

Communication

Seven New Tetrahydroanthraquinones from the Root of *Prismatomeris connata* and Their Cytotoxicity against Lung Tumor Cell Growth

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Abstract: The root of *Prismatomeris connata* has been used in China for centuries as the medicinal herb “*Huang Gen*” (HG), but its phytochemicals or active ingredients are not well understood. In this study, we performed chemical analysis of the ethyl acetate fraction of a HG ethanol extract. We thus isolated seven new tetrahydroanthraquinones, prisconnatanones C–I (compounds 1–7) from the root of *P. connata* and identified their structures using spectroscopic analyses. Their absolute configurations were established by both modified Mosher’s and Mo₂OAc₄ methods, and ORD techniques. Their cytotoxicity was tested in a panel of human lung tumor cells (H1229, HTB179, A549 and H520 cell lines). Prisconnatanone I (7) showed the highest activity, with an IC₅₀ value ranging from 2.7 μM to 3.9 μM in the suppression of tumor cell growth, and the others with chelated phenolic hydroxyls exhibited relatively lower activity (IC₅₀: 8–20 μM). In conclusion, these data suggest that some of the natural tetrahydroanthraquinones in HG are bioactive, and hydroxylation at C-1 significantly increases the cytotoxicity of these compounds against lung tumor cell growth.

Keywords: natural compound; *Prismatomeris connata*; tetrahydroanthraquinones; prisconnatanones C–I; HPLC-NMR; anti-tumor activity

1. Introduction

The root of *Prismatomeris connata*, termed “*Huang Gen*” (HG) in Chinese herbal medicine, has been used in traditional medicine in China for the treatment of hepatitis, anaemia, leucocythemia, and pneumoconiosis [1–3]. Many secondary metabolites, including anthraquinones, anthraquinone glycosides, and iridoids have been identified in HG extracts in earlier studies [4–10], and in our previous study, two tetrahydroanthraquinones (prisconnatanones A and B) have been successfully isolated, and prisconnatanone A showed a significant cytotoxicity against the A549 lung tumor cell line [9]. This study aimed at further chemical analyses of the phytochemicals in the ethyl acetate (EtOAc) fraction of a HG ethanol extract. In addition to the two known tetrahydroanthraquinone compounds, we identified seven new tetrahydroanthraquinones, prisconnatanones C–I (compounds

1–7). Herein, we report the isolation, structure identification, and biological activities of these new natural compounds.

2. Results and Discussion

The 95% EtOH HG extract was successfully fractionated with petroleum ether (PE), ethyl acetate (EtOAc), and *n*-butanol (BuOH). Seven new tetrahydroanthraquinones 1–7, which we have named prisconnatanones C–I, were then purified from the EtOAc soluble fraction by repeated column chromatography on silica gel, Sephadex LH-20, and semipreparative high-performance liquid chromatography (HPLC).

The chemical properties of these prisconnatanones C–I were determined with both ^{13}C -NMR (Table 1) and ^1H -NMR spectroscopy (Table 2). Their specific chemical structures (Figure 1) were identified according to 1D and 2D NMR spectra (Figures 2 and 3).

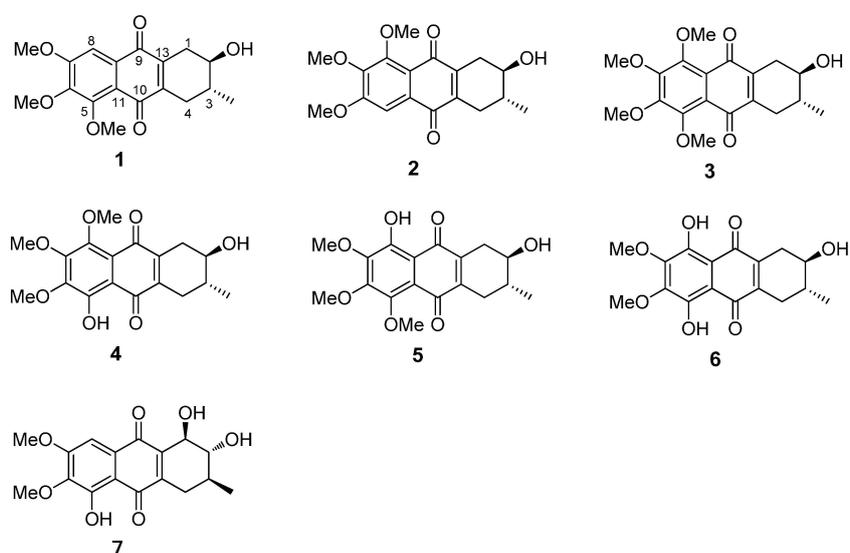


Figure 1. Chemical structure of compounds 1–7.

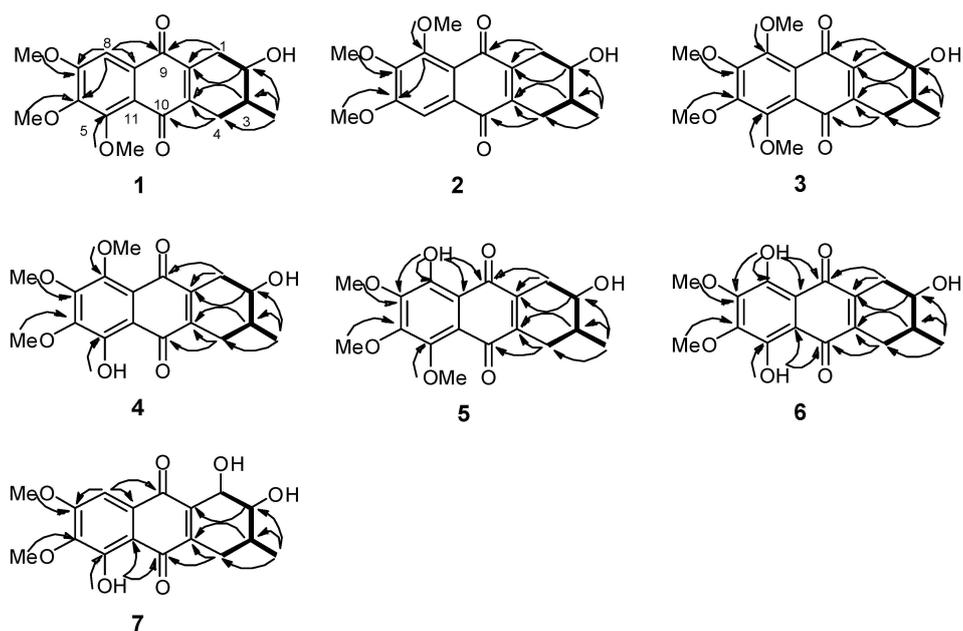


Figure 2. Key ^1H - ^1H COSY (bold) and HMBC (arrows) correlations of 1–7.

