



RESEARCH

Open Access

Functional analysis of the impact of *ORMDL3* expression on inflammation and activation of the unfolded protein response in human airway epithelial cells

Karolynn J Hsu and Stuart E Turvey*

Abstract

Background: The gene *ORMDL3* was shown to be associated with early-onset asthma susceptibility in multiple independent genome-wide and candidate-gene association studies. Asthmatic patients have elevated expression levels of this gene. *ORMDL3* encodes a transmembrane protein localized in the endoplasmic reticulum (ER) that may be involved in ER stress and inflammation. It is essential to validate the genetic associations linking *ORMDL3* with asthma through functional studies that confirm the biological relevance of this gene in disease. We investigated the effects of manipulating *ORMDL3* expression levels *in vitro* in airway cells on innate immune inflammatory responses, ER stress and activation of the unfolded protein response (UPR).

Methods: *ORMDL3* expression levels were manipulated in airway cells using an overexpression plasmid and siRNA technologies. Successful modulation of *ORMDL3* was confirmed at both the gene and protein level. The functional impact of modulation of *ORMDL3* expression levels on inflammatory responses and activation of the UPR were quantified using complementary cellular and molecular immunology techniques.

Results: Cells with altered *ORMDL3* levels responded equally well to innate immune stimuli and produced similar levels of pro-inflammatory cytokines compared to wild-type cells. Treatment with ER stress inducers, thapsigargin and tunicamycin, resulted in activation of the unfolded protein response (UPR). However, we observed no difference in UPR activation in cells with *ORMDL3* knockdown compared to cells with normal *ORMDL3* levels.

Conclusions: Our results suggest that *ORMDL3* variation in the airway epithelium is unlikely to play a significant role in modulating innate immune responses and the UPR in the lung.

Keywords: Immune response, Epithelium, Cytokines, Chemokines, Host defense

Introduction

Asthma and allergic diseases are rapidly becoming the most common chronic diseases in the developed world. Current asthma therapy treats symptoms of the disease, however it is ineffective in up to 25% of patients [1]. Asthma and allergic diseases are complex disorders caused by the interaction of various genetic and environmental factors [2-4].

Genome-wide association studies (GWAS) have been used to identify genes that may be involved in asthma pathogenesis [5]. Moffatt and colleagues first reported that multiple single nucleotide polymorphisms (SNPs) on chromosome 17q21 linked *ORMDL3* (orosomucoid 1-like 3) to the risk of developing childhood asthma [6]. This association has since been reproduced in multiple independent studies [7-14]. However, little work has been done to elucidate the biological and functional relevance of this gene in asthma. The disadvantage of these association studies is that they cannot differentiate between true causal SNPs and non-causal variants simply in linkage disequilibrium with disease-causing genes. It

* Correspondence: sturvey@cfri.ca

Division of Infectious and Immunological Diseases, Department of Pediatrics, BC Children's Hospital and Child & Family Research Institute, University of British Columbia, Vancouver, BC, Canada

is therefore imperative to validate GWAS data through functional studies that confirm the biological relevance of a gene in disease.

SNP variants have also linked *ORMDL3* to inflammatory bowel disease (IBD) and Type I diabetes, suggesting that *ORMDL3* may be involved in dysregulation of the immune system [15,16]. Association of *ORMDL3* in both asthma and IBD is of interest because the lung and gut are composed of similar mucosal surface cells and these tissues are exposed to many potentially harmful antigens and allergens requiring tight regulation of the mucosal immune system [17]. This unique system is responsible for maintaining a delicate equilibrium between antigen responsiveness and tolerance and is therefore responsible for preventing hyper-reactivity [17]. Inappropriate immune responses to foreign components or commensal bacteria can lead to inflammation characteristic of asthma and IBD. Furthermore, the polymorphisms may be involved in regulation of mRNA expression of 17q21 locus genes, including *ORMDL3* [6]. The expression of *ORMDL3* was recently associated with elevated levels of IL-17 secretion [18] and *ORMDL3* was expressed at higher levels in the peripheral blood of patients with recurrent wheeze compared to controls [19]. This correlation further supports the hypothesis that *ORMDL3* is involved in immunity.

The *ORMDL3* gene is a member of a family of conserved endoplasmic reticulum (ER)-localized transmembrane proteins [20]. The functions of the *ORMDL* proteins are currently unknown, but a recent study suggested that *ORMDL3* is involved in ER-mediated Ca^{2+} homeostasis and activation of the unfolded protein response (UPR) – *ORMDL3* may inhibit sarco/endoplasmic reticulum Ca^{2+} ATPase (SERCA) activity [21,22]. Disruptions to ER Ca^{2+} concentrations can cause protein misfolding, and accumulation of these unfolded proteins can lead to ER stress [23,24]. UPR signaling cascades are initiated in response to this stress and have been shown to activate the JNK-AP-1 and NF- κ B-IKK pathways [25-27]. The ER stress response and UPR, caused by changes in *ORMDL3* expression, can initiate inflammation through induction of cytokine production. This mechanism may explain the role of *ORMDL3* in asthma pathogenesis. Indeed, Miller *et al.* have shown that in mice *ORMDL3* is an allergen and cytokine (IL-4 or IL-13) inducible ER gene expressed predominantly in airway epithelial cells, and that it activates the ATF6 pathway of the ER localized UPR regulating expression of metalloprotease, chemokine, and oligoadenylate synthetase genes [28].

Although the symptoms of asthma are largely driven by dysregulated T helper type 2 ($\text{T}_{\text{H}2}$) responses, innate immune responses are also involved in asthma pathogenesis [29,30]. Airway epithelia are central to host defense and immune regulation. These cells are among the first to encounter environmental insults and play an important role

in shaping downstream immune responses. Any dysregulation of the innate immune response can result in hypersensitivity to environmental factors, leading to asthma symptoms.

Given the multiple lines of evidence suggesting that *ORMDL3* is involved in immunity, we investigated the role of the gene in innate immune responses of airway cells. We hypothesized that elevated *ORMDL3* levels result in heightened inflammatory responses that are associated with the asthmatic phenotype. Increased levels of *ORMDL3* protein may in turn disrupt ER homeostasis, leading to ER overload and activation of the UPR, initiating inflammatory responses. Using an *in vitro* model, we manipulated *ORMDL3* expression in airway cells to determine whether a difference in basal *ORMDL3* expression affected inflammatory responses or activation of the UPR before and after stimulation.

