protons. A reciprocal enhancement of the H-10 signal was observed upon irradiation of the H-19 signal. These results support the β -configuration of the C-10 hydroxyl moiety in **256**.

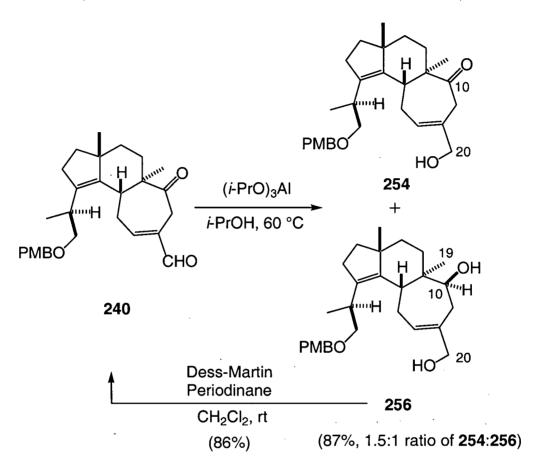
On the basis of the low chemoselectivity displayed by the DIBALH reduction of **240**, an alternative reduction method was investigated. The Meerwein–Ponndorf–Verley (MPV) reduction with aluminum isopropoxide has been shown to reduce unsaturated aldehydes to the corresponding allylic alcohols.⁹⁷ The proposed mechanistic pathway of the aluminum isopropoxide MPV reduction is outlined in Scheme 50.⁹⁸ Treatment of an aldehyde (**257**) with aluminum isopropoxide generates the cyclic coordination complex

258, which undergoes intramolecular hydride transfer to afford the mixed alkoxide **259** and one equivalent of acetone. Hydrolysis of the mixed alkoxide **259** under acidic or basic conditions affords the corresponding alcohol **260**.



Scheme 50

It was hoped that an aluminum isopropoxide-mediated MPV reduction of keto aldehyde **240** would be chemoselective for the aldehyde function. A solution of the keto aldehyde **240** in a 1:1 mixture of benzene and *i*-proponal was treated with one equivalent of aluminum isopropoxide and the resultant mixture was heated to reflux. Work-up and purification of the resultant material by flash chromatography provided a 1:2 ratio of keto alcohol **254** and diol **256** as well as 40% recovered starting material. Gratifyingly, an MPV reduction of **240** performed in 100% *i*-propanol at 60 °C afforded an 87% yield of **254** and **256** with an improved ratio of 1.5:1 in favor of the desired keto alcohol **254** (Scheme 51). The undesired diol **256** was subsequently oxidized with Dess–Martin periodinane reagent⁹⁰ in 86% yield and the resultant keto aldehyde **240** recycled through the MPV reduction. After one cycle, the desired alcohol **254** was obtained in 70% yield from keto aldehyde **240**.

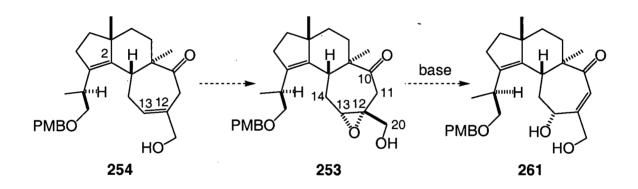


Scheme 51

2.1.4.3. Attempted Preparation of Diol 261

With the keto alcohol **254** in hand, the preparation of diol **261** *via* the epoxide intermediate **253** was investigated (Scheme 52). The first transformation would require an epoxidation method that was (a) chemoselective for the C–12 double bond over the C–

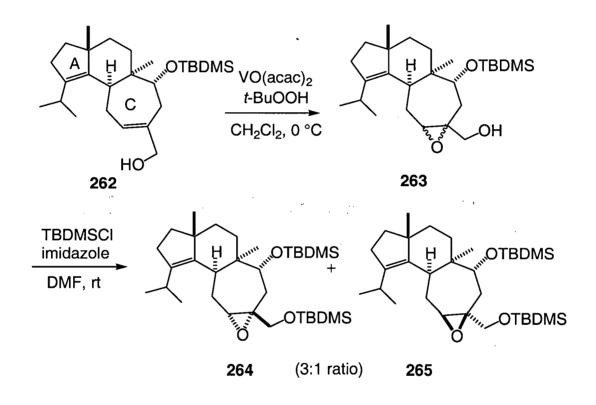
2 double bond in **254**, and (b) stereoselective for the α -face of the C–12 double bond. It was anticipated that the hydroxymethyl appendage at C–12, through coordination with a suitable reagent, might help chemoselectively direct the C–12 alkene epoxidation. Based on molecular models, an approach to the α -face of the C–12 alkene appeared slightly less sterically hindered. Thus, it was hoped that an epoxidation of **254** would favour formation of the α -epoxide **253**. The next step would involve a ring opening of the epoxide function *via* a base-mediated deprotonation α to the C–10 ketone function, with subsequent β -elimination of the epoxide oxygen anion. Work-up should then afford diol **261**.



Scheme 52

An epoxidation similar to that required for our synthesis (*vide supra*) was previously noted by Snider and co-workers in their synthetic studies directed towards the diterpenoid allocyathin B_2 (see Section 2.1.2.1., *vide supra*).⁵⁶ Their account of several abandoned synthetic routes included the selective epoxidation of intermediate **262** (Scheme 53). Treatment of **262** with *t*-BuOOH and a catalytic amount of VO(acac)₂

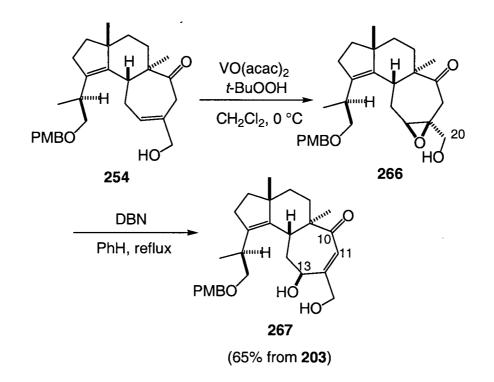
resulted in a hydroxyl-directed epoxidation of the C-ring double bond, providing a mixture of epoxides 263. This material, which was too acid-sensitive for purification by silica gel chromatography, was treated with TBDMSCl to afford the corresponding silyl ether epoxides 264 and 265 in a 3:1 ratio. Evidently, the epoxidation step (262 to 263) was threefold stereoselective for the α -face of the C-ring double bond in 262.



Scheme 53

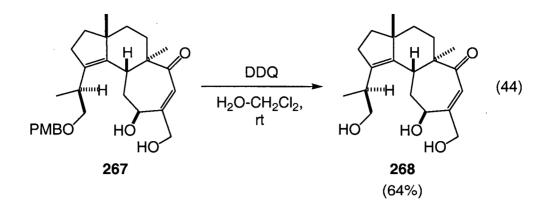
It was hoped that a VO(acac)₂/*t*-BuOOH hydroxyl-directed epoxidation of our intermediate **254** would display both the chemo- (ring C double bond vs ring A double bond) and stereoselectivity (α - vs β -face) reported by Snider and coworkers (*vide supra*). Thus, a cool (0 °C) solution of alkene **254** and *t*-BuOOH in CH₂Cl₂ was treated with a catalytic amount of VO(acac)₂ (Scheme 54).⁹⁹ The resultant mixture was stirred for 2 h at

0 °C. Work-up, followed by attempted purification of a portion of the resultant material by flash chromatography on silica gel, led to product decomposition. ¹H NMR spectral analysis of the remaining crude material revealed the presence of only one major product, which was subsequently shown to possess structure 264, the product of a α -face epoxidation of 254. Successful reaction of the C-12 alkene in 264 was supported by of the ¹H NMR spectroscopic analysis of the product **266**. The ¹H NMR spectrum derived from the latter did not display signals in the olefinic region (δ 5-7). Further evidence supporting the epoxidation was obtained upon analysis of the H–20 methylene signal, observed as a multiplet between $\delta 4.8$ -4.9 in the ¹H NMR spectrum of **266**. The corresponding signal for the H-20 protons in the ¹H NMR spectrum of the starting material 254 had been noted at δ 3.9–4.1. The downfield shift of the signal for the H–20 protons in 266, relative to that for the H–20 protons in 254, was attributed to the newly introduced epoxide function in **266**. Evidence supporting the assigned β -configuration of the epoxide in 266 was later obtained through chemical transformations and spectroscopic correlations (vide infra). Without further purification, a benzene solution of crude 266 was treated with DBN, resulting in a ring opening of the epoxide function. Upon solvent removal under reduced pressure and purification of the resulting material by flash chromatography, the allylic alcohol **267** was obtained in 65% yield from **254**.

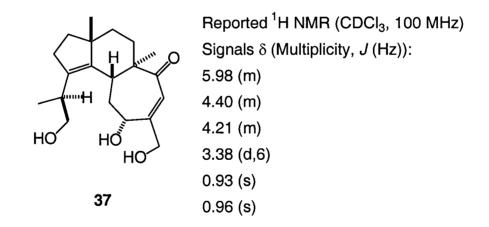


Scheme 54

The constitutional structure of **267** was supported by spectroscopic analysis. The IR spectrum of **267** exhibited a broad absorption at 3402 cm⁻¹ and a carbonyl stretch at 1665 cm⁻¹, attributed to the hydroxyl and ketone functions, respectively. The ¹H NMR spectrum of **267** displayed a singlet (1H) at δ 5.96, representing the H–11 olefinic proton. The ¹³C NMR resonance for the C–10 ketone carbon in **267** was observed at δ 210.1. ¹H NMR NOED experiments preformed on **267**, in hopes of confirming the relative configuration of the C–13 hydroxyl moiety, were not conclusive. Nevertheless, on the basis of spectroscopic analysis of a derivative of **267** (*vide infra*), the C–13 hydroxyl moiety was assigned a β -configuration. The PMB protecting group in **267** was removed by treatment of a solution of this material in aqueous CH₂Cl₂ with DDQ. Standard work-up and purification of the resultant material by flash chromatography provided triol **268**, the C–13 epimer of the target structure, cyathin A₄ (eq 44).



The successful removal of the PMB group in 267 was supported by ¹H NMR analysis of the product, triol 268. The lack of signals in the aromatic region of the ¹H NMR spectrum of 268 showed that the PMB group was no longer present. The signals in the ¹H NMR spectrum derived from triol 268 (Table 6) and those reported¹⁶ for its C–13 epimer, cyathin A₄ (37), were compared. The notable differences in the chemical shifts of corresponding protons in 268 and 37 were attributed to the C–13 epimeric relationship of the two substances.

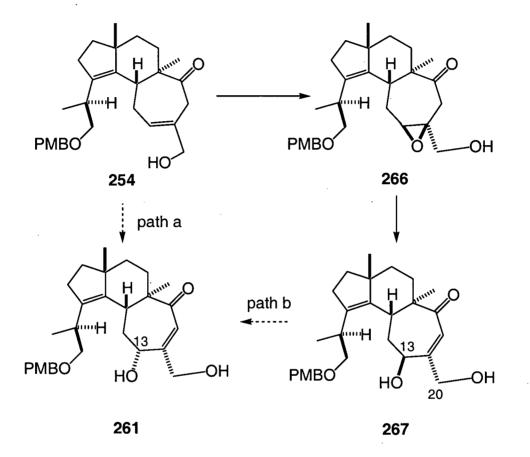


1000 CDC13, 400 WH2).			
¹ H assignment ^a H–x	¹ H NMR (CDCl ₃ , 400 MHz) Signals Displayed by Triol 268 δ (Multiplicity, <i>J</i> (Hz))		
H–17	0.96 (d, 6.7)		
H–19	1.04 (s)		
H–18	1.12 (s)		
· .	1.43–1.51 (m)		
	1.53–1.69 (m)		
	1.81 (dt, 4.9, 13.4)		
	1.97–2.07 (m)		
	2.19–2.37 (m)		
	2.63–2.70 (m)		
	3.07–3.15 (m)		
H–16	3.25–3.31 (m)		
	3.37–3.52 (m)		
H–20	4.24–4.42 (m)		
H–13	4.55-4.59 (m)		
H-11	6.00 (s)		

Table 6. ¹H NMR Data for Synthetic (±)-Triol 268 (CDCl₃, 400 MHz).

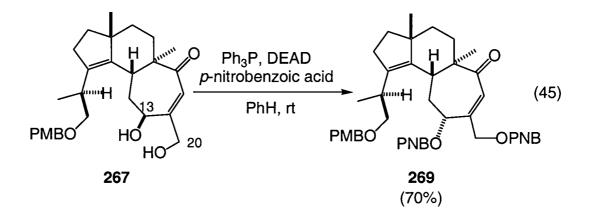
^a Systematic IUPAC-based numbering system.

Evidently, epoxidation of **254** proceeded with high β -face stereoselectivity to give epoxide **266**, which ultimately afforded **267** (Scheme 55). However, the main contributing factors responsible for this undesired selectivity are not fully understood. With only limited amounts of intermediate **254** available, the search for an α -face stereoselective epoxidation method to afford **261** from **254** (path a) *via* an α -epoxide was not pursued. As an alternative, the preparation of **261** was approached *via* a Mitsunobu¹⁰⁰ inversion of the C–13 hydroxyl group in **267** (path b). To minimize the steps required for this transformation, the primary hydroxyl function at C–20 was not protected from the Mitsunobu reaction.

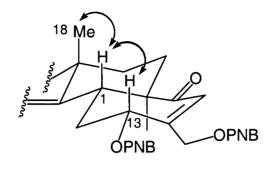


Scheme 55

Following optimized procedures developed by Martin and Dodge,¹⁰¹ a solution of the diol **267** in benzene was treated with Ph_3P and *p*-nitrobenzoic acid. Diethyl azodicarboxylate (DEAD) was slowly added and the mixture was stirred for 18 h at room temperature. The solvent was removed and the resulting material was purified by flash chromatography to give the desired diester **269** in 70% yield.

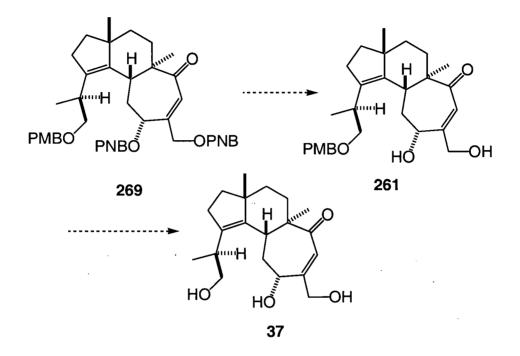


The assigned structure of diester **269** was supported by standard spectrometric analysis. The IR spectrum of **269** exhibited absorptions at 1729 and 1688 cm⁻¹, representing the ester and ketone carbonyl stretches, respectively. Sharp absorptions were also observed at 1530 and 1270 cm⁻¹, characteristic of aromatic nitro groups. The ¹H NMR spectrum of **269** displayed doublets at δ 8.06 (2H, J = 8.2 Hz) and δ 8.16 (2H, J = 8.6 Hz) and a multiplet (4H) between δ 8.20–8.29, attributed to the aromatic protons of the *para*-substituted nitrobenzoate esters. A multiplet (1H) observed between δ 5.70– 5.78 in the ¹H NMR spectrum of **269** was attributed to the allylic proton on C–13. Unfortunately, the C₄₄H₄₄N₂O₁₁ molecular formula of **269** could not be confirmed by mass spectroscopic analysis. Although the mass spectra of **269** were obtained by DCI+ with isobutene and NH₃ and by LSMIS with thioglycerol and CHCl₃ matrix, a parent ion representing a structure with C₄₄H₄₄N₂O₁₁ molecular formula was not detected. The inability to detect the requisite parent molecular ion for **269** was attributed to the labile nature of the *p*-nitrobenzoate ester moieties under ionization conditions.



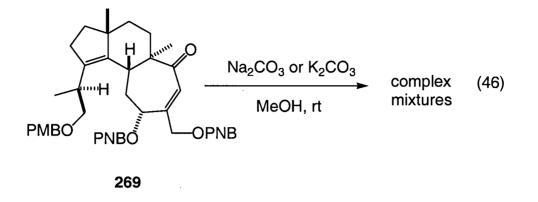
The assigned relative configuration of the *p*-nitrobenzoyl moiety at C-13 in **269** was supported by ¹H NMR NOE difference experiments. Irradiation of the signal at δ 2.95 in the ¹H NMR of **269**, a broad doublet attributed to the angular proton H-1, caused enhancement of a singlet at δ 1.08 and a multiplet between δ 5.70–5.78, assigned to H–18 and H–13, respectively. Irradiation of the H–18 and H–13 signals both caused enhancement of the H–1 signal. These results support the assigned α relative configuration of the *p*-nitrobenzoyl moiety at C–13 in **269**.

The target structure, cyathin A_4 (37), was expected to be available from 269 via saponification of the two PNB esters in 269 and subsequent removal of the PMB group (Scheme 56).



Scheme 56

With only limited amounts of substance **269** in hand, the first step, a saponification reaction, was attempted. To our disappointment, treatment of a MeOH solution of diester **269** with K_2CO_3 afforded, upon work-up, a complex mixture of unidentified products (eq 46). The saponification step was also attempted with Na₂CO₃ in MeOH but these conditions also led to a complex mixture of products.

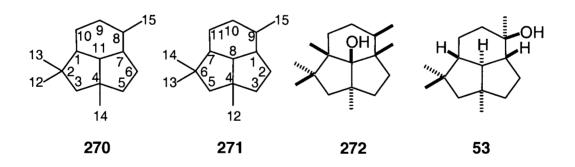


Unfortunately, limitations in available advanced materials prevented further investigations into the preparation of diol **261**. On the basis of time restraints, a scale-up preparation of these advanced intermediates was not pursued and our synthetic study geared towards the natural product cyathin A_4 (**37**) was abandoned.

2.2. Studies Towards Presilphiperfolane Sesquiterpenoid Synthesis

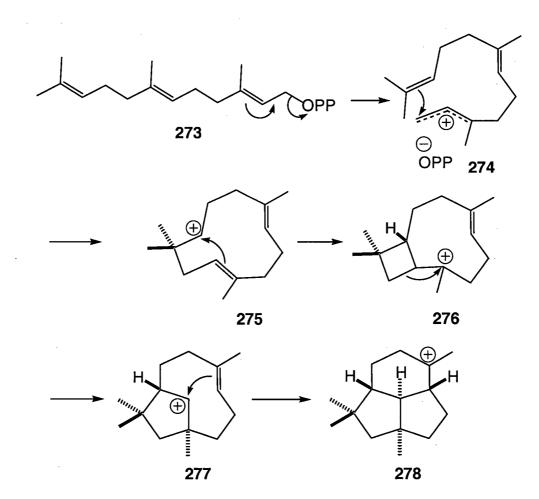
2.2.1. Background: Presilphiperfolane Isolation, Biogenesis and Biological Activity

The presilphiperfolane family of sesquiterpenoids share the common carbon skeleton **270**, which displays an unusual tricyclo[5.3.1.0^{4,11}]undecane framework. The IUPAC-based numbering and presilphiperfolane natural product numbering schemes are illustrated in **270** and **271**, respectively.* In 1981, Bohlmann and coworkers reported the first structural elucidation of a member of the presilphiperfolane family, presilphiperfoan-8-ol (**272**), from the California coastal succulent *Eriophyllum staechadifolium*.¹⁰² In 1996, the research group of Weyerstahl and that of Marco concurrently reported the isolation of presilphiperfolan-9-ol (**53**) from *Artemisia laciniata* and from *Artemisia chamaemelifolia*, respectively.^{20,103}



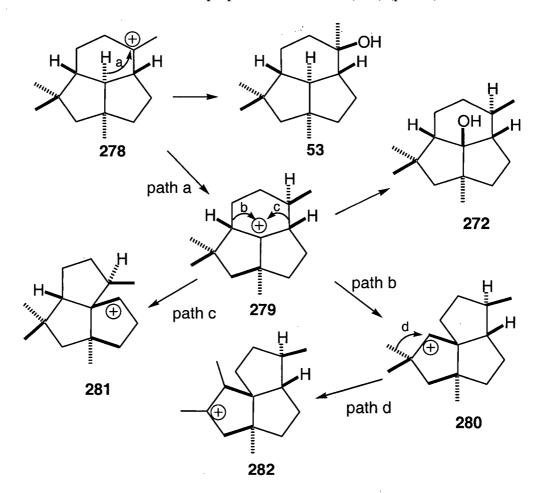
^{*} In the text of this thesis, the presilphiperfolane numbering system 271 is used in most descriptions of presilphiperfolane-related natural products and the IUPAC-based numbering system 270 is used to describe the synthetic intermediates. The experimental section of this thesis contains IUPAC names for the synthetic intermediates.

Bohlmann and coworkers have proposed a biogenetic pathway for the formation of presilphiperfolane natural products from (E,E)-farnesyl diphosphate (273) (Schemes 57 and 58).¹⁰⁴ Sequential cationic cyclizations of salt 274 provide the caryophyllen-8-yl cation 276 via 11-membered macrocycle intermediate 275 (Scheme 57). Cyclobutylcarbinyl ring expansion of 276 generates a cyclopentyl cation with a bridging (E)-3-hexenyl chain, 277. Cationic cyclization of 277 affords the tricyclo $[5.3.1.0^{4,11}]$ undecyl cation **278**, which displays the presilphiperfolane skeleton.





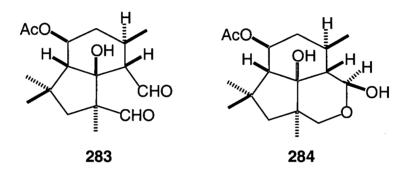
The natural product presilphiperfolan-9-ol (53) presumably arises directly from cation 278 *via* an oxygenation step (Scheme 58). On the other hand, a 1,3-hydride shift in 278 generates cation 279 (path a), the precursor for presilphiperfolan-8-ol (2). The biogenetic pathway leading to the presilphiperfolanes is common to several structurally related families of sesquiterpenes, including the silphiperfolanes, the silphinanes and the camaroonanes.^{102,104,105} The proposed biogenetic pathways to these families share the advanced intermediate 279. 1,2-Alkyl shifts involving cation 279 gives the cameroonane (280) (path b) or silphinane (281) (path c) skeletons. A 1,2-methyl shift in the cameroonane cation 280 affords the silphiperfolane skeleton (282) (path d).



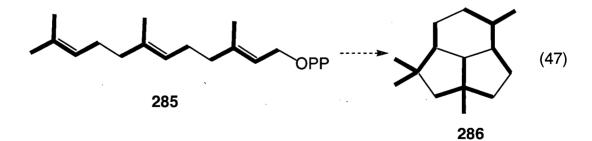
Scheme 58

124

Although several members of the presilphiperfolane and related families of sesquiterpenoids exhibit pleasant odiferous qualities, only a select few display pharmaceutically applicable biological activities. As an example, two sesquiterpenoids isolated from *Botrytis cinerea*, botrydial (**283**) and dihydoxybotrydial (**284**), demonstrate phytotoxic and antibiotic properties.¹⁰⁶

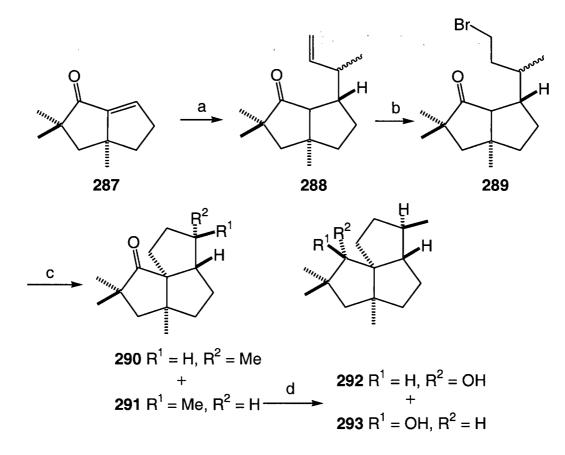


Studies¹⁰⁷⁻¹⁰⁹ by Hanson and coworkers on the metabolites **283** and **284** support the biosynthetic pathway proposed by Bohlmann and coworkers (*vide supra*). The fungus *Botrytis cinerea* was fed $[1-^{13}C]$ - and $[2-^{13}C]$ -acetate as well as $[4,5-^{13}C_2]$ -mevalonate, and the labeling patterns of the resulting metabolites were investigated. The pattern of incorporation of three isoprene units, highlighted in **285**, into the carbon framework of **286** was rationalized *via* a sequence of cationic cyclizations in agreement with those proposed by Bohlmann (eq 47). Coates and coworkers have also reported several studies on the presilphiperfolanes biogenesis. ¹¹⁰⁻¹¹³



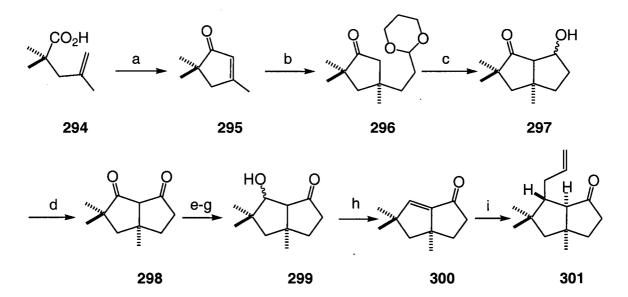
2.2.2. Previous Syntheses

In 2000, Coates and coworkers reported¹¹⁴ the first total synthesis of (\pm) cameroonan-7-ol¹⁰⁵ (293) from the known^{115,116} bicyclic enone 287. The key step in this
synthesis involved an intramolecular alkylation reaction (Scheme 59). Enone 287 was
subjected to a Sakurai reaction¹¹⁷ with (*Z*)-2-buten-1-yltrimethylsilane and TiCl₄,
affording a 2:1 mixture of epimers 288. A photochemical free radical hydrobromination
of this mixture yielded the corresponding bromides 289. Base-mediated cyclization of
these bromides afforded two cyclic ketones, 290 and 291. The desired isomer, ketone
291, was reduced with LiAlH₄, yielding the target structure (\pm)-cameroonan-7-ol 293 and
its epimer, 292, in a 1.4:1 ratio.



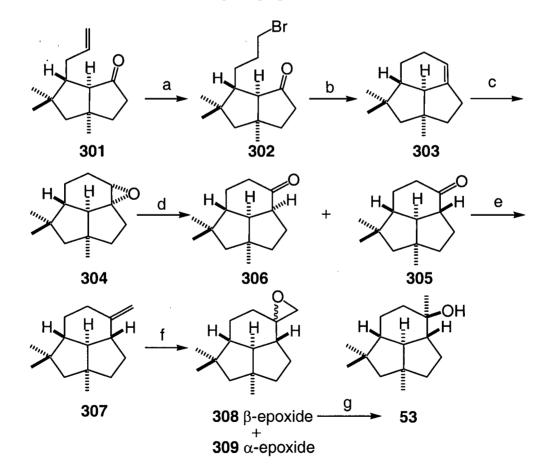
Reagents: a) (Z)-2-buten-1-yltrimethylsilane, TiCl₄; b) HBr, hv; c) t-BuOK; d) LiAlH₄.

In 1996, Weyerstahl and coworkers reported the first total synthesis of (\pm) presilphiperfolan-9-ol (53) (Schemes 60 and 61). The 16-step synthesis of (\pm) -53
commenced with the conversion of the pentenoic acid 294 to the corresponding acid
chloride, which was subsequently treated with AlCl₃ to afford the cyclopentenone 295
(Scheme 60). Copper(I)-catalyzed conjugate addition of the Grignard reagent derived
from 2-(2-bromoethyl)-1,3-dioxane to enone 295 provided ketone 296 in good yield.
Acid hydrolysis of 296, followed by an aldol cyclization of the resultant keto aldehyde,
gave a mixture of hydroxy ketones 297. Jones oxidation of this mixture provided
diketone 298, which was transformed to hydroxy ketone 299 in three steps. Conversion
of 299 to the corresponding mesylates, followed by a DBU-mediated elimination
reaction, gave enone 300. TiCl₄-mediated Sakurai conjugate allylation of enone 300 with
allyltrimethylsilane provided ketone 301.