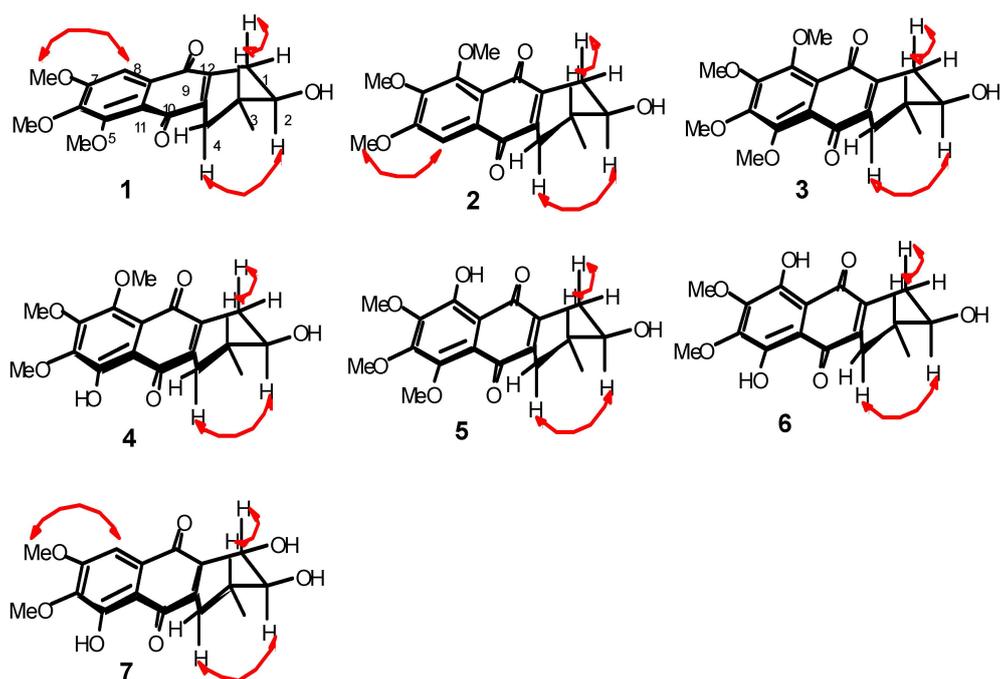


Figure 3. Key ROESY (red arrows) correlations of 1–7.

Table 1. ^{13}C -NMR data for priconnatanones C–I (1–7) ^a.

Position	1 ^b	2 ^b	3 ^b	4 ^b	5 ^b	6 ^b	7 ^b
	δ_{C} , mult.						
1	30.9, CH ₂	31.3, CH ₂	30.9, CH ₂	31.3, CH ₂	30.3, CH ₂	29.8, CH ₂	73.0, CH
2	71.0, CH	71.1, CH	71.0, CH	70.9, CH	70.8, CH	34.1, CH ₂	76.7, CH
3	34.2, CH	34.0, CH	34.0, CH	33.8, CH	34.0, CH	71.0, CH ₂	31.9, CH
4	29.9, CH ₂	29.4, CH ₂	29.4, CH ₂	28.8, CH ₂	29.9, CH ₂	31.4, CH ₂	31.1, CH ₂
5	154.7, C	106.1, C	152.7, C	154.7, C	149.2, C	159.5, C	155.9, C
6	157.3, C	147.9, C	151.2, C ^c	146.7, C	153.7, C	147.9, C	141.9, C
7	147.9, C	157.3, C	151.2, C ^c	153.7, C	146.7, C	148.0, C	158.1, C
8	106.1, C	154.3, C	152.7, C	149.1, C	154.6, C	159.6, C	104.3, CH
9	184.2, C	183.2, C	183.7, C	182.5, C	189.3, C	181.8, C	185.5, C
10	183.3, C	184.2, C	183.7, C	189.4, C	182.6, C	181.9, C	188.7, C
11	129.7, C	119.8, C	121.8, C	110.6, C	118.9, C	107.7, C	111.0, C
12	119.7, C	129.6, C	121.8, C	119.0, C	110.7, C	107.8, C	127.7, C
13	139.8, C	143.6, C	141.8, C	144.7, C	140.3, C	138.3, C	145.1, C
14	145.0, C	141.2, C	143.2, C	141.8, C	146.2, C	137.1, C	142.1, C
CH ₃ (3)	17.5	17.4	17.4	17.4	17.4	17.6	17.4
OCH ₃ (5)	61.8		61.8 ^c		61.6 ^c		
OCH ₃ (6)	56.6	61.6	61.9 ^c	61.9	62.0		61.3
OCH ₃ (7)	61.6	56.6	61.9 ^c	61.2	61.6 ^c	61.9 ^c	56.7
OCH ₃ (8)		61.8	61.8 ^c	61.6		61.9 ^c	

^a The chemical shifts in δ values (ppm) from TMS. ^{13}C multiplicities were determined by HSQC assay; ^b Readout (δ_{C} , mult.) in CDCl_3 at 150 MHz; ^c overlapped.

Table 2. ¹H-NMR Data for prisconnatanones C–I (1–7) ^a.

Position	1 ^b	2 ^b	3 ^b	4 ^b	5 ^b	6 ^b	7 ^b
	δ_H (J in Hz)	δ_H (J in Hz)	δ_H (J in Hz)	δ_H (J in Hz)	δ_H (J in Hz)	δ_H (J in Hz)	δ_H (J in Hz)
1 α	2.97 ddt (19.2, 5.0, 1.8)	2.96 br dd (19.2, 4.8)	2.91 br dd (19.2, 4.8)	3.04 ddt (19.2, 4.8, 1.8)	2.96 ddt (19.2, 4.8, 2.1)	3.01 br dd (19.0, 5.4)	4.69 ddd (7.8, 2.6, 1.1)
1 β	2.44 ddt (19.2, 7.8, 2.0)	2.42 br dd (19.2, 7.2)	2.39 ddt (19.2, 7.2, 3.0)	2.52 ddt (19.2, 7.2, 2.4)	2.45 ddt (19.2, 7.2, 1.8)	2.32 br dd (19.0, 8.4)	
2	3.69 dt (5.0, 7.8)	3.66 dt (4.8, 7.2)	3.64 dt (4.8, 7.2)	3.76 dt (4.8, 7.2)	3.71 dt (5.4, 7.2)	3.5 dt (5.4, 8.4)	3.5 dd (11.4, 7.8)
3	1.85 m	1.81 m	1.80 m	1.92 m	1.86 m	1.85 m	1.85 m
4 α	2.89 ddt (19.5, 6.6, 1.8)	2.82 dd (19.8, 6.0)	2.81 dd (19.8, 6.0)	2.92 ddt (19.8, 6.0, 1.8)	2.89 ddt (19.8, 5.4, 1.8)	3.11 dd (18.6, 5.4)	2.94 ddd (19.8, 4.8, 0.6)
4 β	2.24 ddq (19.5, 7.8, 1.8)	2.18 br dd (19.8, 8.4)	2.18 ddq (19.8, 7.8, 1.8)	2.28 ddt (19.8, 7.8, 2.4)	2.26 ddt (19.8, 8.4, 1.8)	2.55 dd (18.6, 7.8)	2.16 ddd (19.8, 11.4, 3.6)
5		7.39 s					
8	7.42 s					12.9 s	7.24 s
CH ₃ (3)	1.07 br d (7.8)	1.03 d (6.6)	1.01 d (6.6)	1.12 d (6.6)	1.07 d (7.2)	1.07 d (6.6)	1.22 d (6.6)
OH (5)				13.2 s		12.9 s	12.1 s
OCH ₃ (5)	3.90 s		3.94 s ^c		3.85 s		
OCH ₃ (6)	3.97 s	3.88 s	3.84 s ^c	4.07 s	4.01 s	4.05 s	3.99 s
OCH ₃ (7)	3.93 s	3.93 s	3.84 s ^c	4.06 s	4.02 s	4.04 s	4.00 s
OCH ₃ (8)		3.86 s	3.94 s ^c	3.90 s			
OH(8)					13.1 s	12.9 s	