Materials and methods

Cell culture

1HAE⁺ (1HAE) cells (SV40-transformed normal human airway epithelial cells) were cultured in DMEM-high glucose medium with 10% fetal calf serum (FCS), 2 mM L-glutamine, and 1 mM sodium pyruvate (HyClone). A549 cells (adenocarcinomic human alveolar basal epithelial cells) were cultured in F-12K medium supplemented with 10% fetal calf serum (FCS), 2 mM L-glutamine, and 1 mM sodium pyruvate (HyClone). Cells were incubated in a 37°C, 5% CO₂ incubator. All cells were cultured under non-polarizing conditions.

Cloning *ORMDL3* cDNA into pEGFP-N1 vector

The *ORMDL3* gene was amplified from cDNA using forward primer 5'-CTAAGAATTCATGAATGTGGGCACAGCGCAC-3' and reverse primer 5'-TACTGGTACCCC GTACTTATTGATTCCAAAAATCCGGACT-3', introducing *Eco*RI and *Kpn*I restriction endonuclease sites, respectively. The *ORMDL3* PCR product was then inserted into a pEGFP-N1 eukaryotic expression vector (Clontech). *ORMDL3* and *eGFP* are in frame and produce a fusion protein with eGFP expressed at the C-terminus of *ORMDL3*. The construct was verified by sequencing and is denoted as pEGFP-*ORMDL3*. Protein is denoted as *ORMDL3*-eGFP.

Cell transfection

A549 and 1HAE cell lines were transfected with pEGFP-*ORMDL3*, scramble (non-specific) or *ORMDL3*-specific siRNA (pre-designed by Qiagen) using Amaxa[®] Cell Line Nucleofector[®] Kit T (Lonza). Two *ORMDL3*-specific siRNAs were used. Concentrations used for transfection represent pooled siRNA concentration. Cells were seeded into a 24-well plate (BD Biosciences) at a

density of 2×10^5 cells/well for A549 cells or 1×10^5 cells/well for 1HAE cells.

Cell stimulation and immune response quantification

Twenty-four hours post-transfection, cells were stimulated with TNF- α (200 ng/ml) (eBioscience), *E. coli* K12 LPS (100 μ g/ml) (InvivoGen), *S. typhimurium* flagellin (10-200 ng/ml) (InvivoGen), or IL-1 β (200 ng/ml) (eBioscience). Stimulants and their concentrations were chosen based on published literature or past experiments [31-34]. Cells were stimulated for 24 hours. Supernatants were collected and analyzed for cytokine secretion. Pro-inflammatory cytokines, IL-6 and IL-8, were detected and quantified using Human IL-6 and IL-8 Ready-Set-Go![®] ELISA kits (eBioscience). Experiments were repeated three times (n = 3).

ER stress induction and UPR activation

Cells were stimulated with tunicamycin (200 μ g/mL) (Calbiochem) or thapsigargin (10 μ M) (Sigma) for 2 or 4 hours to activate the UPR. For *ORMDL3* knockdown cells, stimulation was performed 24 hours post-transfection. RNA was extracted and expression of genes *XBP-1u*, *XBP-1s*, and *CHOP* were then quantified as markers of UPR activation. For measurement of p-eIF2 α levels by Western blot, lysates from unstimulated cells with *ORMDL3* knockdown were collected 24 hours post-transfection. Experiments were repeated three times (n = 3).

RNA isolation and reverse transcription

RNA was extracted from lysates using E.Z.N.A.[®] Total RNA Kit (Omega Bio-Tek) according to the manufacturer's protocol. Extracted RNA was reverse transcribed into cDNA using the SuperScript[®] VILO[™] cDNA Synthesis Kit (Invitrogen). Complement DNA was diluted to 200 ng/ μ l prior to quantification of gene expression by qPCR. This method was followed for all samples, unless otherwise stated (see PCR Array).

Quantification of *ORMDL3* mRNA expression

Gene expression was calculated relative to *GAPDH* or *PPIA* (encoding cyclophilin A) and was quantified by SYBR Green chemistry (PerfeCTa[™] qPCR SuperMix, Quanta Biosciences) using a 7300 Real Time PCR System (Applied Biosystems). Reactions were performed in triplicate using the following cycling conditions: 50°C for 2 mins, 95°C for 10 mins, [95°C for 15 s, 60°C for 1 min] x 40. The relative expression of the measured gene was calculated by the Pfaffl method [35]. The primers used are listed in Table 1.

Western blot analysis

Cells were lysed in 50 μ l RIPA Buffer + 1x HALT[™] protease inhibitor (Thermo Scientific). Cell debris were removed by centrifugation: 18,000 x g for 10 min at 4°C. Proteins were analyzed by standard Western blotting protocols where they were transferred onto Immobilon[®]-FL transfer membrane (Millipore). Antibodies used for Western blot

Table 1 Quantitative PCR primer sequences

Gene	NCBI accession	Forward (5' → 3')	Reverse (5' → 3')	CDS region [†]	Product size
<i>GAPDH</i>	NM_002046.4	GCACCGTCAAGGCTGAGAACGG	CGACGTACTCAGCGCCAGCATC	c.173-286	114
<i>PPIA</i>	NM_021130	TAAAGCATACGGGCTCTGGCATCT	ATCCAACCACTCAGTCTTGGCAGT	c.269-369	101
<i>ACTB</i>	NM_001101.3	GTTGCGTTACACCCCTTCTT	ACCTTCACCGTTCCAGTTT	c.*16-*162	147
<i>DDIT3</i>	NM_001195053.1	GAAATGAAGAGGAAGAATCA	TTCTCCTTCATGCGCT	c.197-437	241
<i>XBP-1s</i>	NM_001079539.1	ATGGATGCCCTGTTGCTGAAGA	TGCACCTGCTGCGGACTCA	c.415-504	90
<i>XBP-1u</i>	NM_005080.3	AGCACTCAGACTACGTGCACCTCT	CCAGAAATGCCCAACAGGATATCAG	c.495-624	130
<i>ORMDL1</i>	NM_016467.4	AATGGCTGGTCCTTCAAGTGCT	ACCCTCACTGTGATGCCCTTTA	c.*121-*269	149
<i>ORMDL2</i>	NM_014182.4	ACACACTGGGAGCAAAATGGACT	AGTGCGCAGCATCATACTTGGT	c.250-370	121
<i>ORMDL3</i>	NM_139280.2	TCAGGCAGCCAAAGCACTTTAACC	ACCCATCCCACACTTGCTTCCATA	c.*358-*496	139
<i>BCL6</i>	NM_0011706.4	ACAATCCCAGAAGAGGCACGAAGT	GCTCGAAATGCAGGGCAATCTCAT	c.790-952	163
<i>CCL2</i>	NM_002982.3	TCGCTCAGCCAGATGCAATCAATG	TGGAATCCTGAACCCACTTCTGCT	c.65-259	195
<i>CCL5</i>	NM_002985.2	TGCCTGTTTCTGCTTCTTGTGTC	TGTGGTAGAATCTGGGCCCTTCAA	c.*36-*127	92
<i>CSF2</i>	NM_000758.3	AAATGTTTGACCTCCAGGAGCCGA	GGTGATAATCTGGGTTGCACAGGA	c.185-357	173
<i>IL12A</i>	NM_000882.3	ATGATGGCCCTGTGCCCTTAGTAGT	AGGGCTGCATCAGCTCATCAATA	c.457-611	155
<i>IL13RA1</i>	NM_001560.2	GTCCCACTGTAGACCAATGA	CAGTCACAGCAGACTCAGGAT	c.297-391	95
<i>ADRB2</i>	NM_000024.5	TCATCATGGGCACTTTCACCTCT	AGCTCCTGGAAGCAATCCTGAAA	c.830-1016	187
<i>VEGFA</i>	NM_001025366.2	TTCAGGACATTGCTGTGCTTTGGG	TGGGCTGCTTCTTCAACAATGTG	c.*778-*969	192
<i>IL23A</i>	NM_016584.2	TCGGTGAACAACCTGAGGGAACCAA	TGGAATCTCTGCCCACTTCCACTT	c.-140- -54	87