Reagents: a) (COCl)₂; AlCl₃; b) 2-[(1,3)-dioxan-2-yl]-1-ethylmagnesium bromide, CuI, TMEDA, TMSCl; c) acetone, HCl; d) CrO₃, H₂SO₄, H₂O; e) (CH₂OH)₂, HC(OMe)₃, TsOH; f) LiAlH₄; g) dil. HCl; h) MsCl, Et₃N; DBU; i) allyltrimethylsilane, TiCl₄.

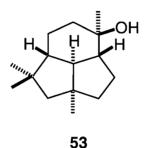
The final synthetic steps leading to (\pm)-presilphiperfolan-9-ol (53) are illustrated in Scheme 61. Benzoyl peroxide-mediated addition of HBr to enone 301 afforded bromide 302. The Wittig salt derived from bromide 302 was treated with base, resulting in a cyclization that gave the tricyclic alkene 303. A stereoselective α -epoxidation of 303, followed by a ZnBr₂-mediated rearrangement of the resultant epoxide 304, afforded norpresilphiperfolan-9-one 305 and its epimer 306. Treatment of 305 with Lombardo's reagent (CH₂Br₂/Zn/TiCl₄) provided the corresponding alkene 307, which was oxidized with mCPBA to give epoxides 308 and 309 in a 3:2 ratio. LiAlH₄ reduction of epoxide 308 yielded the target structure, (\pm)-presilphiperfolan-9-ol (53).



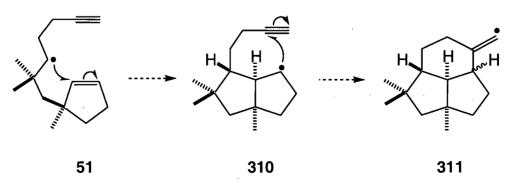
Reagents: a) HBr, Bz₂O₂;b) Ph₃P; NaHMDS; c) mCPBA; d) ZnBr₂; e) Zn, CH₂Br₂, TiCl₄; f) mCPBA; g) LiAlH₄.

2.2.3. Studies Towards a Formal Synthesis of (±)-Presilphiperfolan-9-ol

Our synthetic studies directed towards the formal synthesis of (\pm) presilphiperfolan-9-ol (53) relied on a proposed radical-mediated tandem cyclization¹¹⁸
sequence in the construction of the tricyclo[5.3.1.0^{4,11}]undecane carbon framework (*vide infra*).

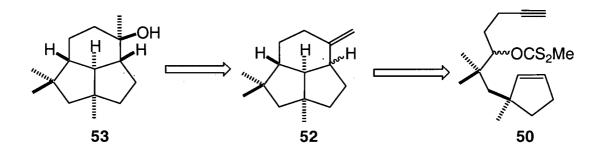


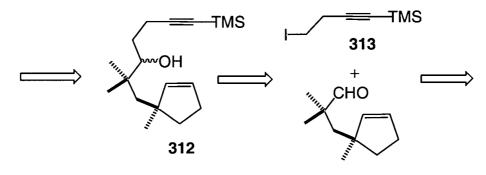
Our proposed tandem cyclization sequence commences with the secondary radical **51**. The first reaction is a 5-*exo* cyclization of the 5-hexenyl-type radical **51**, which was anticipated to result in the *cis*-fused 5-5 bicyclic radical **310** (Scheme 62). The 6-heptynyl-type radical **310** was expected to cyclize in a 6-*exo* fashion, generating the *cis*-fused bridge in **311**. A more detailed analysis of the proposed transition states for the tandem cyclizations and the expected relative configurations for the ring-fusion centers will follow.



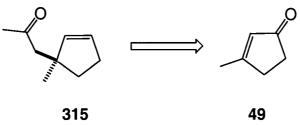
2.2.3.1. Retrosynthetic Analysis

Our retrosynthetic plan for the formal synthesis of (\pm)-presilphiperfolan-9ol (53) is outlined in Scheme 63. The target compound 53 is readily available from the β -isomer of alkenes 52, as reported by Weyerstahl and coworkers (see Section 2.2.2., *vide supra*).²⁰ Treatment of xanthates 50 with Bu₃SnH and AIBN should provide the corresponding secondary alkyl radical, the first intermediate in our proposed radical-mediated tandem cyclization leading to alkenes 52 (*vide supra*). The cyclization precursors, xanthates 50, should be readily derived from alcohols 312. 1,2-Addition of the alkyllithium species derived from the known¹¹⁹ iodide 313 to aldehyde 314 was not expected to be stereoselective and should afford a mixture of alcohols 312. A sequence of chain homologation and α -methylation steps should provide aldehyde 314 from ketone 315. Finally, ketone 315 should be available from enone 49 *via* a sequence of steps that include a Carroll [3,3] rearrangement.¹²⁰







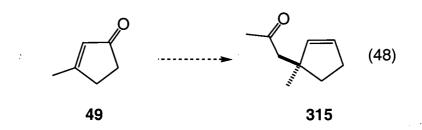




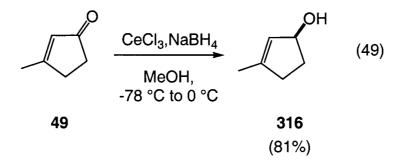
Scheme 63

2.2.3.2. Preparation of Ketone 315

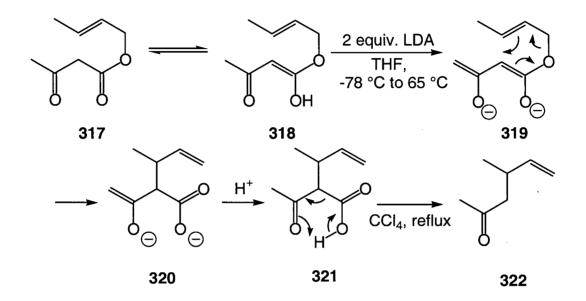
The first synthetic sequence in our study involved the preparation of ketone **315** from 3-methyl-2-cyclopenten-1-one (**49**) (eq 48).



A racemic mixture of the known¹²¹⁻¹²⁵ alcohol **316** was prepared from commercially available enone **49** by selective 1,2-reduction with CeCl₃ and NaBH₄ (eq 49).¹²⁶ Alcohol **316**, obtained in 81% yield upon purification by flash chromatography, was analyzed by standard spectrometric methods. All spectral data derived from **316** were identical to those reported in the literature.¹²⁵

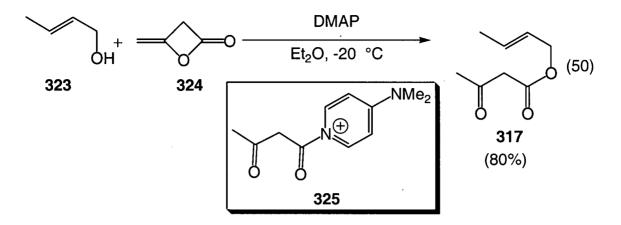


In 1984, Wilson and coworkers reported¹²⁷ a dianionic version of the well-known Carroll rearrangement¹²⁰ of allylic β -keto esters. A THF solution of allylic β -keto ester **317** and its tautomer **318** was treated with two equivalents of LDA (Scheme 64). The resultant solution of dianion **319** was heated to 65 °C, generating, after a suitable work-up procedure, the [3,3] rearrangement product, β -keto acid **321**. Heating a solution of **321** in CCl₄ resulted in a decarboxylation, affording alkenone **322**.

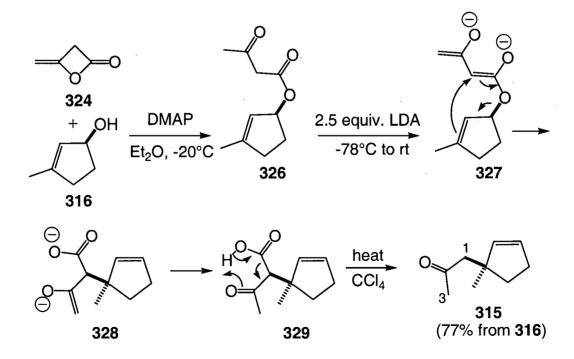


Scheme 64

Wilson and coworkers also developed¹²⁷ an efficient preparation of the allylic β keto ester **317** from the corresponding allylic alcohol **323** (eq 50). Treatment of a cold (– 20 °C) solution of **323** in Et₂O with diketene (**324**), followed by the addition of a catalytic amount of DMAP, provided the corresponding allylic β -keto ester **317** in good yield. The active acylating reagent, compound **325**, is formed *in-situ* by reaction of DMAP and diketene (**324**), followed by protonation.



The sequence developed by Wilson and coworkers for the preparation of allylic esters and their subsequent Carroll rearrangement was applied to alcohol **316** (Scheme 65). An ethereal solution of **316** and diketene (**324**) was treated with a catalytic amount of DMAP and the resultant allylic ester **326** was subsequently treated 2.5 equiv. of LDA. Dianion **327** underwent a [3,3] sigmatropic rearrangement to dianion **328**, providing keto acid **329** after work-up. A solution of the crude keto acid **329** in CCl_4 was refluxed, affording the desired ketone **315**.

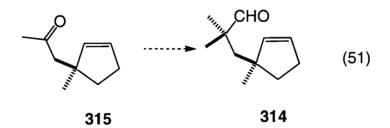


Scheme 65

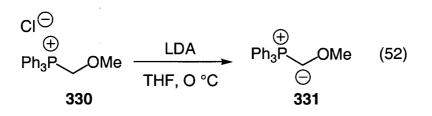
The structure of ketone **315** was supported by standard spectrometric analysis. The IR spectrum of **315** displayed a sharp absorption at 1718 cm⁻¹, characteristic of an aliphatic ketone. The ¹H NMR spectrum of **315** exhibited a 3-proton singlet at δ 2.11 and a 2-proton singlet at δ 2.49, attributed to the H–3 and H–1, respectively. The ¹H NMR spectrum of **315** also exhibited overlapping multiplets between δ 5.61–5.68, attributed to the olefinic protons. The ¹³C NMR resonances for the carbonyl and olefinic carbons of **315** were observed at δ 208.3 and at δ 129.1 and 139.0, respectively.

2.2.3.3. Preparation of Aldehyde 314

With ketone **315** in hand, the preparation of **314** via a sequence of chainhomologation and methylation steps was investigated (eq 51).

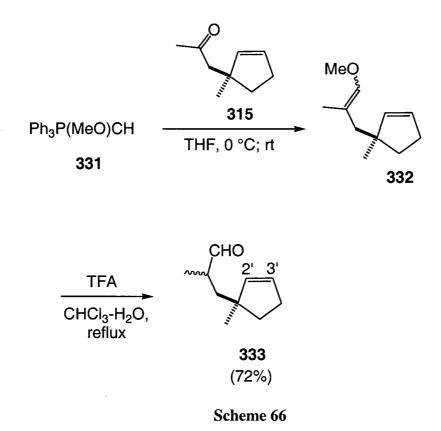


The first transformation of this sequence was attempted with the modified Wittig reagent 331,¹²⁸ which was prepared by treatment of a cold (0 °C) solution of 330 in THF with LDA (eq 52).



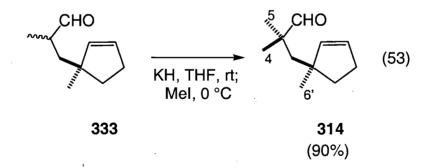
136

A cold (0 °C) solution of **331** was treated with ketone **315**, was stirred for 1 h at 0 °C and then was allowed to warm to room temperature (Scheme 66). After aqueous work-up, the resultant crude mixture of enol ethers **332** was subjected to hydrolysis conditions (TFA in refluxing aqueous CHCl₃), affording a 1:1 mixture of aldehydes **333**, in 72% yield.



Evidence for the successful carbonyl homologation of ketone **315** was obtained by spectroscopic analysis of aldehydes **333**. The IR spectrum of **333** displayed an absorption at 1735 cm⁻¹, characteristic of aldehydic carbonyls. The ¹H NMR spectral signals of the aldehydic protons in **333** were observed as doublets at δ 9.51 (J = 2.9 Hz) and 9.53 (J = 2.7 Hz). Resonances at δ 205.2 and 205.3 in the ¹³C NMR spectrum of **333** were attributed to the aldehydic carbons. The ¹³C NMR resonances for the C–2' and C–3' olefinic carbons in **333** were observed at δ 139.2 and 139.5 and at δ 129.4 and 129.9, respectively.

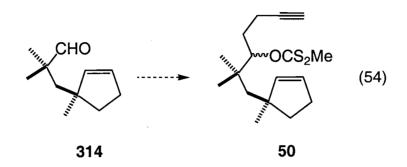
With aldehydes **333** in hand, the preparation of the corresponding α,α -dimethyl aldehyde **314** was straightforward (eq 53). Aldehydes **333** were added to a THF solution of potassium hydride and the resultant enolate solution was treated with methyl iodide at 0 °C.¹²⁹ Work-up and purification of the resulting material by flash chromatography provided the desired aldehyde **314** in 90% yield.



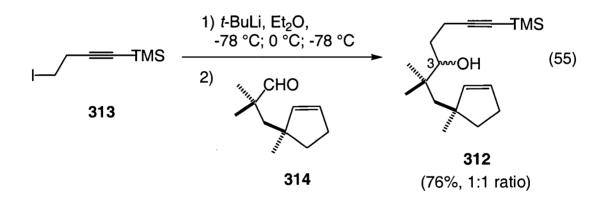
The successful α -methylation of **333** was supported by ¹H NMR spectroscopic analysis of the product **314**. The ¹H NMR spectrum derived from **314** exhibited 3-proton singlets at δ 1.01 and 1.04 attributed to the H–4 and H–5 methyl protons. The ¹H NMR spectral resonance for the aldehydic proton in **333** was observed as a 1-proton singlet at δ 9.44.

2.2.3.4. Preparation of the Tandem Cyclization Precursors, Xanthates 50

Xanthates **50**, the tandem cyclization precursors, were readily prepared in three steps from aldehyde **314** (*vide infra*) (eq 54).



Following a protocol developed¹³⁰ by Negishi and coworkers, a cold (-78 °C) Et₂O solution of the known¹¹⁹ iodide **313** was treated with 2.5 equiv. of *t*-BuLi and the resulting mixture was warmed to 0 °C, generating the corresponding alkyllithium species (eq 55). Upon cooling to -78 °C, the solution of the alkyllithium was treated with aldehyde **314**, affording, after a work-up step, a 1:1 mixture of alcohols **312**.



The 1:1 mixture alcohols **312**, inseparable by silica gel flash chromatography, was analyzed by NMR spectroscopy. The ¹H NMR spectrum of **312** displayed overlapping 1-

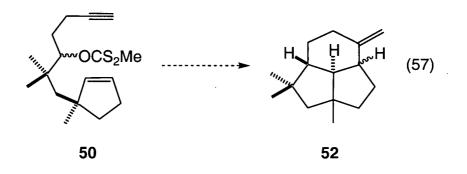
proton multiplets between δ 3.3–3.45, attributed to H–3. The C–3 carbons gave rise to resonances at δ 78.4 and 79.2 in the ¹³C NMR spectrum of **312**.

The preparation of xanthates **50** from alcohols **312** was straightforward. A solution of alcohols **312** in THF was added to a THF solution of NaH and the resultant mixture was treated with imidazole and carbon disulfide (eq 56). The resultant solution of dithiocarboxylate anions was treated with methyl iodide, affording the corresponding xanthates. The TMS function on the alkyne moiety was removed by treatment a THF– MeOH solution of the latter material with TBAF, affording the desilylation products **50**. The structures of **50** were confirmed by ¹H NMR spectroscopic analysis. The ¹H NMR spectrum of xanthates **50** displayed two 3-proton singlets at δ 2.55 and 2.57, representing the methyls in the xanthates moieties. The H–3 protons in xanthates **50** were observed as overlapping multiplets between δ 5.79–5.85, characteristic of secondary xanthates.

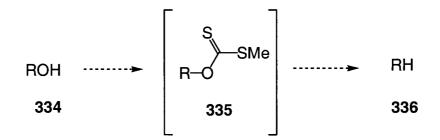
TMS 1) NaH, imidazole, CS₂ THF, reflux; Mel (81%) S₂Me (56)2) TBAF, THF-MeOH. rt (90%) 312 50

2.2.3.5. Tandem Free-radical Cyclization

With xanthates **50** in hand, attention was turned to the proposed tandem radicalmediated cyclization (eq 57).

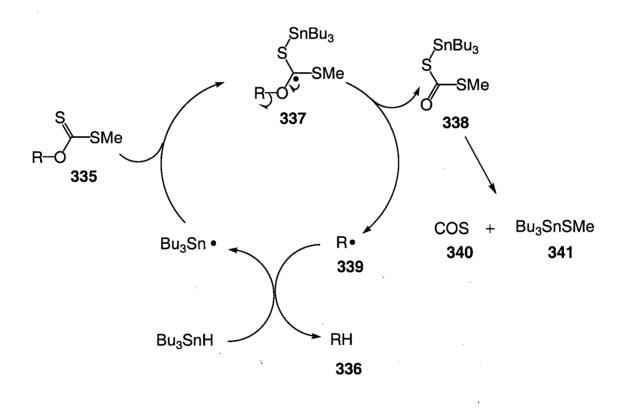


The xanthate functional group has traditionally served as convenient derivative for the Barton–McCombie deoxygenation of alcohols (Scheme 67). In 1975, Barton and McCombie reported¹³¹ that xanthates (**335**), readily derived from the corresponding alcohols (**334**), generate the reduced products (**336**) *via* Bu₃SnH-mediated processes that proceed by way of radical intermediates.



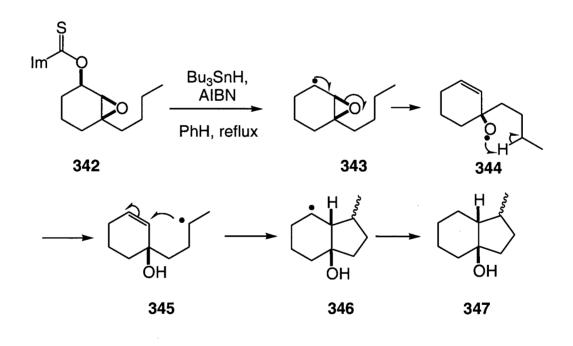
Scheme 67

The proposed mechanistic pathway^{131,132} for the Barton–McCombie radicalmediated reduction of xanthates is illustrated in Scheme 68. The stannyl radical, which displays a high affinity for sulfur, attacks the thiocarbonyl group in xanthate **335**, generating radical adduct **337**. A β -scission of the carbon–oxygen bond in **337** provides the alkyl radical **339** and the co-product **338**, which is unstable with respect to elimination of carbon oxysufide **340**. Abstraction by the alkyl radical **339** of a hydrogen atom from Bu₃SnH generates the desired alkane **336** and regenerates the stannyl radical, which results in propagation of the chain reaction.



Scheme 68

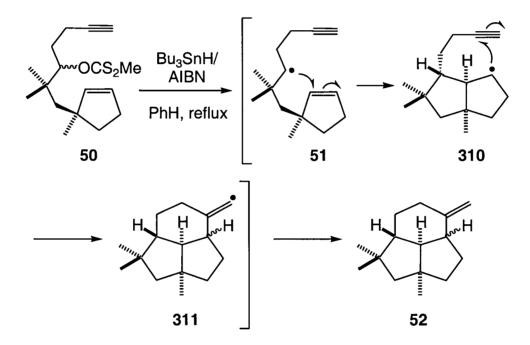
The radical intermediate obtained from treatment of xanthate-type derivatives with Bu₃SnH/AIBN have also been used for reactions other than direct reduction by hydrogen atom abstraction.^{133,134} For example, in 1990, Rawal and coworkers reported¹³⁵ the preparation of carbocyclic systems from xanthates *via* radical-induced epoxide fragmentation (Scheme 69). Xanthate **342** was converted to the corresponding radical **343**, which fragmented to the oxy-radical **344**. Intramolecular hydrogen abstraction converted **344** into the radical **345**. A 5-*exo* cyclization of the 5-hexenyl radical **345** generated a mixture of the *cis*-fused bicyclic intermediates **346**, which yielded **347** upon hydrogen atom abstraction.





The proposed tandem reactions of the radical intermediate 51, obtained by the Barton-McCombie-type reaction of xanthates 50, are outlined in Scheme 70. In

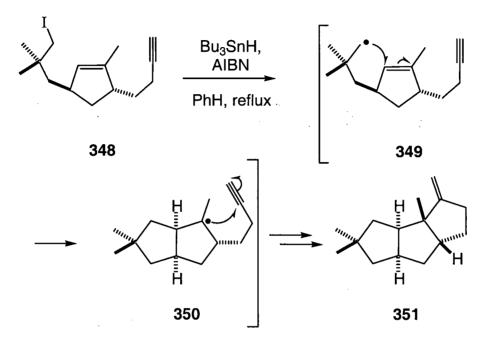
accordance with guidelines¹³⁶ developed for the stereochemical predictions of radicalmediated cyclizations, a 5-*exo* cyclization of the 5-hexenyl-type radical in **51** was anticipated to form the *cis*-fused 5-5 bicyclic intermediate **310**. The second reaction, a 6*exo*-type cyclization of the 6-heptynyl-type radical in **310**, was expected to result in a sixmembered ring, as displayed in **311**. Finally, hydrogen abstraction by **311** would afford the tricycles **52**.





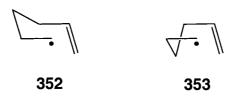
The first cyclization reaction in the proposed sequence (*vide supra*) is analogous to that of a tandem sequence developed by Curran and coworkers for the total synthesis of (\pm)-hirsutene (**351**) (Scheme 71).¹³⁷ Thus, treatment of iodide **348** with Bu₃SnH and AIBN generated the corresponding 5-hexenyl-type radical **349**, which underwent a 5-*exo*

cyclization to give the *cis*-fused 5-5 bicyclic intermediate **350**. A 5-*exo* cyclization of the 5-hexynyl-type radical **350**, followed by hydrogen abstraction, afforded (\pm) -hirsutene (**351**).

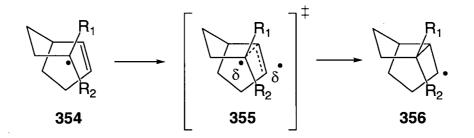


Scheme 71

Predictions and rationalizations of stereoselectivity¹³⁶ in 5-*exo* hexenyl cyclization are generally based on the Beckwith–Houk transition state model.¹³⁸⁻¹⁴¹ In brief, the Beckwith–Houk model involves a combination of the proposed transition structure for bimolecular radical additions to alkenes and the principles of conformational analysis, which support the postulated chair- and boat-like transition state structures, **352** and **353**, for the cyclization of acyclic substrates.

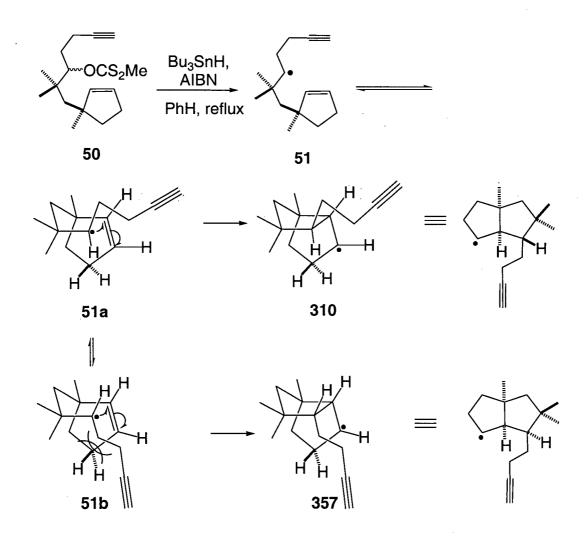


Radical cyclizations resulting in the formation of a 5-5- or smaller fused ringsystems favour chair-like transition-structures and usually form *cis*- rather than *trans*fused products.¹³⁶ The first step of our proposed tandem sequence, represented by the cyclization of **354** to **356** in Scheme 72, was anticipated to involve a transition state resembling **355** in which $R_1 = H$ and $R_2 =$ tethered acetylene or vice versa.



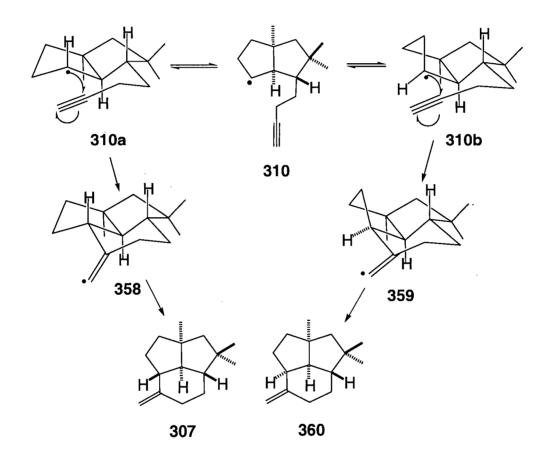
Scheme 72

Upon treatment with $Bu_3SnH/AIBN$, the mixture of xanthates **50** was expected to form the secondary radical intermediate **51** (Scheme 73). On the basis of molecular modeling and conformational analysis, the preferred transition state for the cyclization of **51** was expected to resemble structures **51a** or **51b**, leading to the diastereomeric intermediates, **310** and **357**, respectively. The main difference between **51a** and **51b** is the orientation of the substituents on the radical-bearing carbon. In **51a**, the smaller substituent (hydrogen) is positioned over the cyclopentenyl ring. Conversely, in **51b**, the larger substituent lies over the ring. On the basis of the possible steric interactions between the larger substituent and the ring-protons in **51b**, the transition state for the cyclization of **51** was anticipated to favour a geometry resembling **51a**, leading to radical **310**.



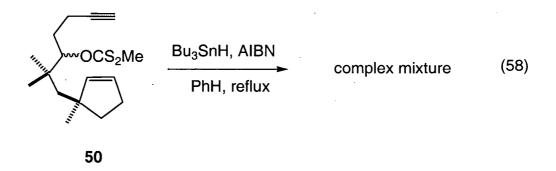


For the second step of the proposed tandem sequence, a 6-*exo* cyclization of 6heptynyl radical **310**, the transition structures leading from **310a** and **310b** to give intermediates **358** and **359**, respectively, were considered (Scheme 74). Conformational analysis and molecular modeling studies of these transition structures suggested that a mixture of isomers, favouring **360** *via* **359**, might be obtained. The most notable difference between the two proposed transition structures was the closer structural proximity of the tethered acetylene to the radical center afforded by the transition structure leading from **310b** to **359** compared to that afforded by the transition structure leading from **310a** to **358**.