^a The chemical shifts in δ values (ppm) from tetramethylsilane (TMS); ^b Readout [δ_H (J in Hz)] in CDCl₃ at 600 MHz; ^c overlapped.

2.1. Chemical Structure of Prisconnatanones C to I (Compounds 1–7)

Compound **1** formed yellow crystals, and its molecular formula was C₁₈H₂₀O₆, as determined by the high-resolution electrospray ionization mass spectrometry (HRESIMS) [M + Na]⁺ ion at *m/z* 355.1152 and ¹³C-NMR spectroscopy (Table 1), showing nine degrees of unsaturation. The infrared (IR) spectrum displayed the presence of hydroxy (3441 cm⁻¹), conjugated carbonyl (1655 cm⁻¹), and aromatic ring groups (1610, 1577, and 1459 cm⁻¹) in this compound. Based on a previous study [11] the UV spectrum suggested that there might be an anthraquinone chromophore, according to the presence of absorption peaks at 210, 268, 362 nm. The ¹³C-NMR spectrum revealed 18 characteristic signals indicating two quinone carbonyl groups (δ_C 184.2, 183.3) on positions 9 and 10, eight aromatic or olefinic carbons (δ_C 157.3, 154.3, 147.9, 145.0, 139.8, 129.7, 119.7, 106.1), five sp³ carbons (δ_C 71.0, 34.2, 30.9, 29.9, 17.5) and three methoxy groups (δ_C 61.8, 61.6, 56.6) (Table 1). The ¹H-NMR data showed an aromatic proton δ_H 7.42 (1H, s), and three methoxyls (δ_H 3.97, 3.93, 3.90, every 3H, s) (Table 2). In the comparison with the published data of prisconnatanone A [9], both the ¹H-NMR and ¹³C-NMR data were similar, suggesting that compound **1** was also a tetrahydroanthraquinone. Analysis of 1D NMR, ¹H-¹H COSY, and HSQC data revealed the presence of one 2-hydroxy-3-methylbutane unit and one pentasubstituted naphthoquinone moiety. The methyl (δ_C 17.4) was confirmed to be at C-3 by ¹H-¹H COSY (Figure 2), cross-peaks (from H-3 to H₃-Me, H-2, H-4, from H₂-1 to H-2) and HMBC correlations (from H₃-Me to C-2, C-3, and C-4) (Figure 2). The aromatic proton δ_H 7.42 (1H, s) showed HMBC correlations with C-9 (δ 184.2), C-12 (δ 119.7), C-7 (δ 157.3), suggesting that C-8 was unsubstituted and the three methoxy groups (δ_C 61.8, 61.6, 56.6) were located at C-5, C-6 and C-7. Therefore, the planar structure of compound **1** was identified as 1,2,3,4-tetrahydro-2-hydroxy-5,6,7-trimethoxy-3-methylanthracene-9,10-dione (Figure 1).

A single crystal of compound **1** was obtained by slow crystallization in methanol and water, and was mounted on a X-ray diffractometer equipped with Cu K α monochromated radiation, which allowed the compound structure to be unequivocally determined with OH-2, Me-3 in a *trans* relationship (Figure 4). The absolute configuration of the chiral centers at C-2 and C-3 of **1** was assigned as 2*R* and 3*R* by using Hooft methods, which were further confirmed by application of the advanced Mosher's method (Figure 5). Therefore, compound **1** was trivially named as prisconnatanone C and unequivocally identified as (2*R*,3*R*)-1,2,3,4-tetrahydro-2 α -hydroxy-

5,6,7-trimethoxy-3 β -methylantracene-9,10-dione (Figure 1). To the best of our knowledge, this is the first time the absolute configuration of a tetrahydroanthraquinone from *P. connata* has been reported.

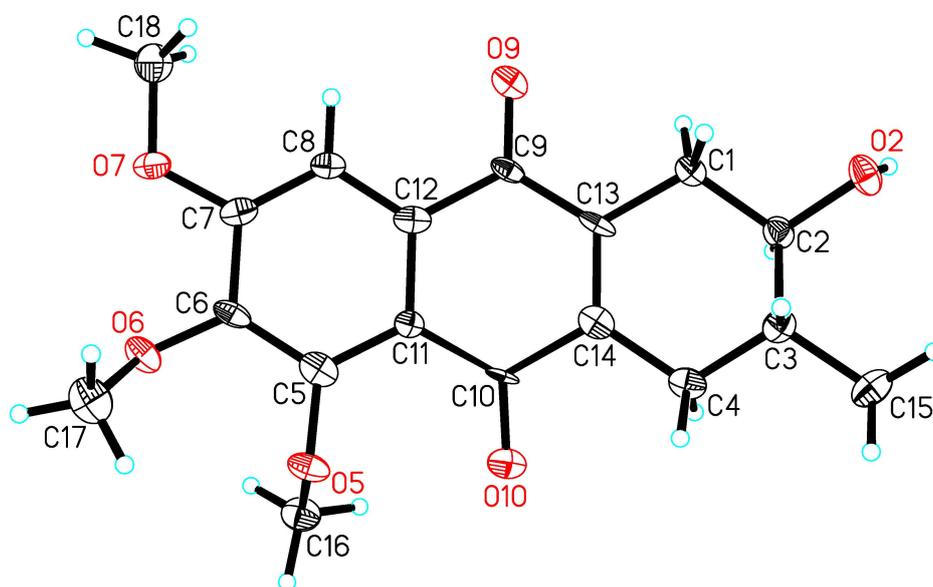


Figure 4. X-ray crystal structure of compound 1.