[†] c.-number if in the 5'UTR, c.*number if in the 3'UTR.

analysis were: monoclonal anti-GFP antibody 1:10,000 (Clontech), anti-ACTB antibody 1:6,000 (Cell Signaling), anti-p-eIF2 α 1:500 (Cell Signaling) and IRDye[®] 680 or 800 secondary antibodies 1:8000 (Li-cor). Western blots were visualized using an Odyssey Infrared Imaging System (Li-cor).

PCR array

1HAE cells co-transfected with pEGFP-ORMDL3 and *ORMDL3* siRNA (low *ORMDL3* expression) were compared to cells co-transfected with pEGFP-ORMDL3 and scramble siRNA (high *ORMDL3* expression) at two time-points (2 and 24 hours) after TNF- α stimulation. Extracted RNA was reverse transcribed into first strand cDNA using the RT² First Strand Kit (SABiosciences, Qiagen). Protocol as described by the manufacturer was followed.

Two RT² Profiler PCR arrays (SABiosciences, Qiagen), profiling expression of 84 genes each, were used: Human Cytokines & Chemokines and Allergy & Asthma (see Additional file 1 for complete list). Complementary DNA template was mixed with RT² SYBR[®] Green qPCR Mastermix (SABiosciences, Qiagen) as follows: 1350 μ L SYBR Green Master Mix, 1248 μ L nuclease-free H₂O, and 102 μ L cDNA (~200 ng/ μ L). Note: these volumes were used as recommended by the manufacturer for use with a 7300 Real Time PCR System (Applied Biosystems). Template was then aliquoted into PCR plates containing pre-dispensed primers. Cycler program as provided by the manufacturer was used. Results were analyzed using the PCR Array Data Analysis Web Portal.

Statistical analysis

Data are shown as mean \pm SEM of three separate experiments. Results were analyzed using one-way ANOVA with Bonferroni post-test. Statistical analysis was performed using GraphPad Prism5 (GraphPad Software, Inc.). Differences with $p < 0.05$ were considered significant.

Results

ORMDL3 modulation in airway cells

In order to determine functional impact of *ORMDL3* modulation, knockdown of the gene was performed using siRNAs. A549 and 1HAE cells were transfected with scramble (non-specific) or *ORMDL3*-specific siRNA. Modeling variation expected to occur in the human population, we achieved 40-70% knockdown of *ORMDL3* gene expression using siRNA concentrations of 50 nM-500 nM. We also confirmed that *ORMDL3*-specific siRNA did not affect expression of genes in the same family, *ORMDL1* or *ORMDL2* (Additional file 2). Sequences of primers used for qPCR are listed in Table 1.

ORMDL3 has 84% and 80% protein sequence homology to *ORMDL1* and *ORMDL2*, respectively. This presented a challenge for confirming knockdown of *ORMDL3* protein

because commercially available antibodies detect all three proteins. Therefore, we were unable to show *ORMDL3* protein knockdown in cells transfected with siRNA alone. To overcome this limitation, we constructed a fusion protein, where *ORMDL3* is tagged with an eGFP protein. Co-transfection of this overexpression plasmid with *ORMDL3*-specific siRNA enabled us to knockdown *ORMDL3* and detect changes at the protein level using an anti-GFP antibody.

Airway cells were co-transfected with both pEGFP-*ORMDL3* and siRNA (scramble or *ORMDL3*). Protein and mRNA were analyzed for gene knockdown. At the transcript level, we observed (Figure 1) a small but significant increase ($p < 0.05$) in *ORMDL3* expression in the cells transfected with pEGFP-*ORMDL3* and scramble siRNA compared to cells alone, confirming successful overexpression of the *ORMDL3* gene. Transfection with pEGFP-*ORMDL3* and increasing concentrations of *ORMDL3* siRNA resulted in a titration effect of increasing *ORMDL3* knockdown. One advantage to constructing a fusion protein, *ORMDL3*-eGFP, is that eGFP is only expressed with expression of *ORMDL3*. Therefore, when cells are co-transfected with pEGFP-*ORMDL3* and siRNA, knockdown of *ORMDL3* protein could subsequently be detected by immunoblot analysis using the anti-GFP antibody. Expression of the *ORMDL3*-eGFP protein was confirmed by

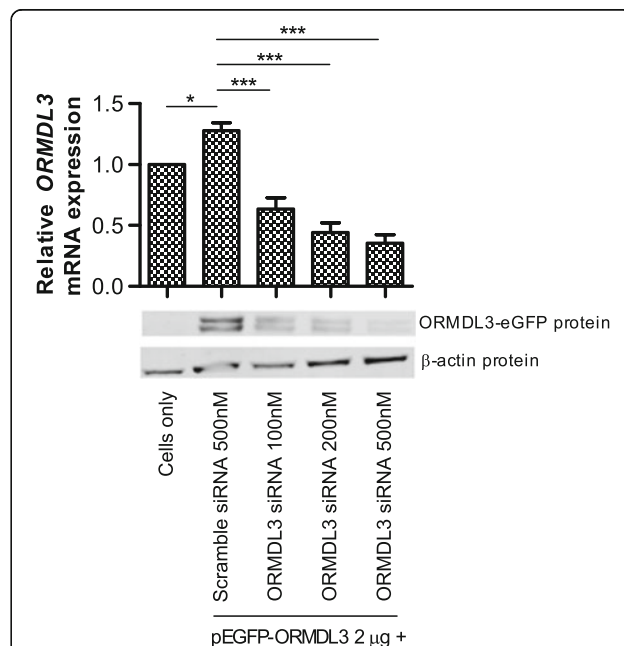


Figure 1 Experimental manipulation of *ORMDL3* expression.