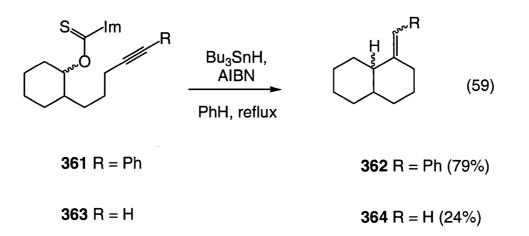


Scheme 74

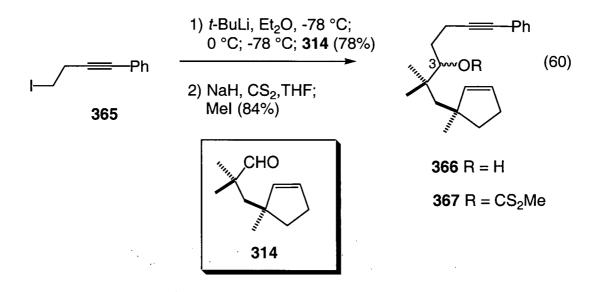
In the event, slow syringe-addition of Bu_3SnH and a catalytic amount of AIBN to a refluxing solution of xanthates **50** in benzene afforded neither tandem cyclization product **307** nor **360** (eq 58). Upon removal of the solvent, a mixture of tin-containing products, representing less than 5% mass balance with respect to xanthates **50**, was obtained. Presumably, the products were volatile and consequently, under the work-up conditions, were lost to the atmosphere.



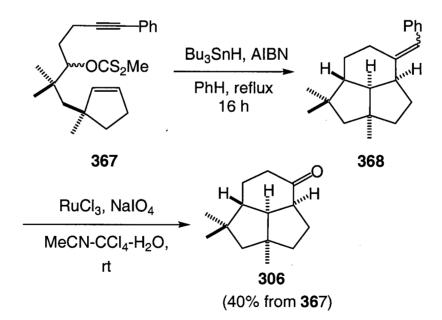
The literature pertaining to radical chemistry was reviewed for possible methods of improving our cyclization reaction. Specifically, alterations to our cyclization substrate that might reduce product loss by evaporation were sought. In 1984, Clive and coworkers reported¹⁴² that the radical cyclization of phenylacetylene **361** provided the corresponding cyclic product **362** in a yield higher than that obtained from the corresponding cyclization of the non-phenyl derivative **363** to **364** (eq 59).



It was expected that phenyl derivatives of our acetylenic substrates would provide less volatile products upon cyclization. Thus, the phenyl-substituted substrates were prepared following the procedures described earlier for xanthates **50**. The known¹⁴³ iodide **365** was treated with 2.1 equiv. of *t*-BuLi and the resultant solution of the required alkyllithium species was treated with aldehyde **314** (eq 60). The alcohol adducts **366**, obtained in 78% yield, were then transformed into the corresponding xanthates **367**. Xanthates **367** were purified by flash chromatography on silica gel and analyzed by standard spectroscopic methods. The ¹H NMR spectrum of **367** displayed multiplets between δ 7.20–7.42, representing the aromatic protons of the phenyl ring. Overlapping doublets at δ 5.91 in the ¹H NMR of **367** were attributed to H–3, the proton on the carbon bearing the xanthate moiety.

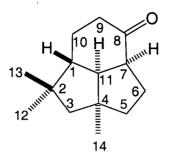


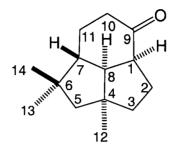
With xanthates **367** in hand, the proposed tandem cyclization reaction to generate the presilphiperfolane skeleton was further investigated. Treatment of a benzene solution of xanthates **367** with Bu₃SnH and AIBN (slow syringe addition over 18 h) provided a mixture of non-polar compounds, which were inseparable by silica gel flash chromatography (Scheme 75). On the basis of spectroscopic analysis (*vide infra*), the two main components of this mixture were tentatively assigned structures **368**. The ¹H NMR spectrum of the mixture displayed two overlapping signals at δ 6.35, attributed to respective olefinic protons in **368**. The ¹³C NMR spectrum of the mixture displayed four olefinic signals between δ 125 and 126, representing the carbons of the double bonds in **368**. The known ketone **306**, reported as 1-epi-9-norpresilphiperfolan-9-one,²⁰ was readily obtained from the mixture of cyclization products **368** upon subjection of the latter to an oxidative cleavage protocol developed⁸¹ by Sharpless and coworkers. Treatment of a solution of **368** in MeCN–CCl₄–H₂O with NaIO₄ and RuCl₃ afforded ketone **306**, obtained in 40% overall yield from **367**. The assigned structure of ketone **306** was supported by comparison of its derived spectral data with those previously reported by Weyerstahl and coworkers for 1-epi-9-norpresilphiperfolan-9-one. Comparisons of the ¹H and ¹³C NMR data are included in **Tables 7** and **8**, respectively.



Scheme 75

Table 7. Comparison of selected (assigned) ¹H NMR (CDCl₃, 400 MHz) Data for Ketone **306** with those Reported²⁰ for (\pm)-1-Epi-9-norpresilphiperfolan-9-one (**306**).



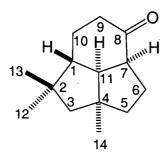


306 showing IUPAC-based numbering

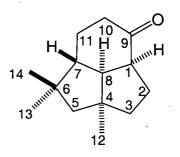
306 showing presilphiperfolane numbering

Assignment H-x (306)	¹ H NMR δ ppm (Multiplicity, J (Hz))	Presilphiper- folane Numbering H-x	Literature ¹ H NMR Assignments δ ppm (Multiplicity, J (Hz))
H–1	1.40 (ddd, 4, 12.5, 13)	H–7	1.39 (ddd, 4, 12.5, 13)
H–11	2.19 (dd, 7, 12.5)	H–8	2.18 (dd, 7,12.5)
H–12	0.88 (s) or 1.01 (s)	H–13	0.88 (s) or 1.01 (s)
H–13	0.88 (s) or 1.01 (s)	H–14	0.88 (s) or 1.01 (s)
H–14	1.07 (s)	H–12	1.07 (s)

Table 8. Comparison of the ¹³C NMR (CDCl₃, 100.6 MHz) Data for Ketone **306** and those Reported²⁰ for (\pm)-1-Epi-9-norpresilphiperfolan-9-one (**306**).



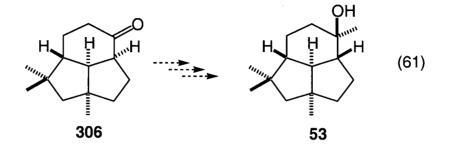
306 showing IUPAC-based numbering



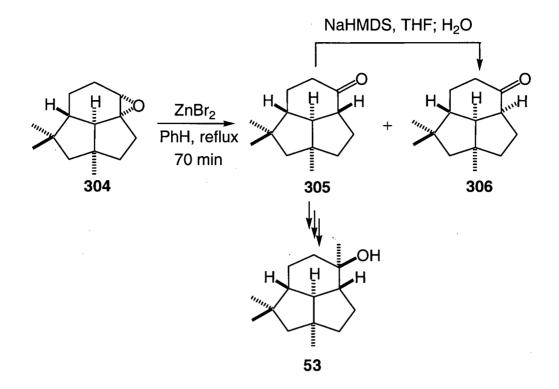
306 showing presilphiperfolane numbering

Assignment C-x (306)	¹³ C NMR δ ppm (Multiplicity, J (Hz))	Presilphiperfolane Numbering C-x (306)	Literature ¹³ C NMR Assignments δ ppm (Multiplicity, J (Hz))
C-1	50.9	C-7	50.9
C-2	40.6	C6	40.6
C-3	58.0	C-5	58.0
C-4	47.5	C-4	47.5
C5	41.9	C-3	41.9
C6	28.2	C-2	28.2
C–7	52.7	C-1	52.6
C8	216.2	C-9	216.2
C9	38.3	C-10	38.3
C-10	21.8	C-11	21.8
C-11	59.1	C-8	59.1
C-12	22.7	C-13	22.7
C-13	28.9 or 29.0	C-14	29.0
C-14	28.9 or 29.0	C-12	29.0

The transformation of ketone 306 into the target structure (±)-presilphiperfolan-9ol (53) was briefly investigated (eq 61).



The prospect of transforming ketone **306** into its C–1 epimer **305**, an advanced intermediate in Weyerstahl and coworkers' total synthesis of presilphiperfolan-9-ol (**53**), was considered. In the study by Weyerstahl and coworkers (see Section 2.2.2 *vide supra*), ketones **305** and **306** were obtained upon treatment of a benzene solution of epoxide **304** with $ZnBr_2$ (Scheme 76). If the reaction was stopped after 40 min, ketone **305** was obtained exclusively. However, prolonged reaction times (70 min) resulted in the formation of **305** and **306** in a 3:1 ratio. The same isomers, **305** and **306**, were obtained in a 1:7 ratio upon equilibration of **305** with 0.9 equiv. of NaHMDS.



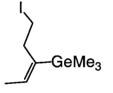
Scheme 76

Although the desired isomer **305** was less favoured under thermodynamically controlled epimerizing conditions (*vide supra*), ketone **306** was nevertheless subjected to base-mediated isomerization conditions (0.9 equiv. LDA /THF, *t*-BuOK/*t*-BuOH or NaOMe/MeOH) in hopes of obtaining a minor portion of **305**. Disappointingly, in each case, **306** was completely recovered.

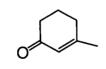
On the basis of time-restraints, our proposed formal synthesis of presilphiperfolan-9-ol (53) *via* direct conversion of ketone 305 to ketone 306 was abandoned. Although an alternate method of preparing the target structure 53 from ketone 306 likely exists, a more concise approach to the presilphiperfolane core, displaying all the correct ring-fusion configurations, was desired. Thus, further investigation into the possibility of effectively controlling the stereoselectivity of the tandem cyclization reaction are necessary.

III. CONCLUSIONS

The work described in the first part of this thesis illustrates a successful application of annulation sequences developed in our laboratories to the construction of the 5-6-7 fused ring system of the cyathanes. The synthetic studies commenced with 3-methyl-2-cyclohexen-1-one (**38**), which eventually became the B-ring of the cyathane carbon skeleton (Scheme 77). An annulation sequence with bifunctional reagent alkenylstannane **39** was employed in the construction of the C-ring. In turn, the A-ring annulation was achieved with alkenylgermane **44**. As a result of the step-wise annulation strategy used in this synthesis, several advanced functionalized intermediates, which display the complete cyathane 5-6-7 fused tricyclic framework **54**, were made readily available.

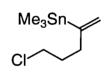






38

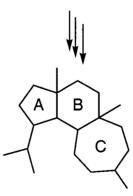
Ring B substrate







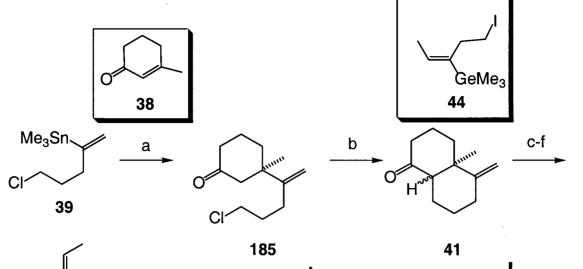
Ring A annulation reagent

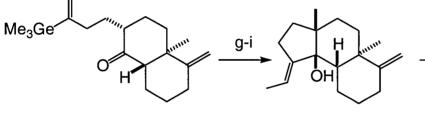




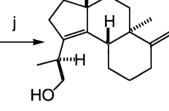
Scheme 77

A summary of the cyathane synthetic studies, including key intermediate structures, is outlined in Schemes 78 and 79. The synthesis commenced with a copper(I)-catalyzed conjugate addition of the Grignard reagent derived from **39** to enone **38**, affording adduct **185** (Scheme 78). Base-mediated cyclization of **185** gave the known bicyclic enones **41**,¹⁸ which represent the B-C ring system of the cyathane core. The subsequent A-ring annulation sequence employed bifunctional reagent **44** in the transformation of **41** to ketone **187**, which ultimately afforded the tricyclic alcohol **155**. Subjection of **155** to the Still-Mitra protocol yielded carbinol **151**.⁶⁹ With the A-ring and its hydroxyl isopropyl appendage in place, attention was turned to elaboration of the C-ring. Substance **151** was converted to iodides **160**, which were subjected to a SmI₂-mediated ring expansion reaction. The resulting product, keto ester **159**, displayed the complete 5-6-7 fused cyathane framework.





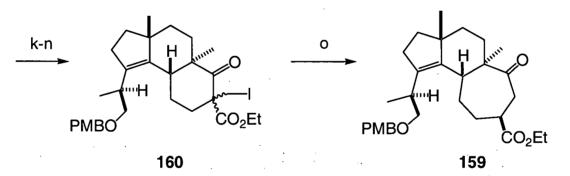
187





.

151

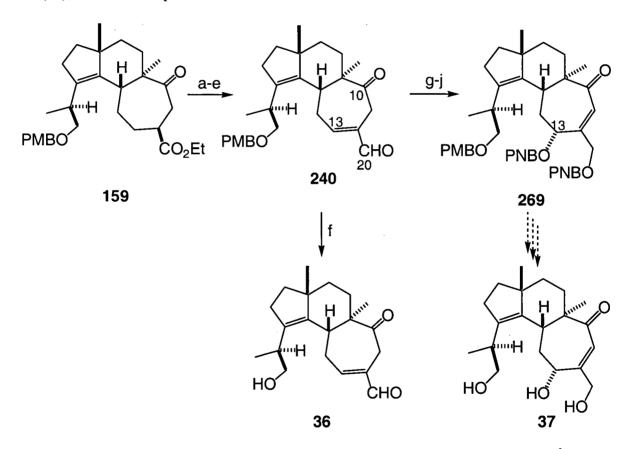


155

Reagents: a) MeLi, THF, -78 °C; MgBr·Et₂O; CuBr·DMS, BF₃·Et₂O, **38**; b) KH, THF, reflux; EtOH; c) Me₂NNH₂, CSA, PhH, reflux; d) KDA, DMPU, THF, -78 °C; **44**; e) AcOH, NaOAc, THF, H₂O, 75 °C; f) KOMe, MeOH, 65 °C; g) Et₂NLi, THF, -78 °C; 0 °C; MeI; h) NIS, CH₂Cl₂, 0 °C; i) BuLi, THF, -78 °C; j) KH, 18-cr-6, THF, rt; Bu₃SnCH₂I; BuLi, -78 °C; k) KH, THF, rt; PMBCl, Bu₄NI; l) OsO₄, KIO₄, *t*-BuOH, NaHCO₃, H₂O, rt; m) KH, NaH, (EtO)₂CO, THF, 65 °C; aq. HCl; n) TBAF, THF; CH₂I₂; o) SmI₂, THF, rt.

Scheme 78

Efforts to transform the advanced intermediate **159** into natural product target structures are summarized in Scheme 79. Keto aldehyde **240** was obtained from **159** in five steps. Removal of the PMB protecting group in **240** represented the final step in the first reported⁹⁴ total synthesis of (±)-sarcodonin G (**36**).¹⁶ Synthetic studies towards a second cyathane target, (±)-cyathin A₄ (**37**),¹⁷ afforded the advanced intermediate **269** in four steps from **240**. On the basis of limited amounts of advanced materials, the final deprotection steps involved in the conversion of **269** to the target structure, (±)-cyathin A₄ (**37**), were not completed.



Reagents: a) DIBALH, Et₂O; b) Dess-Martin periodinane, CH₂Cl₂; c) piperidine, 3Å mol. sieves, PhH, reflux; PhSeCl, THF, -78 °C; H₂O; d) KIO₄, THF, MeOH, H₂O, rt; 3) DBN, PhH, reflux; f) DDQ, CH₂Cl₂-H₂O; g) Al(*i*-PrO)₃, *i*-PrOH, 60 °C; h) VO(acac)₂, *t*-BuOOH, CH₂Cl₂, 0 °C; i) DBN, PhH, reflux; j) PPh₃, *p*-nitrobenzoic acid, DEAD, PhH.

Scheme 79

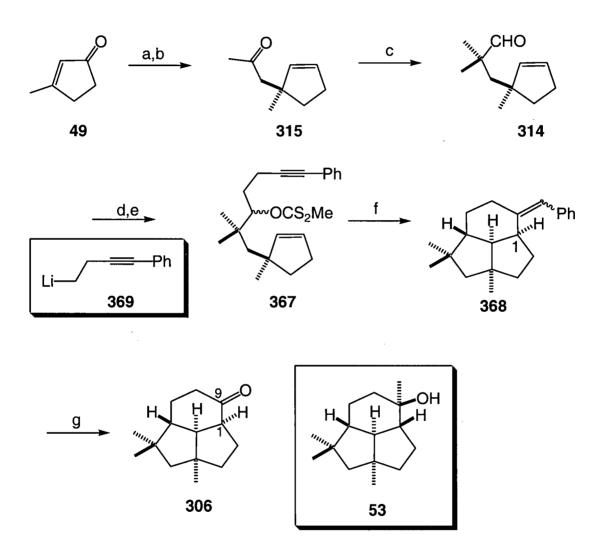
As illustrated by this concise 22-step total synthesis of (\pm) -sarcodonin G (36) from the known ketones 41, our step-wise annulation strategy provides an efficient approach to the cyathane carbon skeleton. In addition, the specific sequence of transformations and the high stereoselectivity of the reactions allowed for excellent control of the relative configuration of all stereogenic centers. Indeed, the anti configuration of the 1,4 angular methyl groups, which was a problematic issue in several previously reported approaches to the cyathanes (see section 2.1.2., vide supra), was well established at an early point in the synthesis. The hydroxyl isopropyl side chain at C-3, which displays an S configuration at C-18, represents a rare structural complexity with regards to cyathane synthetic studies that had yet to be challenged. In our synthesis, the task of fashioning this side chain was successfully accomplished with complete stereoselectivity via a sequence of steps, culminating with a Still-Mitra rearrangement¹⁹ reaction of the cis-fused cyclization product 155. Of note, the use of a SmI₂-mediated ring-expansion reaction to prepare a 5-6-7 tricycle from a 5-6-6 tricycle, was the first reported application of this free-radical reaction, developed by Hasegawa and coworkers,⁸⁹ in natural product synthesis.

The difficulties encountered in our attempt to prepare (\pm)-cyathin A₄ (**37**) from advanced intermediate **240** illustrate the synthetic complexity associated with elaboration of highly functionalized ring-systems. While the selective reduction of the aldehyde function at C-20 over the ketone function at C-10 in **240** was eventually accomplished, the subsequent introduction of an α -face hydroxyl moiety at C-13 represented a serious challenge. In our studies, an intermediate with a β -face hydroxyl moiety at C-13 was obtained and, following a Mitsunobu-type inversion protocol, this substance was transformed into compound **269**, which displays the desired configuration at C–13. Although **269** should provide access to the target structure, \pm -**37** upon removal of the PMB and PNB moieties, further investigations into a more direct method of introducing the α -face C–13 hydroxyl moiety are warranted.

In the second part of this thesis, a tandem radical cyclization was proposed for the assembly of 5-5-6 fused carbocycles. The synthetic study commenced with the commercially available 3-methyl-2-cyclopenten-1-one (**49**), which was converted to ketone **315** in several steps, including a Carroll [3,3] rearrangement reaction (Scheme 80). Subsequent transformation of ketone **315** into aldehyde **314** proved straightforward. A Negishi-type reaction of lithio species **369** with aldehyde **314** afforded a mixture of alcohols, which was converted into the corresponding mixture of xanthates **367**. Bu₃SnH-mediated tandem cyclization of xanthates **367** yielded a mixture of alkenes **368**, which display the 5-5-6 fused tricyclic ring system of the presilphiperfolane framework. Subjection of **368** to oxidative cleavage conditions afforded the known²⁰ ketone (±)-1-epi-9-norpresilphiperfolan-9-one (**306**).

A formal synthesis of the target structure, (\pm) -presilphiperfolan-9-ol (53), from cyclization products 368 *via* ketone 306 was not realized. To obtain 53, the unnatural configuration at C-1, exhibited in 368 and 306, needed to be corrected. Although a substance with the desired relative configuration at C-1 might eventually be obtained in several steps from 306, an alternate strategy that would reverse the undesired stereoselectivity in the second of the tandem cyclizations, which afforded 368, would be ideal. The inherently difficult nature of either of these tasks represents a drawback of our tandem reaction strategy to presilphiperfolane natural product synthesis. Nevertheless, in

the course of this synthetic study, a concise approach to 5-5-6 fused tricyclic ring systems was developed, illustrating the power of tandem free radical cyclizations in carbocycle construction.



Reagents: a) NaBH₄, CeCl₃, MeOH; b) diketene, DMAP, Et₂O, -20 °C; LDA, -78 °C to rt; CCl₄, reflux; c) Ph₃P(OMe)CHLi; THF; 0 °C to rt; TFA, H₂O-CHCl₃, reflux; d) **369**, Et₂O, -78 °C; e) NaH, imidazole, CS₂, THF, reflux; MeI, rt; f) AIBN, Bu₃SnH, PhH, reflux; g) RuCl₃, NaIO₄, MeCN-CCl₄-H₂O.

Scheme 80

IV. EXPERIMENTAL

4.1. General

4.1.1. Data Acquisition and Presentation

Proton nuclear magnetic resonance (¹H NMR) spectra were recorded on Bruker models WH-400 (400 MHz), AM-400 (400 MHz), AV-300 (300 MHz), AV-400 (400 MHz) or AMX-500 (500 MHz) spectrometers using deuteriochloroform (CDCl₃) as a solvent. Signal positions (δ) are given in parts per million (ppm) from tetramethylsilane and were measured relative to that of chloroform (CHCl₃) (δ 7.24 ppm). The multiplicity, number of protons and coupling constants are indicated in parentheses following the chemical shift. In some cases, when mixtures of compounds are present, ratios of integration are given. The abbreviations used in describing multiplicity are: ssinglet, d-doublet, t-triplet, q-quartet, m-multiplet, br-broad. Coupling constants (Jvalues) are given in Hertz (Hz).

Carbon nuclear magnetic resonance (¹³C NMR) spectra were obtained on Bruker models AV-300 (75.5 Hz), AM-400 (100.6 MHz), AV-400 (100.6 MHz), and AMX-500 (125.8 MHz) spectrometers or a Varian model XL-300 (75.5 MHz) spectrometer using CDCl₃ as the solvent. Signal positions (δ) are given in parts per million (ppm) from tetramethylsilane and were measured relative to the signal of CDCl₃ (δ 77.0 ppm). In some cases, the proton and carbon assignments were supported by two-dimensional (¹H, ¹³C)-heteronuclear multiple quantum coherence experiments (HMQC) which were carried out on the Bruker AV-400 and AMX-500 spectrometers. Infrared (IR) spectra were recorded on a Perkin-Elmer model 1710 Fourier transform spectrophotometer with internal calibration on liquid films (sodium chloride plates) or solid pellets (infrared grade potassium bromide). Only selected characteristic absorptions are listed for each compound.

Low and high resolution mass spectra were recorded on a Kratos Concept II HQ or on a Kratos MS 80 mass spectrometer by the UBC MS laboratory. High resolution mass spectra were measured using electron impact ionization (EI) or desorption chemical ionization (DCI) using CH₄ or NH₃. (M⁺) and (M–Me)⁺ ions were detected and analyzed by EI. (M+NH₄)⁺, (M+H)⁺ and (M⁺) ions were detected and analyzed by DCI. Unless otherwise noted, the molecular ion (M⁺) masses are given.

Elemental analysis were performed on a Carlo Erba CHN model 1106 or on a Fisons EA model 1108 elemental analyzer by the Microanalytical Laboratory at UBC.

Unless otherwise stated, all reactions were carried out under an atmosphere of dry argon using glassware that had been oven-dried (~140 °C) and/or flame dried. Glass syringes, stainless steel needles and Teflon[®] cannulae used to handle anhydrous solvents and reagents were oven-dried (~140 °C), cooled in a dessicator and flushed with argon prior to use. Plastic syringes were flushed with argon prior to use. Microsyringes were stored in a dessicator and were flushed with argon prior to use.

Cold temperatures were maintained using the following baths: 0 °C, ice–water; – 10 °C and –20 °C, aqueous calcium chloride–dry ice (17 g, 27 g of CaCl₂/100 mL of H₂O respectively); –78 °C, acetone–dry ice; –98 °C, methanol–dry ice.

Thin layer chromatography (TLC) was performed using commercial aluminum backed silica gel 60 F 254 plates (E. Merck, type 5554, thickness 0.2 mm). Visualization of the chromatograms was accomplished using ultraviolet light (254 nm) followed by heating of the TLC plate after staining with one of the following solutions: (a) vanillin in a sulfuric acid–ethanol mixture (6% vanillin w/v, 4% sulfuric acid v/v, 10% water v/v in EtOH), (b) phosphomolybdic acid in ethanol (20% phosphomolybdic acid w/v, Aldrich), (c) anisaldehyde in a sulfuric acid–ethanol mixture (5% anisaldehyde v/v and 5% sulfuric acid v/v in EtOH). Flash column chromatography was performed using 230–400 mesh silica gel (E, Merck, Silica Gel 60 or Silicycle, Silica Gel).

Gas liquid chromatography (GLC) was performed on Hewlett-Packard models 5880A or 5890 capillary gas chromatographs, both equipped with flame ionization detectors and fused silica columns. The former instrument contained a 25 m \times 0.21 mm column, while the latter chromatograph utilized a 25 m \times 0.20 mm column. Both were coated with HP-5 (crosslinked 5% phenylmethyl silicone).

4.1.2. Solvents and Reagents

All solvents and reagents were purified, dried and/or distilled using standard procedures. Diethyl ether (Et₂O) and tetrahydrofuran (THF) were distilled from sodium/benzophenone, while benzene (C_6H_6) and dichloromethane (CH_2Cl_2) were distilled from calcium hydride, all under an atmosphere of dry argon. Magnesium was added to methanol and, after the mixture had been refluxed, the methanol was distilled from the resulting solution of magnesium methoxide. Solvents were distilled immediately prior to use.

Diisopropylamine, diethylamine, triethylamine, hexamethylphosphoramide (HMPA) and dimethyltetrahydropyrimidinone (DMPU) were distilled from calcium hydride. The reagents were stored in Sure SealTM (Aldrich Chemical Co. Inc.) bottles over 3Å molecular sieves under an atmosphere of argon.

Before use, methyl iodide and deuteriochloroform $(CDCl_3)$ were passed through a short column of basic alumina activity I, which had been over-dried (~140 °C), and then cooled in a dessicator prior to use.

Petroleum ether refers to a mixture of hydrocarbons with a boiling range of 35–60 °C.

Solutions of diisobutylaluminum hydride (DIBALH) in hexanes and *t*butyllithium (*t*-BuLi) in pentane were purchased from Aldrich Chemical Co. Inc. Solutions of methyllithium (MeLi) in diethyl ether and *n*-butyllithium (*n*-BuLi) in hexanes were obtained from Aldrich Chemical Co. Inc. and Acros, and were standardized using diphenylacetic acid as a primary standard using the procedure of Kofron and Baclawski.¹⁴⁴

Potassium hydride (KH) was obtained as a 35% suspension in mineral oil and sodium hydride (NaH) as a 60% dispersion in mineral oil from Aldrich Chemical Co. Inc., and were rinsed free of oil with solvent under a stream of argon prior to use.

Iodine was sublimed and was stored no longer than three months.

Aqueous ammonium chloride–ammonia (NH_4Cl-NH_3) (pH 8) was prepared by the addition of ~50 mL of concentrated aqueous ammonia (28–30%) to 950 mL of a saturated aqueous ammonium chloride (NH_4Cl) solution. Lithium diisopropylamide (LDA) was prepared by the addition of a solution of *n*-butyllithium in hexanes to a solution of diisopropylamine (1.1 equiv.) in dry tetrahydrofuran at -78 °C. The resulting solution was warmed to 0 °C, stirred for 15 min, and cooled back to -78 °C prior to use.

Argon was dried by bubbling it through concentrated sulfuric acid (H_2SO_4) and then passing it through a drying tube packed with potassium hydroxide (KOH) and Drierite[®].