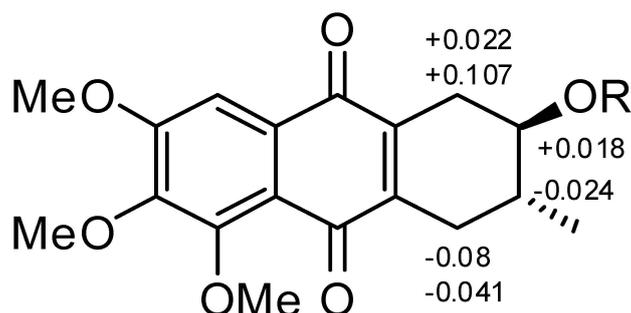


Figure 5. $\Delta\delta$ ($\delta_S - \delta_R$) values (in ppm) for the MTPA esters of 1. 1a R = (S)-MTPA; 1b R = (R) – MTPA.

Compound 2 showed a pseudomolecular $[M + Na]^+$ ion peak at m/z 355.1156, indicating the same molecular formula $C_{18}H_{20}O_6$ as 1 based on the HRESIMS data. By comparison of their 1D NMR (Tables 1 and 2), there were slight differences in two aromatic carbons and two methylene groups (Figure 1): 1 with (δ_C 145.0, 139.8, 30.9, 29.9) and 2 with (δ_C 143.6, 141.2, 31.3, 29.4). Further, HMBC correlations from H-5 to C-6, C-10 and C-11 were observed in 2, revealing that the C-5 was unsubstituted and the three methoxy groups (δ_C 56.6, 61.6, 61.8) were located at C-6, C-7 and C-8 (Figure 2). Besides, a $^3J_{H-2,H-3} = 7.2$ Hz coupling constant, in conjunction with a ROESY correlation were observed between H-1 and H_{ax-3} and between H-2 and H_{ax-4} , indicating the axial-axial relationship for these two protons and both OH-2, Me-3 in equatorial positions (Figure 3). The optical rotation (ORD) of 2, $[\alpha]_D^{21} = -55.7$ ($c = 0.16$, MeOH), had the similar sign and magnitude as seen in 1, $[\alpha]_D^{21} = -61.7$ ($c = 0.2$, MeOH), suggesting the same absolute configurations at C-2 and C-3. Thus, compound 2 was identified as (2R,3R)-1,2,3,4-tetrahydro-2 α -hydroxy-6,7,8-trimethoxy-3 β -methylantracene-9,10-dione (Figure 1) and named prisconnatanone D.

Compound 3 was a yellow powder with a pseudomolecular $[M + Na]^+$ ion at m/z 385.1258, indicating a molecular formula of $C_{19}H_{22}O_7$. The 1H - and ^{13}C -NMR spectra (Tables 1 and 2) were

similar to those of **2**, except for the presence of an additional methoxy group (Figure 1). There was no aromatic proton in the $^1\text{H-NMR}$ spectrum, suggesting that the four positions at aromatic ring of compound **3** were substituted and the extra methoxy was connected to C-8 (Figure 1). The absolute configurations at C-2 and C-3 were determined as *R* by the similar ORD between **2**, $[\alpha]_D^{21} = -55.7$ ($c = 0.16$, MeOH), and **3**, $[\alpha]_D^{21} = -57.8$ ($c = 0.2$, MeOH). By combined analysis of the 2D NMR spectra ($^1\text{H-}^1\text{H}$ COSY, HSQC, HMBC and ROESY) (Figures 2 and 3), the structure of **3** was identified as 1,2,3,4-tetrahydro-2 α -hydroxy-5,6,7,8-tetramethoxy-3 β -methylanthracene-9,10-dione (Figure 1) and it was named prisconnatanone E.

Compound **4** was a red, amorphous powder with a negative ion $[\text{M} - \text{H}]^-$ peak at m/z 347.1137, corresponding to the molecular formula $\text{C}_{18}\text{H}_{20}\text{O}_7$ on the basis of its $^{13}\text{C-NMR}$ spectroscopic and HRESIMS data, and indicating the absence of a methylene as compared with **3**. The $^1\text{H-NMR}$ (Table 2) spectra of **4** were similar to **3** except for a characteristic chelated hydroxyl group observed as a sharp singlet at δ_{H} 13.2 instead of a methoxy. Furthermore, the observation of HMBC correlations (from OH-5 to C-5, C-6, C-10 and C-11) suggested that the phenolic hydroxyl was connected to C-5 (Figure 1). According to the similar ORD between compound **2**, $[\alpha]_D^{21} = -55.7$ ($c = 0.16$, MeOH), and compound **4**, $[\alpha]_D^{21} = -68.4$ ($c = 0.105$, MeOH), the absolute configurations at C-2 and C-3 were deduced as *R*. Thus, this compound was named as prisconnatanone F, and based on combination of $^1\text{H-}^1\text{H}$ COSY, HSQC, HMBC, and ROESY (Figures 2 and 3), it was identified as 1,2,3,4-tetrahydro-2 α ,5-dihydroxy-6,7,8-trimethoxy-3 β -methylanthracene-9,10-dione (Figure 1).

Compound **5** was a red powder with a pseudomolecular ion at m/z 371.1114 ($[\text{M} + \text{Na}]^+$), suggesting a molecular formula of $\text{C}_{18}\text{H}_{20}\text{O}_7$ based on the HRESIMS data, which was the same as that of **4**. From carefully comparison of the differences between **5** and **4** as well as between **1** and **2**, there was just a small distinction between the two aromatic carbons and two methylenes (Figure 1). Based on the data of **2**, the phenolic hydroxy was placed at C-8, which was supported by HMBC correlations (from OH-8 to C-7, C-8, C-9 and C-12), and also there was similar ORD between **2**, $[\alpha]_D^{21} = -55.7$ ($c = 0.16$, MeOH), and **5**, $[\alpha]_D^{21} = -61.3$ ($c = 0.20$, MeOH), suggesting the absolute configurations at C-2 and C-3 were also *R* (Figure 2). The relative configuration of compound **5** was determined to be the same as that of **4** based on ROESY experiments and coupling constants, suggesting that **5** was isomeric with **4**. Thus, compound **5** was named prisconnatanone G and identified as 1,2,3,4-tetrahydro-2 α ,8-dihydroxy-5,6,7-trimethoxy-3 β -methylanthracene-9,10-dione (Figure 1).