Graph (top) shows relative *ORMDL3* transcript levels. Data represents the mean \pm SEM of three experimental repeats. Statistical analysis was performed using one-way ANOVA with Bonferroni post-test. * $p < 0.05$, *** $p < 0.001$. Western blot analysis (bottom) shows protein expression of *ORMDL3*-eGFP and β -actin as a loading control. Blot is representative of three independent experiments.

Western blot of whole cell lysate collected from the cells in each condition. Figure 1 shows knockdown of ORMDL3-eGFP protein, confirming that *ORMDL3* siRNA affects protein expression. The double band may be explained by either variation in mRNA splicing or post-translational modifications to the fusion protein, such as acetylation, methylation, myristylation, phosphorylation, or glycosylation.

***ORMDL3* knockdown does not affect IL-6 or IL-8 production following innate immune activation**

1HAE cells transfected with pEGFP-*ORMDL3* and scramble or *ORMDL3*-specific siRNA were stimulated 24 hours post-transfection and supernatants were collected 48 hours post-transfection. Stimuli used were TNF- α , IL-1 β , LPS, and flagellin. TNF- α and IL-1 β were chosen because both are early response cytokines that perpetuate acute inflammatory processes. LPS and flagellin, in contrast, are common microbial antigens recognized by the innate immune system. Two classic and biologically-relevant NF- κ B-induced cytokines with important roles in innate immunity, interleukin-6 (IL-6) and interleukin-8 (IL-8), were measured by ELISA. Despite confirmation of *ORMDL3* mRNA and protein knockdown, we did not observe any impact on IL-6 or IL-8 production after stimulation as shown in Figure 2A-B. Although the cells have low baseline responsiveness to LPS and flagellin, our results indicate that *ORMDL3* knockdown does not enhance sensitivity to these stimuli. Similar results were

obtained in A549 cells, as well as 1HAE cells transfected with siRNA alone (Additional file 2).

***ORMDL3* knockdown does not enhance UPR activation upon stimulation**

We next explored the effects of *ORMDL3* expression on activation of the UPR. Initiation of the UPR is mediated by one or more of the ER-membrane protein sensors: PKR-like eukaryotic initiation factor 2 α kinase (PERK), inositol requiring enzyme 1 (IRE1), and activating transcription factor-6 (ATF6) [23]. Activation of any of the three pathways initiates signaling cascades that mediate changes to relieve ER stress. The gene *XPB-1* is a substrate for IRE1 ribonuclease [24]. Upon activation of the IRE1 pathway, the IRE1 ribonuclease removes a 26-bp intron from the unspliced variant, *XPB-1u*, which results in the spliced variant, *XPB-1s* [36]. This spliced variant is the active form of the gene that contributes to ER stress responses. *CHOP* transcription, in contrast, can be induced by the PERK and ATF6 pathways [24]. Phosphorylated eIF2 α (p-eIF2 α) is an early marker of PERK pathway activation and is upstream of *CHOP* induction [23]. Expression changes in these three genes that signify UPR activation, *XPB-1u*, *XPB-1s*, and *CHOP*, were determined by qPCR. We also evaluated phosphorylation of eukaryotic initiation factor 2 α (eIF2 α) by Western blot, as modulation of *ORMDL3* expression has been reported to influence eIF2 α phosphorylation [21].

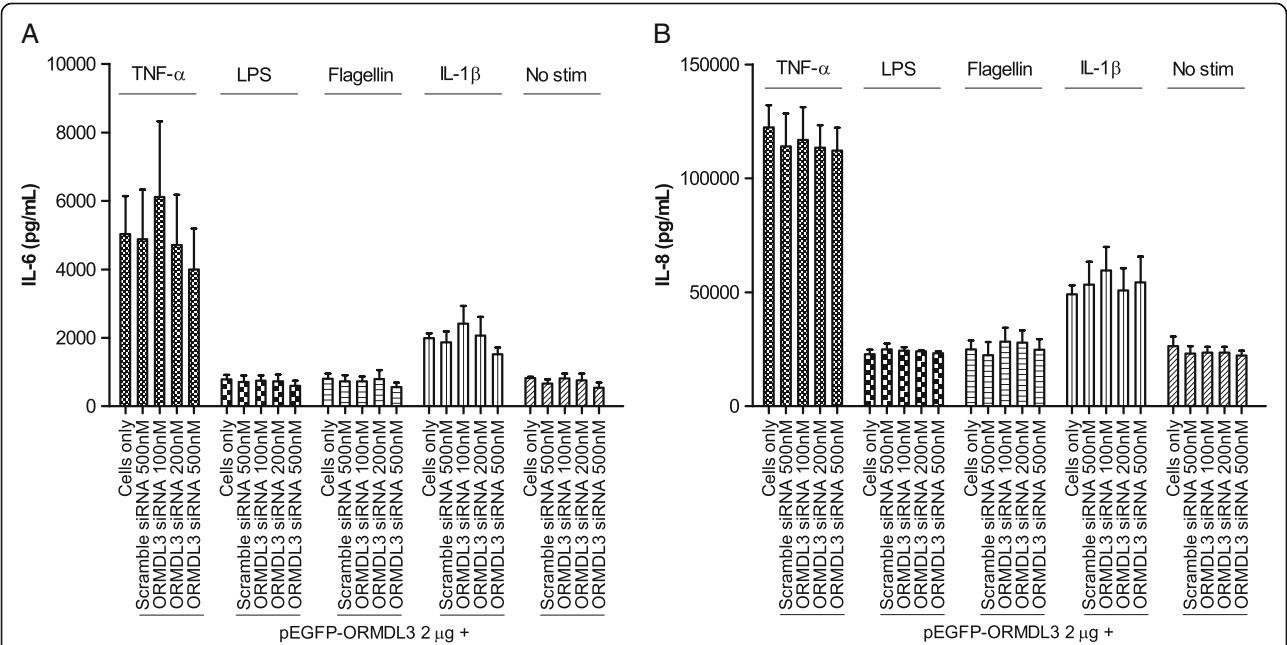


Figure 2 Cytokine production in cells with *ORMDL3* knockdown. A) Secreted IL-6 and B) IL-8 levels after cell stimulation. 1HAE cells were stimulated with TNF- α , LPS, flagellin, or IL-1 β for 24 hours. Data represent the mean \pm SEM of three experimental repeats. Statistical analysis was performed using one-way ANOVA with Bonferroni post-test.