4.2. Total Synthesis of the Cyathane Diterpenoid (±)-Sarcodonin G

Preparation of Ethyl (Z)-3-Trimethylgermyl-2-pentenoate (165)



To stirred cold (-10 °C) dry THF (50 mL) was added cold (0 °C) Me₃GeH (14.1 g, 0.119 mol) *via* a cannula. A solution of *t*-BuLi (1.7 M in pentane, 87.5 mL, 0.149 mol) was added *via* a syringe over 5 min. The resulting yellow solution was stirred at -10 °C for 5 min to afford a colourless solution. A solution of MeLi (1.4 M in Et₂O, 106.0 mL, 0.149 mol) was added and the resulting mixture was transferred *via* a cannula to a cold (-78 °C) stirred suspension of CuCN (13.3 g, 0.149 mol) in dry THF (20 mL). The resulting suspension was stirred a -78 °C for 1 h to afford a colourless solution. A solution of ethyl 2-pentynoate (10.0 g, 0.079 mol) in THF (5 mL) was added *via* a cannula and the reaction mixture was stirred at -78 °C for 45 min. AcOH (12.4 g, 0.206 mol) was added *via* a syringe. The resulting heterogeneous mixture was stirred for 5 min and then poured into stirred aqueous NH₄Cl–NH₃ (pH 8, 500 mL). Et₂O (500 mL) was added and the mixture was deep blue. The layers were separated and the aqueous layer was extracted with Et₂O (2 × 200 mL). The combined organic extracts were washed with H₂O (2 × 200 mL) and

brine (2 × 200 mL), dried over anhydrous MgSO₄, and concentrated under reduced pressure. The crude oil was subjected to flash chromatography (600 g of silica gel, 40:1 petroleum ether–Et₂O) to afford two fractions, which were concentrated under reduced pressure. The first fraction to be eluted afforded 13.7 g (71%) of ethyl (*Z*)-3-trimethylgermyl-2-pentenoate (**165**) as a colourless oil. The second fraction afforded 3.91 g (20%) of ethyl (*E*)-3-trimethylgermyl-2-pentenoate (**174**) as a colourless oil.

Ethyl (Z)-3-trimethylgermyl-2-pentenoate (165) exhibited the following spectral data:

IR (neat): 2968, 1718, 1607 cm^{-1} .

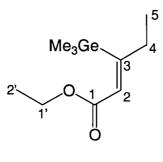
¹H NMR (CDCl₃, 400 MHz) δ: 0.29 (s, 9H), 0.99 (t, 3H, *J* = 7.3 Hz), 1.24 (t, 3H, *J* = 7.0 Hz), 2.30 (q, 2H, *J* = 7.3 Hz), 4.12 (q, 2H, *J* = 7.0 Hz), 6.21 (s, 1H).

¹³C NMR (CDCl₃, 100.6 MHz) δ: -0.4, 13.5, 14.2, 31.9, 59.9, 126.6, 166.8, 171.4.

Exact mass calcd for $C_{10}H_{21}O_2^{-74}$ Ge (M+H)⁺: 247.0753. Found: 247.0762.

Anal. calcd for C₁₀H₂₀O₂Ge : C 49.05, H 8.23. Found: C 48.84, H 8.10.

Table 9. ¹H NMR (CDCl₃, 400 MHz) Data for Ethyl (*Z*)-3-Trimethylgermyl-2-pentenoate (**165**): NOED Experiment.

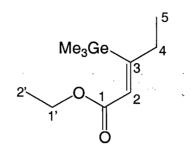


165

Assignments H–x	¹ Η NMR δ ppm (Multiplicity, <i>J</i> (Hz))	Observed NOEs
$H-2^{a}$	6.21 (s)	H4, H5
$H-4^a$	2.30 (q, 7.3)	H–5, H–2, –GeMe ₃
H–5	0.99 (t, 7.3)	
H–1 ^{<i>i</i>a}	4.12 (q, 7.0)	H–2'
H–2'	1.24 (t, 7.0)	
-GeMe ₃ ^a	0.29 (s)	H-4

^a Irradiation of this signal generated the corresponding NOEs in the right hand column

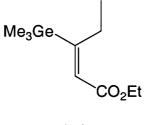
Table 10. ¹³C NMR (CDCl₃, 100.6 MHz) and ¹H NMR (CDCl₃, 400 MHz) Data for Ethyl (*Z*)-3-Trimethylgermyl-2-pentenoate (**165**): HMQC Experiment.



1	(F
	65
_	

Assignments C-x	¹³ C NMR δ ppm	APT	HMQC ¹ H NMR Correlations (δ ppm)
C-1	166.8 or 171.4	C or CH ₂	
C-2	126.6	CH or CH ₃	H–2 (6.21)
C-3	166.8 or 171.4	C or CH ₂	
C-4	31.9	C or CH ₂	H-4 (2.30)
C-5	13.5	CH or CH ₃	H–5 (0.99)
C-1'	59.9	C or CH ₂	H–1' (4.12)
C–2'	14.2	CH or CH ₃	H-2' (1.24)
-GeMe ₃	-0.4	CH or CH ₃	-GeMe ₃ (0.29)

Ethyl (E)-3-trimethylgermyl-2-pentenoate (174) exhibited the following spectral data:





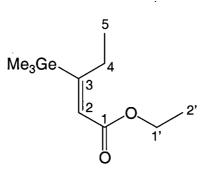
IR (neat): 2975, 1718, 1608 cm⁻¹.

¹H NMR (CDCl₃, 400 MHz) δ: 0.23 (s, 9H), 1.00 (t, 3H, *J* = 7.5 Hz), 1.24 (t, 3H, *J* = 7.2 Hz), 2.73 (q, 2H, *J* = 7.5 Hz), 4.11 (q, 2H, *J* = 7.2 Hz), 5.89 (s, 1H).

¹³C NMR (CDCl₃, 100.6 MHz) δ: -2.1, 13.8, 14.2, 25.4, 59.5, 124.5, 164.9, 171.3.

Exact mass calcd for $C_{10}H_{21}O_2^{-74}$ Ge (M+H)⁺: 247.0753. Found: 247.0758.

Table 11. ¹H NMR (CDCl₃, 400 MHz) Data for Ethyl (*E*)-3-Trimethylgermyl-2-pentenoate (**174**): NOED Experiment.

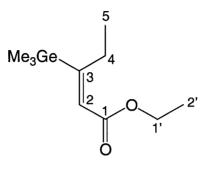


174

Assignments H–x	¹ Η NMR δ ppm (Multiplicity, <i>J</i> (Hz))	Observed NOEs
$H-2^{a}$	5.89 (s)	-GeMe ₃
$H-4^a$	2.73 (q, 7.5)	H–5, –GeMe ₃
H–5	1.00 (t, 7.5)	
H-1 ^a	4.11 (q, 7.2)	H–2'
H–2'	1.24 (t, 7.2)	
GeMe ₃ ^a	0.23 (s)	H–2, H–4, H–5

^{*a*} Irradiation of this signal generated the corresponding NOEs in the right hand column

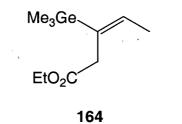
Table 12. ¹³C NMR (CDCl₃, 100.6 MHz) and ¹H NMR (CDCl₃, 400 MHz) Data for Ethyl (*Z*)-3-Trimethylgermyl-2-pentenoate (**174**): HMQC Experiment.



1	7	4

Assignments C-x	¹³ C NMR δ ppm	АРТ	HMQC ¹ H NMR Correlations (δ ppm)
C-1	164.9 or 171.3	C or CH ₂	
C-2	124.5	CH or CH ₃	H–2 (5.89)
C-3	164.9 or 171.3	C or CH ₂	
C-4	25.4	C or CH ₂	H-4 (2.73)
C-5	13.8	CH or CH ₃	H–5 (1.00)
C-1'	59.5	C or CH ₂	H–1' (4.11)
C-2'	14.2	CH or CH ₃	H-2' (1.24)
-GeMe ₃	-2.1	CH or CH ₃	-GeMe ₃ (0.23)

Preparation of Ethyl (*E*)-3-Trimethylgermyl-3-pentenoate (164)



To a cold (-78 °C), stirred solution of LDA (130 mmol) in dry THF (500 mL) was added HMPA (23.3 g, 130 mmol). The mixture was stirred for 30 min and a solution of ethyl (*Z*)-3-trimethylgermyl-2-pentenoate (**165**) (13.0 g, 53.1 mmol) in dry THF (50 mL) was added *via* a cannula. The resulting mixture was stirred for 30 min at -78 °C, warmed to 0 °C for 1 h and then recooled to -78 °C for 15 min. The mixture was transferred, *via* a cannula, to a cold (-98 °C) stirred solution of acetic acid (31.9 g, 0.531 mol) in dry Et₂O (500 mL). The mixture was allowed to warm to room temperature and saturated aqueous NaHCO₃ (500 mL) was added. The layers were separated and the aqueous layer was extracted with Et₂O (3 × 500 mL). The combined organic extracts were washed with brine (300 mL) and dried over anhydrous MgSO₄. The solvent was removed under reduced pressure and the resulting material was purified by flash chromatography (500 g of silica gel, 20:1 petroleum ether-Et₂O) to yield 11.8 g (91%) of ethyl (*E*)-3-trimethylgermyl-3-pentenoate (**164**) as a colourless oil.

IR (neat): 2976, 1737, 1626 cm⁻¹.

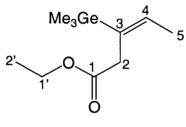
¹H NMR (CDCl₃, 400 MHz) δ: 1.15 (s, 9H), 1.21 (t, 3H, J = 7.2 Hz), 1.69 (d, 3H, J = 6.7 Hz), 3.16 (s, 2H), 4.07 (q, 2H, J = 7.2 Hz), 5.87 (q, 1H, J = 6.7 Hz).

¹³C NMR (CDCl₃, 75.2 MHz) δ: -2.0, 14.1, 14.4, 35.3, 60.4, 135.5, 135.7, 171.8.

Exact mass calcd for $C_{10}H_{21}O_2^{74}$ Ge (M+H)⁺: 247.0753. Found: 247.0764.

Anal. calcd for C₁₀H₂₀O₂GeI: C 49.05, H 8.23. Found: C 49.15, H 8.23.

Table 13. ¹H NMR (CDCl₃, 400 MHz) Data for Ethyl (*E*)-3-Trimethylgermyl-3-pentenoate (**164**): NOED Experiment.

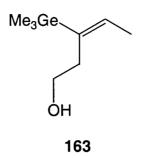


1	.64

_Assignments H–x	¹ H NMR δ ppm (Multiplicity, J (Hz))	Observed NOEs
$H-2^a$	3.16 (s)	H–5, –GeMe ₃
H-4 ^a	5.87 (q, 6.7)	H–5, –GeMe ₃
H–5 ^a	1.69 (d, 6.7)	H–2, H–4
H–1'	4.07 (q, 7.2)	
H–2'	1.21 (t, 7.2)	- ·
-GeMe ₃ ^{<i>a</i>}	1.15 (s)	H–2, H–4

^{*a*} Irradiation of this signal generated the corresponding NOEs in the right hand column

Preparation of (E)-3-Trimethylgermyl-3-penten-1-ol (163)



To a cold (-78 °C), stirred solution of ethyl (*E*)-3-trimethylgermyl-3-pentenoate (**164**) (8.0 g, 32.7 mmol) in dry Et₂O (300 mL) was added a solution of DIBALH (1.0 M in hexanes, 82.5 mL, 82.5 mmol). The resulting mixture was stirred at -78 °C for 1 h, warmed to 0 °C and then stirred for an additional h. Aqueous NH₄Cl-NH₃ (pH 8, 8 mL) was added, and the resulting white slurry was warmed to room temperature and was then stirred for 1 h. Solid anhydrous MgSO₄ (2.0 g) was added and the mixture was stirred for 1 h. The slurry was filtered through a column of Florisil[®] (50 g) and the column was washed with Et₂O (3 × 200 mL). The solvent of the combined eluates was removed under reduced pressure and the resulting material was purified by flash chromatography (400 g of silica gel, 8:1 pentane-Et₂O) to give 5.85 g (88%) of (*E*)-trimethylgermyl-3-penten-1-ol (**163**) as a colourless oil.

IR (neat): 3331, 1622, 1235, 1043, 824, 754, 597, 570 cm⁻¹.

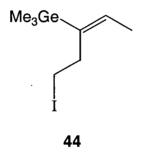
¹H NMR (CDCl₃, 400 MHz) δ: 0.17 (s, 9H), 1.28 (t, 1H, *J* = 6.0 Hz), 1.72 (d, 3H, *J* = 6.6 Hz), 2.51 (t, 2H, *J* = 7.0 Hz), 3.58 (td, 2H, *J* = 7.0, 6.0 Hz), 5.88 (q, 1H, *J* = 6.6 Hz).

¹³C NMR (CDCl₃, 75.2 MHz) δ: -1.7, 14.4, 33.4, 61.7, 135.1, 138.9.

Exact mass calcd for C₇H₁₅⁷⁴GeO (M⁺–Me): 189.0335. Found: 189.0336.

Anal. calcd for C₈H₁₈GeO: C 47.38, H 8.95. Found: C 47.57, H 9.12.

Preparation of (E)-5-Iodo-3-trimethylgermyl-2-pentene (44)



I₂ (6.20 g, 24.4 mmol) was added to a solution of Ph₃P (6.41 g, 24.4 mmol) in dry CH₂Cl₂ (200 mL) and the mixture was stirred for 20 min. (*E*)-3-Trimethylgermyl-3-penten-1-ol (163) (4.13 g, 20.4 mmol) and dry Et₃N (3.4 mL, 24 mmol) were added as a solution in dry CH₂Cl₂ (20 mL) and the resulting mixture was stirred for 18 h. Most of the solvent was removed under reduced pressure and the residual material was diluted with pentane (100 mL). The mixture was filtered through a sintered glass funnel (8 cm diameter) containing Florisil[®] (10 cm deep) on top of a thin layer of Celite[®]. The column was flushed with pentane (600 mL). The solvent of the combined eluates was removed under

reduced pressure. The residual material was purified by distillation (48–54 °C / 0.3 Torr) to give 6.02 g (95%) of (*E*)-5-iodo-3-trimethylgermyl-2-pentene (44) as a colourless oil. IR (neat): 1619, 1236, 1168, 823, 756, 597, 567 cm⁻¹.

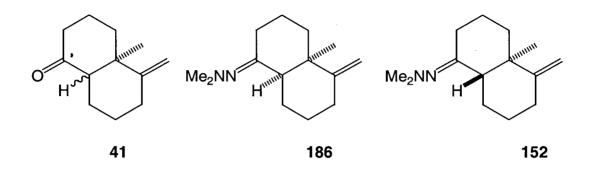
¹H NMR (CDCl₃, 400 MHz) δ : 0.18 (s, 9H), 1.67 (d, 3H, J = 6.7 Hz), 2.78 (t, 2H, J = 8.5 Hz), 3.02 (t, 2H, J = 8.5 Hz), 5.82 (q, 1H, J = 6.7 Hz).

¹³C NMR (CDCl₃, 75.2 MHz) δ: -1.8, 3.9, 14.4, 35.2, 134.5, 142.2.

Exact mass calcd for $C_7H_{14}^{-74}$ GeI (M⁺-Me): 298.9352. Found: 298.9359.

Anal. calcd for C₈H₁₇GeI: C 30.73, H 5.48. Found: C 30.86, H 5.74.

Preparation of (1*S**,6*R**)-6-Methyl-7-methylidenebicyclo[4.4.0]decan-2-one Dimethylhydrazone (152)



1,1-Dimethylhydrazine (7.50 g, 123 mmol) was added to a stirred solution of the bicyclic ketone 41¹⁸ (3.5:1 mixture of the *trans*- and *cis*-fused isomers, 10.9 g, 61.6 mmol) in dry benzene (300 mL). A catalytic amount of 10-camphorsulfonic acid (1.1 g, 4.7 mmol) was added and the mixture was refluxed for 72 h employing a Dean-Stark trap. The excess 1,1-dimethylhydrazine and most of the solvent were removed by distillation under Ar (90 °C). The remaining material was diluted with Et₂O (100 mL) and saturated aqueous NaHCO₃ (100 mL) was added. The resulting layers were separated and the aqueous layer was extracted with Et₂O (3 × 100 mL). The combined organic layers were dried over anhydrous MgSO₄, concentrated under reduced pressure and purified by flash chromatography (600 g of silica gel, 5:1 pentane–Et₂O) to yield 5.82 g (42.9%) of the *trans*-fused bicyclic hydrazone **152** and 5.32 g (39.2%) of the *cis*-fused bicyclic hydrazone **186** as colourless oils.

Isomerization of $(1R^*, 6R^*)$ -6-Methyl-7-methylidenebicyclo[4.4.0]decan-2-one Dimethylhydrazone (186)

1,1-Dimethylhydrazine (3.0 g, 50 mmol) was added to a stirred solution of the *cis*-fused bicyclic hydrazone **186** (5.32 g, 24.1 mmol) in dry benzene (100 mL). A catalytic amount of 10-camphorsulfonic acid (50 mg, 0.2 mmol) was added and the mixture was refluxed for 48 h employing a Dean-Stark trap. The excess 1,1-dimethylhydrazine and most of the solvent were removed by distillation under Ar (90 °C). The remaining material was diluted with Et₂O (100 mL) and saturated aqueous NaHCO₃ (100 mL) was added. The resulting layers were separated and the aqueous layer was extracted with Et₂O (3 × 100 mL). The combined organic layers were dried over anhydrous MgSO₄, concentrated under reduced pressure and purified by flash chromatography (300 g of silica gel, 5:1 pentane–Et₂O) to yield 2.55 g (48.3%) of the *trans*-fused bicyclic hydrazone **186** was recycled twice though the epimerization step to yield a total of 9.61 g (71%) of the *trans*-fused bicyclic hydrazone **152** from 10.9 g of the mixture of ketones **41** (*vide supra*).

The *trans*-fused bicyclic hydrazone **152** exhibited the following spectral data:

IR (neat): 1638, 1446, 1373, 1022, 965, 894 cm⁻¹.

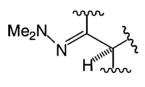
¹H NMR (CDCl₃, 400 MHz) δ: 0.88 (s, 3H), 1.22–1.26 (m, 1H), 1.54–1.79 (m, 6H), 1.79–1.89 (m, 2H), 1.95 (dd, 1H, *J* = 11.7, 3.7 Hz), 2.09–2.16 (m, 1H), 2.25–2.35 (m, 1H), 2.4 (s, 6H), 3.26–3.32 (m, 1H), 4.63–4.68 (m, 2H).

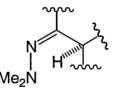
¹³C NMR (CDCl₃, 75.2 MHz) δ: 18.1, 22.1, 23.1, 27.4, 28.5, 32.6, 36.5, 43.1, 47.8, 52.5 (two signals superimposed), 105.3, 157.1, 169.5.

Exact mass calcd for C₁₄H₂₄N₂: 220.1940. Found: 220.1940.

Anal. calcd for C₁₄H₂₄N₂: C 76.31, H 10.98, N 12.71. Found: C 76.16, H 10.86, N 12.72.

The *cis*-fused bicyclic hydrazone **186** exists as a mixture of isomers with respect to the configuration of the imine double bond.





major

minor

The mixture of *cis*-fused bicyclic hydrazone **186** isomers exhibited the following spectral data:

IR (neat): 1635, 1446, 1374, 1021, 967, 892 cm⁻¹.

¹H NMR (CDCl₃, 400 MHz) δ: 1.06 (minor) and 1.08 (major) (s, s, 3H total), 1.2–2.5 (m, 12H), 2.36 (minor) and 2.41 (major) (s, s, 6H total), 3.07–3.17 (major) and 3.19–3.26 (minor) (m, m, 1H total), 4.66–4.73 (m, 2H).

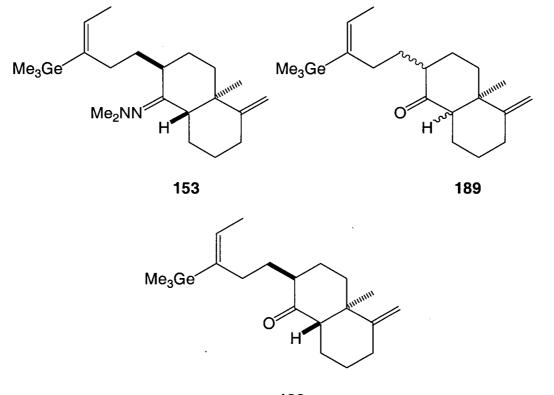
¹³C NMR (CDCl₃, 75.2 MHz) δ: 21.6, 22.7, 23.4, 23.7, 27.2, 27.3, 27.4, 28.7, 31.1, 31.5, 31.7, 32.8, 40.7, 41.0, 45.5, 47.4, 47.7 (two signals superimposed), 52.8 (two signals superimposed), 107.2, 155.1, 155.4, 172.0, 174.4 (three signals are not accounted for).

٥

Exact mass calcd for C₁₄H₂₄N₂: 220.1940. Found: 220.1935.

Preparation of (1S*,3R*,6R*)-6-Methyl-7-methylidene-3-[(3E)-3-trimethylgermyl-3-

pentenyl]bicyclo[4.4.0]decan-2-one (188)



188

To a stirred solution of *t*-BuOK (5.00 g, 44.6 mmol) in dry THF (150 mL) was added dry i-Pr₂NH (4.52 g, 44.6 mmol). The mixture was cooled to -78 °C. A solution of BuLi (1.6 M in hexanes, 27.9 mL, 44.6 mmol) was added and the mixture was stirred for 30 min. The *trans*-fused hydrazone **152** (4.93 g, 22.3 mmol) was added as a solution in dry THF (10 mL) and the mixture was stirred at -78 °C for 1 h. Dry DMPU (5.72 g, 44.6 mmol) and a solution of freshly distilled (*E*)-5-iodo-3-trimethylgermyl-2-pentene (**44**) (10.5 g, 33.5 mmol) in dry THF (10 mL) were sequentially added and the mixture was stirred at -78 °C for 1.5 h. Aqueous NH₄Cl–NH₃ (pH 8, 200 mL) was added and the mixture was added to warm to room temperature. Et₂O (200 mL) was added and the

resulting layers were separated. The aqueous layer was extracted with Et₂O (3×200 mL), and the combined organic layers were washed with brine (200 mL). The solvent was removed under reduced pressure and the resulting crude hydrazone **153** was taken up in THF (30 mL). H₂O (7 mL) and solid NaOAc \exists 3H₂O (13.8 g, 0.101 mol) were added and the mixture was stirred for 5 min. AcOH (43.0 g, 0.716 mol) was added and the mixture was heated to 75 °C for 18 h. The mixture was cooled to room temperature and saturated aqueous NaHCO₃ (60 mL) was added. Solid NaHCO₃ was added until the solution was alkaline to litmus paper. Et₂O (150 mL) was added to the mixture and the resulting layers were separated. The aqueous layer was extracted with Et₂O (3×150 mL). The combined organic layers were washed with brine (150 mL) and dried over anhydrous MgSO₄. The solvent was removed under reduced pressure and the resulting material was purified by flash chromatography (600 g of silica gel, 40:1 pentane–Et₂O) to afford 5.60 g (69.0%) of a mixture of ketones **189**, of which ketone **188** was the major isomer.

For characterization, pure ketone **188** was obtained by flash chromatography (50:1 pentane– Et_2O) of a small amount of the mixture of ketones **189**.

Ketone 188 exhibited the following spectral data:

IR (neat): 1709, 1639, 1628, 1448, 1376, 1234, 952 cm⁻¹.

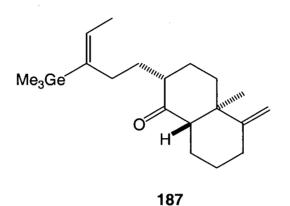
¹H NMR (CDCl₃, 400 MHz) δ: 0.13 (s, 9H), 0.91 (s, 3H), 1.18–1.30 (m, 1H), 1.43–1.56 (m, 1H), 1.56–1.69 (m, includes 3-proton doublet at δ 1.62, *J* = 6.6 Hz, 7H total), 1.82–1.89 (m, 2H), 1.91–2.00 m, 1H), 2.01–2.08 (m, 2H), 2.10–2.40 (m, 5H), 4.65–4.72 (m, 2H), 5.67 (q, 1H, *J* = 6.6 Hz).

¹³C NMR (CDCl₃, 75.2 MHz) δ: -1.7, 13.9, 18.8, 20.9, 26.6 (two signals superimposed), 28.2, 31.4, 32.0, 32.2, 44.9, 50.0, 54.1, 105.8, 132.2, 142.4, 155.7, 215.3.

Exact mass calcd for $C_{20}H_{34}^{-74}$ GeO: 364.1822. Found: 364.1818.

Anal. calcd for C₂₀H₃₄GeO: C 66.16, H 9.44. Found C 66.35, H 9.55.

Preparation of (1*S**,3*S**,6*R**)-6-Methyl-7-methylidene-3-[(3*E*)-3-trimethylgermyl-3pentenyl]bicyclo[4.4.0]decan-2-one (187)



To a stirred solution of NaOMe (7.83 g, 145 mmol) in dry MeOH (300 mL) was added a solution of the mixture of ketones **189** (17.4 g, 48.0 mmol) in dry MeOH (100 mL). The mixture was heated to 65 °C for 3 h and then cooled to room temperature. H₂O (300 mL) and Et₂O (500 mL) were added, and the layers were separated. The aqueous layer was extracted with Et₂O (3×300 mL). The combined organic layers were washed with brine (200 mL) and dried over anhydrous MgSO₄. The solvent was removed under reduced pressure and the resulting material was purified by flash chromatography (1 kg of silica gel, 40:1 pentane–Et₂O) to yield 11.5 g (66.2%) of ketone **187** as well as 3.9 g (22.4%) of a mixture of three other diastereomers. This mixture of isomers was resubmitted to the same isomerization reaction to yield an additional 2.71 g of **187** for a total overall yield of 14.21 g (82%).

IR (neat): 1713, 1638, 1623, 1447, 1376, 1234, 1098, 940, 895, 823, 754 cm⁻¹.

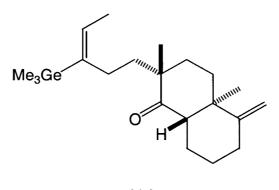
¹H NMR (CDCl₃, 400 MHz) δ: 0.15 (s, 9H), 0.86 (s, 3H), 1.05–1.17 (m, 1H), 1.17–1.31 (m, 2H), 1.53–1.69 (m, includes 3-proton doublet at δ 1.66, *J* = 6.6 Hz, 5H total), 1.74–1.89 (m, 3H), 1.90–2.00 (m, 1H), 2.10–2.32 (m, 7H), 4.69 (m, 2H), 5.68 (q, 1H, *J* = 6.6 Hz).

¹³C NMR (CDCl₃, 75.2 MHz) δ: -1.7, 14.0, 18.9, 21.0, 26.6, 27.7, 29.0, 30.0, 32.2, 36.0, 45.4, 49.7, 58.3, 105.6, 131.9, 143.3, 155.8, 212.7.

Exact mass calcd for $C_{19}H_{31}^{-74}$ GeO (M⁺–Me): 349.1587. Found: 349.1590.

Anal. calcd for C₂₀H₃₄GeO: C 66.66, H 9.44. Found: C 66.25, H 9.38.