Compound **6** was red, amorphous powder with a pseudomolecular ion $[\text{M} + \text{Na}]^+$ at m/z 357.0941, indicating a molecular formula of $\text{C}_{17}\text{H}_{18}\text{O}_7$ based on the HRESIMS. Comparison of its NMR spectra with those of the tetrahydroanthraquinones indicated that the $^1\text{H-}$ and $^{13}\text{C-NMR}$ data of **6** (Tables 1 and 2) were similar to those of **4**, except that a hydroxyl at δ_{H} 12.89 in **6** replaced a methoxyl in **4**. 2D NMR analysis indicated that the hydroxyl group was located at C-5. The absolute configurations 2*R* and 3*R* were determined according to the similar ORD between **1**, $[\alpha]_D^{21} = -61.7$ ($c = 0.20$, MeOH), and **6**, $[\alpha]_D^{21} = -45.3$ ($c = 0.075$, MeOH). Therefore, the structure of **6** was named as prisconnatanone H and identified as 1,2,3,4-tetrahydro-2 α ,5,8-trihydroxy-6,7-dimethoxy-3 β -methylanthracene-9,10-dione (Figure 1).

Compound **7** was a reddish brown, amorphous solid with a pseudomolecular ion $[\text{M} + \text{Na}]^+$ peak at m/z 357.0948, suggesting the molecular formula $\text{C}_{17}\text{H}_{18}\text{O}_7$ based on HRESIMS data. The $^1\text{H-}$ and $^{13}\text{C-NMR}$ (Tables 1 and 2) spectra were similar to those of **6**, except of the absence of a sharp singlet hydroxyl signal (Figure 1). Besides, two *O*-bearing methines δ 76.7 (C-1), and 73.0 (C-2), and only one methylene group δ 31.1 (C-4) were detected at a high field in the $^{13}\text{C-NMR}$, indicating the absence of the hydroxyl moved to C-1 (Figure 1). The HMBC correlations (from OH-5 to C-5, C-6, C-10, and C-11, from H-8 to C-7, C-9 and C-12) implied that phenolic hydroxyl δ_{C} 12.1 was located at C-5, and two methoxy groups (δ_{C} 61.3, 56.7) were attached to C-6 and C-7 (Figure 2). Meanwhile, the three substituents (OH-1, OH-2, Me-3) were identified at equatorial locations by the two coupling constants $^3J_{\text{H-1,H-2}} = 7.8$ Hz and

$^3J_{H-2,H-3} = 11.2$ Hz. Based on these data, compound 7 was named prisconnatanone I and identified as 1,2,3,4-tetrahydro-1 α ,2 β ,5-trihydroxy-6,7-dimethoxy-3 β -methylantracene-9,10-dione (Figure 1). The absolute configurations of this compound were further determined by using circular dichroism (CD) with the Mo₂OAc₄ method (Figures 6 and 7). A negative Cotton effect (CE) at 312 nm in the CD spectrum of 7 (Figure 7) with Mo₂OAc₄ was observed, suggesting that the absolute configuration of C-1, C-2 and C-3 could be assigned as *R,R,S*, respectively.

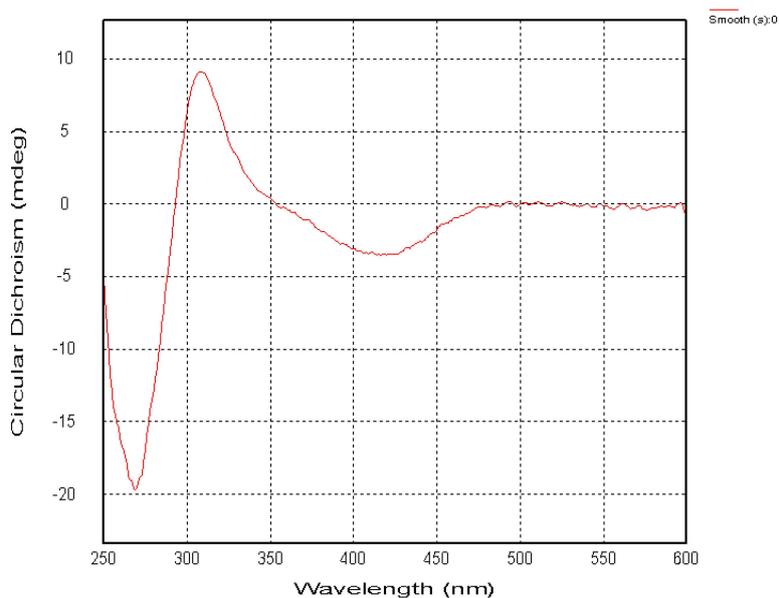


Figure 6. The circular dichroism (CD) spectrum of compound 7 in DMSO.

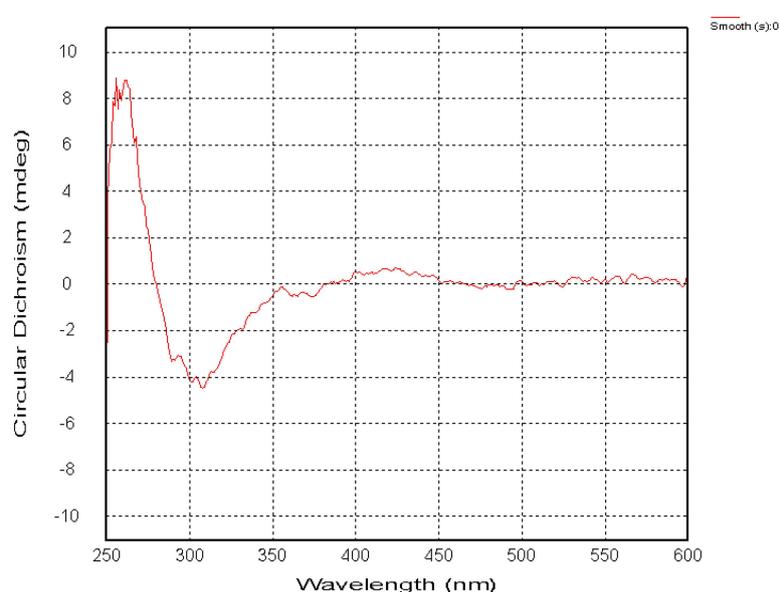


Figure 7. The circular dichroism (CD) spectrum of compound 7 with Mo₂OAc₄ in DMSO.

2.2. Structure-Dependent Cytotoxicity of Prisconnatanones C to I (1–7) against Tumor Cell Growth

The current study led to the identification of seven new compounds 1–7, which belonged to a class of tetrahydroanthraquinones by their chemical structure. Finally, the cytotoxicity of all of these compounds 1–7 at a range of up to 10 μ M was tested against a panel of four non-small cell lung tumor cell lines (H1229, HTB179, A549 and H520). As shown in Figure 8, compounds 1–3

did not show any inhibitory activity at all to any of the cell lines, while the inhibitory activities of compounds 4–6 were relatively low, and compound 7 showed the most potent cytotoxicity among these compounds against the growth of these lung tumor cells. Based on the IC_{50} values of these compounds against the growth of lung tumor cells (Table 3), their cytotoxicity could be ranked in the order of decreasing inhibitory activity as follows: Compound 7 > 6 = 4 > 5 > 1 = 2 = 3. The activity profiles of these compounds implied that the positions of hydroxyl groups (C-5 and C-8) might be necessary for their antitumor potency, and hydroxylation at C-1 could significantly enhance its cytotoxic activity; however, the underlying mechanisms remain further investigation. Cisplatin is one of first-line drugs in the treatment of lung cancer [12]. Comparing the cytotoxicity of this drug (IC_{50} : >20 μ M in H1229; 7.5 μ M in HTB179; 14.5 μ M in H549; and 8 μ M in H520) with those of the isolated compounds (Figure 8, Table 3), the cytotoxicity of 7 was higher than that of cisplatin against this panel of lung tumor cells.