As positive controls for UPR activation, we stimulated 1HAE cells with tunicamycin and thapsigargin. Both are inducers of ER stress – tunicamycin inhibits N-linked glycosylation and thapsigargin inhibits the SERCA pump causing ER calcium stores to be depleted [37]. Quantitative measurement of transcript levels showed that both *XBP-1s* and *CHOP* increased, while *XBP-1u* decreased upon stimulation with either tunicamycin or thapsigargin (Figure 3A-B). This confirmed the utility of measuring these genes to monitor UPR activation.

At baseline (unstimulated cells), variation in *ORMDL3* expression did not induce UPR activation in A549 or 1HAE cells (Additional file 2). 1HAE cells with *ORMDL3* knockdown were also stimulated with tunicamycin or thapsigargin (Figure 3C-D). In both conditions, *ORMDL3* knockdown did not show increased UPR activation compared to the negative control. Furthermore, levels of phosphorylated-eIF2 α were indistinguishable between *ORMDL3* knockdown cells and controls (Figure 3E).

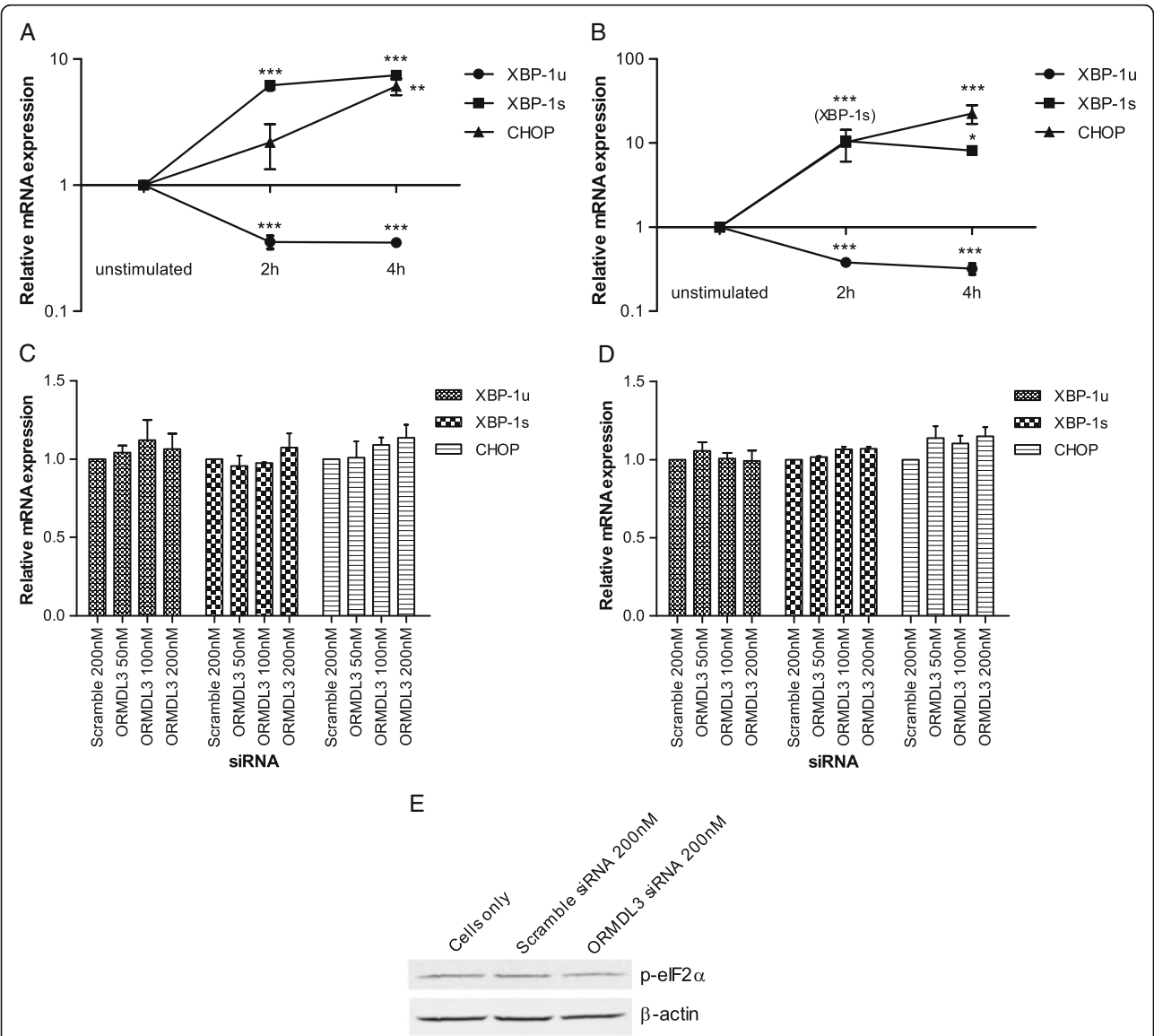


Figure 3 Unfolded protein response activation in cells with *ORMDL3* knockdown. ER stress was induced in 1HAE cells by stimulation with **A**) tunicamycin or **B**) thapsigargin for 2 or 4 hours. Relative expression levels of *XBP-1u*, *XBP-1s*, and *CHOP* were quantified and compared to unstimulated cells. 1HAE cells with *ORMDL3* knockdown were stimulated with **C**) tunicamycin or **D**) thapsigargin for 4 hours. Relative gene expression levels were quantified and compared to cells transfected with scramble (a non-specific) siRNA. Data represent the mean \pm SEM of three experimental repeats. Statistical analysis was performed using one-way ANOVA with Bonferroni post-test. *** $p < 0.001$, ** $p < 0.01$, * $p < 0.05$. **E**) Western blot analysis of p-eIF2 α with β -actin loading control. Lysates are from unstimulated 1HAE cells 24 hours after *ORMDL3* knockdown. Blot is representative of three independent experiments.

Impact of *ORMDL3* knockdown on the expression of multiple genes involved in inflammation, asthma & allergy

To expand our search for immune functions potentially altered by *ORMDL3*, the expression of 168 genes was determined at two time points (2 and 24 hours) following stimulation with TNF- α . We performed PCR arrays to profile expression of cytokines, chemokines, and key genes involved in asthma and allergy (a complete list of genes that were studied can be found in Additional file 1).