Preparation of (1*S**,3*S**,6*R**)-3,6-Dimethyl-7-methylidene-3-[(3*E*)-3-trimethylgermyl-3-pentenyl]bicyclo[4.4.0]decan-2-one (194)





To a cold (-78 °C), stirred solution of dry Et_2NH (4.35 g, 59.5 mmol) in dry THF (100 mL) was added a solution of BuLi (1.6 M in hexanes, 30.2 mL, 48.3 mmol). The mixture was warmed to 0 °C for 30 min and cooled to -78 °C. A solution of ketone **187** (13.5 g, 37.2 mmol) in dry THF (20 mL) was added and the resulting mixture was warmed to 0 °C and stirred for 1 h. MeI (107 g, 750 mmol) was added and the mixture was warmed to room temperature. The mixture was stirred for 1.5 h and aqueous NH_4Cl-NH_3 (pH 8, 100 mL) was added. The resulting layers were separated and the aqueous layer was extracted with Et_2O (3 × 100 mL). The combined organic layers were washed with brine (100 mL) and dried over anhydrous MgSO₄. The solvent was removed under reduced pressure and the resulting crude material was purified by flash chromatography (500 g of silica gel, 50:1 pentane- Et_2O) to give 11.86 g (84.7%) of ketone **194** as a colourless oil.

IR (neat): 1704, 1637, 1623, 1457, 1379, 1234, 1118, 896, 824, 754, 597, 569 cm⁻¹.

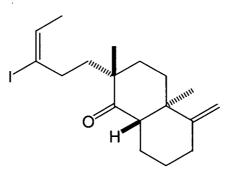
¹H NMR (CDCl₃, 400 MHz) δ: 0.17 (s, 9H), 0.86 (s, 3H), 1.17 (s, 3H), 1.18–1.31 (m, 1H), 1.31–1.48 (m, 2H), 1.48–1.61 (m, 1H), 1.63–1.79 (m, includes 3-proton doublet at δ 1.70, *J* = 6.6 Hz, 6H total), 1.83–1.96 (m, 2H), 2.04–2.33 (m, 5H), 2.44 (dd, 1H, *J* = 11.9, 3.5 Hz), 4.68–4.74 (m, 2H), 5.67 (q, 1H, *J* = 6.6 Hz).

¹³C NMR (CDCl₃, 75.2 MHz) δ: -1.7, 14.0, 18.7, 21.2, 24.1 (two signals superimposed), 26.7, 32.1, 32.2, 33.4, 38.0, 44.6, 46.8, 53.7, 105.7, 131.6, 143.5, 155.9, 215.6.

Exact mass calcd for $C_{21}H_{36}^{-74}$ GeO: 378.1978. Found: 378.1986.

Anal. calcd for C₂₁H₃₆GeO: C 66.89, H 9.62. Found: C 66.63, H 9.69.

Preparation of $(1S^*, 3S^*, 6R^*)$ -3-[(3E)-3-Iodo-3-pentenyl]-3,6-dimethyl-7-methylidenebicyclo[4.4.0]decan-2-one (154)





To a cold (0 °C), stirred solution of ketone **194** (11.4 g, 30.5 mmol) in dry CH₂Cl₂ (250 mL) was added solid *N*-iodosuccinimide (8.24 g, 36.6 mmol) in one portion and the mixture was stirred for 1 h. This solution was then poured into a cold (0 °C) mixture of aqueous Na₂S₂O₃ (1.0 M, 100 mL) and saturated aqueous NaHCO₃ (100 mL). The biphasic mixture was stirred for 0.5 h and the layers were separated. The aqueous layer was extracted with Et₂O (3 × 15 mL) and the combined organic layers were dried over anhydrous MgSO₄. After removal of the solvent under reduced pressure, the remaining crude material was purified by flash chromatography (500 g of silica gel, 40:1 pentane–Et₂O) to give 10.55 g (91%) of the iodide **154** as a colourless oil.

IR (neat): 1703, 1637, 1448, 1380, 895 cm⁻¹.

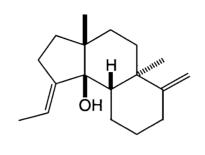
¹H NMR (CDCl₃, 400 MHz) δ: 0.85 (s, 3H), 1.17 (s, 3H), 1.17–1.30 (m, 1H), 1.48–1.76 (m, includes 3-proton doublet at δ 1.67, *J* = 7.0 Hz, 9H total), 1.81–1.91 (m, 2 H), 2.04–

2.17 (m, 2H), 2.21–2.32 (m, 1H), 2.35–2.51 (m, 3H), 4.67–4.73 (m, 2H), 6.18 (q, 1H, *J* = 7.0 Hz).

¹³C NMR (CDCl₃, 75.2 MHz) δ: 16.2, 18.7, 21.2, 24.1, 26.6, 32.0, 32.1, 33.6, 33.8, 37.6, 44.8, 46.4, 53.7, 103.0, 105.8, 135.5, 155.7, 215.5.

Exact mass calcd for $C_{18}H_{27}IO$: 386.1107. Found: 386.1101.

Anal. calcd for C₁₈H₂₇IO: C 55.96, H 7.04. Found: C 56.09, H 7.01.





To a cold (-78 °C), stirred solution of iodide **154** (10.4 g, 26.9 mmol) in dry THF (200 mL) was added a solution of BuLi (1.6 M in hexanes, 25.3 mL, 40.4 mmol) *via* syringe. After the mixture had been stirred for 40 min, H₂O (100 mL) and Et₂O (100 mL) were added. The biphasic mixture was allowed to warm to room temperature over 0.5 h and the layers were separated. The aqueous layer was extracted with the Et₂O ($3 \times 100 \text{ mL}$) and the combined organic layers were washed with brine (100 mL) and dried over anhydrous MgSO₄. The solvent was removed under reduced pressure and the resulting crude oil was purified by flash chromatography (500 g of silica gel, 20:1 pentane–Et₂O) to give 5.99 g (86%) of the alcohol **155** as a colourless oil.

IR (neat): 3478, 1660, 1635, 1448, 1375, 1034, 1000, 986, 949, 909, 891, 864, 819 cm⁻¹.

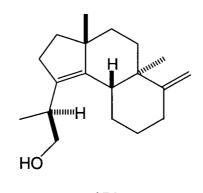
¹H NMR (CDCl₃, 400 MHz) δ : 0.92 (s, 3H), 1.01 (s, 3H), 1.17 (s, 1H, exchanges with D₂O), 1.15–1.31 (m, 2H), 1.31–1.42 (m, 2H), 1.52–1.78 (m, 4H), 1.59 (d, 3H, J = 6.7

Hz), 1.81–1.95 (m, 2H), 2.07–2.18 (m, 2H), 2.27–2.40 (m, 3H), 4.56–4.58 (m, 2H), 5.88 (q, 1H, *J* = 6.7 Hz).

¹³C NMR (CDCl₃, 75.2 MHz) δ: 15.2, 19.3, 19.8, 22.6, 26.0, 29.3, 30.7, 32.4, 33.4, 36.4, 41.1, 47.0, 50.4, 81.5, 104.3, 121.5, 149.7, 158.9.

Exact mass calcd for C₁₈H₂₈O: 260.2140. Found: 260.2144.

Preparation of $(1S^*, 6S^*, 9R^*)$ -3-[(S^*) -2-Hydroxy-1-methylethyl]-6,9-dimethyl-10methylidenetricyclo[7.4.0.0^{2,6}]tridec-2-ene (151)



151

To a stirred suspension of KH (3.98 g, 34.7 mmol) in dry THF (80 mL) was added a solution of alcohol **155** (4.52 g, 17.4 mmol) in dry THF (40 mL). The mixture was stirred for 1 h and a solution of 18-cr-6 ether (9.17 g, 34.7 mmol) in THF (40 mL) was added. After the mixture had been stirred 0.5 h, a solution of Bu₃SnCH₂I (18.71 g, 43.4 mmol) in THF (40 mL) was added *via* a cannula. The resulting thick white slurry was stirred for 1 h at room temperature and then was cooled to -78 °C. A solution of BuLi (1.6 M in hexanes, 48.8 mL, 78.1 mmol) was added, and the mixture was allowed to warm to room temperature over 30 min. H₂O (120 mL) and Et₂O (120 mL) were added, and the resulting layers were separated. The aqueous layer was extracted with Et₂O (3 × 120 mL), and the combined organic layers were dried over anhydrous MgSO₄. The solvent was removed under reduced pressure, and the residual material was purified by flash chromatography (200 g of silica gel, 7:1 pentane–Et₂O) to yield 4.10 g (86%) of the alcohol **151** as a colourless oil.

IR (neat): 3388, 1633, 1455, 1373, 1032, 894, 788 cm⁻¹.

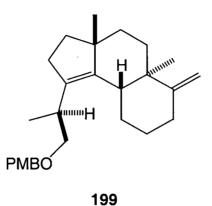
¹H NMR (CDCl₃, 400 MHz) δ: 0.92 (s, 3H), 0.96 (d, 3H, *J* = 6.8 Hz), 1.03 (s, 3H), 1.16– 1.31 (m, 2H), 1.48–1.59 (m, 4H), 1.59–1.68 (m, 1H), 1.75–1.91 (m, 3H), 1.91–1.99 (m, 1H), 2.10–2.36 (m, 5H), 3.18–3.29 (m, 1H), 3.38–3.45 (m, 2H), 4.64–4.68 (m, 2H).

¹³C NMR (CDCl₃, 75.2 MHz) δ: 16.1, 18.4, 24.7, 27.8, 28.5, 28.6, 33.1, 34.0, 34.8, 37.1, 38.8, 42.1, 46.6, 49.3, 66.0, 105.6, 134.5, 144.0, 157.6.

Exact mass calcd for C₁₉H₃₀O: 274.2297. Found: 274.2289.

Anal. calcd for C₁₉H₃₀O: C 83.15 H 11.02. Found: C 82.76 H 11.33.

Preparation of $(1S^*, 6S^*, 9R^*)$ -3-[(S^*) -2-(4-Methoxyphenyl)methoxy-1-methylethyl]-6,9-dimethyl-10-methylidenetricyclo[7.4.0.0^{2,6}]tridec-2-ene (199)



To a stirred suspension of KH (354 mg, 8.81 mmol) in dry THF (40 mL) was added a solution of alcohol **151** (1.72 g, 6.28 mmol) in dry THF (40 mL) *via* a cannula. After the mixture had been stirred for 1 h, 4-methoxybenzyl chloride (1.38 g, 8.81 mmol) and solid Bu₄NI (465 mg, 1.26 mmol) were sequentially added. The mixture was stirred overnight at room temperature. H₂O (60 mL) and Et₂O (60 mL) were added and the resulting layers were separated. The aqueous layer was extracted with Et₂O (3×50 mL) and the combined organic layers were washed with brine (50 mL) and dried over anhydrous MgSO₄. The solvent was removed under reduced pressure and the crude oil was purified by flash chromatography (50 g of silica gel, 25:1 pentane–Et₂O) to yield 2.25 g (91%) of ether **199** as a colourless oil.

IR (neat): 2933, 2857, 1631, 1613, 1513, 1247 cm⁻¹.

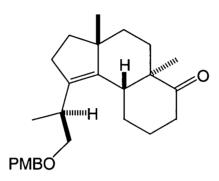
¹H NMR (CDCl₃, 400 MHz) δ: 0.92 (s, 3H), 1.00 (d, 3H, *J* = 6.4 Hz), 1.01 (s, 3H), 1.41– 1.68 (m, 6H), 1.75–1.92 (m, 3H), 1.95–2.03 (m, 1H), 2.12–2.40 (m, 5H), 3.22–3.40 (m, 3H), 3.78 (s, 3H), 4.38–4.46 (m, 2H), 4.62–4.70 (m, 2H), 6.86 (d, 2H, *J* = 8.9 Hz), 7.23 (d, 2H, *J* = 8.9 Hz).

¹³C NMR (CDCl₃, 75.2 MHz) δ: 17.1, 18.5, 24.0, 27.2, 28.6, 29.4, 32.5, 33.2, 34.2, 37.5, 39.3, 41.9, 46.7, 48.8, 55.2, 72.5, 74.2, 105.3, 113.7 (two signals superimposed), 129.0 (two signals superimposed), 131.0, 136.0, 140.1, 157.9, 159.1.

Exact mass calcd for $C_{27}H_{39}O_2(M+H)^+$: 395.2950. Found: 395.2952.

Anal. calcd for C₂₇H₃₈O₂: C 82.12, H 9.71. Found C 82.12, H 9.74.

Preparation of $(1R^*, 6R^*, 9R^*)$ -3-[(S*)-2-(4-Methoxyphenyl)methoxy-1-methylethyl]-6,9-dimethyltricyclo[7.4.0.0^{2,6}]tridec-2-en-10-one (161)





To a stirred solution of alkene **199** (2.10 g, 5.32 mmol) in 5:1 *t*-BuOH–H₂O (120 mL) at room temperature were added KIO₄ (7.34 g, 31.9 mmol) and NaHCO₃ (4.47 g, 53.2 mmol). A catalytic amount of OsO₄ (25 mg, 0.01 mmol) was added and the resulting brown mixture was stirred for 72 h. Et₂O (100 mL) and H₂O (100 mL) were added and the resulting layers were separated. The aqueous layer was extracted with Et₂O (3×100 mL) and the combined organic layers were washed with brine (50 mL) and dried over anhydrous MgSO₄. The solvent was removed under reduced pressure and the crude oil was purified by flash chromatography (100 g of silica gel, 20:1 toluene–Et₂O) to yield 1.37 g (64.9%) of ketone **161** as a colourless oil.

IR (neat): 2938, 2860, 1703, 1613, 1514, 1248 cm⁻¹.

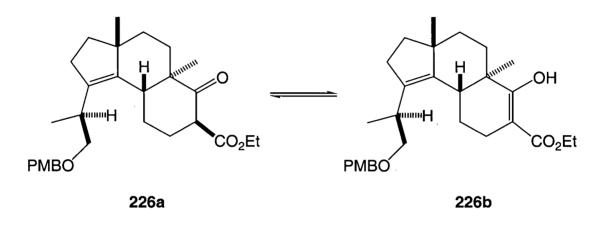
¹H NMR (CDCl₃, 400 MHz) δ: 0.99 (s, 3H), 1.02 (d, 3H, *J* = 7.3 Hz), 1.03 (s, 3H), 1.35– 1.66 (m, 5H), 1.76 (dt, 1H, *J* = 4.3, 14.3 Hz), 1.96–2.30 (m, 7H), 2.42–2.49 (m, 1H), 2.60 (dt, 1H, *J* = 6.1, 14.3 Hz), 3.20–3.35 (m, 3H), 3.78 (s, 3H), 4.35–4.45 (m, 2H), 6.84 (d, 2H, *J* = 8.9 Hz), 7.20 (d, 2H, *J* = 8.9 Hz).

¹³C NMR (CDCl₃, 75.2 MHz) δ: 17.1, 17.2, 23.9, 25.8, 26.7, 29.4, 29.9, 32.6, 36.6, 37.8, 39.1, 46.7, 48.5, 50.9, 55.5, 72.6, 74.0, 113.7 (two signals superimposed), 129.0 (two signals superimposed), 130.8, 137.7, 138.2, 159.1, 216.3.

Exact mass calcd for $C_{26}H_{40}O_3N (M+NH_4)^+$: 414.3008. Found 414.2997.

Anal. calcd for C₂₆H₃₆O₃: C 78.75, H 9.15. Found C 78.63, H 9.19.

Preparation of a 1:1 Mixture of $(1R^*, 6R^*, 9R^*, 11S^*)$ -3-[(S^*) -2-(4-Methoxyphenyl)methoxy-1-methylethyl]-6,9-dimethyl-10-oxotricyclo[7.4.0.0^{2,6}]trideca-2-ene-11-carboxylic Acid Ethyl Ester (226a) and $(1R^*, 6R^*, 9R^*)$ -3-[(S^*) -2-(4-Methoxyphenyl)methoxy-1-methylethyl]-6,9-dimethyl-10-hydroxytricyclo[7.4.0.0^{2,6}]trideca-2,10-diene-11-carboxylic Acid Ethyl Ester (226b)



To a stirred suspension of NaH (132 mg, 5.50 mmol) in dry THF (40 mL) was added a solution of ketone **161** (725 mg, 1.83 mmol) in dry THF (10 mL). (EtO)₂CO (425 mg, 3.6 mmol) was added *via* syringe and the mixture was heated to reflux for 20 min. A catalytic amount of KH (5 mg, 0.13 mmol) was added and the mixture was refluxed for a further 18 h. The solution was cooled to room temperature and dilute aqueous HCl (0.5 M, 30 mL) was added. Et₂O (50 mL) was added and the resulting layers were separated. The aqueous layer was extracted with Et₂O (3 × 50 mL) and the combined organic layers were washed with brine (50 mL) and dried over anhydrous MgSO₄. The solvent was removed under reduced pressure and the crude oil was purified by flash chromatography (25 g of silica gel, 10:1 pentane–Et₂O) to yield 625 mg (74%) of a mixture of keto ester **226a** and enol ester **226b** as a colourless oil.

IR (neat): 2937, 2859, 1743, 1709, 1649, 1613, 1514, 1245 cm⁻¹.

A solution of 1:1 mixture of keto ester **226a** and enol ester **226b** in $CDCl_3$ exhibited the following NMR data:

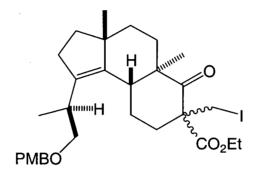
¹H NMR (CDCl₃, 400 MHz) δ: 0.95–1.1 (m), 1.23–1.32 (m), 1.35–1.96 (m), 2.00–2.40 (m), 2.52 (bd, *J* = 12 Hz), 3.13–3.36 (m), 3.68 (dd, *J* = 5.5, 13.5 Hz), 3.78 (s), 4.12–4.26 (m), 4.35–4.45 (m), 6.82–6.88 (m), 7.18–7.25 (m), 12.4 (s).

¹³C NMR (CDCl₃, 75.2 MHz) δ: 14.2, 14.3, 16.8, 16.9, 17.1, 18.0, 22.6, 23.3, 23.8, 24.1, 24.7, 29.2, 29.4, 29.5, 29.8, 31.7, 32.3, 32.7, 36.4, 37.0, 39.1, 39.3, 41.5, 43.1, 46.8, 48.3, 48.4, 51.2, 53.1, 55.2, 60.2, 60.9, 72.5, 72.6, 73.8, 74.0, 94.8, 113.7 (four signals superimposed), 128.9 (four signals superimposed), 131.0 (two signals superimposed), 136.3, 137.6, 138.3, 139.2, 159.1, 170.6, 173.5, 178.2 (three signals not accounted for).

Exact mass calcd for $C_{29}H_{41}O_5 (M+H)^+$: 469.2954. Found: 469.2950.

Anal. calcd for C₂₉H₄₀O₅: C 74.33, H 8.60. Found: C 74.19, H 8.60.

Preparation of a 3:1 Mixture of (1*R**,6*R**,9*R**)-11-(Iodomethyl)-3-[(*S**)-2-(4methoxy-phenyl)methoxy-1-methylethyl]-6,9-dimethyl-10-oxotricyclo[7.4.0.0^{2,6}]tridec-2-ene-11-carboxylic Acid Ethyl Esters (160)



160

To a stirred solution of a mixture of keto ester **226a** and enol ester **226b** (139 mg, 0.297 mmol) in dry THF (40 mL) was added a solution of TBAF (1.0 M in THF, 0.6 ml, 0.6 mmol). The mixture was stirred for 1 h and neat CH_2I_2 (800 mg, 3.0 mmol) was added. After the mixture had been stirred for an additional 1 h, Et_2O (30 mL) and saturated aqueous NaHCO₃ (30 mL) were added. The resulting layers were separated and the aqueous layer was washed with Et_2O (3 × 30 mL). The combined organic layers were dried over anhydrous MgSO₄, concentrated and purified by flash chromatography (5 g of silica gel, 5:1 petroleum ether– Et_2O) to yield 141 mg (78.0%) of a 3:1 mixture of diasteriomeric iodides **160** as a colourless oil.

The relative configurations of the isomers were not determined. The 3:1 mixture of diasteriomeric iodides **160** exhibited the following spectral data:

IR (neat): 2937, 2859, 1735, 1708, 1614, 1514 cm⁻¹.

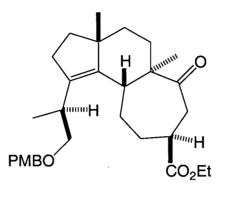
¹H NMR (CDCl₃, 400 MHz) (signals assigned to major isomer) δ: 0.94 (s, 3H), 0.98 (s, 3H), 0.99 (d, 3H, *J* = 5.5 Hz), 1.27 (t, 3H, *J* = 7.0 Hz), 1.35–2.55 (m, 12H), 2.67 (br d, 1H, *J* = 12.8 Hz), 3.13–3.35 (m, 3H), 3.52 (m, 2H), 3.79 (s, 3H), 4.20 (dq, 2H, *J* = 2.1, 7.0 Hz), 4.36–4.45 (m, 2H), 6.86 (d, 2H, *J* = 8.6 Hz), 7.22 (d, 2H, *J* = 8.6 Hz).

¹³C NMR (CDCl₃, 75.2 MHz) (signals assigned to major isomer) δ: 9.5, 14.0, 17.0, 17.1,
20.7, 23.6, 29.4, 29.8, 32.0, 32.6, 36.1, 39.2, 40.3, 48.1, 49.6, 55.2, 57.9, 62.0, 72.5, 73.8,
113.6 (two signals superimposed), 129.0 (two signals superimposed), 130.8, 137.8, 137.9,
159.0, 169.3, 210.

Exact mass calcd for fragment C₂₂H₃₂O₄I (M-MeOPhCH₂)⁺: 487.1346. Found: 487.1337

Exact mass calcd for fragment C₈H₉O (MeOPhCH₂⁺): 121.0653. Found: 121.0654

Preparation of $(1R^*, 6R^*, 9R^*, 12S^*)$ -3-[(S*)-2-(4-Methoxyphenyl)methoxy-1-methylethyl]-6,9-dimethyl-10-oxotricyclo[7.5.0.0^{2,6}]tetradec-2-ene-12-carboxylic Acid Ethyl Ester (159)



159

To a stirred solution of a 3:1 mixture of iodides **160** (161 mg, 0.265 mmol) in dry THF (30 mL) was added a solution of SmI₂ (0.1 M in THF, 7.9 mL, 0.79 mmol). The resulting mixture was stirred for 1 h at room temperature. Dilute aqueous HCl (0.5 M, 40 mL) and Et₂O (40 mL) were added and the resulting layers were separated. The aqueous layer was extracted with Et₂O (3×40 mL) and the combined organic layers were washed with brine (40 mL) and dried over anhydrous MgSO₄. The solvent was removed under reduced pressure and the crude oil was purified by flash chromatography (5 g of silica gel, 40:1 petroleum ether–Et₂O) to yield 91 mg (71%) of keto ester **159** as a colourless oil.

IR (neat): 2933, 2859, 1733, 1698, 1514, 1248 cm⁻¹.

¹H NMR (CDCl₃, 400 MHz) δ : 0.97 (d, 3H, J = 6.8 Hz), 1.00 (s, 3H), 1.07 (s, 3H), 1.18– 1.23 (m, 1H), 1.24 (t, 3H, J = 7.1 Hz), 1.40–1.72 (m, 5H), 1.81 (dt, 1H, J = 5.2, 13.3 Hz), 1.85–2.30 (m, 5H), 2.65 (br d, 1H, J = 11.2 Hz), 2.77–2.84 (m, 2H), 3.05 (dd, 1H, J =9.5, 14.4 Hz), 3.10–3.20 (m, 1H), 3.23–3.34 (m, 2H), 3.78 (s, 3H), 4.08–4.15 (m, 2H), 4.36–4.44 (m, 2H), 6.85 (d, 2H, J = 8.6 Hz), 7.21 (d, 2H, J = 8.6 Hz).

¹³C NMR (CDCl₃, 75.2 MHz) δ: 14.1, 14.6, 16.6, 24.2, 26.6, 29.2, 29.9, 33.0, 33.1, 36.0, 38.3, 38.7, 39.3, 43.3, 49.0, 53.6, 55.2 60.8, 72.5, 73.8, 113.7 (two signals superimposed), 128.9 (two signals superimposed), 130.8, 136.6, 139.4, 159.0, 174.5, 215.0.

Exact mass calcd for $C_{30}H_{43}O_5 (M+H)^+$: 483.3110. Found: 483.3114.

Anal. calcd for C₃₀H₄₂O₅: C 74.66, H 8.77. Found: C 74.77, H 8.99.

18 ,^{,,,,,,,,}19 8 6 2 9 3 10 15 H 17 3 12 16 ^{/////}H 20 6' 2" 5' 4' Ο 3 8 159

Assignments H–x	¹ H NMR δ ppm (Multiplicity, <i>J</i> (Hz))	Observed NOEs
H-1 ^a	2.65 (br d, 11.2)	Η–18, Η–11(β)
H-4	part of m at 1.85–2.30, part of m at 1.40–1.72	11 10, 11 11(p)
H5	part of m at 1.40–1.72	
H–7	part of m at 1.40–1.72,	
H–8	1.81 (dt, 5.2, 13.3), 1.18–1.23 (m)	
Η-11(α)	part of m at 2.77–2.84	
Η–11(β)	3.05 (dd, 9.5,14.4)	
H-12	part of m at 2.77–2.84	
H-13	part of m at 1.85–2.30	
H–14	part of m at 1.85–2.30	
H–15	3.10–3.20 (m)	
H–16	3.23–3.34 (m)	
H–17	0.97 (d, 6.8)	
H–18 ^{<i>a</i>}	1.07 (s)	H–1
H–19 ^a	1.00 (s)	$H-11(\alpha)$
H–1'	4.08-4.15 (m)	
H–3', H–7'	7.21 (d, 8.6)	
H-4', H-6'	6.85 (d, 8.6)	
H–8'	3.78 (s)	
H–1"	4.36–4.44 (m)	
H–2"	1.24 (t, 7.1)	

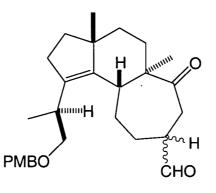
^a Irradiation of this signal generated the corresponding NOEs in the right hand column

Table 14. ¹H NMR (CDCl₃, 400 MHz) Data for Keto Ester 159: NOED Experiment.

	4 <u>3</u> 2 H 9		
	Ý	1 10 H	
	15	"H /14 11	
	1/	$H_{13} \frac{12}{12}$	
	\sim $\frac{1}{\sim}$ $\frac{1}{\sim}$	6 ····································	
١	6' 7' 2' O	20 1"	
8'	5' 4' 3'	0 2"	
0	4	159	
Assignments	¹³ C NMR	Observed HMQC Correlations	
Cx	δppm	(δ ppm)	
C-1	43.3	H–1 (2.65)	
C-2	139.4		
C-3	136.6		
C-4	29.2	H–4 (part of m at 1.40–1.72,	
	29.2	part of m at 1.85–2.30)	
C-5	38.3	H-5 (part of m at 1.40–1.72)	
C6	49.0		
C-7	36.0	H-7(part of m at 1.40-1.72)	
C-8	33.1	H-8 (1.81, part of m at 1.18-1.23)	
C9	53.6		
C-10	215.0		
C-11	39.3	H–11 (part of m at 2.77–2.84, 3.05)	
C-12	38.7	H–12 (part of m at 2.77–2.84)	
C-13	29.9	H–13 (part of m at 1.85–2.30)	
C-14	26.6	H–14 (part of m at 1.85–2.30)	
C-15	33.0	H-15 (3.10-3.20)	
C-16	73.8	H–16 (3.24–3.34)	
C-17	16.6	H-17 (0.97)	
C–18	24.2	H–18 (1.07)	
C-19	14.6	H–19 (1.00)	
C-20	174.5		
C-1'	60.8	H–1' (4.08–4.15)	
C-2'	130.8	•	
C-3', C-7'	128.9	H–3', H–7' (7.21)	
C-4', C-6'	113.7	H–4', H–6' (6.85)	
C-5'	159.0		
C-8'	. 55.2	H–8' (3.78)	
C-1"	72.5	H–1" (4.36–4.44)	
C-2"	14.1	H–2" (1.24)	

Table 15. ¹³C NMR (CDCl₃, 100.6 MHz) and ¹H NMR (CDCl₃, 400 MHz) Data for Keto Ester **159**: HMQC Experiment.

Preparation of a 1:1 Mixture of $(1R^*, 6R^*, 9R^*)$ -3-[(S*)-2-(4-Methoxyphenyl)methoxy-1-methylethyl]-6,9-dimethyl-10-oxotricyclo[7.5.0.0^{2,6}]tetra-dec-2-ene-12carboxaldehydes (158)





To a cold (0 °C) stirred solution of keto ester **159** (181 mg, 0.374 mmol) in dry Et₂O (20 mL) was added a solution of DIBALH (1.0 M in hexanes, 1.8 mL, 1.80 mmol) and the resulting mixture was stirred for 1 h. After this time, the mixture was warmed to room temperature and was stirred for an additional 1 h. Saturated aqueous NH_4Cl-NH_3 (pH 8, 180 *u*L) was added and the mixture was stirred for 1 h. Anhydrous $MgSO_4$ (100 g) was added and the mixture was stirred for 1 h. EtOH (2 mL) was added and the mixture was stirred for 1 h. EtOH (2 mL) was added and the mixture was stirred for 1 h. EtOH (2 mL) was added and the mixture was stirred for 1 h. The mixture was filtered through a sintered glass funnel (4 cm diameter) containing Florisil[®] (4 cm depth) on top of a thin layer of Celite[®]. The column was washed with 20:1 EtOH–Et₂O (50 mL). The solvent of the combined eluates was removed under reduced pressure to give a crude mixture of the diols.

The crude mixture of diols was taken up in dry CH_2Cl_2 (5 mL) and the solution was added to a solution of Dess-Martin's periodinane reagent (510 mg, 1.2 mmol) in dry CH_2Cl_2 (30 mL). After the mixture had been stirred for 1 h, saturated aqueous NaHCO₃ (100 mL) and saturated aqueous Na₂S₂O₃ (100 mL) were added. The biphasic mixture was stirred for 1 h and the layers were then separated. The aqueous portion was extracted with Et_2O (3 × 10 mL) and the combined organic layers were dried over anhydrous MgSO₄. The solvent was removed under reduced pressure and the crude oil was purified by flash chromatography (10 g of silica gel, 2:1 petroleum ether– Et_2O) to give 141 mg (86%) of a 1:1 mixture of epimeric keto aldehydes **158** as a colourless oil.

The 1:1 mixture of epimeric keto aldehydes 158 exhibited the following spectral data:

IR (neat): 2932, 2857, 1726, 1697, 1514, 1248, 1089 cm⁻¹.

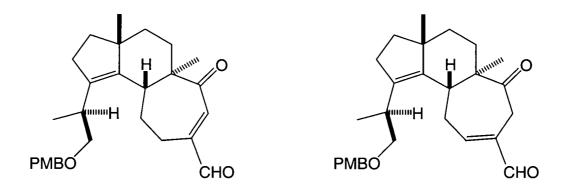
¹H NMR (CDCl₃, 400 MHz) δ: 0.96 and 0.99 (s, s, ratio 1:1, 3H), 0.96–1.00 (m, 3H), 1.08 and 1.10 (s, s, ratio 1:1, 3H), 1.14–1.24 (m, 2H), 1.42–1.92 (m, 6H), 2.05–2.46 (m, 5H), 2.53–2.79 (m, 2H), 2.88–3.15 (m, 2H), 3.23–3.35 (m, 2H), 3.78 (s, 3H), 4.35–4.44 (m, 2H), 6.82–6.87 (m, 2H), 7.18–7.23 (m, 2H), 9.60 and 9.63 (s, s, ratio 1:1, 1H).

¹³C NMR (CDCl₃, 75.2 MHz) δ: 14.0, 14.3, 16.3, 16.7, 24.1, 24.4, 25.9, 26.7, 27.4, 29.0, 29.1, 29.2, 32.7, 33.0, 33.1, 35.7, 35.8, 36.0, 37.5, 43.0, 43.1, 45.1, 48.9, 49.2, 49.3, 53.4, 54.5, 55.2, 72.5, 73.7, 73.8, 113.6 (two signals superimposed), 113.7 (two signals superimposed), 128.8 (two signals superimposed), 128.9 (two signals superimposed), 130.7, 130.8, 135.9, 137.0, 138.7, 139.7, 159.0, 201.0, 201.5, 214.9, 215.8 (six signals not accounted for).

Exact mass calcd for $C_{28}H_{42}O_4N(M+NH_4)^+$: 456.3114. Found: 456.3122.

Anal. calcd for C₂₈H₃₈O₄: C 76.68, H 8.73. Found: C 77.00, H 8.81.

Preparation of $(1R^*, 6R^*, 9R^*)$ -3-[(S*)-2-(4-Methoxyphenyl)methoxy-1-methylethyl]-6,9-dimethyl-10-oxotricyclo[7.5.0.0^{2,6}]tetradeca-2,11-diene-12-carboxaldehyde (249)



249

240

To a stirred solution of a 1:1 mixture of keto aldehydes **158** (106 mg, 0.242 mmol) in dry benzene (12 mL) over 3Å mol sieves was added piperidine (103 mg, 1.21 mmol). The mixture was heated to reflux for 3 h and cooled to room temperature. The solvent and excess piperidine were removed under reduced pressure and the remaining material was taken up in dry THF (12 mL). The mixture was cooled to -78 °C and solid PhSeCl (232 mg, 1.21 mmol) was added. After the mixture had been stirred for 5 min, Et₂O (15 mL) and H₂O (15 mL) were added. The biphasic mixture was warmed to room temperature and stirred for 3 h. The phases were separated and the aqueous phase was extracted with Et₂O (3 × 10 mL). The combined organic layers were concentrated and taken up in MeOH (10 mL). Saturated aqueous KHCO₃ (2 mL) and solid KIO₄ (557 mg, 2.42 mmol) were sequentially added, and the mixture was stirred for 20 min. Et₂O (20 mL) and H₂O (20 mL) were added, and the resulting layers were separated. The aqueous layer was extracted with Et₂O (3 × 20 mL). The combined organic layers were washed with brine

(20 mL) and dried over anhydrous MgSO₄. The solvent was removed under reduced pressure and the crude oil was purified by flash chromatography (5 g of silica gel, 4:1 petroleum ether–Et₂O) to yield 79.5 mg (75.2%) of the α , β -unsaturated ketone **249** and 2.4 mg (2.3%) of the β , γ -unsaturated ketone **240** as colourless oils

 α , β -Unsaturated ketone **249** exhibited the following spectral data:

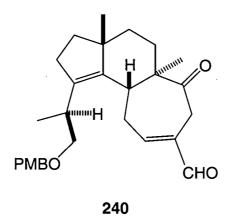
IR (neat): 2932, 2152, 1697, 1514, 1248 cm⁻¹.

¹H NMR (CDCl₃, 400 MHz) δ: 0.99 (d, 3H, *J* = 6.7 Hz), 1.03 (s, 3H), 1.10 (s, 3H), 1.22– 1.64 (m, 5H), 1.83–1.97 (m, 2H), 2.23–2.43 (m, 4H), 2.59 (dt, 1H, *J* = 18.6, 4.3 Hz), 2.67 (br d, 1H, *J* = 9.7 Hz), 3.02–3.13 (m, 1H), 3.24–3.36 (m, 2H), 3.80 (s, 3H), 4.34–4.45 (m, 2H), 6.63 (m), 6.86 (d, 2H, *J* = 8.6 Hz), 7.21 (d, 2H, *J* = 8.6 Hz), 9.56 (s, 1H).

¹³C NMR (CDCl₃, 75.2 MHz) δ: 15.9, 16.5, 24.2, 25.2, 26.6, 29.2, 33.1, 33.2, 35.5, 37.6, 43.0, 49.4, 55.0, 55.3, 72.6, 73.7, 113.7 (two signals superimposed), 128.9 (two signals superimposed), 130.7, 136.9, 139.6, 144.2, 146.1, 159.1, 194.7, 209.6.

Exact mass calcd for $C_{28}H_{40}O_4N (M+NH_4)^+$: 454.29575. Found: 454.29515

Preparation of $(1R^*, 6R^*, 9R^*)$ -3-[(S*)-2-(4-Methoxyphenyl)methoxy-1-methylethyl]-6,9-dimethyl-10-oxotricyclo[7.5.0.0^{2,6}]tetradeca-2,12-diene-12-carboxaldehyde (240)



To a stirred solution of ketone **249** (51.6 mg, 0.118 mmol) in dry benzene (5 mL) was added DBN (29.3 mg, 0.236 mmol). The mixture was heated to reflux for 12 h and cooled to room temperature. The solvent was removed under reduced pressure and the crude oil was purified by flash chromatography (2 g of silica gel, 3:1 petroleum ether- Et_2O) to yield 49 mg (95.8%) of ketone **240** as a colourless oil.

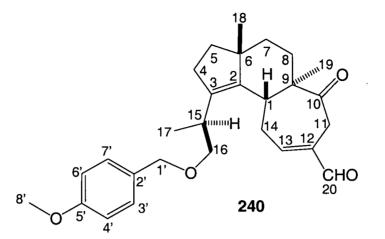
IR (neat): 2931, 2854, 1704, 1688, 1513, 1248 cm⁻¹.

¹H NMR (CDCl₃, 500 MHz) δ: 0.99 (d, 3H, *J* = 6.7 Hz), 1.02 (s, 3H), 1.13 (s, 3H), 1.20– 1.30 (m, 1H), 1.50–1.70 (m, 4H), 1.94 (dt, 1H, *J* = 5.3, 13.1 Hz), 2.15–2.25 (m, 2H), 2.66–2.75 (m, 1H), 3.05–3.10 (m, 1H), 3.14 (dd, 1H, *J* = 6.4, 19.8 Hz), 3.25–3.35 (m, 3H), 3.46 (d, 1H, *J* = 14.0 Hz), 3.72 (br d, 1H, *J* = 14.0 Hz), 3.78 (s, 3H), 4.35–4.46 (m, 2H), 6.63–6.68 (m, 1H), 6.85 (d, 2H, *J* = 8.6 Hz), 7.20 (d, 2H, *J* = 8.6 Hz), 9.33 (s, 1H). ¹³C NMR (CDCl₃, 75.2 MHz) δ: 12.8, 16.4, 24.2, 29.1, 31.7, 32.9, 33.3, 34.1, 35.9, 38.1, 39.6, 49.4, 55.0, 55.3, 72.7, 73.6, 113.7 (two signals superimposed), 128.9 (two signals superimposed), 130.7, 135.7, 137.0, 137.9, 153.7, 159.2, 192.3, 210.6.

Exact mass calcd for $C_{28}H_{40}O_4N(M+NH_4)^+$: 454.29575. Found: 454.29541.

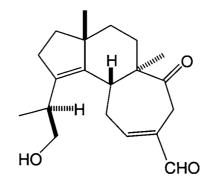
Anal. calcd for C₂₈H₃₆O₄: C 77.03, H 8.31. Found: C 77.03, H 8.36.

Table 16. ¹³C NMR (CDCl₃, 100.6 MHz) and ¹H NMR (CDCl₃, 400 MHz) Data for Ketone **240**: HMQC Experiment.



Assignments	¹³ C NMR	Observed HMQC Correlations
Č–x	δ ppm	(δ ppm)
C-1	39.6	H–1 (part of m at 3.25–3.35)
C-2	135.7	
C-3	137.0	
C-4	29.1	H-4 (2.15-2.25)
C-5	35.9	H-1 (part of m at 1.50-1.70)
C-6	49.4	
C-7	38.1	H–7 (part of m at 1.50–1.70)
C-8	32.9	H–8 (1.94, 1.20–1.30)
C-9	55.0	
C-10	210.6	
C-11	34.1	H–11 (3.46, 3.72)
C-12	137.9	
C-13	153.7	H–13 (6.63–6.68)
C-14	31.7	H–14 (2.65–2.75, 3.14)
C-15	33.3	H–15 (3.05–3.10)
C-16	73.6	H–16 (part of m at 3.25–3.35)
C-17	16.4	H–17 (0.99)
C-18	24.2	H–18 (1.13)
C-19	12.8	H–19 (1.02)
C-20	192.3	H–20 (9.33)
C-1'	72.7	H-1' (4.35-4.46)
C-2'	130.7	
C-3', C-7'	128.9	H–3', H–7' (7.20)
C4', C6'	113.7	H-4', H-6' (6.85)
C-5'	159.2	
C-8'	55.3	H–8' (3.78)

Preparation of (1*R**,6*R**,9*R**)-3-[(*S**)-2-Hydroxy-1-methylethyl]-6,9-dimethyl-10oxotricyclo[7.5.0.0^{2,6}]tetradeca-2,12-diene-12-carbaldehyde [(±)-Sarcodonin G] (36)



36

To a stirred solution of ketone **240** (6.2 mg, 0.014 mmol) in a biphasic mixture of 20:1 $CH_2Cl_2-H_2O$ (5.25 mL) was added DDQ (4.8 mg, 0.021 mmol). The mixture was stirred at room temperature for 0.5 h and H₂O (5 mL) was added. The layers were separated and the aqueous layer was extracted with CH_2Cl_2 (3 × 10 mL). The combined organic layers were washed with brine (10 mL) and dried over anhydrous MgSO₄. The solvent was removed under reduced pressure and the residual material was purified by flash chromatography (1 g of silica gel, 1:1 petroleum ether–Et₂O) to yield 4.1 mg (92%) of (±)-sarcodonin G (**36**) as an amorphous solid.

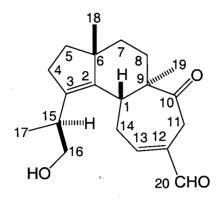
IR (KBr): 3445, 1703, 1640, 1450, 1376 cm^{-1} .

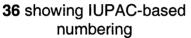
¹H NMR (CDCl₃, 400 MHz) δ : 0.97 (d, 3H, J = 6.9 Hz), 1.03 (s, 3H), 1.14 (s, 3H), 1.24– 1.30 (m, 1H), 1.46 (s, 1H), 1.49–1.55 (m, 1H), 1.58–1.70 (m, 2H), 1.95 (dt, 1H, J = 5.0, 13.4 Hz), 2.21–2.40 (m, 3H), 2.69–2.80 (m, 1H), 2.98–3.08 (m, 1H), 3.10–3.20 (m, 1H), 3.36 (br d, 1H, *J* = 12.6 Hz), 3.41–3.51 (m, 3H), 3.71–3.75 (m, 1H), 6.70–6.73 (m, 1H), 9.35 (s, 1H).

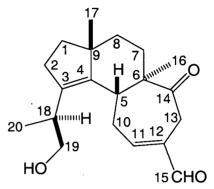
¹³C NMR (CDCl₃, 75.2 MHz) δ: 12.7, 15.6, 24.8, 28.6, 32.1, 32.7, 34.1, 35.2, 35.5, 37.7,
39.7, 49.8, 55.3, 65.8, 135.9 (two signals superimposed), 141.3, 153.2, 192.2, 210.3.

Exact mass calcd for C₂₀H₂₈O: 316.2039. Found: 316.2038.

Table 17. Comparison of ¹H NMR Data for Synthetic $(1R^*, 6R^*, 9R^*)$ -3-[(S*)-2-Hydroxy-1-methylethyl]-6,9-dimethyl-10-oxotricyclo[7.5.0.0^{2,6}]tetradeca-2,12-diene-12-carbox-aldehyde [(±)-Sarcodonin G] (**36**) (CDCl₃, 400 MHz) with those Reported¹⁶ for Natural (–)-Sarcodonin G (**36**) (CDCl₃, 250 MHz).





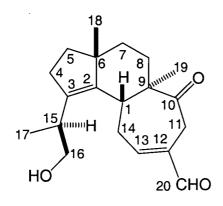


36 showing IUPAC-based numbering

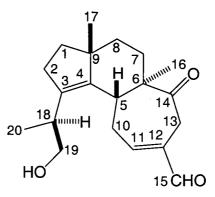
¹ H NMR (CDCl ₃ , 400 MHz) Signals Displayed ^a by Synthetic (±)-Sarcodonin G (36) δ ppm (Multiplicity, <i>J</i> (Hz))	¹ H Assignment H–x	Reported ¹ H NMR (CDCl ₃ , 250 MHz) Signals Displayed by Natural (–)-Sarcodonin G (36) δ ppm (Multiplicity, <i>J</i> (Hz))	Cyathane Numbering
0.97 (d, 6.9)	H–17	0.96 (d, 7.3)	H–20
1.03 (s)	H–19	1.01 (s)	H–16
1.14 (s)	H–18	1.12 (s)	H–17
1.24–1.30 (m)		1.24 (dt, 3.9, 13.2)	
1.46 (s)		part of m at 1.59	
1.49–1.55 (m)		part of m at 1.59	
1.58–1.70 (m)		part of m at 1.59	
1.95 (dt, 5, 13.4)		1.94 (dt, 5.3, 13.3)	
2.21–2.40 (m)		2.31 (m)	
2.69–2.80 (m)		2.74 (m)	
2.98-3.08 (m)	H–15	3.02 (sextet, 7.3)	H–18
3.10–3.20 (m)		3.16 (dd, 13.4, 6.4)	
3.36 (br d, 12.6)		3.34 (d, 12)	
3.41–3.51 (m)		3.42 (d, 13.9), 3.45 (d, 7.3)	
3.71-3.75 (m)		3.72 (br d, 13.6))	
6.70–6.73 (m)	H–13	6.71 (m)	H–11
9.35 (s)	H–20	9.31	H–15

^{*a*} The difference between observed and reported δ is likely due to the CDCl₃ reference.

Table 18. Comparison of ¹³C NMR Data for Synthetic $(1R^*, 6R^*, 9R^*)$ -3-[(S*)-2-Hydroxy-1-methylethyl]-6,9-dimethyl-10-oxotricyclo[7.5.0.0^{2,6}]tetradeca-2,12-diene-12-carboxaldehyde [(±)-Sarcodonin G] (**36**) (CDCl₃, 400 MHz) with those Reported¹⁶ for Natural (–)-Sarcodonin G (**36**) (CDCl₃, 62.5 MHz).



36 showing IUPAC-based numbering

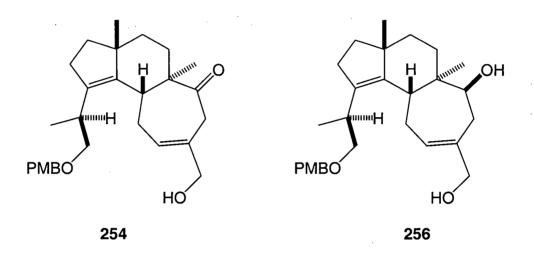


36 showing IUPAC-based numbering

¹³ C NMR (CDCl ₃ , 100.6 MHz) Signals Displayed by Synthetic (±)-Sarcodonin G	¹³ C assignment	Reported ¹³ C NMR (CDCl ₃ , 62.5 MHz) Signals Displayed by Natural	Cyathane
(36)	C–x	(–)-Sarcodonin G (36)	Numbering
δppm		δppm	
12.7	C-19	12.7	C-16
15.6	C-15	15.6	C-20
24.8	C-18	24.8	C-17
28.6		28.6	
32.1		32.1	
32.7		32.7	
34.1		34.1	
35.2		35.2	
35.5		35.5	
37.7		37.7	
39.7		39.5	
49.8		49.8	
55.3		55.3	
65.8	C-16	65.8	C-19
135.9		135.8, 135.9	
(two signals superimposed)		135.8, 155.9	
141.3		141.2	
153.2	C-13	153.5	C-11
192.2	C-20	192.4	C-15
210.3	C-10	210.6	C-14

4.3. Synthetic Studies Directed Towards Cyathane Diterpenoid (±)-Cyathin A4

Preparation of $(1R^*, 6R^*, 9R^*)$ -12-Hydroxymethyl-3-[$(1S^*)$ -2-(4-methoxyphenyl)meth-oxy-1-methylethyl]-6,9-dimethyltricyclo[7.5.0.0^{2,6}]tetradeca-2,12-dien-10-one (254)



To a stirred solution of keto aldehyde **240** (200 mg, 0.458 mmol) in *i*-PrOH (10 mL) was added (*i*-PrO)₃Al (140 mg, 0.687 mmol). The mixture was heated to 60 °C for 8 h and cooled to room temperature. Et₂O (10 mL) and aqueous NH₄Cl–NH₃ (pH 8, 0.1 mL) were added and the resulting slurry was stirred for 18 h. MgSO₄ (100 mg) was added and the mixture was stirred for 1 h. The slurry was filtered through a column of Florisil[®] (1 g) and the column was washed with a mixture of 1:1 Et₂O–EtOH (10 mL). The solvent of the eluate was removed under reduced pressure and the resulting material was purified by flash chromatography (5 g of silica gel, 9:1 Et₂O–petroleum ether) to give 110 mg (55%) of keto alcohol **254** and 65 mg (32%) of diol **256** as colourless oils.

The diol **256** (65 mg, 0.147 mmol) was taken up in dry CH_2Cl_2 (2 mL) and the solution was added to a solution of Dess-Martin's periodinane reagent (250 mg, 0.6 mmol) in dry CH_2Cl_2 (15 mL). After the mixture had been stirred for 1 h, saturated aqueous NaHCO₃ (50 mL) and saturated aqueous Na₂S₂O₃ (50 mL) were added. The biphasic mixture was stirred for 1 h and the layers were separated. The aqueous portion was extracted with Et_2O (3 × 5 mL) and the combined organic layers were dried over anhydrous MgSO₄. The solvent was removed under reduced pressure and the crude oil was purified by flash chromatography (5 g of silica gel, 3:1 petroleum ether– Et_2O) to give 55 mg (86%) of keto aldehyde **240** as a colourless oil. Keto aldehyde **240** was resubmitted to the reduction reaction to yield an additional 31 mg of keto alcohol **254** for a total overall yield of 141 mg (70%).

Keto alcohol **254** exhibited the following spectral data:

IR (neat): 3436, 2932, 2855, 1702, 1613, 1513 cm⁻¹.

¹H NMR (CDCl₃, 400 MHz) δ : 0.99 (d, 3H, J = 6.7 Hz), 1.03 (s, 3H), 1.12 (s, 3H), 1.45– 1.65 (m, 6H), 1.91 (dt, 1H, J = 5.0, 12.8 Hz), 2.10–2.35 (m, 2H), 2.45–2.53 (m, 1H), 2.68–2.82 (m, 2H), 3.06–3.15 (m, 1H), 3.21–3.36 (m, 3H), 3.79 (s, 3H), 3.92 (br d, 1H, J = 12.8 Hz), 3.95–4.08 (m, 2H), 4.32–4.49 (m, 2H), 5.60–5.69 (m, 1H), 6.85 (d, 2H, J = 8.6 Hz), 7.21 (d, 2H, J = 8.6 Hz). ¹³C NMR (CDCl₃, 75.2 MHz) δ: 12.9, 16.5, 24.3, 29.2, 30.0, 32.8, 33.0, 36.1, 38.2, 39.3, 40.3, 49.3, 54.6, 55.3, 68.8, 72.5, 73.8, 113.7 (two signals superimposed), 126.6, 128.8 (two signals superimposed), 130.8, 131.7, 136.5, 138.5, 159.0, 211.3.

Exact mass calcd for $C_{28}H_{39}O_4$ (M+H)⁺: 439.28482. Found: 439.28435.

Diol **256** exhibited the following spectral data:

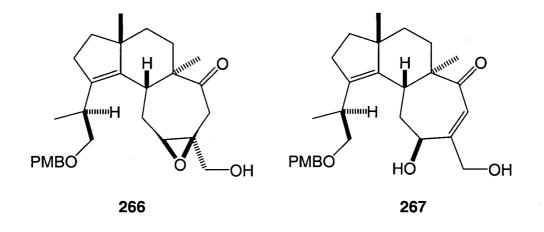
IR (neat): 3368, 2931, 2860, 1613, 1514 cm⁻¹.

¹H NMR (CDCl₃, 400 MHz) δ: 0.89 (s, 3H), 0.99 (d, 3H, *J* = 6.8 Hz), 1.05 (s, 3H), 1.32– 1.58 (m, 8H), 1.72–1.90 (m, 2H), 2.02–2.16 (m, 2H), 2.21–2.64 (m, 3H), 3.03–3.13 (m, 1H), 3.24–3.38 (m, 2H), 3.43 (d, 1H, *J* = 8.1 Hz), 3.79 (s, 3H), 3.92–3.99 (m, 2H), 4.38– 4.47 (m, 2H), 5.85 (d, 1H, *J* = 7.0 Hz), 6.85 (d, 2H, *J* = 8.7 Hz), 7.21 (d, 2H, *J* = 8.7 Hz).

¹³C NMR (CDCl₃, 75.2 MHz) δ: 16.3, 17.1, 24.5, 28.2, 29.1, 30.3, 31.8, 33.1, 35.8, 37.6, ...
39.0, 45.2, 49.8, 55.2, 68.4, 72.4, 74.0, 75.3, 113.7 (two signals superimposed), 128.9 (two signals superimposed), 130.0, 130.9, 134.5, 137.6, 142.0, 159.0.

Exact mass calcd for $C_{28}H_{44}O_4N(M+NH_4)^+$: 458.32703. Found: 458.32817.

Preparation of $(1R^*, 6R^*, 9R^*, 13S^*)$ -13-Hydroxy-12-hydroxymethyl-3-[(S^*) -2-(4methoxyphenyl)methoxy-1-methylethyl]-6,9-dimethyltricyclo[7.5.0.0^{2,6}]tetradeca-2,11-dien-10-one (267)



To a cool (0 °C), stirred solution of keto alcohol **256** (110 mg, 0.250 mmol) in dry CH_2Cl_2 (6 mL) was added *t*-BuO₂H (70% in water, 51 uL, 0.375 mmol) *via* syringe. $VO(acac)_2$ (7 mg, 0.026 mmol) was added and the mixture was stirred for 2 h. The solution was poured into a cold (0 °C) mixture of aqueous Na₂S₂O₃ (1.0 M, 3 mL) and saturated aqueous NaHCO₃ (3 mL) and the mixture was stirred for 0.5 h. The resulting layers were separated and the aqueous layer was extracted with CH_2Cl_2 (3 × 5 mL). The combined organic layers were washed with brine (5 mL) and dried over anhydrous MgSO₄. The solvent was removed under reduced pressure and the residual material, crude keto epoxide **266**, was taken up in dry benzene (10 mL). DBN (62 mg, 0.50 mmol) was added and the mixture was heated to reflux for 6 h. The mixture was cooled to room temperature and the solvent was removed under reduced pressure. The crude oil was

purified by flash chromatography (5 g of silica gel, 50:1 Et_2O -petroleum ether) to yield 74 mg (65%) of keto diol **267** as a colourless oil.

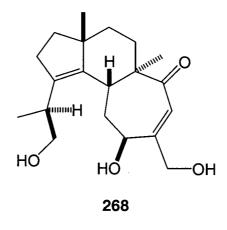
IR (neat): 3402, 2932, 1665, 1514 cm⁻¹.

¹H NMR (CDCl₃, 400 MHz) δ: 0.98 (d, 3H, *J* = 6.8 Hz), 1.03 (s, 3H), 1.09 (s, 3H), 1.20– 1.32 (m, 1H), 1.42–1.67 (m, 5H), 1.78 (dt, 1H, *J* = 4.9, 13.0 Hz), 1.98–2.09 (m, 1H), 2.13–2.34 (m, 3H), 2.42–2.57 (m, 1H), 3.09–3.20 (m, 2H), 3.32 (d, 2H, *J* = 7.9 Hz), 3.79 (s, 3H), 4.16–4.45 (m, 5H), 5.96 (s, 1H), 6.86 (d, 2H, *J* = 8.6 Hz), 7.21 (d, 2H, *J* = 8.6 Hz).

¹³C NMR (CDCl₃, 75.2 MHz) δ: 14.8, 16.5, 24.1, 39.0, 33.0, 33.9, 34.6, 36.0, 36.9, 38.1, 49.4, 55.1, 65.7, 65.8, 70.5, 72.8, 73.9, 113.8 (2 signals superimposed), 124.9, 129.3 (2 signals superimposed), 130.4, 135.6, 140.3, 150.4, 159.3, 210.1.

Exact mass calcd for C₂₈H₃₈O₅: 454.27191. Found: 454.27163.

Preparation of $(1R^*, 6R^*, 9R^*, 13S^*)$ -13-Hydroxy-12-hydroxymethyl-3-[(S^*) -2-hydroxy-1-methylethyl]-6,9-dimethyltricyclo[7.5.0.0^{2,6}]tetradeca-2,11-dien-10-one (268)



To a stirred solution of keto diol **267** (11 mg, 0.024 mmol) in a biphasic mixture of 20:1 $CH_2Cl_2-H_2O$ (10.5 mL) was added DDQ (11 mg, 0.048 mmol). The mixture was stirred at room temperature for 1 h and H₂O (10 mL) was added. The layers were separated and the aqueous layer was extracted with CH_2Cl_2 (3 × 10 mL). The combined organic layers were washed with brine (10 mL) and dried over anhydrous MgSO₄. The solvent was removed under reduced pressure and the residual material purified by flash chromatography (1 g of silica gel, 10:1 Et₂O–MeOH) to yield 5.4 mg (64%) of keto triol **268** as an amorphous solid.

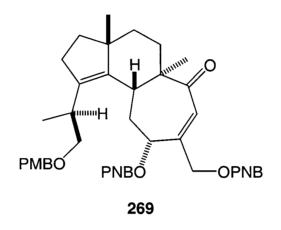
IR: not obtained

¹H NMR (CDCl₃, 400 MHz) δ: 0.96 (d, 3H, *J* = 6.7 Hz), 1.04 (s, 3H), 1.12 (s, 3H), 1.20– 1.34 (m, 2H), 1.43–1.51 (m, 1H), 1.53–1.69 (m, 4H), 1.81 (dt, 1H, *J* = 4.9, 13.4 Hz), 1.97–2.07 (m, 1H), 2.19–2.37 (m, 2H), 2.63–2.70 (m, 1H), 3.07–3.15 (m, 1H), 3.25–3.31 (m, 1H), 3.37–3.52 (m, 3H), 4.24–4.42 (m, 2H), 4.55–4.59 (m, 1H), 6.00 (s, 1H).

¹³C NMR: not obtained

Exact mass calcd for C₂₀H₃₀O₄: 334.21442. Found: 334.21369.

Preparation of $(1R^*, 6R^*, 9R^*, 13R^*)$ -3-[(S*)-2-(4-Methoxyphenyl)methoxy-1-methylethyl]-6,9-dimethyl-13-(4-nitro-benzoyl)oxy-12-(4-nitro-benzoyl)oxymethyltricyclo-[7.5.0.0^{2,6}]tetradeca-2,11-dien-10-one (269)



To a stirred solution of keto diol **267** (20 mg, 0.044 mmol) in dry benzene (2 mL) were added PPh₃ (57 mg, 0.217 mmol) and *p*-nitrobenzoic acid (33 mg, 0.193 mmol). Neat DEAD (38 mg, 0.217 mmol) was added *via* syringe and the mixture was stirred for 18 h at room temperature. The solvent was removed under reduced pressure and the crude oil was purified by flash chromatography (1 g of silica gel, 1:1 Et₂O-petroleum ether) to give 23 mg (70%) of the keto diester **269** as an amorphous solid.

IR (neat): 2920, 2850, 1729, 1688, 1608, 1529 cm⁻¹.

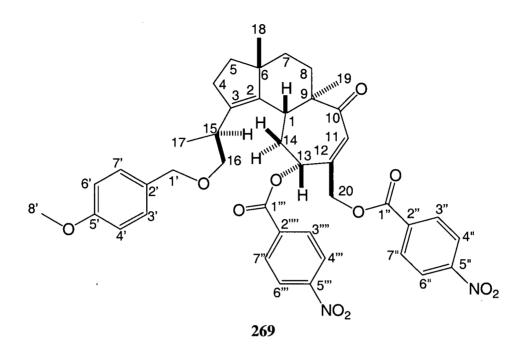
¹H NMR (CDCl₃, 400 MHz) δ: 0.98 (d, 3H, *J* = 6.7 Hz), 1.08 (s, 3H), 1.21 (s, 3H), 1.34– 1.80 (m, 6H), 2.16–2.38 (m, 3H), 2.81–2.87 (m, 1H), 2.93–2.98 (m, 1H), 3.03–3.10 (m, 1H), 3.22–3.35 (m, 2H), 3.75 (s, 3H), 4.31–4.38 (m, 2H), 5.02 (s, 2H), 5.70–5.78 (m, 1H), 6.31 (s, 1H), 6.77 (d, 2H, *J* = 8.6 Hz), 7.12 (d, 2H, *J* = 8.6 Hz), 8.06 (d, 2H, *J* = 8.2 Hz), 8.16 (d, 2H, *J* = 8.6 Hz), 8.20–8.29 (m, 4H).

¹³C NMR (CDCl₃, 75.2 MHz) δ: 14.5, 16.7, 24.0, 29.5, 31.9, 33.4, 34.7, 36.2, 38.2, 39.2, 49.3, 54.1, 55.2, 66.0, 72.6, 73.2, 73.9, 113.7, 123.6, 123.7, 128.7, 128.9, 130.7, 130.8, 130.9, 134.7, 134.8, 137.3, 138.0, 141.7, 150.8 (2 signals superimposed), 159.1, 163.8, 164.0, 208.6.

Exact mass calcd for C₄₂H₄₄N₂O₁₁: 752.2945. Parent ion was not detected.

Melting point: amorphous material displayed a broad melting point range: 40–60 °C.





Assignments H-x	¹ H NMR δ ppm (Multiplicity, J (Hz))	Observed NOEs
H–1 ^a	2.93–2.98 (m)	H–13, H–18
H4	part of m at 2.16–2.38	
H–5	part of m at 1.34–1.80	
H–7	part of m at 1.34–1.80	
H–8	part of m at 1.34-1.80	
H-11 ^a	6.31 (s)	H–20
H–13 ^a	5.70–5.78 (m)	H–1, H–14(β), H–20
Η–14(α)	part of m at 2.16–2.38	
$H-14(\beta)^a$	2.81–2.87 (m)	H–14(a), H–1, H–15
H–15 ^a	3.30–3.10 (m)	H–14, H–16, H–17
H–16 ^a	3.22–3.35 (m)	H–15, H–17
H–17 ^a	0.98 (d, 6.7)	H–15, H–16
H–18 ^a	1.08 (s)	H–1

		1
H–19	1.21 (s)	
H–20 ^{<i>a</i>}	5.02 (s)	H–11
H–1'	4.31-4.38	
H–3', H–7'	7.12 (d, 8.6)	
H4', H6'	6.77 (d, 8.6)	
H–8'	3.75 (s)	· · ·
H–3", H–7"	8.06 (d, 8.2) or 8.16 (d, 8.6)	
H4", H6"	part of m at 8.20-8.29	
H–3''', H–7'''	8.06 (d, 8.2) or 8.16 (d, 8.6)	
H–4''', H–6'''	part of m at 8.20-8.29	
^{<i>a</i>} Irradiation of this signal	gnal generated the corresponding	NOEs in the right hand column

^a Irradiation of this signal generated the corresponding NOEs in the right hand column

.

•

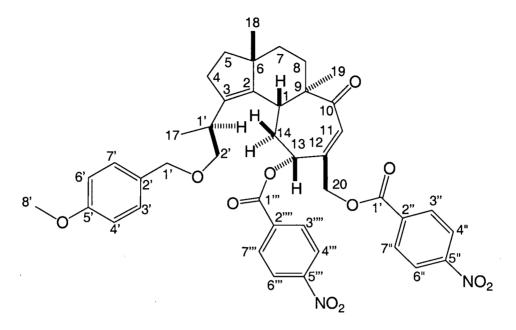


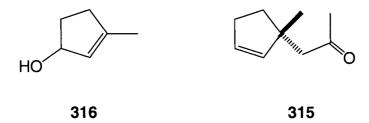
Table 20. ¹³C NMR (CDCl₃, 100.6 MHz) and ¹H NMR (CDCl₃, 400 MHz) Data for Diester **269**: HMQC Experiment.

Assignments C-x	¹³ C NMR δ ppm	Observed HMQC Correlations (δ ppm)
C-1	39.2	H–1 (2.93–2.98)
C-2	138.0	
C-3	137.3	
C-4	29.5	H-4 (part of m at 2.16-2.38)
C-5	36.2	H-5 (part of m at 1.34-1.80)
C-6	49.3	
C–7	38.2	H-7 (part of m at 1.34-1.80)
C-8	34.7	H-8 (part of m at 1.34-1.80)
C–9	54.1	
C-10	208.6	
C–11	128.9	H–11 (6.31)
C–12	141.7	
C–13	73.2	H–13 (5.70–5.78)

C-14	31.9	H–14 (2.81–2.87, part of m at 2.16–2.38)
C–15	33.4	H–15 (3.30–3.10)
C–16	73.9	H–16 (3.22–3.35)
C–17	16.7	H–17 (0.98)
C–18	24.0	H–18 (1.08)
C–19	14.5	H–19 (1.21)
C-20	66.0	H–20 (5.02)
C-1'	72.6	H–1' (4.31–4.38)
C-2'	130.7	
C-3', C-7'	128.7	H–3', H–7' (7.12)
C-4', C-6'	113.7	H4', H6' (6.77)
C-5'	159.1	
C-8'	55.2	H-8' (3.75)
C–1"	163.8 or164.0	
C–2"	134.7 or 134.8	
C–3", C–7"	130.8 or 130.9	H–3", H–7" (8.06 or 8.16)
C-4", C-6"	123.6 or 123.7	H-4", H-6" (part of m at 8.20-8.29)
C-5"	150.8	
C–1'''	163.8 or164.0	
C–2'''	134.7 or 134.8	
C–3''', C–7'''	130.8 or 130.9	H–3"", H–7"" (8.06 or 8.16)
C–4''', C–6'''	123.6 or 123.7	H–4"", H–6"" (part of m at 8.20–8.29)
C–5'''	150.8	

4.4. Synthetic Studies Directed Towards the (±)-Presilphiperfolan-9-ol

Preparation of 1-(1-Methyl-cyclopent-2-enyl)-propan-2-one (315)



To a cold (-20 °C), stirred solution of (\pm)-3-methylcyclopent-2-en-1-ol (**316**) (16.51 g, 0.168 mol) in dry Et₂O (250 mL) was added a solution of diketene (16.99 g, 0.202 mol) in dry Et₂O (25 mL) *via* cannula. A catalytic amount of DMAP (204 mg, 1.68 mmol) was added and the resulting yellow mixture was stirred for 1 h. The mixture was warmed to room temperature, stirred for 18 h and cooled to -78 °C. A cold (-78 °C) solution of LDA (0.73 M in THF, 575 mL, 0.420 mmol) was added over 15 min *via* cannula and the resulting mixture was stirred for 1 h. The mixture was warmed to 0 °C for 30 min and then to room temperature for 5 h. The mixture was extracted with aqueous NaOH (2N, 3 × 150 mL). The combined aqueous extracts were acidified by dropwise addition of concentrated HCl and back-extracted with Et₂O (3 × 300 mL). The combined ethereal extracts were washed with brine (100 mL) and dried over anhydrous MgSO₄. The solvent was removed under reduced pressure and the remaining material was taken up CCl₄ (50 mL). The mixture was heated to reflux for 12 h and cooled to room temperature. Most of the solvent was removed under reduced pressure and the residual

material was purified by flash chromatography (150 g of silica gel, 10:1 pentane $-Et_2O$) to give 17.8 g (77%) of the ketone **315** as a clear colourless oil.

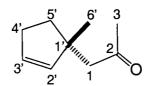
IR (neat): 3050, 2952, 1718, 1452, 1358 cm⁻¹.

¹H NMR (CDCl₃, 400 MHz) δ: 1.12 (s, 3H), 1.63–1.74 (m, 1H), 1.78–1.87 (m, 1H), 2.11 (s, 3H), 2.26–2.37 (m, 2H), 2.49 (s, 2H), 5.61–5.68 (m, 2H).

¹³C NMR (CDCl₃, 75.2 MHz) δ: 26.2, 31.2, 31.7, 37.2, 47.2, 54.0, 129.1, 139.0, 208.3.

Exact mass calcd for C₉H₁₄O: 138.10446. Found: 138.10502.

Table 21. ¹³C NMR (CDCl₃, 100.6 MHz) and ¹H NMR (CDCl₃, 400 MHz) Data for 1-(1-Methyl-cyclopent-2-enyl)-propan-2-one (**315**): HMQC Experiment.



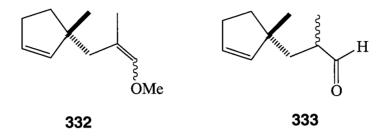
3	1	5

Assignments C–x	¹³ C NMR δ ppm	HMQC ¹ H NMR Correlations (δ ppm)
C-1	54.0	H–1 (2.49)
C-2	208.3	
C-3	31.7	H–3 (2.11)
C–1'	47.2	
C–2'	139.0	H–2' (part of m at 5.61–5.68)
C–3'	129.1	H–3' (part of m at 5.61–5.68)
C-4'	31.2	H–4' (2.26–2.37)
C–5'	37.2	H–5' (1.63–1.74, 1.78–1.87)
C–6'	26.2	H6' (1.12)

.

234

Preparation of a 1:1 Mixture of 2-Methyl-3-(1-methyl-cyclopent-2-enyl)-propionaldehydes (333)



To a cool (0 $^{\circ}$ C), stirred solution of (methoxymethyl)triphenylphosphonium chloride (84.3 g, 0.246 mol) in dry THF (1 L) was added a cold (-78 °C) solution of LDA (0.33 M in THF, 943 mL, 0.312 mol) via cannula. The resulting purple solution was stirred for 1 h. A cool (0 °C) solution of ketone **315** (17.0 g, 0.123 mol) in dry THF (100 mL) was added via cannula and the mixture was stirred for 1 h. The mixture was warmed to room temperature and stirred for 2 h. Saturated aqueous NaHCO₃ (300 mL) was added and resulting layers were separated. The aqueous layer was extracted with Et₂O (3×300 mL). Most of the solvent of the combined organic layers was removed under reduced pressure and the residual material, a crude mixture of enol ethers 332, was taken up in CHCl₃ (200 mL). TFA (10 mL) and H_2O (10 mL) were added and the resulting mixture was heated to reflux for 18 h. The solution was cooled to room temperature and H₂O (200ml) was added. The resulting layers were separated and the organic layer was washed with aqueous NaOH (2N, 2×100 mL). The organic layer was then washed with brine (200 mL) and dried over anhydrous MgSO4. The solvent was removed under reduced pressure and the crude oil was purified by flash chromatography (500 g of silica gel, 2:1 petroleum ether– CH_2Cl_2) to yield a 13.4 g (72%) of a 1:1 mixture of epimeric aldehydes **333** as clear colourless oils.

The 1:1 mixture of epimeric aldehydes 333 exhibited the following spectral data:

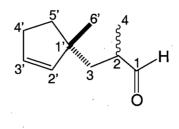
IR (neat): 2952, 1735, 1457 cm⁻¹.

¹H NMR (CDCl₃, 400 MHz) δ: 1.02 and 1.03 (s, s, ratio 1:1, 3H), 1.04 and 1.05 (d, *J* = 7.1 Hz, d, *J* = 7.1 Hz, ratio 1:1, 3H), 1.25 and 1.32 (dd, *J* = 4.8, 14.2 Hz, dd, *J* = 3.9, 14.3 Hz, ratio 1:1, 1H), 1.53–1.72 (m, 2H), 1.87–1.99 (m, 1H), 2.22–2.41 (m, 3H), 5.37–5.44 (m, 1H), 5.56–5.63 (m, 1H), 9.51 and 9.53 (d, *J* = 2.9 Hz, d, *J* = 2.7 Hz, ratio 1:1, 1H).

¹³C NMR (CDCl₃, 75.2 MHz) δ: 15.9, 16.2, 27.1, 27.6, 31.7, 31.9, 36.8, 37.0, 42.2, 42.9, 43.6, 43.7, 48.4 (two signals superimposed), 129.4, 129.9, 139.2, 139.5, 205.2, 205.3.

Exact mass calcd for C₁₀H₁₆O: 152.12012. Found: 152.12068.

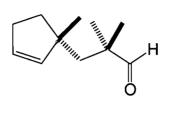
Table 22. ¹³C NMR (CDCl₃, 100.6 MHz) and ¹H NMR (CDCl₃, 400 MHz) Data for the 1:1 Mixture of 2-Methyl-3-(1-methyl-cyclopent-2-enyl)-propionaldehydes (**333**): HMQC Experiment.



2	2	2
- 4	- 4	. 4
-	-	-

Assignment C-x	¹³ C NMR ^a δ ppm	HMQC ¹ H NMR Correlations ^a δ ppm (Multiplicity, J (Hz))
C-1	205.2, 205.3	H–1 (9.51, 9.53)
C-2	31.7, 31.9	H-2 (part of m at 2.22-2.41)
C-3	36.8, 37.0	H–3 (1.53–1.72)
C4	15.9, 16.2	H-4 (1.04, 1.05)
C-1'	48.4	
C-2'	139.2, 139.5	H–2' (5.37–5.44)
C-3'	129.4, 129.9	H–3' (5.56–5.63)
C-4'	43.6, 43.7	H-4' (part of m at 2.22-2.41)
C-5'	42.2, 42.9	H–5' (1.25, 1.32, 1.87–1.99)
C6'	27.1, 27.6	H–6' (1.02, 1.03)

^{*a*} The respective signals of both isomers of 333 are noted.



314

To a stirred suspension of KH (3.81 g, 0.095 mol) in dry THF (500 mL) was added a solution of a 1:1 mixture of epimeric aldehydes **333** (13.0 g, 0.086 mol) in dry THF (50 mL). The resulting yellow mixture was stirred at room temperature for 1 h and cooled to 0 °C. Neat MeI (13.5 g, 0.095 mol) was added and the mixture was stirred for 1 h. The resulting slurry was slowly poured into a stirred mixture of 1:1 Et₂O–H₂O (0.5 L). The resulting layers were separated and the aqueous layer was extracted with Et₂O (3 × 200 mL). The combined organic layers were washed with brine (100 mL) and dried over anhydrous MgSO₄. The solvent was removed under reduced pressure and the crude oil was purified by flash chromatography (300 g of silica gel, 15:1 pentane–Et₂O) to yield 12.8 g (90%) of aldehyde **314** as a clear colourless oil.

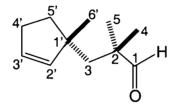
IR (neat): 3050, 2956, 1727, 1456 cm⁻¹.

¹H NMR (CDCl₃, 400 MHz) δ: 0.93 (s, 3H), 1.01 (s, 3H), 1.04 (s, 3H), 1.50–1.83 (m, 4H), 2.20–2.30 (m, 2H), 5.45–5.49 (m, 2H), 5.53–5.58 (m, 1H), 9.44 (s, 1H).

¹³C NMR (CDCl₃, 75.2 MHz) δ: 22.9, 23.7, 27.7, 31.3, 39.1, 46.6, 48.4, 49.3, 128.9, 140.4, 206.6.

Exact mass calcd for $C_{11}H_{18}O$: 166.13577. Found 166.13591.

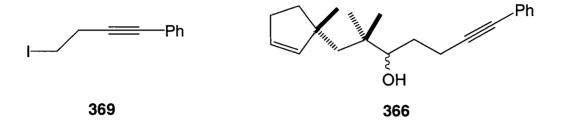
Table 23. ¹³C NMR (CDCl₃, 100.6 MHz) and ¹H NMR (CDCl₃, 400 MHz) Data for 2,2-Dimethyl-3-(1-methyl-cyclopent-2-enyl)-propionaldehyde (**314**): HMQC Experiment.



314

Assignment C-x	¹³ C NMR δ ppm	HMQC ¹ H NMR Correlations δ ppm (Multiplicity, J (Hz))	
C-1	206.6	H–1 (9.44)	
C-2	46.6		
C-3	49.3	H–3 (part of m at 1.50–1.83)	
C-4	22.9	H–4 (1.04)	
C-5	23.7	H–5 (1.01)	
C-1'	48.4		
C-2'	140.4	H-2' (5.45-5.49)	
C-3'	128.9	H–3' (5.53–5.58)	
C-4'	31.3	H-4' (2.20-2.30)	
C-5'	39.1	H-5' (part of m at 1.50-1.83)	
C6'	27.7	H-6' (0.93)	

Preparation of a 1:1 Mixture of 2,2-Dimethyl-1-(1-methyl-cyclopent-2-enyl)-7phenyl-hept-6-yn-3-ols (366)



To a cold (-78 °C), stirred solution of iodide 369^{81} (1.71 g, 6.69 mmol) in dry Et₂O (40 mL) was added a solution of *t*-BuLi (1.7 M in pentane, 8.26 mL, 14.05 mmol) *via* syringe over 2 minutes. The mixture was stirred for 1 h, warmed to 0 °C for 15 min and then recooled to -78 °C for 30 min. A solution of aldehyde **314** (1.0 g, 6.02 mmol) in dry Et₂O (10 mL) was added *via* cannula and the mixture was stirred for 30 min. H₂O (25 mL) was added and the mixture was warmed to room temperature. The resulting layers were separated and the aqueous layer was extracted with Et₂O (3 × 30 mL). The combined organic layers were washed with brine (30 mL) and dried over anhydrous MgSO₄. The solvent was removed under reduced pressure and the residual material was purified by flash chromatography (40 g of silica gel, 5:1 petroleum ether–Et₂O) to give 1.40 g (78%) of a 1:1 mixture of alcohols **366** as a clear colourless oil.

The 1:1 mixture of alcohols **366** exhibited the following spectral data:

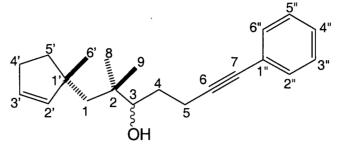
IR (neat): 3500, 3030, 2952, 2230, 1490 cm⁻¹.

¹H NMR (CDCl₃, 400 MHz) δ: 0.96 and 0.98 (s, s, ratio 1:1, 3H), 0.99 (s, 3H), 1.11 and 1.12 (s, s, ratio 1:1, 3H), 1.32–1.89 (m, 7H), 2.27–2.35 (m, 2H), 2.46–2.65 (m, 2H), 3.42–3.51 (m, 1H), 5.51–5.59 (m, 1H), 5.60–5.72 (m, 1H), 7.22–7.29 (m, 3H), 7.35–7.39 (m, 2H).

¹³C NMR (CDCl₃, 75.2 MHz) δ: 17.1 (two signals superimposed), 24.6, 24.7, 24.8 (two signals superimposed), 28.9, 29.0, 30.2, 30.3, 31.2, 31.4, 38.9 (two signals superimposed), 39.5, 40.3, 48.1 (two signals superimposed), 48.7, 48.9, 78.6, 79.3, 81.1 (two signals superimposed), 90.0, 90.1, 123.8, 123.9, 127.6, 128.2, 128.3 (four signals superimposed), 131.5 (two signals superimposed), 141.3, 141.7.

Exact mass calcd for C₂₁H₂₈O: 296.21402. Found 296.21390.

Table 24. ¹³C NMR (CDCl₃, 100.6 MHz) and ¹H NMR (CDCl₃, 400 MHz) Data for a 1:1 Mixture of 2,2-Dimethyl-1-(1-methyl-cyclopent-2-enyl)-7-phenyl-hept-6-yn-3-ol (**366**): HMQC Experiment.

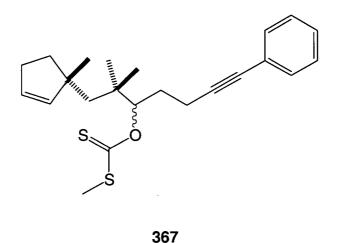


366

Assignment C-x	¹³ C NMR ^{<i>a</i>} δ ppm	HMQC ¹ H NMR Correlations ^{<i>a</i>} δ ppm (Multiplicity, <i>J</i> (Hz))	
C-1	48.1	H-1 (part of m at 1.32-1.89)	
C-2	38.9		
C-3	78.6, 79.3	H–3 (3.42–3.51)	
C-4	30.2, 30.3	H-4 (part of m at 1.32-1.89)	
C-5	17.1	H–5 (2.46–2.65)	
C6	90.0, 90.1		
C-7	81.0		
C-8	24.6, 24.7	H–8 (0.96, 0.98)	
C-9	24.8	H–9 (0.99)	
C-1'	39.5, 40.3		
C-2'	141.3, 141.7	H-2' (5.60-5.72)	
C-3'	127.6, 128.2	H-3' (5.51-5.59)	
C-4'	31.2, 31.4	H-4' (2.27-2.35)	
C-5'	48.7, 48.9	H-5' (part of m at 1.32-1.89)	
C6'	28.9, 29.0	H–6' (1.11, 1.12)	
C-1"	123.8, 123.9		
C-2", C-6"	131.5	H–2", H–6" (7.35–7.39)	
C-3", C-5"	128.3	H-3", H-5" (part of m at 7.22-7.29)	
C-4"	128.3	H-4" (part of m at 7.22-7.29)	

^a The respective signals for both isomers of the 1:1 mixture of **366** were noted.

Preparation of a 1:1 Mixture of Dithiocarbonic Acid *O*-{1-[1,1-Dimethyl-cylopent-2enyl]-5-phenyl-pent-4-ynyl} Ester S-Methyl Esters (367)



To a stirred suspension of NaH (49 mg, 2.04 mmol) in dry THF (40 mL) was added a solution of a 1:1 mixture of alcohols **366** (291 mg, 1.021 mmol) in dry THF (1 mL). Carbon disulfide (233 mg, 3.06 mmol) was added *via* syringe followed by a catalytic amount of imidazole (3.5 mg, 0.05 mmol). The mixture was heated to reflux for 3 h and then cooled to room temperature. MeI (724 mg, 5.11 mmol) was added and the mixture was stirred for 30 min. H₂O (20 mL) and Et₂O (20 mL) were added and the biphasic mixture was stirred for 10 min. The resulting layers were separated and the aqueous layer was extracted with Et₂O (3 × 30 mL). The combined organic layers were washed with brine (30 mL) and dried over anhydrous MgSO₄. The solvent was removed under reduced pressure and the residual material was purified by flash chromatography (20 g of silica gel, 20:1 petroleum ether–Et₂O) to give 332 mg (84%) of a 1:1 mixture of xanthates **367** as a clear red oil.

The 1:1 mixture of xanthates 367 exhibited the following spectral data:

IR (neat): 3050, 2221, 1715, 1598, 1490, 1230, 1050 cm⁻¹.

¹H NMR (CDCl₃, 300 MHz) δ: 1.02 (s, 3H), 1.05 (s, 3H), 1.11 and 1.12 (s, s, ratio 1:1, 3H), 1.45–1.69 (m, 3H), 1.94–2.15 (m, 3H), 2.24–2.36 (m, 2H), 2.37–2.46 (m, 2H), 2.54 (s, 3H), 5.52–5.58 (m, 1H), 5.61–5.68 (m, 1H), 5.91 (br d, *J* = 10 Hz, 1H), 7.20–7.30 (m, 3H), 7.32–7.42 (m, 2H).

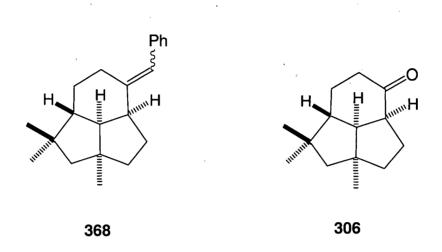
¹³C NMR (CDCl₃, 75.2 MHz) δ: 16.8, 18.9, 24.7, 24.8, 25.1, 28.7, 28.2, 29.7, 31.3, 39.5, 40.1, 48.6, 48.7, 48.9, 80.9, 89.5, 90.4, 123.8, 127.6, 128.0, 128.2, 131.6, 141.2, 141.3. Most of the ¹³C signals of the respective xanthate isomers **367** were superimposed on each other.

Exact mass calcd for $C_{23}H_{31}OS_2 (M+H)^+$: 387.18163. Found 387.18155.

Anal. calcd for C₂₃H₃₀OS₂: C 71.45, H 7.81. Found: C 71.38, H 7.81.

•

Preparation of $(1S^*, 4R^*, 7R^*, 11R^*)$ -2,2,4-Trimethyltricyclo[5,3,1,0^{4,11}]undecan-8-one (306)



To a stirred, refluxing solution of a 1:1 mixture of xanthates **367** (100 mg, 0.258 mmol) in dry, degassed benzene (3 mL) were added solutions of Bu₃SnH (0.31 M in benzene, 1 mL, 0.31 mmol) and AIBN (0.061 M in benzene, 1 mL, 0.061 mmol) *via* syringes on a double barrel syringe pump over 16 hrs. The mixture was cooled to room temperature and the solvent was removed under reduced pressure. The residual material, a mixture of olefins **368** (ratio not determined), was taken up in a 1:1:1.5 mixture of MeCN–CCl₄– H₂O (3.5 mL). NaIO₄ (150 mg, 0.69 mmol) and a catalytic amount of RuCl₃ (2 mg, 0.01 mmol) were added and the resulting black mixture was stirred at room temperature for 2 h. CH₂Cl₂ (2 mL) was added and the mixture was stirred for 10 min. The layers were separated and the aqueous layer was extracted with CH₂Cl₂ (3 × 2 mL). The combined organic layers were washed with brine (2 mL) and dried over anhydrous MgSO₄. Most of the solvent was removed under reduced pressure and the residual material was diluted

with Et_2O (5 mL). The mixture was filtered through a short column of Celite[®] and the column was flushed with Et_2O (5 mL). The solvent of the combined eluates was removed under reduced pressure and the crude oil was purified by flash chromatography (1 g of silica gel, 9:1 petroleum ether- Et_2O) to give 21.3 mg (40%) of the ketone **306** as a clear colourless oil.

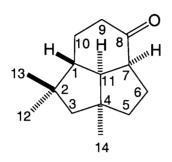
Ketone **306** exhibited the following spectral data:

IR (neat): 2950, 1719 cm⁻¹.

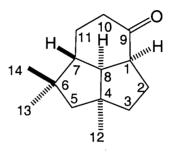
¹H NMR (CDCl₃, 400 MHz) δ: 0.88 (s, 3H), 1.01 (s, 3H), 1.07 (s, 3H), 1.40 (ddd, 1H, *J* = 4, 12.5, 13 Hz), 1.45–1.69 (m, 5H), 1.72–1.84 (m, 1H), 1.95–2.05 (m, 2H), 2.19 (dd, 1H, *J* = 7, 12.5 Hz), 2.25–2.53 (m, 2H), 2.71–2.86 (m, 1H).

¹³C NMR (CDCl₃, 100.6 MHz) δ: 21.8, 22.7, 28.2, 28.9, 29.0, 38.3, 40.6, 41.9, 47.5, 50.9, 52.7, 58.0, 59.1, 216.2

Exact mass calcd for $C_{14}H_{22}O$: 206.16707. Found 206.16727.



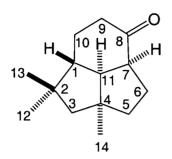
306 showing IUPAC-based numbering

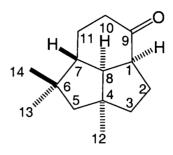


306 showing presilphiperfolane numbering

Assignment H-x (306)	¹ H NMR δ ppm (Multiplicity, J (Hz))	Presilphiper- folane Numbering H-x (306)	Literature ¹ H NMR Assignments δ ppm (Multiplicity, J (Hz))
H–1	1.40 (ddd, 4, 12.5, 13)	H–7	1.39 (ddd, 4, 12.5, 13)
H3	part of m at 1.45–1.69	H–5	1.58 and 1.68 (AB, 13)
H–5	part of m at 1.45–1.69	H–3	1.52 (m) and 1.65 (m)
H6	1.95-2.05 (m)	H–2	2.01 (m)
H–7	2.71–2.86 (m)	H–1	2.81 (ddd, 9, 8, 7)
H–9	2.25–2.53 (m)	H–10	2.31 and 2.47 (AB, 16)
H-10	part of m at 1.45–1.69 and 1.72–1.84 (m)	H–11	1.55 (m) and 1.78 (m)
H–11	2.19 (dd, 7, 12.5)	H–8	2.18 (dd, 7.12.5)
H–12	0.88 (s) or 1.01 (s)	H–13	0.88 (s) or 1.01 (s)
H–13	0.88 (s) or 1.01 (s)	H-14	0.88 (s) or 1.01 (s)
H–14	1.07 (s)	H–12	1.07 (s)

Table 26. Comparison of the ¹³C NMR (CDCl₃, 100.6 MHz) Data for $(1R^*, 4R^*, 7S^*, 8R^*)$ -4,6,6-Trimethyltricyclo[5.3.1.0^{4,11}]undecan-9-one (**306**) and those Reported²⁰ for (±)-1-Epi-9-norpresilphiperfolan-9-one (**306**).





306 showing IUPAC-based numbering

306 showing presilphiperfolane numbering

Assignment C-x (306)	¹³ C NMR δ ppm (Multiplicity, J (Hz))	Presilphiperfolane Numbering C-x (306)	Literature ¹³ C NMR Assignments δ ppm (Multiplicity, J (Hz))
C-1	50.9	C-7	50.9
C-2	40.6	C6	40.6
C-3	58.0	C-5	58.0
C-4	47.5	C-4	47.5
C-5	41.9	C-3	41.9
C-6	28.2	C-2	28.2
C–7	52.7	C-1	52.6
C-8	216.2	C-9	216.2
C9	38.3	C-10	38.3
C-10	21.8	C-11	21.8
C-11	59.1	C-8	59.1
C-12	22.7	C-13	22.7
C-13	28.9 or 29.0	C-14	29.0
C-14	28.9 or 29.0	C-12	29.0

REFERENCES AND FOOTNOTES

- (1) Ahmad, V. U.; Zahid, M.; Ali, M. S.; Ali, Z.; Jassbi, A. R.; Abbas, M.; Clardy, J.; Lobkovsky, E.; Tareen, R. B.; Iqbal, M. Z. J. Org. Chem. **1999**, 64, 8465.
- (2) Stork, G.; Burgstahler, A. W. J. Am. Chem. Soc. 1955, 77, 5068.
- (3) Eschenmoser, A.; Ruzicka, L.; Jeger, O.; Arigoni, D. *Helv. Chim. Acta* **1955**, *38*, 1890.
- (4) Xing, X.; Demuth, M. Eur. J. Org. Chem. 2001, 537.
- (5) Parsons, P. J.; Penkitt, C. S.; Shell, A. J. Chem. Rev. 1996, 96, 195.
- (6) Curran, D. P.; Kuo, S. C. J. Am. Chem. Soc. **1986**, 108, 1106.
- (7) Schwartz, C. E.; Curran, D. P. J. Am. Chem. Soc. 1990, 112, 9272.
- (8) Trost, B. M. Acc. Chem. Res. 1978, 11, 453.
- (9) Danheiser, R. L.; Gee, S. K.; Sard, H. J. Am. Chem. Soc. 1982, 104, 7670.
- (10) Seebach, D. Angew. Chem. Int. Edit. Engl. 1990, 29, 1320.
- (11) Ramachandran, S.; Newman, M. S. Org. Synth. 1961, 41, 38.
- (12) Wieland, P.; Miescher, K. Helv. Chim. Acta 1950, 33, 2215.
- (13) Cornforth, J. W.; Robinson, R. J. Chem. Soc. 1949, 1855.
- (14) Piers, E.; Renaud, J. J. Org. Chem. 1993, 58, 11.
- (15) Piers, E.; Renaud, J.; Rettig, S. J. Synthesis 1998, 590.
- (16) Shibata, H.; Takunaga, T.; Karasawa, D.; Hirota, A.; Nakayama, M.; Nozaki, H.; Tada, T. Agric. Biol. Chem. **1989**, 53, 3373.
- (17) Ayer, W. A.; Browne, L. M.; Mercer, J. R.; Taylor, D. R.; Ward, D. E. *Can. J. Chem.* **1978**, *56*, 717.
- (18) Piers, E.; Yeung, B. W. A.; Fleming, F. F. Can. J. Chem. 1993, 71, 280.
- (19) Still, W. C.; Mitra, A. J. Am. Chem. Soc. 1978, 100, 1927.

- (20) Weyerstahl, P.; Marschall, H.; Schulze, M.; Schwope, I. *Liebigs Ann.* **1996**, 799.
- (21) Brodie, H. J. Can. J. Bot. 1966, 44, 1235.
- (22) Olchowecki, A. Ph.D. Thesis, University of Alberta, Edmonton, 1967.
- (23) Albutt, A. D.; Ayer, W. A.; Brodie, H. J.; Johri, B. N.; Taube, H. Can. J. *Microbiol.* **1971**, *17*, 1401.
- (24) Ayer, W. A.; Lee, S. P.; Nakashima, T. T. Can. J. Chem. 1979, 57, 3338.
- (25) Kenmoku, H.; Kato, N.; Shimada, M.; Omoto, M.; Mori, A.; Mitsuhashi, W.; Sassa, T. *Tetrahedron Lett.* 2001, 42, 7439.
- (26) Cassidy, M. P.; Ghisalberti, E. L. J. Nat. Prod. 1993, 56, 1190.
- (27) Ayer, W. A.; Taube, H. Tetrahedron Lett. 1972, 19, 1917.
- (28) Ayer, W. A.; Carstens, L. L. Can. J. Chem. 1973, 51, 3157.
- (29) Ayer, W. A.; Lee, S. P. Can. J. Chem. 1979, 57, 3332.
- (30) Cassidy, M. P.; Ghisalberti, E. L.; Jefferies, P. R.; Skelton, B. W.; White, A. H. Aust. J. Chem. 1985, 38, 1187.
- (31) Kawagishi, H.; Shimada, A.; Shirai, R.; Okamoto, K.; Ojima, F.; Sakamoto, H.; Ishiguro, Y.; Furukawa, S. *Tetrahedron Lett.* **1994**, *35*, 1569.
- (32) Kawagishi, H.; Simada, A.; Shizuki, K.; Mori, H.; Okamoto, K.; Sakamoto, H.; Furukawa, S. *Heterocycl. Commun.* **1996**, *2*, 51.
- (33) Kawagishi, H.; Shimada, A.; Hosokawa, S.; Mori, H.; Sakamoto, H.; Ishiguro, Y.; Sakemi, S.; Bordner, J.; Kojima, N.; Furukawa, S. *Tetrahedron Lett.* **1996**, *37*, 7399.
- (34) Kenmoku, H.; Sassa, T.; Kato, N. Tetrahedron Lett. 2000, 41, 4389.
- (35) Hecht, H. J.; Hofle, G.; Steglich, W.; Anke, T.; Oberwinkler, F. J.C.S. Chem. Comm. 1978, 38, 665.
- (36) Ayer, W. A.; Yoshida, T.; Van Schie, D. M. J. *Can. J. Chem.* **1978**, *56*, 2113.

- (37) Sennett, S. H.; Pomponi, S. A.; Wright, A. E. J. Nat. Prod. **1992**, 55, 1421.
- (38) Ohta, T.; Kita, T.; Kobayashi, N.; Obara, Y.; Nakahata, N.; Ohizumi, Y.; Takaya, Y.; Oshima, Y. *Tetrahedron Lett.* **1998**, *39*, 6229.
- (39) Kita, T.; Takaya, Y.; Oshima, Y.; Ohta, T.; Aizawa, K.; Hirano, T.; Inakuma, T. *Tetrahedron* **1998**, *54*, 11877.
- (40) Obara, Y.; Nakahata, N.; Kita, T.; Takaya, Y.; Kobayashi, H.; Hosoi, S.; Kiuchi, F.; Ohta, T.; Oshima, Y.; Ohizumi, Y. Eur. J. Pharmacol. 1999, 370, 79.
- (41) Green, D.; Goldberg, I.; Stein, Z.; Ilan, M.; Kashman, Y. Nat. Prod. Lett. I 1992, 193.
- (42) The relative configuration of C-18 in structures 37, 89 and 91 was not reported. However, on the basis of the S configuration reported for C-18 in 36 (see ref. 16), analogous configurations have been tentatively assigned to the structurally-related substances 37, 89 and 91.
- (43) Hseu, T. H.; Wang, J. L.; Tang, C. P. Acta Cryst. 1980, b36, 2802.
- (44) Inchausti, A.; Yaluff, G.; deArias, A. R.; Torres, S.; Ferreira, M. E.; Nakayama, H.; Schinini, A.; Lorenzen, K.; Anke, T.; Fournet, A. *Phytother. Res.* **1997**, *11*, 193.
- (45) Saito, T.; Aoki, F.; Hirai, H.; Inagaki, T.; Matsunaga, Y.; Sakakibara, T.;
 Sakemi, S.; Suzuki, Y.; Watanabe, S.; Suga, O.; Sujaku, T.; Smogowicz,
 A. A.; Truesdell, S. J.; Wong, J. W.; Nagahisa, A.; Kojima, Y.; Kojima, N.
 J. Antibiot. 1998, 51, 983.
- Lee, E. W.; Shizuki, K.; Hosokawa, S.; Suzuki, M.; Suganuma, H.;
 Inakuma, T.; Li, J. X.; Ohnishi-Kameyama, M.; Nagata, T.; Furukawa, S.;
 Kawagishi, H. *Biosci. Biotechnol. Biochem.* 2000, 64, 2402.
- (47) Pert, C. B.; Snyder, S. H. Science **1973**, *179*, 1011.
- (48) Pasternak, G. W. *The opiate receptors*; Human Press.: Clifton, NJ, 1988.
- (49) Stein, C.; Millan, M. J.; Shippenberg, T. S.; Peter, K.; Herz, A. J. *Pharmacol. Exp. Ther.* **1989**, 248, 1269.
- (50) Obara, Y.; Kobayashi, H.; Ohta, T.; Ohizumi, Y.; Nakahata, N. Mol. *Pharmacol.* **2001**, *59*, 1287.

- (51) Connor, B.; Dragunow, M. Brain Res. Rev. 1998, 27, 1.
- (52) Tamai, I.; Tsuji, A. Adv. Drug Deliv. Rev. 1996, 19, 401.
- (53) Saltzman, W. M.; Mak, M. W.; Mahoney, M. J.; Duenas, E. T.; Cleland, J. L. Pharm. Res. 1999, 16, 232.
- (54) Wright, D. L.; Whitehead, C. R. Org. Prep. Proced. Int. 2000, 32, 307.
- (55) Snider, B. B.; Vo, N. H.; O'Neil, S. V.; Foxman, B. M. J. Am. Chem. Soc. 1996, 118, 7644.
- (56) Snider, B. B.; Vo, N. H.; O'Neil, S. V. J. Org. Chem. 1998, 63, 4732.
- (57) Tori, M.; Toyoda, N.; Sono, M. J. Org. Chem. 1998, 63, 306.
- (58) Piers, E.; Boulet, S. L. Tetrahedron Lett. 1997, 38, 8815.
- (59) Boulet, S. L.; Ph.D. Thesis, University of British Columbia, Vancouver, 1998.
- (60) Ayer, W. A.; Ward, D. E.; Browne, L. M.; Delbaere, L. T. J.; Hoyano, Y. *Can. J. Chem.* **1981**, *59*, 2665.
- (61) Ward, D. E. Can. J. Chem. 1987, 65, 2380.
- (62) Ward, D. E.; Gai, Y. Z.; Qiao, Q. Org. Lett. 2000, 2, 2125.
- (63) Dahnke, K. R.; Paquette, L. A. J. Org. Chem. 1994, 59, 885.
- (64) Wright, D. L.; Whitehead, C. R.; Sessions, E. H.; Ghiviriga, I.; Frey, D. A. Org. Lett. **1999**, *1*, 1535.
- (65) Magnus, P.; Shen, L. *Tetrahedron* **1999**, *55*, 3553.
- (66) Takeda, K.; Nakane, D.; Takeda, M. Org. Lett. 2000, 2, 1903.
- (67) Wender, P. A.; Bi, F. C.; Brodney, M. A.; Gosselin, F. Org. Lett. 2001, 3, 2105.
- (68) Piers, E.; Cook, K. L. J. Chem. Soc., Chem. Commun. 1996, 1879.
- (69) Cook, K. L.; Ph.D.Thesis, University of British Columbia, Vancouver, 1999.
- (70) Piers, E.; Gavai, A. V. J. Org. Chem. 1990, 55, 2374.

- (71) Piers, E.; Lemieux, R. M. Organometallics 1998, 17, 4213.
- (72) Kende, A. S.; Toder, B. H. J. Org. Chem. 1982, 47, 163.
- (73) Boerth, D. W.; Streitweisser, A., Jr. J. Am. Chem. Soc. 1978, 100, 750.
- (74) Nakai, T.; Mikami, K. Org. React. 1994, 46, 105.
- (75) Oikawa, Y.; Yoshioka, T.; Yonemitsu, O. Tetrahedron Lett. 1982, 23, 885.
- (76) Criegee, R. Angew. Chem. Int. Edit. Engl. 1975, 14, 745.
- (77) Odinokov, V. N.; Tolsyikov, G. A. Russ. Chem. Rev. 1981, 14, 636.
- (78) Bailey, P.; Lane, A. G. J. Am. Chem. Soc. 1967, 89, 4473.
- (79) Veysoglu, T.; Mitscher, L. A.; Swayze, J. K. Synthesis 1980, 808.
- (80) Lee, D. G.; Van den Engh, M. In *Oxidation in Organic Chemistry*; Trahanovsky, W. S., Ed.; Academic Press: New York, 1973, pp 177.
- (81) Carlsen, H. J.; Katsuki, T.; Martin, V. S.; Sharpless, K. B. J. Org. Chem. 1981, 46, 3936.
- (82) Furber, M.; Mander, L. N. J. Am. Chem. Soc. 1988, 110, 4084.
- (83) Corey, E. J. J. Am. Chem. Soc. 1954, 76, 175.
- (84) Alderdice, M.; Sum, F. W.; Weiler, L. Org. Synth. 1990, 62, 14.
- (85) Clark, J. H.; Miller, J. M. J. Chem. Soc., Perkin Trans. 1 1977, 1743.
- (86) Dowd, P.; Choi, S.-C. J. Am. Chem. Soc. 1987, 109, 3493.
- (87) Dowd, P.; Choi, S.-C. *Tetrahedron* **1989**, *45*, 77.
- (88) Beckwith, A. L. J.; O'Shea, D. M.; Gerba, S.; Westwood, S. W. J. Chem. Soc., Chem. Commun. 1987, 666.
- (89) Hasegawa, E.; Kitazume, T.; Suzuki, K.; Toseka, E. *Tetrahedron Lett.* **1998**, *39*, 4059.
- (90) Dess, D. B.; Martin, J. C. J. Org. Chem. 1983, 48, 4155.
- (91) Williams, D. R.; Nishitani, K. Tetrahedron Lett. 1980, 21, 4417.

- (92) Clinton, R. O.; Manson, A. J.; Stonner, F. W.; Clarke, R. L.; Jennings, K. F.; Shaw, P. E. J. Org. Chem. 1962, 27, 1148.
- (93) Mease, R. C.; Hirsch, J. A. J. Org. Chem. 1984, 49, 2925.
- (94) Piers, E.; Gilbert, M.; Cook, K. L. Org. Lett. 2000, 2, 1407.
- (95) The original publication by Shibata and coworkers (see ref. 16) included a misrepresentative drawing of sarcodonin G with an *R* configuration of C-18. However, the report also includes a perspective drawing obtained by crystallographic analysis, which clearly depicts an *S* configuration at C-18.
- (96) Carey, F. A.; Sunberg, R. J. Advanced Organic Chemistry; 3rd ed.; Plenum Press: New York, 1990; Vol. A.
- (97) Wilds, A. L. Org. React. 1944, 2, 178.
- (98) Woodward, R. B.; Wendler, N. L.; Brutschy, F. J. Am. Chem. Soc. 1945, 67, 1425.
- (99) Sharpless, K. B.; Michaelson, R. C. J. Am. Chem. Soc. 1973, 95, 6136.
- (100) Mitsunobu, O. Synthesis 1981, 1.
- (101) Martin, F. S.; Dodge, J. A. Tetrahedron Lett. 1991, 32, 3017.
- (102) Bohlmann, F.; Zdero, C.; Jakupovic, J.; Robinson, H.; King, R. M. *Phytochemistry* **1981**, *20*, 2239.
- (103) Marco, J. A.; Sanz-Cervera, J. F.; Morante, M. D.; GarciaLliso, V.; Valles-Xirau, J.; Jakupovic, J. *Phytochemistry* **1996**, *41*, 837.
- (104) Bohlmann, F.; Jakupovic, J. Phytochemistry 1980, 19, 259.
- (105) Weyerstahl, P.; Marschall, H.; Seelmann, I.; Jakupovic, J. Eur. J. Org. Chem. 1998, 1205.
- (106) Fehlhaber, H.-W.; Geipal, R.; Merker, H.-J.; Tschesche, R.; Welmar, K. *Chem. Ber.* **1974**, *107*, 3332.
- (107) Hanson, J. R. Pure Appl. Chem. 1981, 53, 1155.
- (108) Bradshaw, A.; Hanson, J. R.; Nyfeler, R. J. Chem. Soc., Perkin Trans. 1 1981, 1469.

- (109) Bradshaw, A. P. W.; Hanson, J. R.; Nyfeler, R.; Sadler, I. H. J. Chem. Soc., Perkin Trans. 1 1982, 2187.
- (110) Klobus, M.; Zhu, L. J.; Coates, R. M. J. Org. Chem. 1992, 57, 4327.
- (111) Coates, R. M.; Ho, Z. Q.; Klobus, M.; Wilson, S. R. J. Am. Chem. Soc. 1996, 118, 9249.
- (112) Coates, R. M.; Ho, J. Z.; Klobus, M.; Zhu, L. J. J. Org. Chem. **1998**, 63, 9166.
- (113) Shankar, S.; Coates, R. M. J. Org. Chem. 1998, 63, 9177.
- (114) Davis, C. E.; Duffy, B. C.; Coates, R. M. Org. Lett. 2000, 2, 2717.
- (115) Paquette, L. A.; Leone-Bay, A. J. Am. Chem. Soc. 1983, 105, 7352.
- (116) Piers, E.; Renaud, J. Synthesis 1992, 74.
- (117) Hosomi, A.; Sakurai, H. J. Am. Chem. Soc. 1977, 99, 1673.
- (118) McCarroll, A.; Walton, J. C. Angew. Chem. Int. Ed. Engl. 2001, 40, 2224.
- (119) Negishi, E.; Boardman, L. D.; Sawada, H.; Baheri, V.; Stoll, A. T.; Tour, J. M.; Rand, C. L. J. Am. Chem. Soc. 1988, 110, 5383.
- (120) Carroll, M. J. Chem. Soc. 1940, 1226.
- (121) Firouzababi, H.; Zeynizadeh, B. Bull. Chem. Soc. Jpn. 1997, 70, 155.
- (122) Ravikumar, K. S.; Baskaran, S.; Chandrasekaran, S. J. Org. Chem. 1993, 58, 5981.
- (123) Kim, S.; Moon, Y.; Ahn, K. H. J. Org. Chem. 1982, 47, 3311.
- (124) Krishnamurthy, S.; Brown, H. J. Org. Chem. 1977, 42, 1197.
- (125) De Clercq, P.; Zhou, X. Tetrahedron: Asymmetry 1995, 6, 1551.
- (126) Luche, J.-L. J. Am. Chem. Soc. 1978, 100, 2226.
- (127) Wilson, S. R.; Price, M. F. J. Org. Chem. 1984, 49, 722.
- (128) Levine, S. G. J. Am. Chem. Soc. **1958**, 80, 6150.

- (129) Groenewegen, P.; Kallanberg, H.; van der Gen, A. *Tetrahedron Lett.* **1978**, 5, 491.
- (130) Negishi, E.; Swanson, D. R.; Rousset, C. J. J. Org. Chem. 1990, 55, 5406.
- (131) Barton, D. H.; McCombire, S. W. J. Chem. Soc.-Perkin Trans. 1 1975, 1674.
- (132) Barton, D. H.; Parekh, S. I.; Tse, C.-L. Tetrahedron Lett. 1993, 34, 2733.
- (133) Hartwig, W. Tetrahedron 1983, 39, 2609.
- (134) Crich, D.; Quintero, L. Chem. Rev. 1989, 89, 1413.
- (135) Rawal, V. H.; Newton, R. C.; Krishnamurthy, V. J. Org. Chem. 1990, 55, 5181.
- (136) Curran, D. P.; Porter, N. A.; Giese, B. Stereochemistry of radical reactions : concepts, guidelines, and synthetic applications; Vch: New York, 1996.
- (137) Curran, D. P.; Rakiewicz, D. M. Tetrahedron 1985, 41, 3943.
- (138) Beckwith, A. L. J.; Easton, C. J.; Serelis, A. K. J. Chem. Soc., Chem. Commun. 1980, 482.
- (139) Beckwith, A. L. J.; Easton, C. J.; Lawrence, T.; Serelis, A. K. Aust. J. Chem. **1983**, *36*, 545.
- (140) Beckwith, A. L. J.; Schiesser, C. H. Tetrahedron Lett. 1985, 26, 373.
- (141) Spellmeyer, D. C.; Houk, K. N. J. Org. Chem. 1987, 52, 959.
- (142) Clive, D. L. J.; Beaulieu, P. L.; Set, L. J. Org. Chem. 1984, 49, 1314.
- (143) Fisher, M. J.; Overman, L. E. J. Org. Chem. 1990, 55, 1447.
- (144) Kofron, W. G.; Baclawaski, L. M. J. Org. Chem. 1976, 41, 1879.