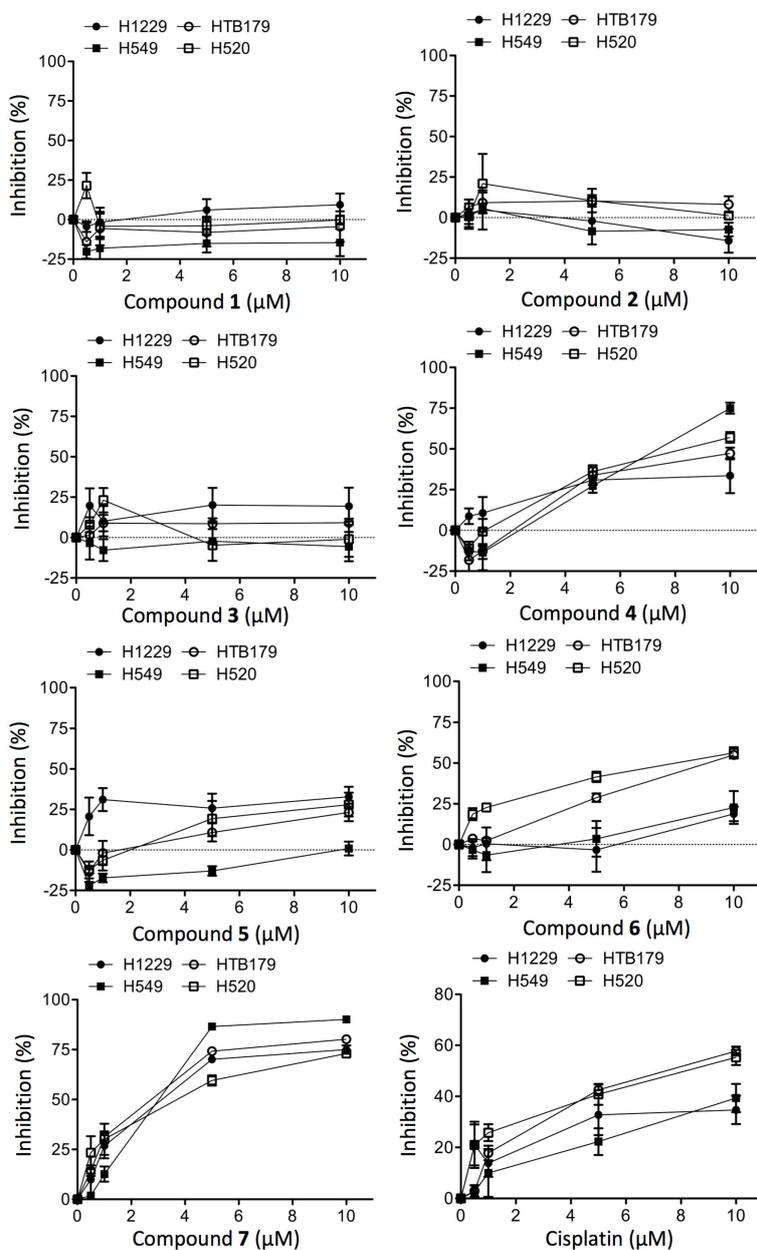


Figure 8. Inhibition of cell growth by compounds 1–7 as compared to cisplatin control in cultured human lung tumor.

Table 3. Cytotoxic activity (mean value of IC₅₀ at μM) of compounds 1–7.

Compound	H1229	HTB179	H549	H520
1	Nil	Nil	Nil	Nil
2	Nil	Nil	Nil	Nil
3	Nil	Nil	Nil	Nil
4	>20	9.8	>20	9.5
5	>20	>20	>20	>20
6	>20	13	8	9.5
7	3.0	2.7	3.0	3.9

Data are presented as a mean value (IC₅₀ at μM) of 2–3 separate experiments. Nil: no effect was found.

The tumor cell cultures were incubated in the absence (untreated control) or presence of 0.5, 1, 5 and 10 μM of a purified compound or cisplatin for 48 h. The cell viabilities at the end of incubation were determined with MTT assay. Data are presented as means ± standard derivation (SD) of six determinants in a typical experiment that was repeated 2–3 times. Dot line: x -axis ($y = 0$).

3. Experimental Section

3.1. General Experimental Procedures

The chemical analyses of purified compounds were performed as follows: optical rotations were examined using a Jasco P-1020 polarimeter (Jasco Analytical Instruments, Easton, MD, USA) in either MeOH or CHCl₃, the UV spectra using a Shimadzu UV-2401PC (Shimadzu China Co., Beijing, China) with MeOH, the CD spectra using a Chirascan spectropolarimeter (Applied Photophysics, Surrey, UK), the IR spectra using a Bruker Tensor 27 Fourier transform infrared spectrometer with KBr pellets (Bruker Beijing Scientific Tech Co., Beijing, China), the ¹H (600 MHz), ¹³C (150 MHz) and 2D NMR using Bruker AM-400 and DR-600 instruments (Bruker Beijing Scientific Tech Co.) with TMS as an internal standard in CDCl₃, the ESIMS using a Bruker HCT/E spectrometer (Bruker Beijing Scientific Tech Co.) and the HREIMS using Waters Autospec Premier P776 spectrometer (Waters Co., Milford, MA, USA). Column chromatography was performed on both silica gel (60–80; 200–300; 300–400 mesh) (Qingdao Marine Chemical Group Co., Qingdao, China) and Sephadex LH-20 (GE Healthcare LifeSciences, Little Chalfont, UK), in which the fractions were monitored by TLC (GF254, Qingdao Marine Chemical Co.). X-ray diffraction data were collected using an Agilent Technologies Gemini A Ultra system (Agilent Tech, Santa Clara, CA, USA).

3.2. Plant Material and Drug

HG was collected in June, 2011 from Nanning, Guangxi, China, and identified by Dr. Tao Chen, one of the co-authors. An authenticated voucher specimen (No. CT20110601) was deposited in the herbarium collection (SZG) at the Fairy Lake Botanical Garden (Chinese Academy of Sciences, Shenzhen, China). Cisplatin was purchased from the Pharmacy Services at Vancouver General Hospital (Vancouver, BC, Canada).

3.3. Extraction, Isolation and Identification

HG extract was prepared from the dried and powdered HG (15 kg) with 95% EtOH (3 × 40 L). The solvent was evaporated under reduced pressure until a paste-like extract was formed. The remaining residue (280 g) was suspended in H₂O, and respectively partitioned using petroleum ether, EtOAc, and BuOH. Next, the EtOAc part (94 g) was submitted to silica gel column chromatography (200–300 mesh) with a gradient elution of PE–EtOAc (from 5:1 to 1:1), and fourteen fractions (Fr.) were collected. All compounds 1–7 were purified from Fr. 12 to Fr. 14.

Fr. 12 (containing 1 and 2) was fractionated using a silica gel column (200–300 mesh) with chloroform–methanol (1000:8) to give fractions 11A–C, followed by further separation of

11A using Sephadex LH-20 column (MeOH), pressurized silica gel column (300–400 mesh) with methanol–acetone–petroleum ether (1:10:95), and then preparative HPLC (MeCN–H₂O, 60:40) to obtain **1** (73.2 mg, *t_R* 28.32 min) and **2** (17.7 mg, *t_R* 32.12 min).

Fr. 13 (containing **4** and **5**) was subjected to silica gel column chromatography (200–300 mesh) with chloroform–methanol (1000:10) to give fractions 13A–F. Fraction 13B was further separated using a Sephadex LH-20 column (MeOH) and then a pressurized silica gel column (300–400 mesh) with methanol–acetone–petroleum ether (1:10:85) to obtain **4** (34.4 mg). Fraction 13D was further purified by using a Sephadex LH-20 column (MeOH), and a pressurized silica gel column (300–400 mesh) with methanol–acetone–petroleum ether (1:10:80), followed by preparative HPLC (MeCN–H₂O, 30:70) to obtain **5** (9.1 mg, *t_R* 14.12 min).

Fr. 14 (containing **6**, **7** and **3**) was fractionated by using silica gel column chromatography (200–300 mesh) with chloroform–methanol (1000:13) as eluents to afford fractions 14A–F, of which fraction 14B was further purified by using Sephadex LH-20 column chromatography (MeOH), pressurized silica gel column chromatography (300–400 mesh) with methanol–acetone–petroleum ether (1:10:75), and then by preparative HPLC (MeOH–H₂O, 70:30) to obtain **6** (10.3 mg, *t_R* 15.32 min). Fraction 14F was purified by using Sephadex LH-20 column chromatography (MeOH), and pressurized silica gel column chromatography (300–400 mesh) with methanol–acetone–petroleum ether (1:10:70) to obtain **7** (45.9 mg). Similarly, **3** (7.7 mg) was obtained from fraction 14C by using Sephadex LH-20 column chromatography with MeOH as eluent.

3.4. Structural Analyses

1,2,3,4-Tetrahydro-2-hydroxy-5,6,7-trimethoxy-3-methylanthracene-9,10-dione (Prisconnatanone C, **1**, C₁₈H₂₀O₆): yellow acicular crystals; $[\alpha]_D^{21} = -61.7$ (*c* = 0.2, MeOH); UV (MeOH) λ_{\max} (log ϵ) 362 (2.81), 268 (3.68), 210 (3.80) nm; IR (KBr) ν_{\max} 3441, 1655, 1577, 1459 cm⁻¹; ¹H- and ¹³C-NMR, see Tables 1 and 2; ESIMS (positive) *m/z* 355 [M + Na]⁺; HREIMS (positive) *m/z* 355.1152 [M + Na]⁺ (calculated for C₁₈H₂₀O₆, 355.1158).

1,2,3,4-Tetrahydro-2 α -hydroxy-6,7,8-trimethoxy-3 β -methylanthracene-9,10-dione (Prisconnatanone D, **2**, C₁₈H₂₀O₆): yellow powder; $[\alpha]_D^{21} = -55.7$ (*c* = 0.16, MeOH); UV (MeOH) λ_{\max} (log ϵ) 361 (2.56), 267.5 (3.43), 209.5 (3.55) nm; IR (KBr) ν_{\max} 3452, 1653, 1577, 1484 cm⁻¹; ¹H- and ¹³C-NMR, see Tables 1 and 2; ESIMS (positive) *m/z* 355 [M + Na]⁺; HREIMS (positive) *m/z* 355.1156 [M + Na]⁺ (calculated for C₁₈H₂₀O₆, 355.1158).

1,2,3,4-Tetrahydro-2 α -hydroxy-5,6,7,8-tetramethoxy-3 β -methylanthracene-9,10-dione (Prisconnatanone E, **3**, C₁₉H₂₂O₇): yellow powder; $[\alpha]_D^{21} = -57.8$ (*c* = 0.2, MeOH); UV (MeOH) λ_{\max} (log ϵ) 375 (2.59), 262 (3.25), 215 (3.45) nm; IR (KBr) ν_{\max} 3433, 1654, 1546, 1465 cm⁻¹; ¹H- and ¹³C-NMR, see Tables 1 and 2; ESIMS (positive) *m/z* 355 [M + Na]⁺; HREIMS (positive) *m/z* 385.1258 [M + Na]⁺ (calculated for C₁₉H₂₂O₇, 385.1263).

1,2,3,4-Tetrahydro-2 α ,5-dihydroxy-6,7,8-trimethoxy-3 β -methylanthracene-9,10-dione (Prisconnatanone F, **4**, C₁₈H₂₀O₇): red powder; $[\alpha]_D^{21} = -68.4$ (*c* = 0.105, MeOH); UV (MeOH) λ_{\max} (log ϵ) 433 (2.95), 289 (3.17), 262.5 (3.32), 224 (3.73), 194.5 (3.46) nm; IR (KBr) ν_{\max} 3449, 1640, 1611, 1454 cm⁻¹; ¹H- and ¹³C-NMR, see Tables 1 and 2; ESIMS (positive) *m/z* 371 [M + Na]⁺; HREIMS (negative) *m/z* 347.1137 [M – H]⁺ (calculated for C₁₈H₂₀O₇, 347.1131).

1,2,3,4-Tetrahydro-2 α ,8-dihydroxy-5,6,7-trimethoxy-3 β -methylanthracene-9,10-dione (Prisconnatanone G, **5**, C₁₈H₂₀O₇): red powder; $[\alpha]_D^{21} = -61.3$ (*c* = 0.20, MeOH); UV (MeOH) λ_{\max} (log ϵ) 430.5 (2.72), 289.5 (2.94), 261.5 (3.08), 224 (3.51) nm; IR (KBr) ν_{\max} 3432, 1641, 1611, 1454 cm⁻¹; ¹H- and ¹³C-NMR, see Tables 1 and 2; ESIMS (positive) *m/z* 371 [M + Na]⁺; HREIMS (positive) *m/z* 371.1114 [M + Na]⁺ (calculated for C₁₈H₂₀O₇, 371.1107).

1,2,3,4-Tetrahydro-2 α ,5,8-trihydroxy-6,7-dimethoxy-3 β -methylanthracene-9,10-dione (Prisconnatanone H, **6**, C₁₇H₁₈O₇): red powder; $[\alpha]_D^{21} = -45.3$ ($c = 0.075$, MeOH); UV (MeOH) λ_{\max} (log ϵ) 572 (2.45), 532.5 (2.83), 498.5 (2.88), 313.5(2.84), 233(3.36), 209.5(3.32) nm; IR (KBr) ν_{\max} 3443, 1630, 1598, 1454 cm⁻¹; ¹H- and ¹³C-NMR, see Tables 1 and 2; ESIMS (positive) m/z 357 [M + Na]⁺; HREIMS (positive) m/z 357.0941 [M + Na]⁺ (calculated for C₁₇H₁₈O₇, 357.0950).

1,2,3,4-Tetrahydro-1 α ,2 β ,5-trihydroxy-6,7-dimethoxy-3 β -methylanthracene-9,10-dione (Prisconnatanone I, **7**, C₁₇H₁₈O₇): reddish brown solid; $[\alpha]_D^{21} = -360$ ($c = 0.11$, CHCl₃); UV (MeOH) λ_{\max} (log ϵ) 416 (2.80), 267.5 (3.48), 218.5 (3.66) nm; IR (KBr) ν_{\max} 3439, 1631, 1579, 1451 cm⁻¹; ¹H- and ¹³C-NMR, see Tables 1 and 2; ESIMS (positive) m/z 357 [M + Na]⁺; HREIMS (positive) m/z 357.0948 [M + Na]⁺ (calculated for C₁₇H₁₈O₇, 357.0950).

3.5. Crystal Data for 1

4(C₁₈H₂₀O₆)·CH₄O·2(H₂O), $M = 1397.43$, orthorhombic, $a = 14.2508(3)$ Å, $b = 44.4928(11)$ Å, $c = 10.7064(3)$ Å, $\alpha = 90.00^\circ$, $\beta = 90.00^\circ$, $\gamma = 90.0^\circ$, $V = 6788.5(3)$ Å³, $T = 100(2)$ K, space group $P21212$, $Z = 4$, $\mu(\text{CuK}\alpha) = 0.874$ mm⁻¹, 30337 reflections measured, 11554 independent reflections ($R_{\text{int}} = 0.1114$). Here were the final values: $R_1 = 0.1143$ ($I > 2\sigma(I)$), $wR(F^2) = 0.2875$ ($I > 2\sigma(I)$), and from all data: $R_1 = 0.1913$, and $wR(F^2) = 0.3513$. The goodness of fit on F^2 was 1.056. Flack parameter = 0.2(4). The Hooft parameter was 0.03(16) for 4723 Bijvoet pairs. The supplementary crystallographic data for this paper was deposited in the Cambridge Crystallographic Data Centre (No: 1419549). These data can be obtained free of charge via <http://www.ccdc.cam.ac.uk/conts/retrieving.html> (or from the CCDC, 12 Union Road, Cambridge CB2 1EZ, UK; Fax: +44 1223 336033; E-mail: deposit@ccdc.cam.ac.uk).

3.6. Esterification of Prisconnatanone C (1) with (R)- and (S)-MTPA

Two portions of prisconnatanone C (**1**), 12 mg, 0.036 mmol each) were treated with (R)- and (S)-MTPA in C₂H₂ with dimethylaminopyridine and *N,N'*-dicyclohexylcarbodiimide at room temperature and pressure, respectively. These two reactions were conducted in parallel, and their progress was monitored by using TLC. After the reaction was completed, and then the solvent was evaporated, the resultant materials were submitted to silica gel column chromatography (200–300 mesh) with PE–EtOAc (2:1), and further purified by using preparative HPLC (MeCN–H₂O, 80:30), from which the (S)-MTPA (**1a**, t_R 13.21 min) and (R)-MTPA (**1b**, t_R 12.54 min) esters of prisconnatanone C (**1**) were collected.

3.7. Tumor Cell Cultures

All the lung tumor cell lines were obtained from the Vancouver Prostate Centre (Vancouver, BC, Canada) and were cultured at 37 °C under 5% of CO₂ atmosphere in following culture media (Thermo Scientific, Rockford, IL, USA): H1299, H520 and A549 were cultured in Dulbecco's modified Eagle's Medium (DMEM), and HTB-179 was grown in Roswell Park Memorial Institute (RPMI) 1640 medium. Both DMEM and RPMI 1640 media were supplemented with heat-inactivated fetal bovine serum (FBS, 10%) and antibiotic mixture (100 U/mL penicillin and 100 µg/mL streptomycin) (Sigma-Aldrich Canada, Oakville, ON, Canada).

3.8. Cell Viability Assay

The effects of all of the purified compounds **1–7** on lung tumor cell growth or viability, expressed as the percentage of inhibition, were determined using a MTT (3-[4,5-dimethylthiazol-2-yl]-2,5-diphenyltetrazolium bromide, Sigma-Aldrich Canada) assay as described previously [13]. In brief, tumor cells in 96-well plates at a density of 2.5–4.0 × 10³ cells/well depending cell types were incubated in the absence (untreated control) or the presence of various concentrations of the compounds (0.5 to 10 µM) for 48 h, MTT solution (5.0 mg/mL in PBS) was added (10 µL/well),

and plates were incubated for another 4 h at 37 °C. The purple formazan crystals were dissolved in 100 µL/well of dimethyl sulfoxide (DMSO, Sigma-Aldrich Canada) for 5 min, and the absorbance (OD) was quantified at a 560 nm wavelength using an ELx808 Ultra Microplate Reader (BioTek, Winooski, VT, USA). Each experiment was repeated 2–3 times. Cell growth inhibition in drug-treated cultures against nondrug-treated control was calculated as follows: Inhibition (%) = (Control – Drug-treated)/Control × 100%. The half maximal inhibitory concentration (IC₅₀) was calculated based on the cytotoxicity curves as described previously [14].

4. Conclusions

In conclusion, this study described the structural analyses of prisconnatanones C to I (compounds 1–7) isolated from the root of *P. connata* and confirmed that these new natural compounds belonged to the rare tetrahydroanthraquinone structural class. In the cytotoxicity assay with lung tumor cells, the relationship of the chemical structure of these compounds with their activities suggested that the chelated phenolic hydroxyls might be the key functional group for their inhibitory activity as compounds 4–7 with chelated phenolic hydroxyls showed activity, while those without the group (compounds 1–3) didn't have any activity. Furthermore, because the most potent activity was found in compound 7, it was suggested that the activity might be enhanced by the numbers of oxygen substituent at the benzene ring of the compounds.

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Sample Availability: Samples of the compounds 1–7 are available from Dr. Tao Chen upon request and certain restrictions may apply.



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