Gene expression was compared between 1HAE cells with high (plasmid + 500 nM scramble siRNA) and low (plasmid + 500 nM of *ORMDL3* siRNA) *ORMDL3* expression that were stimulated for 24 hours with TNF- α . Stimulation with TNF- α induced a robust inflammatory response in the cells, enabling us to observe whether variation in basal *ORMDL3* levels impacts the immune response. These arrays identified eight genes (*CCL2*, *TSLP*, *CSF2*, *CCL5*, *VEGFA*, *ADRB2*, *IL1RL1*, and *IL13RA1*), shown in Figure 4, that appeared to be differentially regulated by more than 1.5 fold and that were expressed at relatively high levels (average threshold cycle <30). However, upon replication to validate these results, we determined that the differences were not statistically significant.

The same arrays were performed on cells stimulated for 2 hours with TNF- α . The same conditions for “high” and “low” *ORMDL3* expression were used. From our results, we identified only one gene, *IL23A*, that was differentially regulated by more than 1.5 fold (fold regulation -2.37)

and amplified at cycle <30. Replication and comparison of *IL23A* expression between knockdown conditions yielded no significant difference. No other genes, as identified in the previous arrays, were found to be differentially regulated at this time point. These results suggest that *ORMDL3* variation does not have a meaningful impact on expression of a large panel of immune-related genes in airway epithelial cells.

Discussion

Asthma is a complex disease affecting many individuals in the developed world. Genome-wide association studies have recently been used to identify genetic causes for such complex diseases. One particular gene, *ORMDL3*, is of interest because of its association with asthma, IBD, and Type I diabetes – all of which are caused by immune-mediated pathology [6,10,22,38,39]. The gene *ORMDL3* is an ER-membrane protein and is potentially involved in Ca²⁺-signaling in the ER and sphingolipid synthesis [20,21,40]. It has also been correlated to activation of the UPR, though the mechanisms remain unclear [21]. Activation of the UPR may be biologically relevant, as ER stress, the UPR, and inflammation have all been linked [23]. However, the functional role *ORMDL3* in the pathogenesis of asthma has yet to be elucidated.

Airway epithelial cells play an important role in innate immunity and in the development of asthma. Current findings in literature indicate that *ORMDL3* is involved in immunity and that asthmatics have higher expression of the gene than non-asthmatics [18,21,22]. A recent study by Miller *et al.* also investigated the role of *ORMDL3* in airway epithelial cells. They reported that *in vitro* overexpression of *ORMDL3* activated the ATF6 pathway of the UPR and induced expression of several genes with potential importance in the pathogenesis of asthma [28]. Our investigation, in contrast, focuses on the effect of variation of *ORMDL3* expression levels, at baseline, on the innate immune responsiveness of airway epithelial cells. By manipulating *ORMDL3* expression *in vitro* to mimic differences in gene expression established between asthmatics and healthy individuals, we aimed to understand the role of *ORMDL3* on the innate immune response and UPR activation status in airway epithelial cells. This method ensured control and the confidence that any effect on the innate immune response was in fact correlated with a change in *ORMDL3* expression levels. If the same experiments were performed on *ex vivo* airway cells of patients, genetic and other differences between individuals could have affected the results.

After knockdown of *ORMDL3* *in vitro*, cells were stimulated with cytokines (TNF- α , IL-1 β) or common microbial components (LPS, flagellin). We monitored production of interleukin-6 (IL-6) and interleukin-8 (IL-8) (alias CXCL8), two pro-inflammatory cytokines produced by airway cells

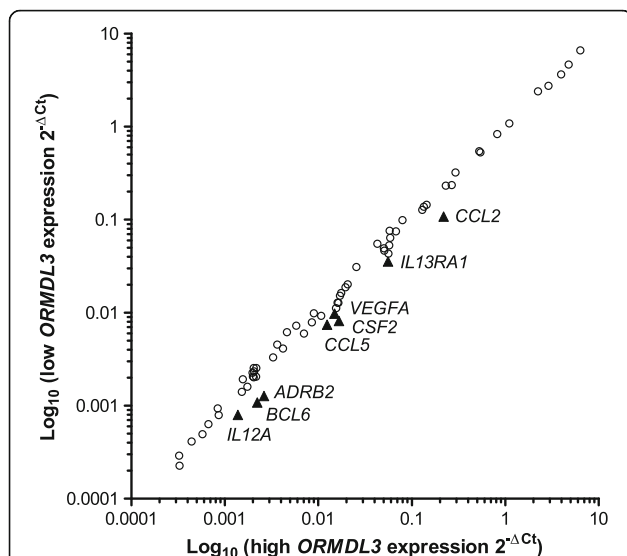


Figure 4 PCR array. Results shown are 168 genes profiled from two PCR arrays: Allergy & Asthma and Human Cytokines & Chemokines (Qiagen). 1HAE cells with high or low *ORMDL3* expression were stimulated with TNF- α for 24 hours and differences in gene expression were compared. Comparing low *ORMDL3* expression to high *ORMDL3* expression, circles are genes with less than 1.5 fold change and triangles are genes down-regulated by more than 1.5 fold.

that are relevant in asthma pathogenesis. Specifically, IL-6 is elevated in individuals with asthma [41] and is also regulated by ATF6 during activation of the UPR [42]. Similarly, transfection of *ORMDL3* into human airway epithelial cells triggers ATF6 activation and IL-8 secretion [28]. However, in our experimental system, variation in *ORMDL3* expression levels did not affect NF- κ B-induced innate immune production of IL-6 and IL-8 in airway epithelial cells.

We next explored the effects of *ORMDL3* expression on activation of the UPR. UPR signaling cascades are initiated in response to ER stress, and restoration of homeostasis is achieved by attenuating translation, restoring protein folding, or degrading misfolded proteins [24]. Although often associated with abnormal physiological conditions, the UPR plays a central beneficial role in normal physiology; as illustrated by the role of the UPR in terminal B cell differentiation which requires a massive increase in the biosynthetic capacity to synthesize antibodies in response to infection [43]. However, the ER stress response and UPR can also initiate inflammation through induction of cytokine production or activation of transcriptional regulators of inflammatory genes. Cytokines IL-6 and IL-8 are examples of genes that may be induced by UPR activation [23]. ER stress and the UPR have been implicated in many immune-related diseases including IBD, diabetes, chronic obstructive pulmonary disease (COPD), arthritis, and neurodegenerative inflammatory diseases [44]. It is poorly understood whether ER stress is an underlying cause of disease or if its induction is a result of chronic inflammation. Indeed, it is possible that environment factors such as infection or inhalation of smoke particles can activate the UPR, triggering the onset of lung disease in genetically predisposed individuals [45]. However, it is also possible that ER stress is exacerbated by inflammation and contributes to the perpetuation of the disease.

Cantero-Recasens *et al.* previously reported that *ORMDL3* overexpression activated the PERK pathway, but did not affect the IRE1 pathway of the UPR [21]. In contrast, Miller *et al.* reported that *ORMDL3* overexpression activated the ATF6 pathway, but not the PERK or IRE1 pathways [28]. In our study, we chose four markers of UPR activation: *XPB-1u*, *XPB-1s*, *CHOP*, and p-eIF2 α . With activation of the UPR, we expect downregulation of *XPB-1u* and upregulation of *XPB-1s* and *CHOP*. However, our results demonstrate that knockdown of *ORMDL3* does not activate the UPR, in either unstimulated or stimulated cells. Immunoblot analysis also showed no change in p-eIF2 α levels with *ORMDL3* knockdown. Furthermore, downstream markers of UPR activation, IL-6 and IL-8 cytokines, were produced at similar levels in unstimulated cells with varying *ORMDL3* levels. This further supports our results that *ORMDL3* does not activate the UPR. Differences in our results compared to previous work might be due to the different types of cells,

conditions, or markers used. It is possible that the effects of variation in *ORMDL3* expression are a cell type-dependent phenomenon. While no effect on the inflammatory response was detected in airway cells, other cell types such as dendritic cells or T cells may be affected by altered *ORMDL3* expression. Observations made by Lluís *et al.* suggest that the 17q21 locus may potentially play a role in T-cell development [18].

Taking a broader approach, PCR arrays looking at expression of 168 common immunity genes were performed. We reasoned that although *ORMDL3* levels may not affect the production of IL-6 or IL-8 cytokines, perhaps they were impacting gene expression of other important immune genes, such as *IL-33*, *IL-25* and *TSLP*, which have all been implicated in asthma pathogenesis [46]. Verification of differential expression of these genes at a transcript level, however, did not show any significant changes between the *ORMDL3* knockdown conditions. This suggests that altering *ORMDL3* expression does not have a profound effect on the expression of innate immune genes upon stimulation in the airway epithelia. However, there may be other genes that are affected that were not investigated in this study. Pfeifer *et al.* recently showed that IL-17C cytokine is expressed by human bronchial epithelial cells and is induced by bacterial infection [47]. It may be worthwhile in future experiments to investigate a broader range of immune-related genes. Interestingly, we did not observe changes to expression of the genes reported by Miller *et al.*, *MMP-9*, *CCL-20*, *CXCL-10*, *CXCL-11*, or *IL-8*. This variance may be explained by differences in experimental conditions. Our study examined outcomes in gene expression after stimulation of cells co-transfected with *ORMDL3* and *ORMDL3*-specific siRNA, while the other study used a different experimental approach.

Although this study focused exclusively on the potential role of *ORMDL3* in asthma pathogenesis, it is possible that neighboring genes such as *GSDML* contribute to disease susceptibility at this locus. Many groups consider *ORMDL3* as an 'asthma gene'; however, it should be acknowledged that the identified SNPs associating this gene to asthma susceptibility are not located in the gene itself. Even so, these polymorphisms have been consistently correlated with increased odds of asthma risk, highlighting the importance of this locus in disease susceptibility [6-11,13,14].

Our data show that variation in *ORMDL3* expression is not correlated with differential innate immune responses to stimuli or activation of the UPR *in vitro* in airway epithelial cells. Taken together, our results are biologically relevant because they suggest that normal human variation of *ORMDL3* expression is not likely an important factor in increasing the innate immune response of airway cells we observe in asthmatics. Despite these results, this

gene remains an important candidate for asthma susceptibility. More research is required to elucidate its role in asthma pathogenesis and its potential role as an initial trigger of inflammation. By increasing our understanding of the mechanisms responsible for allergic and atopic disease development, new treatments can then be developed. Thus, we can reduce inflammatory responses by targeting the potential triggers, rather than the symptoms, of the disease. In doing so, we will ultimately reduce the morbidity, mortality, and socio-economic burden of asthma and related allergic diseases.

Additional files

Additional file 1: Genes analyzed by PCR array.

Additional file 2: Additional figures.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

KJH performed the research. All authors designed the research, analyzed the data, and drafted the manuscript. Both authors read and approved the final manuscript.

Acknowledgements

KJH was funded by the Child & Family Research Institute and AllerGen Networks of Centers of Excellence. SET holds the Aubrey J Tingle Professorship in Pediatric Immunology and is a clinical scholar of the Michael Smith Foundation for Health Research. This work was supported by funding from the AllerGen Networks of Centers of Excellence.

Received: 3 December 2012 Accepted: 7 January 2013
Published: 1 February 2013

References

- Cho SH: Pharmacogenomic approaches to asthma treatment. *Allergy Asthma Immunol Res* 2010, **2**(3):177-182.
- Davies DE, Djukanovic R, Holgate ST: Application of functional genomics to study of inflammatory airways disease. *Thorax* 1999, **54**(1):79-81.
- Gu ML, Dong XQ, Zhao J: New insight into the genes susceptible to asthma. *J Asthma* 2010, **47**(2):113-116.
- Ober C, Hoffman S: Asthma genetics 2006: the long and winding road to gene discovery. *Genes Immun* 2006, **7**(2):p. 95-p. 100.
- Vercelli D: Discovering susceptibility genes for asthma and allergy. *Nat Rev Immunol* 2008, **8**(3):169-182.
- Moffatt MF, et al: Genetic variants regulating ORMDL3 expression contribute to the risk of childhood asthma. *Nature* 2007, **448**(7152):470-473.
- Galanter J, et al: ORMDL3 gene is associated with asthma in three ethnically diverse populations. *Am J Respir Crit Care Med* 2008, **177**(11):1194-1200.
- Halapi E, et al: A sequence variant on 17q21 is associated with age at onset and severity of asthma. *Eur J Hum Genet* 2010, **18**(8):902-908.
- Hirota T, et al: Genetic polymorphism regulating ORM1-like 3 (*Saccharomyces cerevisiae*) expression is associated with childhood atopic asthma in a Japanese population. *J Allergy Clin Immunol* 2008, **121**(3):769-770.
- Moffatt MF, et al: A large-scale, consortium-based genomewide association study of asthma. *N Engl J Med* 2010, **363**(13):1211-1221.
- Ferreira MA, et al: Association between ORMDL3, IL1RL1 and a deletion on chromosome 17q21 with asthma risk in Australia. *Eur J Hum Genet* 2011, **19**(4):458-464.
- Sleiman PM, et al: ORMDL3 variants associated with asthma susceptibility in North Americans of European ancestry. *J Allergy Clin Immunol* 2008, **122**(6):1225-1227.
- Tavendale R, et al: A polymorphism controlling ORMDL3 expression is associated with asthma that is poorly controlled by current medications. *J Allergy Clin Immunol* 2008, **121**(4):860-863.
- Wu H, et al: Genetic variation in ORM1-like 3 (ORMDL3) and gasdermin-like (GSDML) and childhood asthma. *Allergy* 2009, **64**(4):629-635.
- Barrett JC, et al: Genome-wide association study and meta-analysis find that over 40 loci affect risk of type 1 diabetes. *Nat Genet* 2009, **41**(6):703-707.
- Barrett JC, et al: Genome-wide association defines more than 30 distinct susceptibility loci for Crohn's disease. *Nat Genet* 2008, **40**(8):955-962.
- Neurath MF, Finotto S, Glimcher LH: The role of Th1/Th2 polarization in mucosal immunity. *Nat Med* 2002, **8**(6):567-573.
- Lluis A, et al: Asthma-associated polymorphisms in 17q21 influence cord blood ORMDL3 and GSDMA gene expression and IL-17 secretion. *J Allergy Clin Immunol* 2011, **127**(6):p. 1587-p. 1594. e6.
- Jin R, et al: Mechanisms elevating ORMDL3 expression in recurrent wheeze patients: role of Ets-1, p300 and CREB. *Int J Biochem Cell Biol* 2012, **44**(7):1174-1183.
- Hjelmqvist L, et al: ORMDL proteins are a conserved new family of endoplasmic reticulum membrane proteins. *Genome Biol* 2002, **3**(6):p. RESEARCH0027.
- Cantero-Recasens G, et al: The asthma-associated ORMDL3 gene product regulates endoplasmic reticulum-mediated calcium signaling and cellular stress. *Hum Mol Genet* 2009, **19**(1):111-121.
- McGovern DP, et al: Genome-wide association identifies multiple ulcerative colitis susceptibility loci. *Nat Genet* 2010, **42**(4):332-337.
- Hotamisligil GS: Endoplasmic reticulum stress and the inflammatory basis of metabolic disease. *Cell* 2010, **140**(6):900-917.
- McGuckin MA, et al: ER stress and the unfolded protein response in intestinal inflammation. *Am J Physiol Gastrointest Liver Physiol* 2010, **298**(6):G820-G832.
- Cullinan SB, Diehl JA: Coordination of ER and oxidative stress signaling: the PERK/Nrf2 signaling pathway. *Int J Biochem Cell Biol* 2006, **38**(3):317-332.
- Deng J, et al: Translational repression mediates activation of nuclear factor kappa B by phosphorylated translation initiation factor 2. *Mol Cell Biol* 2004, **24**(23):10161-10168.
- Hu P, et al: Autocrine tumor necrosis factor alpha links endoplasmic reticulum stress to the membrane death receptor pathway through IRE1alpha-mediated NF-kappaB activation and down-regulation of TRAF2 expression. *Mol Cell Biol* 2006, **26**(8):3071-3084.
- Miller M, et al: ORMDL3 is an inducible lung epithelial gene regulating metalloproteases, chemokines, OAS, and ATF6. *Proc Natl Acad Sci USA* 2012, **109**(41):16648-16653.
- Kim HY, DeKruyff RH, Umetsu DT: The many paths to asthma: phenotype shaped by innate and adaptive immunity. *Nat Immunol* 2010, **11**(7):577-584.
- Umetsu DT, et al: Asthma: an epidemic of dysregulated immunity. *Nat Immunol* 2002, **3**(8):715-720.
- Palmberg L, et al: Induction of IL-8 production in human alveolar macrophages and human bronchial epithelial cells in vitro by swine dust. *Thorax* 1998, **53**(4):260-264.
- Shanks KK, et al: Interleukin-8 production by human airway epithelial cells in response to *Pseudomonas aeruginosa* clinical isolates expressing type a or type b flagellins. *Clin Vaccine Immunol* 2010, **17**(8):1196-1202.
- Yang Y, et al: Regulation of interleukin-1beta and interleukin-1beta inhibitor release by human airway epithelial cells. *Eur Respir J* 2004, **24**(3):360-366.
- Nagaraju K, et al: A variety of cytokines and immunologically relevant surface molecules are expressed by normal human skeletal muscle cells under proinflammatory stimuli. *Clin Exp Immunol* 1998, **113**(3):407-414.
- Pfaffl MW: A new mathematical model for relative quantification in real-time RT-PCR. *Nucleic Acids Res* 2001, **29**(9):e45.
- Isler JA, Skalet AH, Alwine JC: Human cytomegalovirus infection activates and regulates the unfolded protein response. *J Virol* 2005, **79**(11):6890-6899.
- Samali A, et al: Methods for monitoring endoplasmic reticulum stress and the unfolded protein response. *Int J Cell Biol* 2010, **2010**:830307.
- Lees CW, et al: New IBD genetics: common pathways with other diseases. *Gut* 2011, **60**(12):1739-1753.
- Saleh NM, et al: Genetic association analyses of atopic illness and proinflammatory cytokine genes with type 1 diabetes. *Diabetes Metab Res Rev* 2011, **27**(8):838-843.

40. Han S, et al: **Orm1 and Orm2 are conserved endoplasmic reticulum membrane proteins regulating lipid homeostasis and protein quality control.** *Proc Natl Acad Sci USA* 2010, **107**(13):5851–5856.
41. Broide DH, et al: **Cytokines in symptomatic asthma airways.** *J Allergy Clin Immunol* 1992, **89**(5):958–967.
42. Shi Y, et al: **Role of GRP78/BIP degradation and ER stress in deoxynivalenol-induced interleukin-6 upregulation in the macrophage.** *Toxicol Sci* 2009, **109**(2):247–255.
43. Ma Y, Hendershot LM: **The stressful road to antibody secretion.** *Nat Immunol* 2003, **4**(4):310–311.
44. Hasnain SZ, et al: **The interplay between endoplasmic reticulum stress and inflammation.** *Immunol Cell Biol* 2012, **90**(3):260–270.
45. Adair-Kirk TL, Atkinson JJ, Senior RM: **Smoke particulates stress lung cells.** *Nat Med* 2008, **14**(10):1024–1025.
46. Paul WE, Zhu J: **How are T(H)2-type immune responses initiated and amplified?** *Nat Rev Immunol* 2010, **10**(4):225–235.
47. Pfeifer P, et al: **IL-17C is a Mediator of Respiratory Epithelial Innate Immune Response.** *Am J Respir Cell Mol Biol* 2012, [Epub ahead of print].

doi:10.1186/1710-1492-9-4

Cite this article as: Hsu and Turvey: Functional analysis of the impact of *ORMDL3* expression on inflammation and activation of the unfolded protein response in human airway epithelial cells. *Allergy, Asthma & Clinical Immunology* 2013 **9**:4.

Submit your next manuscript to BioMed Central and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit

