

**EFFECTS OF EXERCISE INTENSITY ON ANTERIOR
CEREBRAL PERFUSION IN PREPUBERTAL CHILDREN.**

by

Ryan Simair

B.H.K., The University of British Columbia, 2015

A THESIS SUBMITTED IN PARTIAL FULFILLMENT OF
THE REQUIREMENTS FOR THE DEGREE OF

MASTER OF SCIENCE

in

THE COLLEGE OF GRADUATE STUDIES
(Interdisciplinary Studies)
THE UNIVERSITY OF BRITISH COLUMBIA
(Okanagan)

November 2017

© Ryan Simair, 2017

The following individuals certify that they have read, and recommend to the College of Graduate Studies for acceptance, a thesis entitled:

EFFECTS OF EXERCISE INTENSITY ON ANTERIOR CEREBRAL PERFUSION IN PREPUBERTAL CHILDREN

Submitted by Ryan Simair in partial fulfillment of the requirements of the degree of Master of Science.

Dr. Ali McManus, Faculty of Health & Social Development

Supervisor

Dr. Glen Foster, Faculty of Health & Social Development

Supervisory Committee Member

Dr. Phil Ainslie, Faculty of Health & Social Development

Supervisory Committee Member

Dr. Paul Shipley, Faculty of Chemistry

University Examiner

Abstract

Introduction: Although high intensity interval exercise (HIIE) has been well characterized in adults as providing superior systemic vascular adaptations compared to moderate intensity exercise (MIE), there is evidence that HIIE poses a challenge to cerebral hemodynamics. Only one study has investigated the child's middle cerebral artery (MCA) response to incremental exercise to maximum, and we do not currently understand the cerebrovascular response to either HIIE or continuous MIE in the child.

Methods: Nine children (aged 7-11 y; 5 girls) completed either HIIE (six 1-minute cycle ergometer sprints at 90% peak power (W_{max}) with 1-minute recovery at 20% W_{max}) or MIE (15 minutes at 44% W_{max} on a cycle ergometer, matched to HIIE for external work). MCA velocity (MCAv), partial pressure of end tidal carbon dioxide (PETCO₂) and mean arterial pressure (MAP) were measured at baseline, immediately post-, 30 minutes post- and 60 minutes post-exercise. MCAv was also continuously recorded throughout exercise. A repeated measures ANOVA was used to compare changes from baseline across the time-points and between the exercise conditions.

Results: The MCAv peaked during the first two minutes in both HIIE (minute 1: $+12.10 \pm 2.70\%$ & minute 2: $+12.65 \pm 6.20\%$; $p < 0.05$) and MIE (minute 1: $+7.46 \pm 5.37\%$ & minute 2: $+9.78 \pm 6.40\%$; $p < 0.05$). In HIIE, MCAv significantly decreased below baseline for the final 3 sprints ($P < 0.05$), oscillating back to baseline for the remaining rest intervals. In MIE, the MCAv returned to baseline after the 4th minute of exercise, where it remained under exercise cessation. The cerebrovascular resistance index (CVRI) significantly increased ($P < 0.05$) during the final minute of both MIE ($+7.0 \pm 6.70\%$) and HIIE ($+21.5 \pm 7.60\%$), with a significant difference between intensities ($P < 0.05$). Immediately post HIIE, the MCAv

uncoupled from PETCO₂, whereas immediately post MIE the MCAv and PETCO₂ remained at baseline values.

Discussion: The decline in MCAv during the latter half of both HIIE and MIE may reflect adequate cerebral oxygenation because of the elevated MCAv baseline values in children compared to adults. The increase in CVRi in the final minute of HIIE is likely driven via hyperventilatory hypocapnia. Finally, it appears that PETCO₂ has a diminished regulatory effect over MCAv in children following HIIE.

Preface

Work described in Chapter 2 and 3 (Methods & Results) was approved by UBC's Clinical Research Ethics Board (H15-02104) and was in accord with the Declaration of Helsinki.

Work was conducted in the Pediatric Exercise Research Laboratory by Dr. AM McManus and Ryan Simair. I was responsible for planning and scheduling the experiment as well as data collection. I wrote this thesis with extensive feedback from Dr. AM McManus and Dr. PN Ainslie.

Table of Contents

Abstract.....	iii
Preface.....	v
Table of Contents	vi
List of Tables	viii
List of Figures.....	iv
List of Symbols & Abbreviations.....	x
Acknowledgements	xii
Dedication	xiii
Chapter 1 Introduction	1
1.1 Literature review.....	3
1.1.1 Cerebral anatomy.....	3
1.1.2 CBF in the child.....	5
1.1.3 CBF during exercise.....	8
1.1.4 Regulation of CBF at rest and during exercise.....	10
1.1.5 High intensity exercise in the child.....	15
1.1.6 Exercise recovery.....	17
1.1.7 High intensity exercise and CBF.....	19
1.1.8 Summary.....	21
Chapter 2 Methods.....	23
2.1 Experimental design and procedures.....	23
2.2 Measurements.....	24
2.2.1 History of CBF measurements.....	24
2.2.2 Principles of TCD.....	25
2.2.3 Technique of TCD.....	26
2.2.4 CBFv acquisition.....	27
2.2.5 Anthropometric measurement.....	27
2.2.6 Maximal exercise test.....	27

2.2.7 HR and arterial blood pressure.....	28
2.2.8 CVRi acquisition.....	28
2.2.9 PETCO ₂ acquisition.....	29
2.3 Statistical analyses.....	29
Chapter 3 Results.....	29
3.1 Descriptive.....	29
3.2 HR and MCAv responses during HIIE and MIE.....	30
3.3 MCAv and PETCO ₂ responses following HIIE and MIE.....	35
3.4 MAP, MCAv and CVRi during the final minute of HIIE and MIE and during recovery.....	36
Chapter 4 Discussion.....	39
4.1 Experimental findings.....	39
4.1.1 Cerebrovascular responses during HIIE and MIE.....	39
4.1.2 Cerebrovascular responses following HIIE and MIE.....	41
4.1.3 Experimental considerations.....	41
4.1.4 Strengths and limitations.....	42
4.2 Conclusion.....	43
References.....	45

List of Tables

Table 3.1	MCAv during HIIE and MIE.....	32
Table 3.2	MAP and MCAv and CVRi at baseline, during and following HIIE and MIE...	37

List of Figures

Figure 1.1	Anatomy of aortic origins of the extracranial vessels.	3
Figure 1.2	Anatomy of the extra- and intracranial vessels feeding to the circle of Willis....	4
Figure 1.3	Anatomy of the intracranial vessels.....	4
Figure 1.4	CBF decreases with age.....	5
Figure 1.5	Reference values for middle cerebral artery velocity (MCAv) from 1 to 14 years of age.....	5
Figure 1.6	Trajectory of CBF development by age and sex.	7
Figure 1.7	Relationship between MCAv and PETCO ₂ up to the ventilatory threshold.....	9
Figure 1.8	The MCAv and PETCO ₂ responses to maximal exercise in children (black) and adults (white).....	11
Figure 1.9	The %CMRO ₂ response to light, moderate and maximal exercise.....	12
Figure 1.10	VO ₂ recovery in children and adults.....	18
Figure 2.1	Example of the Doppler shift.....	26
Figure 3.1	Twelve minutes of absolute minute-by-minute average HR (bpm) values during HIIE (grey line) and MIE (black line).....	31
Figure 3.2	The relative change in MCAv (%) during MIE (panel A) and HIIE (panel B)	34
Figure 3.3	The response of MCAv (A) and PETCO ₂ (B) recovery from baseline.....	35
Figure 3.4	The %CVRi change from baseline.....	38

List of Symbols and Abbreviations

*; significant difference from rest $P < 0.05$; within intensity.

†; significant difference between HIIE and MIE $P < 0.05$.

ACA; Anterior cerebral artery

BA; Basilar artery

CA; Cerebral autoregulation

CBF; Cerebral blood flow

CBFv; Cerebral blood flow velocity

CMRO₂; Cerebral metabolic rate for oxygen

CVC; Cerebrovascular conductance

CVRi; Cerebral vascular resistance index

DBP; Diastolic blood pressure

ECA; External carotid artery

HIIE; High intensity interval exercise

HR; Heart rate

HRmax; Heart rate maximum

ICA; Internal carotid artery

La; Lactate

MAP; Mean arterial pressure

MCAv; Middle cerebral artery velocity

MIE; Moderate intensity exercise

MRI; Magnetic resonance imaging

PaCO₂; Partial pressure of arterial carbon dioxide

PCA; Posterior carotid artery

PETCO₂; Partial pressure of end tidal carbon dioxide

pVO₂ kinetics; Pulmonary oxygen uptake kinetics

SBP; Systolic blood pressure

TCD; Transcranial Doppler ultrasound

VO₂max; Maximal Oxygen consumption

VA; Vertebral artery

Wmax; Maximum external work

Acknowledgements

I would not have made it through my program if it were not for the constant support of Dr. Ali McManus. Dr. McManus challenged me and pushed me to be my best by relating to me on a very real human level. I have had the pleasure of working with Dr. McManus for the passed 3.5 years and although not every day was easy, it certainly was a pleasure. Her passion in her field organically drives her ambition, which is contagious. As my mentor, Dr. McManus helped to engrain in me the scientific process and a plethora of information around our particular field of pediatric cerebrovascular health; And, as my friend she provided constant general life advice and support, watching me grow into the man that I am and playing a significant model role. I am truly grateful for the time spent with Dr. Ali McManus and this incredible chapter of my life.

I am so grateful for my amazing lab mates Victoria Armstrong, Laura Morris, Nathan Sletten, and Christine Tallon who not only supported me through my studies and experiments but also created the laboratory ambiance making each day slightly more manageable with Frisbee, distracting Youtube videos and constant laughs.

Finally, I thank my parents for the constant moral support and love as well as financial contributions. From the home cooked meals on Sunday (and leftovers), repairing the car when needed and massive decompression talks over the many stresses of my day, my parents were the backbone to my education and it would not have been possible without them.

Dedication

This project is dedicated to children of North America to encourage physical activity at a young age and an active lifestyle throughout their lifespan.

Chapter 1 Introduction

Exercise can reduce the risk of cerebrovascular and neurodegenerative diseases and has been associated with extended longevity (Lautenschlager *et al.* 2012), yet only 7% of Canadian children meet current physical activity guidelines (Tremblay *et al.*, 2011B). Instead, most children spend about 60% of their waking hours sedentary, with approximately eight hours a day spent sitting without otherwise moving (Tremblay *et al.*, 2011B). It is concerning that children spend a majority of their day sedentary, given the evidence that sedentary aging is associated with a longitudinal decline in cerebral blood flow (CBF) (Ainslie *et al.*, 2008; Barnes *et al.*, 2013), cerebrovascular reactivity (Bakker *et al.*, 2004), and cerebral autoregulation (CA) (Bailey *et al.*, 2013). Together, these latter changes likely increase the risk of cognitive decline, and stroke (Okazawa *et al.*, 2003).

Physical activity is the number one recommendation for reducing the risk of cardiovascular, cerebrovascular and neurodegenerative diseases across the life-span (Strong *et al.*, 2005). Regular exercise improves cerebral hemodynamics in older adults (Ainslie *et al.*, 2008) and higher levels of aerobic fitness are also associated with improved hippocampal perfusion and cognitive function in children (Chaddock-Heyman *et al.*, 2016; Moore *et al.*, 2013). It is believed that these improvements are linked to the increases in cerebral perfusion during exercise, but there is limited evidence on the impact that an acute bout of exercise has upon the cerebral hemodynamics in children.

While traditional exercise interventions for children have principally involved continuous moderate intensity exercise (MIE) (McMurray *et al.*, 2002; Hansen *et al.*, 1991; Meyer *et al.*,

2006), the relevance of such programs to the child's habitual sporadic play patterns has been questioned (Bailey et al., 1995). For example, movement patterns in children are intermittent, with multiple low to moderate intensity bursts of movement interspersed with occasional vigorous movement (Bailey et al., 1995). High-intensity interval exercise (HIIE) has emerged as a time efficient and effective alternative to MIE, providing superior metabolic, cardiac and systemic vascular adaptations in adults (Weston *et al.* 2014). Indeed, in the healthy children, HIIE has been shown to improve cardiorespiratory fitness in comparison to lower intensity exercise (Baquet et al., 2010; Baquet et al., 2002; McManus et al., 2005), maximal running velocity (Baquet et al., 2010), peak and submaximal cardiovascular outcomes (McManus et al., 2005), and ventilatory function (Nourry et al., 2005). Regardless of the convenient nature of HIIE and the many reported positive effects, the lack of evidence to date examining how HIIE affects the cerebrovasculature has been outlined (Lucas *et al.* 2015) and there is some evidence, albeit in adults, that HIIE may overly challenge the cerebrovasculature (Curtelin et al. 2017). For example, using a single bout Wingate test Curtelin et al (2017) revealed that adults elicit marked elevations in mean arterial pressure (MAP) and middle cerebral artery velocity (MCAv) during the onset of exercise but also showed significant reductions in MAP immediately post exercise; Perhaps, it is possible that these large instantaneous alterations are overbearing for the cerebrovasculature.

The purpose therefore of this thesis is to compare the cerebrovascular responses during continuous MIE and HIIE, as well as during recovery from both exercise conditions, in prepubertal children. The thesis begins with a review of the literature (Chapter 1), introducing cerebral anatomy and the developmental changes that occur in CBF, focusing upon the cerebrovascular response to exercise. Chapter 2 describes the methods used and

Chapter 3 reveals the main results found. Chapter 4 is a discussion of the results including strengths & limitations and future direction in the field.

1.1 Literature review

1.1.1 Cerebral anatomy

The path that blood travels to supply the anterior brain originates at the heart, emerging from the ascending aorta. The left carotid emerges directly from the aorta, while the right branches from the brachiocephalic trunk. The vertebral arteries (VA) emerge from the right and left

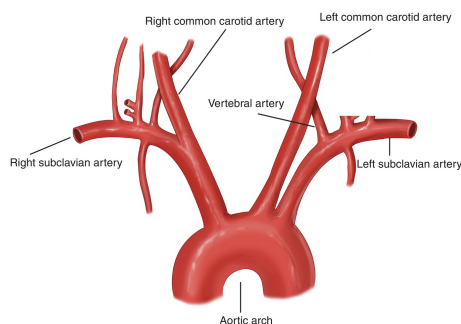


Figure 1.1 Anatomy of aortic origins of the extracranial vessels. Image adapted from Google.

subclavian arteries, although the left VA may also branch directly from the aortic arch in about 15% of individuals (Einstein et al. 2016; see figure 1.1).

The carotid artery (left & right) split at the level of the mastoid, into the external carotid artery (ECA) and the internal carotid artery (ICA). The ECA provides the blood supply to the facial region. The ICA does not branch until joining the Circle of Willis and is often thought of as the origin of cerebral perfusion, contributing about 76% of global CBF at rest (Schöning et al, 1994). The ICA undergoes considerable growth during childhood. It is similar in size to the ECA until about 9 years of age (Seong et al., 2005). During maturation, the ICA remodels at the root or carotid sinus, with no corresponding change in the ECA. The resulting arterial remodelling results in the prominent carotid bulb noted in adults, and coincides with maturation of the brain. The carotid sinus

acts as a stretch receptor providing a direct sensor for blood pressure changes, as well as chemoreception (sensing changes in oxygen and carbon dioxide tension). Although the precise reason for this ICA remodelling is not known, this may be a means of providing more mature baro- and chemoreception, and/or may act as a pressure dampener with the carotid sinus slowing flow to protect the maturing brain from surges in pressure (Seong et al., 2005). Either way the carotid sinus most likely supports the maturing needs of the cerebrovascular

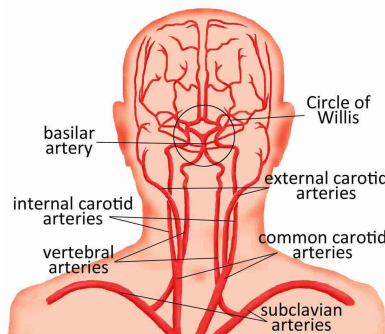


Figure 1.2 Anatomy of the extra- and intracranial vessels feeding to the circle of Willis. Image adapted from Google.

system.

The left and right VA join together at the level of the pons forming the basilar artery (BA), which connects the Circle of Willis. The VA delivers the remaining ~24% of global CBF to the the circle of Willis (Schöning et a., 1994; see figure 1.2).

The Circle of Willis then branches into the basal arteries: the anterior cerebral artery (ACA), the middle cerebral artery (MCA) and the posterior cerebral artery (PCA) (See Figure 1.3). The MCA is the largest intra-cranial artery and supplies a portion of the frontal lobe and the lateral surface of the temporal and parietal lobes, including the primary motor and sensory areas and the areas for speech. The MCA supplies these respective areas with 60-80% of blood flow (Verlhac, 2011).

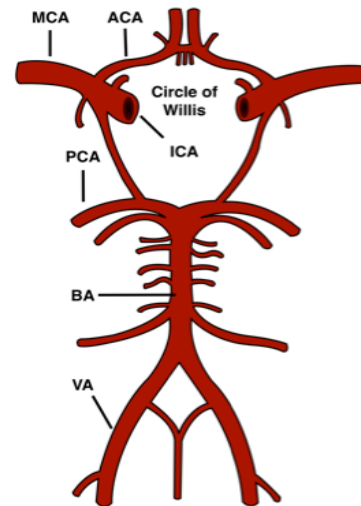


Figure 1.3 Anatomy of the intracranial vessels. Original Image.

1.1.2 CBF in the child

As the child's brain undergoes progressive structural and functional development, alterations to cerebral haemodynamics support these changing needs (Tortori-Donati & Rossi, 2005; Biagi *et al.* 2007; Labarthe *et al.* 2009; Hales *et al.* 2014). Previous work demonstrates that grey matter and white matter CBF is highest during the first two decades of life peaking

around 6-10 years of age (Leung *et al.*, 2016; see figure 1.4).

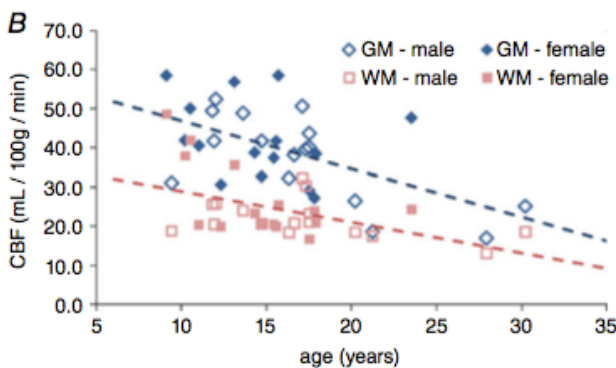


Figure 1.4 CBF decreases with age (GM-grey matter; WM-white matter). Retrieved from Leung *et al.*, 2016 with permission.

Similarly, early cross-sectional work using transcranial Doppler ultrasound (TCD) to measure MCAv analyzed 1,200 examinations from 620 children from birth to 15 years of age, and revealed a rapid increase in MCAv from

2 months of age to 6-8 years, whereby MCAv begins to decline gradually towards adult values by the age of 18 years (Bode & Eden, 1989; see figure 1.5). More recently, Demirkaya *et al.* (2008) have confirmed the earlier work by Bode and Eden (1989) in a group of 30

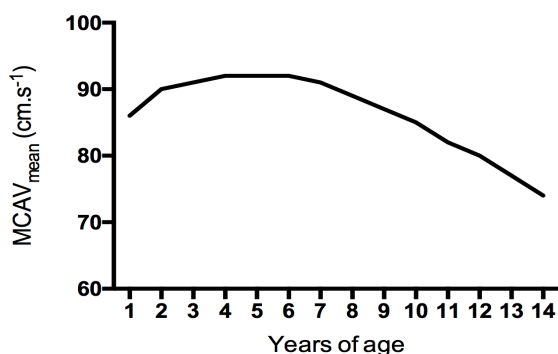


Figure 1.5 Reference values for MCAv from 1 to 14 years of age, adapted from Bode & Eden, 1989.

males and 33 females aged 6-69 years. They demonstrate a 28.8 cm.s⁻¹ decrease in MCAv at rest from 75.7 cm.s⁻¹ in 5-10 year olds, declining to a mean value of 46.9 cm.s⁻¹ for the over 60's (Demirkaya, Uluc, Bek, & Vural, 2008).

The elevated resting CBF in childhood is nearly double that of an adult (Chugani, 1998) and reflects the much higher cerebral metabolism of the developing brain (Biagi *et al.* 2007). Indeed, cerebral oxygen consumption is estimated to be about 50% of total body oxygen consumption in the child (Kennedy and Sokoloff, 1957), nearly 1.3 times higher than noted in adults (Chugani 1998).

Sex differences have been examined for resting CBF. Studies using TCD to derive cerebral blood flow velocity (CBFv) demonstrate that adolescent girls have a higher MCAv compared to age matched boys (Vavilala, Kincaid, Muangman, Suz, Rozet & Lam, 2005). Further work has shown that sex differences in CBFv also exist prior to puberty (Tontisirin *et al.* 2007). Assessing 48 pre-pubertal children aged 4-8 years (24 girls; 24 boys), Tontisirin and colleagues (2007) found that both the anterior and posterior circulations were higher in girls than boys. Values for MCAv were $99 \pm 11 \text{ cm.s}^{-1}$ and $91 \pm 13 \text{ cm.s}^{-1}$ in girls and boys respectively. The BA velocity was $70 \pm 10 \text{ cm.s}^{-1}$ versus $61 \pm 9 \text{ cm.s}^{-1}$ in girls and boys respectively. There are a number of possible explanations for these documented sex differences including differences in haematocrit and therefore blood viscosity, divergent sex hormones, differing cerebral metabolism, cerebrovascular resistance index (CVRi) and vessel size. However, only CVRi and vessel size show sex divergence prior to puberty, with CVRi (See Equation 1.1) lower in 4 to 8 year-old girls than boys (Tontisirin *et al.*, 2007) and limited evidence of smaller ICA and MCA diameters in females compared with males (Gabrielsen & Greiz, 1970).

Equation 1.1.
$$\text{CVRi} = \text{MAP/MCAv}$$

Where CVRi = Cerebral Vascular Resistance index; MAP = Mean Arterial Pressure; MCAv = Middle Cerebral Artery velocity.

In contrast, recent work utilizing magnetic resonance imaging (MRI) (Satterthwaite et al., 2014), reported a nonlinear age-by-sex interaction for CBF in multiple brain regions, with CBF values lower in girls compared to boys before 13 years of age. Boys, however, showed a markedly more rapid age-related decline in CBF resulting in higher CBF values in girls than in boys by mid-adolescence. By late adolescence, CBF increases with age in girls, whereas CBF in boys continues to decline (see figure 1.6).

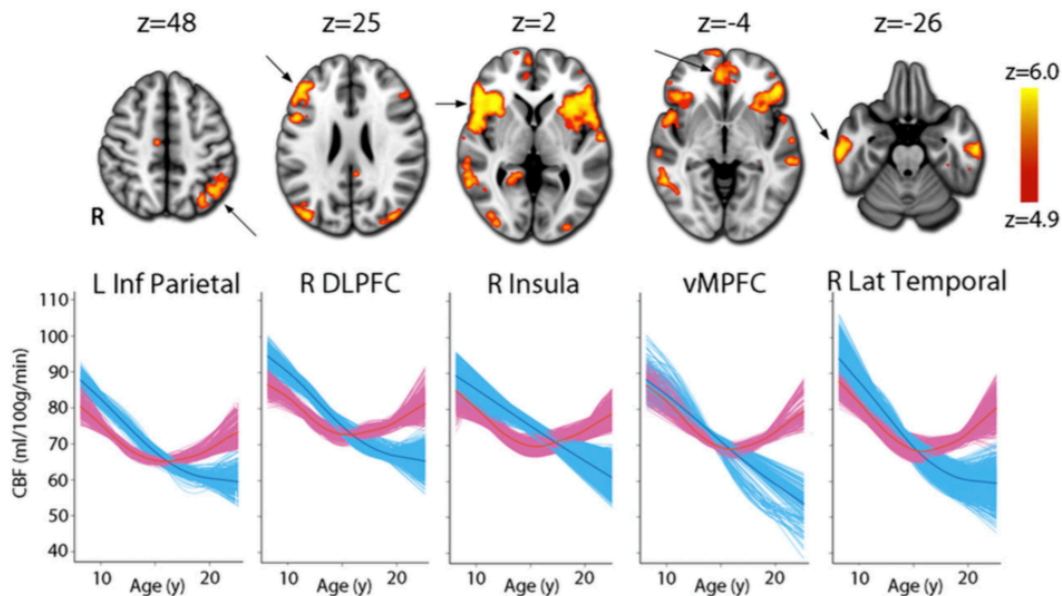


Figure 1.6 Trajectory of CBF development by age and sex. Retrieved from: PNAS in Satterthwaite et al., 2014, no permission required.

When differences in maturational trajectories were accounted for however, the decline in CBF was similar between girls and boys in early puberty (Satterthwaite et al., 2014).

Cerebral perfusion diverges by mid-puberty (See Figure 1.6), with greater declines noted in boys persisting into late puberty and resulting in higher levels of cerebral perfusion in girls than boys by late adolescence (Satterthwaite et al., 2014).

The disparity in findings between the work of Tontisirin et al. (2007) and Satterthwaite et al (2014) may result from the differing techniques as Satterthwaite et al used MRI to index CBF and Tontisirin et al used TCD. There is very limited data regarding sex difference in intra- or extra-cranial vessel diameters in children, but it is possible that if girls have a smaller vessel diameter than boys, female flow velocities may be over-estimated with TCD. Such over estimations may account for the higher CBFv values reported by Tontisirin et al. (2007) with TCD, compared to the lower CBF values acquired with arterial spin labelling MRI from Satterthwaite et al (2014) where vessel diameter is accounted for.

1.1.3 CBF during exercise

Much of the information available on the cerebrovascular response to exercise is currently adult-based. The adult CBFv response to incremental aerobic exercise parallels increases in exercise intensity until ~60% of the maximal rate of oxygen consumption (VO_2max), whereby CBFv then either plateaus (Smith et al., 2014) or declines towards baseline values (Ellis et al., 2017; Fisher et al., 2013; Smith et al., 2012). To date, only one study has documented the impact of acute exercise on CBFv in children (Ellis et al., 2017). Ellis and colleagues (2017) assessed the MCAv response to incremental cycle ergometer exercise to max VO_2 in both children and adults. A similar pattern of change in MCAv was noted across the exercise test in the children and adults, but the increase in MCAv was attenuated in

children. For example, the peak increase above baseline noted in MCAv was about 10% in children and occurred by 50% of the ventilatory threshold. The MCAv remained elevated ~10% above baseline up until the respiratory compensation point, where a decline was then noted (See Figure 1.8). These data highlight that exercise intensity from low through to vigorous intensity provide similar gains in CBF in the child. In adults, MCAv was elevated 20% above baseline at the ventilatory threshold, declining thereafter, highlighting that in adults the greatest gains in CBF occur around MIE. Interestingly, there were no sex differences in the response to incremental exercise (Ellis et al., 2017).

In adults, as exercise approaches maximal intensity, hyperventilation causes hypocapnia and MCAv declines back toward or below baseline values (Oogh et al. 2008). In children, Ellis and colleagues (Ellis et al., 2017) demonstrated that partial pressure of end tidal carbon dioxide (PETCO₂) and MCAv are not correlated, and the earlier decline in PETCO₂ noted in children is not accompanied by a decline in MCAv (Figure 1.7).

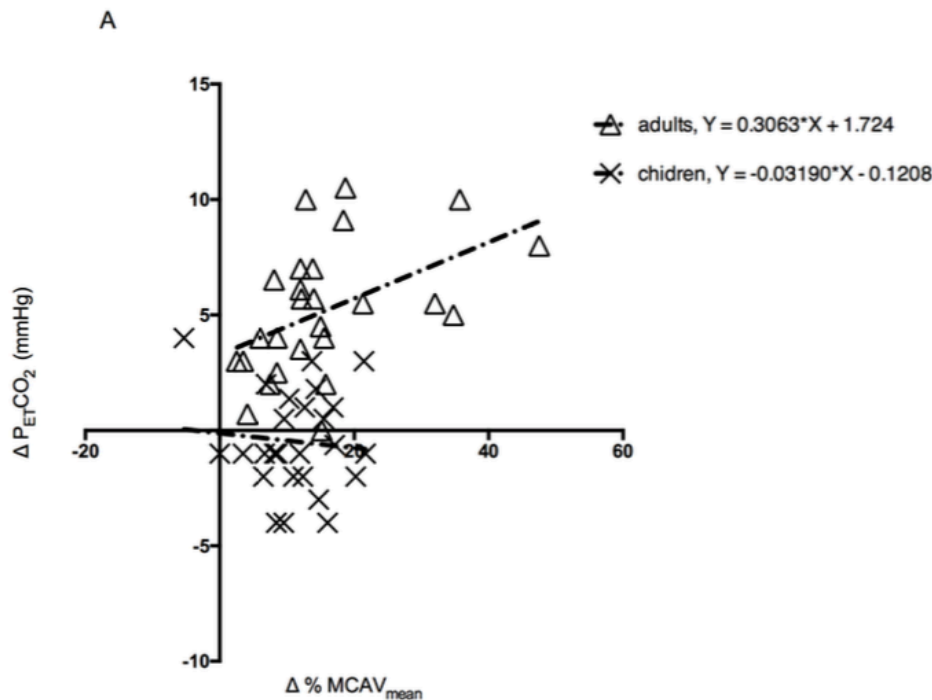


Figure 1.7 Relationship between MCAv and PETCO₂ up to the ventilatory threshold. Retrieved from: AJP Heart in Ellis et al., 2017, no permission required.

These data suggest that the regulation of CBF during exercise is developmentally divergent and the impact that exercise of varying intensities may have on the cerebrovasculature is most likely dependent on both age and maturation.

1.1.4 Regulation of CBF at rest and during exercise

Key factors regulating CBF at rest or during exercise include CO_2 (Ainslie and Burgess 2008), metabolism (Iadecola and Nedergaard 2007; Paulson et al. 2011) and CPP (Panerai et al. 1999; Lucas et al. 2010). Of these dominating factors, Giardino et al (2007) suggested that CO_2 might have the greatest influence on CBF, larger than the other intrinsic physiological factors. Indeed, in a more recent review by Smith & Ainslie (2017), they outline the differences of CBF regulation between rest and exercise, identifying the following primary regulatory factors at rest in order of importance: arterial blood gases (particularly the partial pressure of arterial carbon dioxide; PaCO_2), cerebral metabolism, arterial blood pressure, neurogenic activity and cardiac output. It appears that PaCO_2 is the primary influencing factor during exercise as well (Smith & Ainslie 2017), confirming earlier suggestions by Giardino et al (2007).

In adults, global CBF is positively correlated to PaCO_2 in an approximately linear relationship, within the wide physiological range of 20–60 mmHg PaCO_2 (Giardino et al, 2007). Within this range, every 1 mmHg change in PaCO_2 elicits a corresponding directional change in CBF of approximately 1–2 ml/100g/min, or 2–5% (Poulin et al., 1996; Ide et al., 2003). So, even a small alteration in alveolar ventilation and hence PaCO_2 can produce significant changes in global CBF in adults.

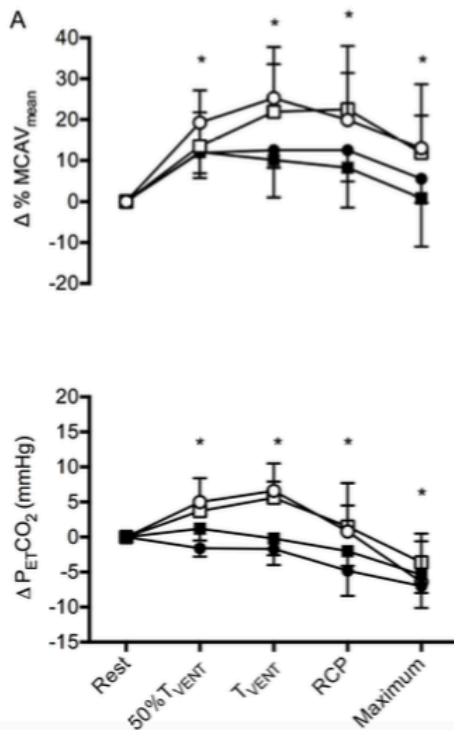


Figure 1.8 The MCAv and PETCO₂ responses to maximal exercise in children (black) and adults (white). Retrieved from: AJP Heart in Ellis et al., 2017, no permission required.

The mechanism by which PaCO₂ influences CBF is dependent on perturbations in extracellular and perivascular pH, altering the properties of vascular smooth muscle (Gotoh et al., 1961; Kontos, et al., 1977). CO₂ is hydrated in the presence of carbonic anhydrase to form carbonic acid and its dissociation by-products, bicarbonate and a proton. The resultant acidic extracellular environment enhances the vasodilatory effect of adenosine (Fenton et al., 1981) and increases potassium ion conductance across vascular smooth muscle (Karaki & Weiss, 1981), resulting in blood vessel dilation, reduced CVR_i and thus increased CBF (Grubb et al., 1974). However,

Ellis et al. (2017) recently found that PETCO₂ is not coupled to the MCAv response during exercise in children (Figure 1.8). This may be due in part to children having a lower cerebrovascular reactivity to CO₂ (Leung et al., 2016). It is possible that the increased level of resting MCAv in children limits the ability for further increases in blood supply as a potentially protective mechanism against hyperperfusion. Alongside the apparent blunted cerebrovascular reactivity to CO₂ (Leung et al., 2016), children also exhibit a lower ventilatory threshold in response to CO₂ (Gratas-Delamarche et al., 1993).

Cerebral metabolic rate of oxygen (CMRO₂) represents the uptake and utilization of oxygen across the arterial and venous circulations in the brain. In 1957, Kennedy and Sokoloff estimated that the child brain uses approximately 50% of total body oxygen consumption at

rest, which Chugani reported (1998) is nearly 1.3 times higher than adults. Ainslie et al later described that the adult brain, despite only weighing 2% of total body mass, still demands an inequitable 20% of basal metabolic oxygen consumption (Ainslie et al., 2016). This increase CMRO_2 in the child is therefore supported by an elevated global CBF, nearly doubling that of the adult during rest (Chugani, 1998).

Little is known regarding the response of CBF and CMRO_2 during exercise in children, and available studies focus on adult participants. Adult data are equivocal based on exercise intensity performed, with reports of CMRO_2 decreasing (Madsen et al., 1993), not changing (Nybo et al., 2003) or increasing (Fisher et al., 2013) with a concomitant drop in CBF during exercise (Figure 1.9).

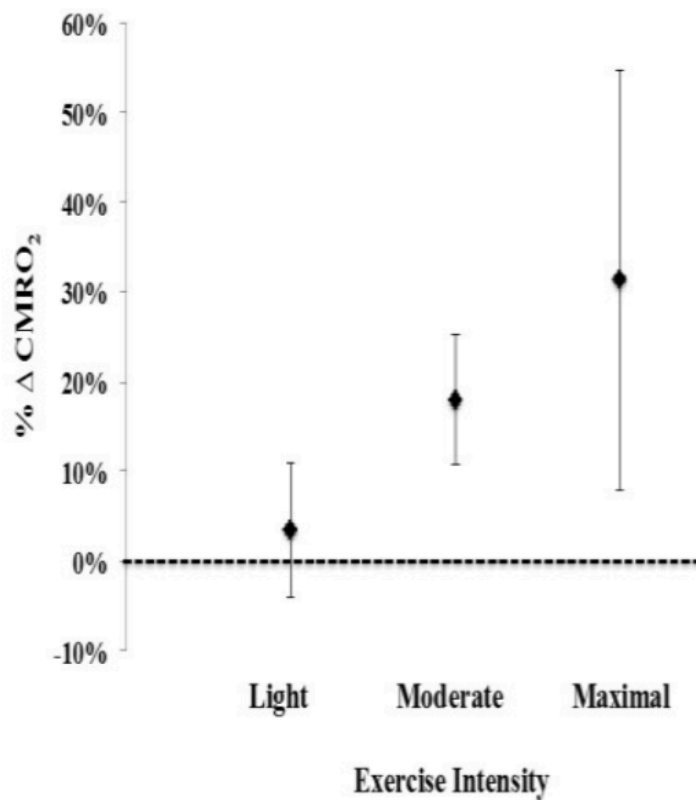


Figure 1.9 The $\% \text{CMRO}_2$ response to light, moderate and maximal exercise. Retrieved from: Experimental Physiology in doi: 10.1113/EP086249, no permission required.

Perhaps the difference in reported responses can be attributed to the use of different exercise intensities and duration or the methodology used to assess CBF and CMRO₂. Regardless, the literature reveals interactions between global CBF, cerebral oxygen delivery and cerebral O₂ extraction, and highlights that it is the balance between these factors that influences the CMRO₂ response to exercise.

In addition to CO₂ and cerebral metabolism, pressure regulation in adults also elicits changes in CBF by altering CVR_i (See Equation 1.1). It is thought that PaCO₂ and pressure autoregulation compete for the control of vessel diameter (Harper and Glass 1965), but the relationship between CBF and blood pressure alone during incremental exercise is weak. The relationship between only CBF and MAP appears to exist at rest (Willie et al., 2014), and MAP is likely still a factor within the multifaceted balance of CBF along with the other primary regulators described (Smith et al., 2017). For example, Smith et al (2017) summarized that although MAP increases with incremental exercise by 20-30% up to maximal intensities, CBF increases to a lesser extent (+15-25%) and only up to ~60% of maximal intensity, at which point CBF returns to baseline (Ogoh & Ainslie, 2009) uncoupling from MAP elevations. It does, however, seem that the pressure flow relationship remains important during HIIE when blood pressure becomes markedly elevated (Curtelin et al., 2017). The recent work by Curtelin et al (2017) suggests that despite an increase in MAP and MCAv at the onset of an all-out sprint, CBF is quickly adjusted and reduced to limit the potential risk of a hyperperfusion injury (described in detail below).

Far less is known regarding CBF regulation in children compared to adults, but what has been studied mainly describes the pressure-flow relationship in the child. Vavilala et al

(2003B) expanded our knowledge of CBF regulation in children by using metrics such as the lower limit of CA, lower limit reserve and the autoregulatory range to probe their pressure-flow relationship. The lower limit of CA is of importance since MAP decreasing below the lower limit of CA potentially increases the risk of cerebral ischemia (Vavilala et al., 2003B), and the brain seems better suited at buffering hypertension than hypotension (Tzeng et al., 2010).

Determining the lower limit of CA involves inspecting the MCAv/MAP data to determine where CBF no longer remains constant during reductions in MAP (Artru et al., 1989; Drummond, 1997). The lower limit of CA alone however cannot sufficiently estimate the risk of cerebral hypoperfusion without the concepts of lower limit reserve and autoregulatory range. The lower limit reserve is an index representing the safety margin against cerebral hypoperfusion and is derived as the difference between baseline MAP and the lower limit of CA (lower limit reserve = Baseline MAP – lower limit of CA). The relative risk of cerebral hypoperfusion must take into account the relationship of the lower limit of CA to baseline MAP. To express the margin of safety as a function of baseline MAP, the autoregulatory reserve index (autoregulatory range %) is derived from the lower limit reserve and baseline MAP (autoregulatory range % = [lower limit reserve/Baseline MAP] × 100).

Vavilala et al (2003B) set out to determine whether or not differences exist between the adult and child values of the lower limit of CA. They reported the lower limit of CA in sick children for the first time, finding: 1) the range of the lower limit of CA is similar between children under 2 years of age and adults (46–70 mmHg), 2) the lower limit of CA is not related to age, 3) older children have a greater lower limit reserve compared with younger

children, and 4) they found the autoregulatory range to be greater in older children compared with younger children (Vavilala et al., 2003B).

It is important to note that Vavilala et al (2003B) reported that baseline MAP is lower in children compared with adults. Consequently, it is often assumed that the lower limit of CA is also lower in children along with their decreased baseline MAP, allowing them to tolerate hypotension more so than adults. However, early hypotension (Vavilala et al., 2003A) and impaired CA (Kennedy & Sokoloff, 1957) are associated with poor outcome after pediatric traumatic brain injury, yet little is known regarding CA in healthy children (Tontisirin et al., 2007). In adults, a marked decline in MAP was noted immediately following sprint exercise (Curtelin et al., 2017), but it is unknown if a similar decline in MAP occurs following sprint or HIIE in children, which may challenge CA.

1.1.5 High intensity exercise in the child

It appears that HIE is well tolerated in children (Hebestreit, Mimura, & Bar-Or, 1993); however, maturational and developmental disparities exist between children and adults, which may influence the acute response to this type of exercise. Insight into the metabolic response to exercise in the child has been gained from pulmonary oxygen uptake kinetics ($\dot{V}O_2$ kinetics). The child's faster $\dot{V}O_2$ response time or tau (t) during MIE (Fawkner et al. 2002) and the attenuated slow-component during high-intensity exercise (Fawkner & Armstrong, 2004) provides evidence of an enhanced oxidative metabolism in the child. It seems that children have a higher arterio-venous oxygen difference (a-v O_2 diff) compared to adults (Prado, Dias, & Trometta, 2006), indicating a better balance of oxygen delivery and oxygen utilization. Indeed, children show faster muscle de-oxyhemoglobin kinetics

compared to adults, reflecting a faster local fractional oxygen extraction (Leclaire et al. 2013). The adjustment of local blood flow to exercise is also faster in the child (Koch & Eriksson, 1973; Leclaire et al. 2013), supporting an enhanced oxygen delivery during moderate and intense exercise (Koch, 1984), thereby increasing blood-myocyte oxygen flux and oxidative phosphorylation (McDonough et al. 2005). Combined, the enhanced oxygen extraction and utilization in children (Leclaire et al. 2013) is likely supported by the larger type I fibre composition (Pringle et al. 2003), enhanced oxidative enzymes (Poole & Jones, 2012), and improved local oxygen delivery (Koch, 1984; Murias et al., 2014).

It is worth clarifying the range of intensities that might encompass 'high intensity'. High intensity could be defined as any exercise within a certain percentage of VO_2max , a percentage of heart rate maximum (HRmax), percentage of the maximum external work (Wmax) achieved in the maximal exercise test, or from the gas exchange (also known as ventilatory) threshold (see Jones and Poole, 2012 for a detailed description of intensity derived from gas exchange threshold). High intensity, however, could equally mean an all-out bout of anaerobic exercise such as the 30s Wingate test. The difference in the protocol is important. If the exercise is high intensity *interval* - such as that used by Bond et al., (2015), an interval protocol of multiple short intervals (eg.. 8 x 1 minute intervals), usually at an intensity derived from a VO_2 max test, such as Wmax or HRmax (e.g., 90% Wmax) is used, and these are interspersed with active recovery periods (e.g, 75s at 20W). This interval style is similar to the start-stop activity free-living activity patterns in children (McManus et al., 2011). In contrast, when a single sprint bout is used (Curtelin et al. 2017), such the Wingate test, the intensity is much higher. This type of high intensity exercise is less likely to be emulated in free-living scenarios, more likely to be found in elite training programs.

There is some evidence that during HIIE children have faster blood H⁺ removal (Ratel et al. 2002) and faster cellular lactate (La) clearance (Ratel et al. 2003) compared to adults, which allows for blood pH to be maintained at higher levels during exercise. The notion that children can maintain blood pH at higher levels during exercise may relate to the preferential recruitment of type I fibres leading to an attenuated increase in Pi/PCr and decrease in pH (Mizuno et al., 1994), and enhanced oxidation of La to replenish ATP (Ratel et al. 2003). Therefore, children may not produce less La *per se*, but they may be more efficient at either utilizing it as an energy source or removing it through higher rates of ventilation, effectively regulating acid-base balance through PaCO₂ regulation (Ratel et al. 2002). The outcome of an enhanced oxidative metabolic engine in the child means they can exercise at intensities close to cardiopulmonary maximum without producing high levels of La resulting in a potentially faster recovery (Tolfrey & Armstrong, 1995).

1.1.6 Exercise recovery

Hill and Lupton (1923) observed that even upon the cessation of exercise, VO₂ remains elevated above resting values. This has been termed the excess post-exercise oxygen consumption (Gaesser & Brooks, 1984), which normally occurs within 1 hour of terminating exercise and works to resynthesize ATP and PCr, aids La removal, and returns ventilatory and circulatory levels to baseline.

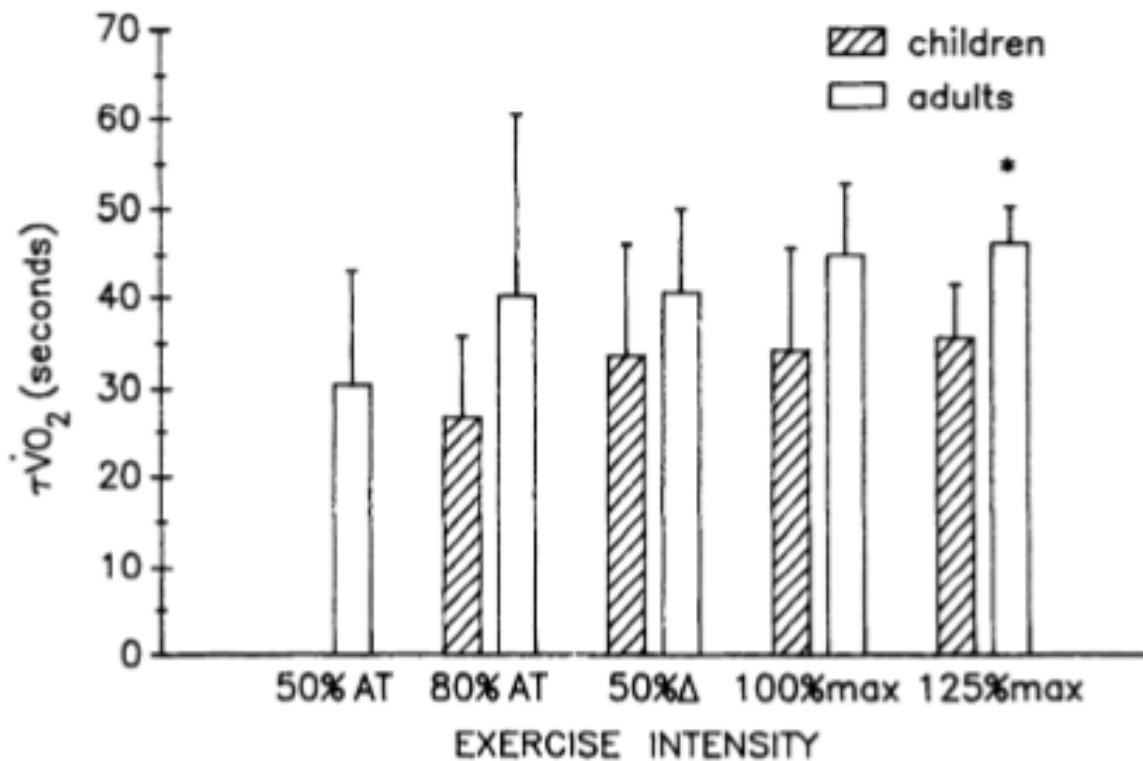


Figure 1.10 The VO_2 recovery in children and adults. The recovery of VO_2 , denoted by the speed of t , is faster in children at the highest intensity of exercise compared to adults (* $P < 0.001$). Reproduced from Zancanato, Cooper, & Armon, 1991, permission not required.

Children recover faster following high intensity exercise than adults and this has been demonstrated using a variety of modalities of high intensity exercise. For example, following a 30s Wingate test, pre-pubertal boys (9-12 y) have faster recovery of ventilation, heart rate (HR), and VO_2 compared to men (Hebestreit, Mimura, & Bar-Or, 1993). Ratel and colleagues (2006) also show the recovery of oxygen uptake is quicker in children at 125% of VO_2 max (figure 1.10). This was accompanied by a smaller decline in peak power, and running velocity. The faster recovery may reflect a more limited capacity for glycolytic metabolism because of the lower type II muscle fibres in the child (Falk & Dotan, 2006). But, this is also compensated for by the enhanced oxidative metabolism in the child. However, this developmentally divergent metabolic response to high intensity exercise ultimately may impact the adaptive responses and we know little about the impact high-

intensity exercise has in terms of eliciting rapid increases in arterial blood pressure (BP) during the high intensity exercise, or substantial declines after, which both may adversely impact CA.

In adults, exercise above 70% $\text{VO}_{2\text{max}}$ results in hyperventilation-induced-hypocapnia and hence cerebrovascular vasoconstriction and subsequent declines in CBF values (Moraine et al., 1993; Subudhi et al., 2008; Smith et al., 2012). This reduction in CBF may be a neuroprotective response to prevent flow disturbances and hyperperfusion injury. Although only found during rest, CBF reductions have been associated with improved CBF regulation during changes in blood pressure (Brassard et al., 2012; Maggio et al., 2013; Aaslid et al., 1989). Ellis et al. (2017) demonstrated in children that during exercise, hyperventilation-induced hypocapnia has minimal impact on MCAv; However, it should be noted that the peak CBFv response to exercise was about 50% of the response in adults. Additionally, Ellis et al (2017) note that although CVRi was lower in children compared to adults, the delta change in MAP with exercise was not related to the delta change in MCAv in children.

1.1.7 High intensity exercise and CBF

There is sparse evidence of CBF during high intensity exercise and to date, none documenting the CBF response to HIIE in children. Previous work has shown that the MCAv response to incremental exercise was attenuated in children compared to adults (Ellis et al., 2017), but values were still elevated 8-10% above baseline at the respiratory compensation point, before returning to baseline as exercise intensity approached maximum.

There is one adult study documenting the CBF and MAP responses to a single 30-s Wingate

test (Curtelin et al. 2017). During the sprint, mean systolic blood pressure (SBP) increased to 200 mmHg, while diastolic blood pressure (DBP) was maintained at pre-sprint values resulting in a 16 mmHg increase in MAP from 115 to 131 mmHg. When examining the time-course, Curtelin and colleagues (2017) showed a downward curvilinear response in CBF, indexed by MCAv. After 7.5s into the Wingate test, adult MCAv peaked to 16% above baseline values, falling thereafter to 10% below baseline by the end of the 30s sprint.

A cerebrovascular conductance (CVC) score of 1.00 indicates impaired cerebrovascular conductance (CBF/MAP). Willie et al (2014) reported a normal CVC range of 0.21 to 0.81 indicating regular CVC. Indeed, Curtelin et al (2017) reported a CVC value of 1.08 during the single all out Wingate sprint, suggesting the cerebral circulation may have been exposed to the potentially harmful effects of high blood pressure combined with increased CBF (hyperperfusion). It is important to consider that immediately following the sprint a fast decline in MAP ensued, and it is possible that the resultant hypotension poses also an additional challenge to cerebral hemodynamics. Within the first 2.5s of recovery Curtelin et al (2017) reported a 26% decline in MAP with a concomitant 18% decline in MCAv from exercise values. Compared to baseline values, MCAv was still reduced by 10% ($P < 0.05$) upon sprint cessation, but despite these marked declines in MAP and MCAv, frontal lobe oxygenation during the first 15 seconds of recovery remained at a similar level to that reached during the last 5 seconds of the sprint, suggesting cerebral oxygenation is preserved despite such hypoperfusive autoregulatory challenges. How HIIE may affect cerebral perfusion in the child is not currently known.

1.1.8 Summary

To summarise, HIIE has emerged as a time efficient and effective alternative to current exercise regimes, providing superior cardiorespiratory adaptations (Baquet *et al.*, 2010; Baquet *et al.*, 2002; McManus *et al.*, 2005) and appears to be better tolerated in children compared to continuous exercise (Timmons & Bar-Or, 2003). Regardless of the reported positive effects and suitability of HIIE, the lack of evidence to date examining how HIIE affects cerebrovascular function raises concerns about its global promotion (Lucas *et al.* 2015).

Evidence in adults suggests that high intensity exercise creates a considerable hemodynamic impact, with marked increases in systolic blood pressure during the sprint and rapid declines in the immediate recovery period challenging cerebrovascular regulation (Curtelin *et al.*, 2017). Children support high intensity exercise with a greater oxidative metabolic contribution compared to adults (Chia *et al.*, 1997), making them more fatigue-resistant to repeated bouts of high intensity exercise (Hebbestreit *et al.*, 1993). It is unclear if a similar cerebrovascular response to a high intensity exercise bout would be noted in children. Previous work has shown that the MCAv response to incremental exercise was attenuated in the child compared to adults (Ellis *et al.*, 2017), but remained 8-10% elevated above baseline up until the respiratory compensation point, before returning to baseline. In addition, these alterations in MCAv during incremental exercise were not correlated with PETCO₂ (and unrelated to changes in MAP (Ellis *et al.*, 2017).

The purpose of this study was to compare the impact HIIE and continuous MIE have on cerebrovascular function in children. To achieve this, a randomized cross-over trial was completed with 7-11 year old boys and girls, completing six high intensity one-minute sprints, as well as continuous MIE matched for external work. The MCAv and PETCO₂ were assessed at baseline and during recovery following both exercise conditions (immediately post-, 30 minutes post- and 60 minutes post HIIE and MIE). In addition, MAP was collected at baseline, during the final minute of exercise as well as 30 minutes post- and 60 minutes post HIIE and MIE. The MCAv was also measured continuously throughout HIIE and MIE.

The hypotheses were

- (i) Increases in MCAv would be greater in response to MIE compared to HIIE
- (ii) MAP will be lower in the final minute of MIE compared to HIIE, resulting in a lower CVRi in MIE compared to HIIE.
- (iii) The MCAv would be lower than baseline immediately post HIIE but not MIE, recovering to baseline values by 30 minutes post exercise;
- (iv) PETCO₂ will be lower immediately post HIIE compared to MIE.

Chapter 2 Methods

Nineteen children aged 7-11 years volunteered to participate in this study. Participants were pre-pubertal (Tanner 1) determined by parental assessment of Tanner staging (Rasmussen et al., 2015). Each participant had a health history clear of cardiorespiratory and cerebrovascular diseases and none had any medical conditions that would influence cerebral perfusion or the ability to exercise. Children were excluded from participating if training >10 hours/week for sport or competition. This study was approved by the Clinical Research Ethics Board of the University of British Columbia (H15-02104). Written consent was obtained from the parents and written assent from the child.

2.1 Experimental design and procedures

A crossover trial with two experimental conditions was employed, including HIIE and MIE both performed on a cycle ergometer.. HIIE consisted of a 3-minute unloaded pedalling warm-up (0 W) followed by six 1 min sprints pedaling at 90% W_{max} , separated by active rest intervals of 1 min of pedaling at 20% W_{max} . This protocol was selected based on the work by Bond et al in 2015 who used 90% W_{max} and 20% W_{max} for exercise and rest intervals, respectively, although they incorporated 8 sprint intervals separated by 75s recovery periods. We found that completing 8 sprints was not well tolerated in this age group following pilot testing, so we reduced the protocol to 6 sprints, but limited the recovery period to 60s. The MIE was matched to HIIE for external mechanical work completed, beginning with a 3-minute unloaded warm-up (0 W) and entailed 15 min of cycling at ~44% W_{max} . The order of completion was randomized.

Each participant visited the laboratory on 3 occasions separated by a 48 h break. Each visit occurred at the same time of day and participants were asked to arrive 2 hours fasted, well hydrated and having abstained from vigorous exercise and caffeine for 24 hours before participating. A low-fat snack was allowed while participants waited their turn and the same snack was brought for remaining visits. The first visit was used for familiarisation and to complete anthropometric measurements and a maximal exercise test. During the two experimental visits participants completed baseline measurements of HR, MAP, PETCO₂ and MCAv. The MCAv was assessed continuously throughout HIIE and MIE and MAP was taken during the final minute of exercise in both conditions. Following exercise, PETCO₂ measurement resumed for 5 minute intervals immediately post-, 30-minutes post- and 60 minutes post-exercise, and BP was taken at 30 minutes post- and 60 minutes post-exercise.

2.2 Measurements

2.2.1 History of CBF measurement

In 1945 Seymour Kety and Carl Schmidt were the first to measure CBF in humans using a low concentration of nitrous oxide inhalation (15%) along with internal jugular venous and femoral artery samples. Flow was calculated from a modified Fick equation (Equation 2.1.):

Equation 2.1.
$$CBF = 100 Q_t (A-V)/t$$

Where: CBF is cerebral blood flow (in ml/100g brain mass/minute); Q_t is the quantity of oxygen consumed over time (expressed as ml/100g brain mass); A and V represent the arterial and cerebral venous oxygen content as a percent of volume; t is time in minutes.

Previously described techniques of measuring CBF (Kety & Schmidt, 1945; Kety & Schmidt, 1948; Gibbs et al., 1946; Lassen & Munck, 1955; Hoedt-Rasmussen, 1966; Ingvar DH, Lassen, 1961; Ingvar et al., 1965; Olesen et al., 1971) revealed the dynamic changes in CBF since the regional and global CBF measures required 10-20 minutes per measure. In 1982, Aaslid and coworkers (Aaslid et al., 1982) used non-invasive TCD to evade the time scale issue for estimation of CBF with a real-time index.

2.2.2 Principles of TCD

TCD is based on the classic principles of Doppler ultrasound, whereby sound waves are transmitted through the acoustic windows of the cranium to target certain cerebral vessels, namely the MCA for this investigation. The Doppler probe emits a 1.5-2 MHz wave, which provides optimal resolution given the depth of the deeper cerebral arteries (Willie et al., 2011). The transmitted signal from the probe reaches red blood cells within the target vessel and reflects the receiving signal back to the transducer, where the difference is recorded (Doppler Principles; see equation 2.2) (Willie et al., 2011).

Equation 2.2.
$$\text{Doppler shift} = 2 \times Ft \times V \times \cos\theta / C$$

Where: Ft is the transmitted frequency (2 MHz); V is the velocity of the reflector (red blood cells); $\cos\theta$ is the correction factor based on the angle of insonation; and C is the speed of sound in the blood (1540m/s).

The frequency data is then converted to the time domain through a fast Fourier transform for tangible velocity measures. The minimum angle of insonation required for TCD must be

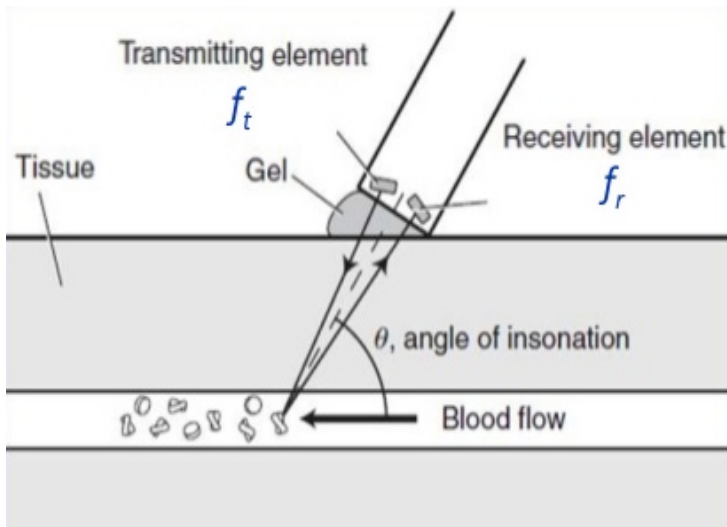


Figure 2.1. Example of the Doppler shift. The frequency difference between the transmitted signal (f_t) and the receiving signal (f_r) is used to determine the direction and velocity of the red blood cells.

below 60 degrees (cosine of 0.50), although it is encouraged to reduce this angle to between 0 and 30 degrees (cosine range 1.00 to 0.87) for optimal quality (See Figure 2.1) (Wintermark et al., 2005). The MCA is particularly in line with the probes typically resulting in an insonation angle below 15 degrees (Wintermark et al., 2005).

2.2.3 Technique of TCD

The process of collecting MCAv measures with TCD begins with a generous application of acoustic gel to the temporal window and probe for optimal signal conduction. An adjustable headband stabilizes probe positioning and maintains the signal quality during data collection. It is imperative that the sonographer has an understanding of the basal structure of the circle of Willis and the common insonation depths associated with each vessel (MCA: 30-65 mm; PCA: 55-75 mm) (Alexandrov et al., 2007). Techniques to confirm the insonated vessel include carotid artery occlusion resulting in a diminished MCAv and unaltered PCAv and an occipital lobe stimulation (eyes open and closed) where the PCAv will increase by up to 20% with eyes open whereas velocity in the MCA will increase much less (<5%).

2.2.4 CBFv acquisition

In this study, MCAv was measured unilaterally (left side) using a 2 MHz TCD ultrasound (Spencer Technologies, Colorado). The MCA was insonated through the trans-temporal window using previously described techniques (Willie et al., 2011). The coefficient of variation for day-to-day reproducibility of MCAv was 6.2% for this study. MCAv was recorded continuously during the VO_2max test, as well as during and after the two experimental exercise conditions.

2.2.5 Anthropometric measurement

Body mass was measured to the nearest 0.1Kg with bioelectrical impedance (Tanita TBF-410GS, Arlington Heights, Illinois). Stature was measured to the nearest 0.1cm with a stadiometer (Seca 213 Portable, Hamburg, Germany). Only healthy weight participants ($\text{BMI } 16.34 \pm 2.09$) were included in the data analyses defined as those with a body mass index (BMI) within 1 standard deviation (\pm) of the World Health Organization (WHO) growth standard median (Onis et al., 2007).

2.2.6 Maximal exercise test

Participants performed a ramp maximal test with supramaximal verification to determine VO_2max and Wmax . The test began with an unloaded 3 minute pedaling warm-up followed by a stature specific workload increase ($<110\text{cm } 5 \text{ W}\cdot\text{min}^{-1}$, $110\text{-}125\text{cm } 10 \text{ W}\cdot\text{min}^{-1}$, $125\text{-}150\text{cm } 15 \text{ W}\cdot\text{min}^{-1}$, $>150\text{cm } 20 \text{ W}\cdot\text{min}^{-1}$) until volitional fatigue. Participants were asked to maintain a pedal cadence between 70-80 rpm during exercise and volitional fatigue was

defined as a cadence drop below 60 rpm for five consecutive seconds. Participants completed 5 minutes of unloaded active cool down following exercise then 10 minutes of passive rest prior to a supramaximal verification test. In brief, following a 2-minute warm-up, children completed a 'step' transition to 105% of the maximum power achieved during the ramp test. Participants cycled at this intensity until exhaustion. $\text{VO}_{2\text{max}}$ was verified if the VO_2 slope of increase between the maximal and supramaximal tests increased by $\leq 5\%$ (Barker et al., 2011). Respiratory gas exchange was assessed breath-by-breath throughout the duration of the ramp and supramaximal tests using an online gas analyzer (Oxycon Pro, Viasys Jaeger; Hoechberg, Germany) first calibrated with gas of a known concentration and flow volume. Data were interpolated to 1s and averaged every 15s.

2.2.7 HR and arterial blood pressure

A three-lead ECG was used to assess beat-by-beat HR. SBP and DBP were measured 3 consecutive times at rest, averaging the last 2 measurements for the resting value, and then in the final minute of exercise, 30-minutes post and 60-minutes post exercise with a manual sphygmomanometer (Prestige Medical, 79-BLK Standard Aneroid, Northridge CA). Values were recorded to the nearest mmHg. DBP was defined as the point of disappearance of Korotkoff sound (5th phase) and MAP was subsequently calculated.

2.2.8 CVRi acquisition

The CVRi was calculated as MAP divided by MCAv ($\text{mmHg}/\text{cm}\cdot\text{s}^{-1}$) at rest, the final minute of exercise in both conditions, and during each of the recovery time points.

2.2.9 PETCO₂ acquisition

Samples of PETCO₂ were collected using a calibrated online gas analyzer (ML206; ADInstruments, Colorado Springs, CO) before and after the two experimental trials (pre, post, 30-minutes post and 60-minutes post exercise), each for 5 minutes in duration. PETCO₂ was not assessed during HIIE or MIE because children did not tolerate the mouthpiece/noseclip during HIIE.

2.3 Statistical analyses

Descriptive data are expressed as mean \pm SD. Changes in MCAv during exercise is presented as both an absolute (Δ) and relative change ($\Delta\%$) from baseline. The response of the main outcome measures (MCAv, PETCO₂, MAP & CVRi) during and after MIE and HIIE were examined using repeated measures analyses of variance (ANOVA), with follow-up t-tests where necessary using Bonferroni corrections. The Δ MCAv and Δ PETCO₂ responses were assessed at each recovery point. An alpha level was set a priori at 0.05. Statistical analyses were performed using SPSS (version 24, SPSS Inc., Chicago, IL, USA).

Chapter 3 Results

3.1 Descriptive data

Data are presented on 9 of the 19 participants recruited. One child was not included in the analyses because they were already pubertal (Tanner 3 for both pubic hair and breast development). One child did not want to return after the maximal exercise test, one child ate a bacon sandwich prior to testing, one child performed vigorous exercise within 24 hours and

the MCA could not be insonated or resulted in inconsistent day-to-day measures in the remaining 6. Of the 9 participants included in the analyses, 4 were boys (age 9.0 ± 2.2 y; stature 137.1 ± 15.2 cm; body mass 30.6 ± 10.3 kg) and 5 were girls (age 10.4 ± 0.9 y; stature 148.8 ± 3.8 cm, body mass 37.4 ± 6.2 kg). All children completed a $\text{VO}_{2\text{max}}$ test with a maximum wattage of 124 ± 27 W (range 67-156W); HRmax of $193 \text{ beats} \cdot \text{min}^{-1}$ (range 183-195 $\text{beats} \cdot \text{min}^{-1}$); maximum MAP of 94.1 ± 11.7 (range 81.3-118.0); the MCAv at maximum VO_2 was $80.4 \pm 7.8 \text{ cm} \cdot \text{s}^{-1}$ (range 66.8-91.6 $\text{cm} \cdot \text{s}^{-1}$).

3.2 HR and MCAv responses during HIIE and MIE

Continuous HR was collected with 3-lead ECG throughout the entire protocol. For congruency with other data, HR is displayed in minute-by-minute averages as a descriptive of the cardiovascular effects of the different exercise conditions (See Figure 3.1). When HR was expressed in absolute terms, there was a main effect for time ($F(11, 110) = 42.715$, $P < 0.001$, $\eta^2 = 987.891$) and intensity ($F(11, 110) = 12.915$, $P < 0.001$, $\eta^2 = 298.694$; see Figure 3.1).

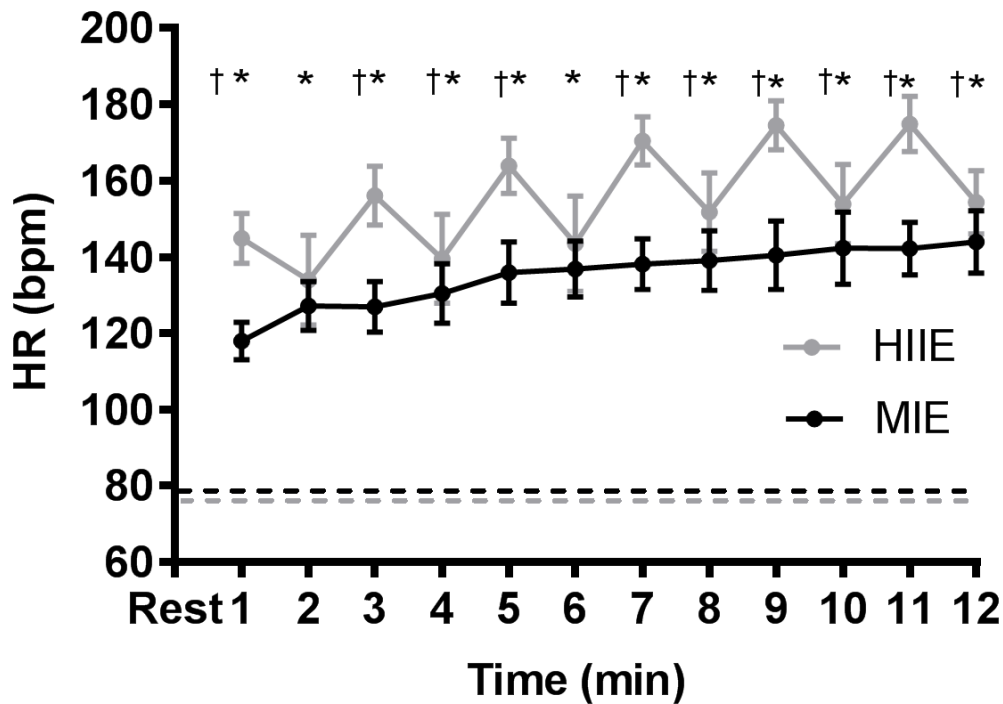


Figure 3.1. Minute-by-minute HR values during HIIE (grey line) and MIE (black line). For the HIIE condition every odd minute (1,3,5 etc.) represents an exercise interval, whereas even minutes (2,4,6 etc.) represent active rest intervals. The dotted lines represent baseline HR values. * significant difference from baseline $P < 0.05$; † significant difference between conditions.

Absolute values for MCAv during HIIE and MIE are provided in Table 3.1 and expressed as minute by minute averages to unveil the difference between rest and exercise intervals during HIIE. The MIE condition was extracted using the same minute to minute technique for the purpose of comparison. When expressed as a percentage change from baseline ($\% \Delta \text{MCAv}$) there was a main effect for time ($F(12, 192) = 29.096$, $P < 0.001$, $\eta^2 = 0.645$), in an intensity specific manner ($F(12, 192) = 3.647$, $P < 0.001$, $\eta^2 = 0.186$; see Figure 3.2). Follow-up analyses revealed that MCAv increased and remained elevated above baseline values ($p < 0.05$) for the first 3 minutes of HIIE and 4 minutes of MIE, before returning toward baseline (see Table 3.1 & Figure 3.2).

Table 3.1. MCAv during HIIE and MIE.

HIIE (n=9)		MIE (n=9)	
Time (min)	MCAv (cm.s ⁻¹)	Time (min)	MCAv (cm.s ⁻¹)
Base	74.20 ± 5.17	Base	72.70 ± 2.01
1	*83.49 ± 6.99	1	*78.18 ± 6.27
2	*83.82 ± 7.34	2	*79.86 ± 6.56
3	*78.38 ± 5.46	3	*77.79 ± 7.23
4	77.11 ± 6.81	4	*76.72 ± 5.96
5	72.96 ± 5.47	5	75.62 ± 5.14
6	73.44 ± 6.90	6	74.81 ± 5.56
7	*70.98 ± 5.33	7	73.61 ± 4.95
8	74.62 ± 7.61	8	74.68 ± 5.14
9	*70.83 ± 5.28	9	73.63 ± 4.08
10	73.54 ± 7.60	10	72.14 ± 4.46
11	*70.38 ± 4.93	11	71.50 ± 5.00
12	72.57 ± 8.20	12	72.08 ± 4.50

MCAv, middle cerebral artery velocity (cm/s); Base, baseline. Table includes twelve minutes of minute-by-minute absolute values. In the HIIE condition every odd minute (1,3,...) represents an exercise interval average and each even minute (2,4,...) represents a rest interval average. MIE is minute-by-minute averages.

*significant difference from baseline P<0.05.

The MCAv remained above baseline from the onset of exercise until the 4th minute of MIE (6.9% and 5.4%, respectively; p 's <0.05) before declining back to baseline for the remainder of the MIE trial (see figure 3.2). In the HIIE condition, follow-up analyses revealed a 12.1% increase in MCAv during the first sprint interval (P <0.001), remaining elevated above baseline during the first rest interval (12.7%; P <0.05). During the second sprint interval, MCAv remained 5.4% above baseline (P <0.05), but declined thereafter, with significant reductions below baseline by the 4th (-4.6%), 5th (-4.8%) and 6th sprint intervals (-5.3%). During the rest intervals between sprints, MCAv returned to baseline (see figure 3.2).

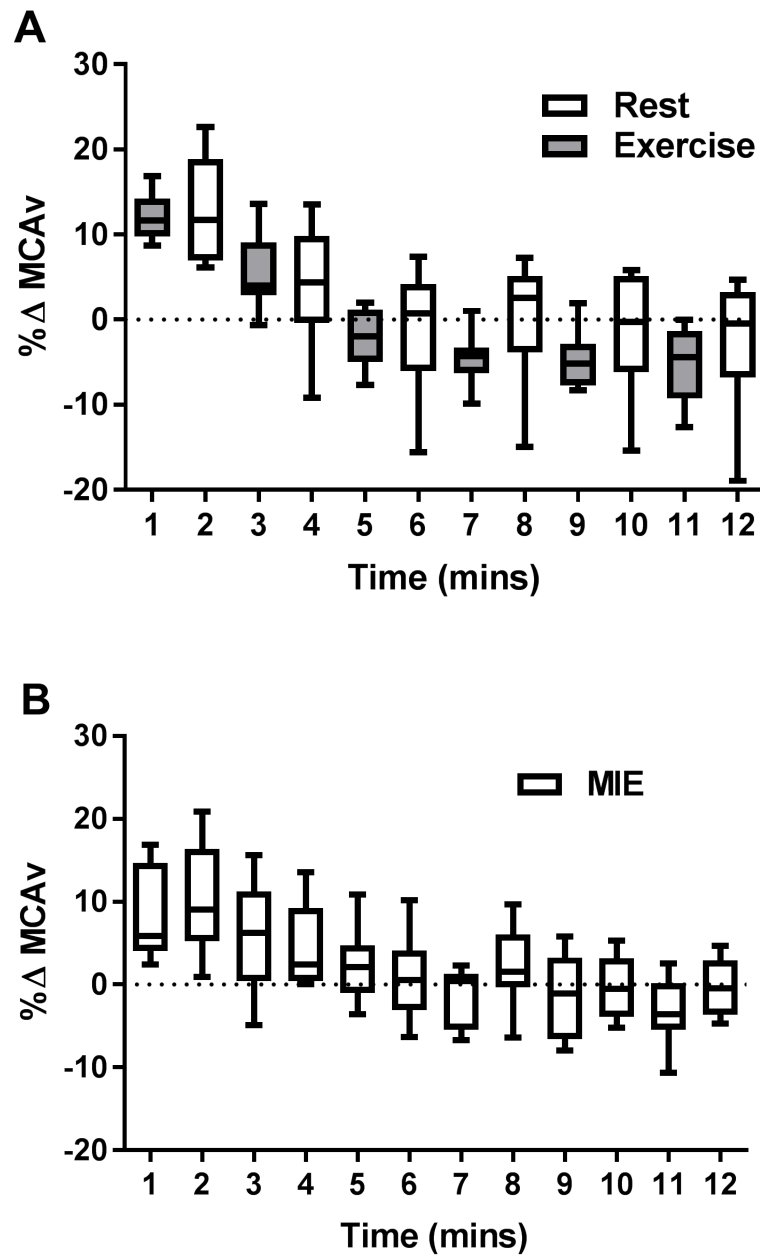


Figure 3.2 The relative change in MCAv (%) during HIIE (panel A) exercise (Ex) and rest (rest) intervals and MIE (panel B). Individual data are provided within the bars (black dots). * significant difference from baseline $P < 0.05$.

It seems that during exercise the MCAv responses are similar between HIIE and MIE at the onset of exercise until the 4th HIIE sprint, thereafter MCAv oscillated significantly below

baseline during HIIE and returned to baseline during rest, whereas MCAv was maintained at baseline values beyond the 4th minute of MIE.

3.3 MCAv and PETCO₂ responses following HIIE and MIE

The MCAv is illustrated in figure 3.3 (panel A) at baseline, immediately post exercise, 30 minutes post- and 60 minutes post exercise. There was no main effect for time or an interaction, with MCAv remaining at baseline values after both exercise conditions.

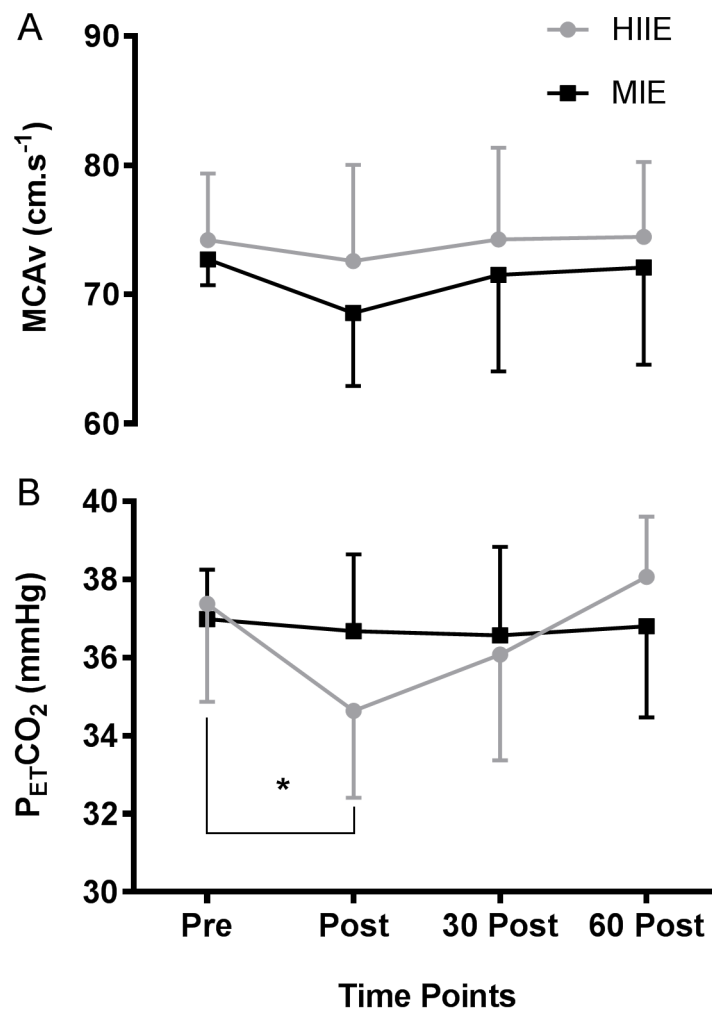


Figure 3.3 The response of MCAv (A) and PETCO₂ (B) at baseline (pre), immediately following HIIE and MIE (Post), 30 minutes post exercise (30 Post) and 60 minutes post HIIE and MIE (60 Post). The grey lines represent HIIE and the black lines represent MIE. *significant difference from baseline P<0.05.

PETCO₂ at baseline, at the end of HIIE and MIE, as well as 30- and 60-minutes post exercise are presented in figure 3.3, panel b. There was a main effect of time ($F(3,48) = 6.339$, $P < 0.001$, $\eta^2 = 0.284$), and an interaction ($F(3,48) = 4.765$, $P < 0.05$, $\eta^2 = 0.229$). Simple effects revealed a 7.1% decline in PETCO₂ after HIIE, which returned to baseline values within 30 minutes post exercise. The PETCO₂ remained unchanged from baseline following MIE and at all recovery time points.

3.4 MAP, MCA_v and CVR_i during the final minute of HIIE and MIE and during recovery.

MAP was assessed at baseline, at the end of exercise and during recovery. MAP changed with time ($F(3, 48) = 45.184$, $P < 0.001$, $\eta^2 = 0.738$), but there was no interaction (see Table 3.2). In both exercise conditions, MAP was increased to a similar extent at the end of exercise (p 's < 0.001), before returning to baseline at 30 minutes and 60 minutes post exercise.

Table 3.2. MAP and MCAv and CVRi at baseline, during and following HIIE and MIE.

	Time	MAP (mmHg)	MCAv (cm.s ⁻¹)	CVRi
HIIE	Base	81.71 ± 3.78	76.78 ± 5.17	1.07 ± 0.09
	Ex	*90.63 ± 3.82	*70.21 ± 4.59	*1.30 ± 0.10
	30 post	82.31 ± 6.70	75.95 ± 5.10	1.07 ± 0.11
	60 post	81.79 ± 5.29	77.06 ± 4.97	1.07 ± 0.10
MIE	Base	81.26 ± 4.18	70.93 ± 6.65	1.15 ± 0.11
	Ex	*87.96 ± 4.35	71.88 ± 7.13	*1.23 ± 0.13
	30 post	81.97 ± 4.15	71.17 ± 7.32	1.17 ± 0.11
	60 post	80.74 ± 4.63	71.89 ± 5.80	1.13 ± 0.09

MAP, mean arterial pressure; MCAv, middle cerebral artery velocity; CVRi, cerebral vascular resistance index; Base, baseline; Ex, final minute of exercise; 30 post, 30 minutes post exercise; 60 post, 60 minutes post exercise.

*significant difference from baseline P<0.05.

Figure 3.4 illustrates CVRi expressed as a percentage change. There was a main effect of time ($F(3,48) = 50.852$, $P < 0.001$, $\eta^2 = 0.761$) and an interaction ($F(3,48) = 12.606$, $P < 0.001$, $\eta^2 = 0.441$).

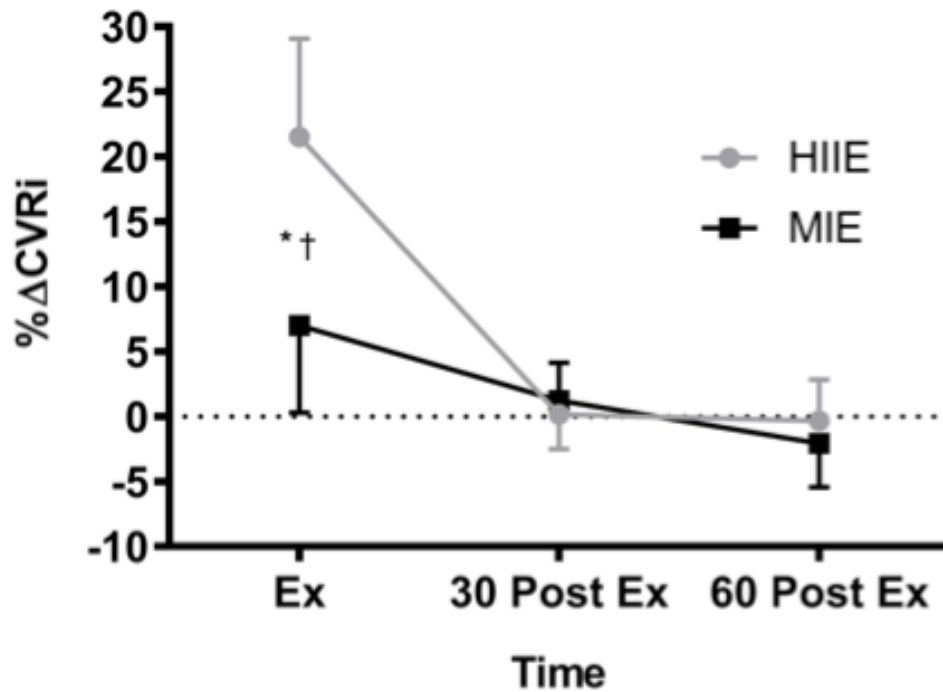


Figure 3.4 The response of the relative change in CVRi to exercise and during recovery. Ex, final minute of exercise; 30 Post Ex, 30 minutes post exercise; 60 Post Ex, 60 minutes post exercise. * significant difference ($P < 0.05$) from baseline; † significant difference ($P < 0.05$) between HIIE and MIE.

The $\% \Delta \text{CVRi}$ significantly increased from baseline during the final minute in both exercise conditions (p 's < 0.05), although the magnitude of this increase was greater during HIIE, with CVRi elevated 21.5% above baseline during the final minute of HIIE, and only 7.0% during MIE. CVRi returned to baseline values 30 minutes post exercise in both conditions (see figure 3.4).

Chapter 4 Discussion

4.1 Experimental findings

The main findings of the current study were as follows: HIIE and MIE elicited a similar MCAv response during the first 3 minutes of exercise, with MCAv elevated for the first 3 minutes of HIIE and for the first 4 minutes of MIE. Although MCAv was maintained at baseline values throughout the remaining duration of MIE, this differed from the HIIE condition whereby MCAv decreased significantly below baseline for the final 3 sprints and oscillated back to baseline in the remaining rest intervals. The oscillatory nature of MCAv during HIIE may be due in part to the rapid redistribution of blood to the working muscles during exercise resulting in decreased MCAv at that time. The CVRi was higher in HIIE compared to MIE during the final minute of both exercise intensities ($P < 0.05$). Although this difference was perhaps mediated via the selective hypocapnia resulting from hyperventilation during HIIE, we did not collect PETCO₂ during exercise. The changes in CVRi were normalized at 30 minutes post- and 60 minutes post exercise in both conditions. Immediately post HIIE, MCAv returned to baseline uncoupling from the PETCO₂ response, which fell significantly below baseline ($P < 0.05$). Following MIE, however, MCAv and PETCO₂ exhibited a similar response remaining at baseline immediately post MIE (See Figure 3.3). The following discussion outlines these findings and highlights strengths and limitations associated with our experimental design.

4.1.1 Cerebrovascular responses during HIIE and MIE

Contrary to our first hypothesis, we did not find a significantly greater increase in MCAv during MIE compared to HIIE. In previous work investigating the child's cerebrovascular response during maximal exercise, Ellis et al (2017) found no significant change in the child's %MCAv from baseline, despite modest increases of ~10% up until the respiratory compensation point, before MCAv began to decline back toward baseline. Similar to the work by Ellis et al (2017) we found that during HIIE, the MCAv increased above baseline in the initial 3 minutes of HIIE before declining to and below baseline for the remainder of the protocol. Regarding the effects of MIE on MCAv, Ogoh & Ainslie (2009) summarized in adults that MCAv remains elevated above baseline throughout the duration of a continuous bout of MIE, although no child data other than those presented here is currently available. We found that the child's MCAv remained elevated only for the first four minutes of continuous MIE before returning to baseline values for the remainder of the test. Perhaps, this is in part due to the already elevated resting MCAv levels in children compared to adults, which is sufficient for the child's cerebral oxygen demands.

Finally, during the last minute of exercise in HIIE and MIE, the CVRi was calculated to investigate a metric considering both MAP and MCAv (see Equation 1.1). We found that CVRi significantly increased in the final minute of exercise in both HIIE and MIE ($P < 0.05$), but this was exacerbated in HIIE ($P < 0.05$). The elevated CVRi response noted in HIIE compared to MIE may be partly due to the hyperventilation-induced hypocapnia resulting from a harder work rate associated with HIIE compared to MIE. The PETCO₂ was only collected during recovery (discussed below), however, so no further conclusions can be

made since we did not monitor PETCO₂ during the final minute of exercise. Although MAP significantly increased from baseline during the final minute of exercise in both conditions, MAP did not significantly differ between intensities. Interestingly, MCAv during the final minute of exercise was at baseline in MIE and below baseline in HIIE, suggesting that perhaps HIIE does not pose any risk of hyperperfusion to the child compared to MIE.

4.1.2 Cerebrovascular responses following HIIE and MIE

As a surrogate measure of PaCO₂, intermittent samples of PETCO₂ were collected at baseline as well as throughout acute recovery following both HIIE and MIE, and findings support the final hypothesis. Immediately post HIIE, the PETCO₂ values fell significantly below baseline, which may correspond with the harder work rate and likely increased ventilation during HIIE compared to MIE. The decline in PETCO₂ immediately post HIIE was not coupled with a decrease in MCAv. The CVRi significantly increased during the last minute of exercise in HIIE, and perhaps this was mediated via a hypocapnic response associated with HIIE eliciting the decline in MCAv in the final minute of HIIE. Thereafter, the CVRi returned to baseline in HIIE following 30 minutes of recovery, as did PETCO₂. The MCAv and PETCO₂ values immediately following MIE remained at baseline values and did not change throughout the hour of recovery.

4.1.3 Experimental considerations

A similar approach to Bond et al (2015) was used to determine the work rates of HIIE. Exercise intervals in HIIE were performed at 90% W_{max}, lasting 1 minute in duration as Bond et al (2015) did. The protocol differed from that used by Bond et al (2015) in rest

intervals, which were calculated as 20% Wmax rather than a set 20W resistance, and the recovery time was reduced from 75 seconds to 60 seconds. Work completed during rest intervals was altered so it was relative to the individual child and a reduced resting period time compared to Bond et al (2015) was incorporated compensate for decreasing the 8 exercise intervals used down to 6 intervals in our experiment. We reduced the number of intervals due to lack of completion by the first 3 participants despite lots of encouragement. Our participants achieved an average HR of 80% HR max across the entire HIIE protocol, although this is an average and was not achieved in every participant. Across all exercise intervals the average HR increased to 85% HR max indicating hard working efforts. There is, however, an issue in the derivation of such values from an initial maximum test that may not sufficiently stress the child's anaerobic system. Previous findings demonstrate that children completing a standard VO₂max test do not experience significant increases in blood La (Armstrong et al., 1996). Armstrong and colleagues (1996) found that blood La can be further increased in children when completing a supramaximal exercise test. We specifically chose a high-intensity, interval modality as this is closer to the free-living physical activity patterns in children (Berman et al., 1998), compared to a one-off Wingate all-out bout of exercise (Curtelin et al., 2017). It is likely our findings are indicative of free-living 'high-intensity' exercise, but do not reflect response to all-out exercise such as the Wingate test.

4.1.4 Strengths and limitations

Limitations to this experiment include a reduced sample size, not separating for sex differences, not providing continuous measures of MAP or PETCO₂, not collecting La and using velocity measures as opposed to true CBF. Initially 19 participants were recruited to

participate in this study and sex differences would have been explored. The main reason for exclusion was an inability to repeat the day-to-day MCAv baseline values across all three visits.

The use of TCD has been under scrutiny since recent MRI work revealed MCAv diameter increases in response to increases in PETCO₂ greater than ~8 mmHg (Coverdale et al., 2014; Verbree et al., 2014). A large reduction in PETCO₂ may result in constriction of the MCA diameter by ~4% (Coverdale et al., 2014). However, the changes in PETCO₂ during HIIE and MIE in the current experiment were relatively minor and are unlikely to have elicited either constriction or dilation in the MCA.

The protocol used in this experiment, although eliciting a higher cardiovascular response, was not as metabolically taxing as expected. Perhaps future work investigating the effects of HIIE on the child's cerebrovasculature should consider the recent protocol used by Curtelin et al (2017), who tested the cerebrovascular response in adults to an all-out Wingate (30s) test. The time-course of CBFv, PETCO₂ and MAP could be explored, and it might also be prudent to collect blood La, although acquiring beat to beat blood pressure and La during HIIE would likely prove to be too invasive in child.

4.2 Conclusion

To conclude, although the current findings reveal noteworthy differences in the cerebrovascular responses to HIIE and MIE (See Figure 3.2), it appears that HIIE poses no additive risk of hyperperfusion injury compared to MIE in the child. The regulatory effects of PaCO₂ (PETCO₂) may be diminished in children immediately following HIIE, although this differs from MIE as PETCO₂ and MCAv remained coupled at baseline. The uncoupling

between MCAv and PETCO₂ noted immediately following HIIE corroborates recent work by Ellis et al. (2017), supporting the contention that the regulatory mechanisms of the CBFv response to high intensity exercise may be developmentally divergent.

The MCAv decline back to baseline in the latter half of MIE is perhaps an outcome of the child's elevated baseline MCAv compared to adults. This elevated MCAv sufficiently supplies the child's cerebral oxygen demands during exercise. In HIIE, however, significant MCAv reductions below baseline occurred in the final 3 sprints, which could pose a hypoperfusive threat to the cerebrovasculature. It is likely that the increased MAP during HIIE likely countered the effects of the decreased MCAv. This increase in MAP during HIIE did not differ from that of MIE, so MAP is elevated during exercise to counter potential decreases in CBFv, or whether these are not related remains to be determined.

To conclude, Canadian children are not currently meeting physical activity guideline recommendations (Tremblay et al., 2011B). The similar cerebrovascular responses at the onset of HIIE and MIE means HIIE is an equally attractive exercise modality for the child and one that is time efficient and congruent with their naturally sporadic movement characteristics (Bailey et al., 1995).

References

1. Aaslid R, Lindegaard KF, Sorteberg W, Nornes H. Cerebral autoregulation dynamics in humans. *Stroke*. 1989; 20:45–52.
2. Aaslid R, Markwalder TM, Nornes H. Noninvasive transcranial Doppler ultrasound recording of flow velocity in basal cerebral arteries. *Journal of Neurosurgery*. 1982; 57:769–774.
3. Ainslie PN, Cotter JD, George KP, Lucas S, Murrell C, Shave R, Thomas KN, Williams MJ & Atkinson G. Elevation in cerebral blood flow velocity with aerobic fitness throughout healthy human ageing. *The Journal of Physiology*. 2008; 586(16):4005-4010.
4. Ainslie PN & Hoiland RL. Transcranial Doppler ultrasound: Valid, invalid, or both? *Journal of Applied Physiology*. 2014; 117:1081-1083.
5. Ainslie PN, Hoiland RL & Bailey DM. Lessons from the laboratory; Integrated regulation of cerebral blood flow during hypoxia. *Experimental Physiology*. 2016; 101(9):1160-1166.
6. Ainslie PN & Burgess KR. Cardiorespiratory and cerebrovascular responses to hyperoxic and hypoxic rebreathing: Effects of acclimatization to high altitude. *Respiratory Physiology and Neurobiology*. 2008; 161:201–209.
7. Armstrong N, Kirby BJ, McManus AM & Welsman JR. Aerobic fitness of prepubescent children. *Annals of Human Biology*. 1995; 22:427-441.
8. Armstrong N, Welsman J & Winsley R. Is peak VO_2 a maximal index of children's aerobic fitness? *International Journal of Sports Medicine*. 1996; 17(5):356-359.
9. Armstrong N, Williams J, Balding I, Gentle P & Kirby B. The peak oxygen uptake of British children with reference to age, sex and sexual maturity. *European Journal of Applied Physiology*. 1991; 62:369-375.

10. Artru AA, Katz RA, Colley PS. Autoregulation of cerebral blood flow during normocapnia and hypocapnia in dogs. *Anesthesiology*. 1989; 70:288–292.
11. Astrand PO & Rodahl I. Textbook of Work Physiology. New York, McGraw-Hill, 1986.
12. Azad N, Pitale S, Barnes WE & Friedman N. Testosterone treatment enhances regional brain perfusion in hypogonadal man. *The Journal of Clinical Endocrinology and Metabolism*. 2003; 88:3064–3.
13. Bailey RC, Olson J, Pepper SL, Porszasz J, Barstow TJ & Cooper DM. The level and tempo of children's physical activities: An observational study. *Medicine and Science in Sports and Exercise*. 1995; 27(7):1033-1041.
14. Bakker SL, de Leeuw FE, den Heijer T, Koudstaal PJ, Hofman A & Breteler MM. Cerebral haemodynamics in the elderly: The Rotterdam study. *Neuroepidemiology*. 2004; 23:178–184.
15. Baquet G, Berthoin S, Dupont G, Blondel N, Fabre C & Van Praagh E. Effects of high intensity intermittent training on peak VO₂ in prepubertal children. *International Journal of Sports Medicine*. 2002; 23(06):439-444.
16. Baquet G, Gamelin FX, Mucci P, Thévenet D, Van Praagh E & Berthoin S. Continuous vs. interval aerobic training in 8 to 11 year old children. *The Journal of Strength & Conditioning Research*. 2010; 24(5):1381-1388.
17. Baquet G, Stratton G, Van Praagh E & Berthoin S. Improving physical activity assessment in prepubertal children with high frequency accelerometry monitoring. *Preventive Medicine*. 2007; 44:143–147.
18. Barker AR, Williams CA, Jones AM & Armstrong N. Establishing maximal oxygen uptake in young people during a ramp cycle test to exhaustion. *British Journal of Sports Medicine*. 2001; 45(6):498-503.

19. Barnes JN, Taylor JL, Kluck BN, Johnson CP & Joyner MJ. Cerebrovascular reactivity is associated with maximal aerobic capacity in healthy older adults. *Journal of Applied Physiology*. 2013; 114(10):1383-1387.
20. Berman N, Bailey R, Barstow T & Cooper D. Spectral and bout detection analysis of physical activity patterns in healthy, prepubertal boys and girls. *American Journal of Human Biology*. 1998; 10:289 – 297.
21. Biagi L, Abbruzzese A, Bianchi MC, Alsop DC, Del Guerra A & Tosetti M. Age dependence of cerebral perfusion assessed by magnetic resonance continuous arterial spin labeling. *Journal of Magnet Resonance Imaging*. 2007; 25:696–702.
22. Bill A & Linder J. Sympathetic control of cerebral blood flow in acute arterial hypertension. *Acta Physiologica Scandinavica*. 1976; 96:114–121.
23. Bishop CCR, Powell S & Rutt D. Transcranial Doppler measurement of middle cerebral artery blood flow velocity: A validation study. *Stroke*. 1986; 17(5):913-915
24. Boddy LM, Stratton G, Hackett AF & George KP. The effectiveness of a ‘short, sharp, shock’ high intensity exercise intervention in 11 and 12 year-old Liverpool schoolgirls. *Archives of Exercise in Health and Disease*. 2010; 1(1):19-25.
25. Bode H & Eden A. Transcranial Doppler sonography in children. *Journal of Child Neurology*. 1989; 4(1):S68-S76.
26. Bolduc V, Thorin-Trescases N & Thorin E. Endothelium-dependent control of cerebrovascular functions through age: Exercise for healthy cerebrovascular aging. *American Journal of Physiology – Heart and Circulatory Physiology*. 2013; 305:H620–H633.
27. Bond B, Hind S, Williams CA & Barker AR. The acute effect of exercise intensity on vascular function in adolescents. *Medicine and Science in Sports and Exercise*. 2015. DOI:

<http://dx.doi.org/10.1249/MSS.0000000000000715>

28. Brassard P, Kim Y-S, van Lieshout J, Secher NH, Rosenmeier JB. Endotoxemia reduces cerebral perfusion but enhances dynamic cerebrovascular autoregulation at reduced arterial carbon dioxide tension. *Critical Care Medicine*. 2012; 40:1873–1878.
29. Brugniaux JV, Marley CJ, Hodson DA, New KJ & Bailey DM. Acute exercise stress reveals cerebrovascular benefits associated with moderate gains in cardiorespiratory fitness. *Journal of Cerebral Blood Flow & Metabolism*. 2014; 34(12):1873-1876.
30. Burley CV, Bailey DM, Marley CJ & Lucas SJ. Brain train to combat brain drain; Focus on exercise strategies that optimize neuroprotection. *Experimental Physiology*. 2016; 101(9):1178-1184.
31. Calbet JAL, González-Alonso J, Helge JW, Søndergaard H, Munch-Andersen T, Saltin B & Boushel R. Central and peripheral hemodynamics in exercising humans: Leg vs arm exercise. *Scandinavian Journal of Medicine & Science in Sports*. 2015; 25(S4):144-157.
32. Carroll TJ, Teneggi V, Jobin M, Squassante L, Treyer V, Hany TF, Burger C, Wang L, Bye A, Schulthess Von GK & Buck A. Absolute quantification of cerebral blood flow with magnetic resonance, reproducibility of the method, and comparison with positron emission tomography. *Journal of Cerebral Blood Flow and Metabolism*. 2002; 22:1149–1156.
33. Chaddock-Heyman L, Erickson KI, Chappell MA, Johnson CL, Kienzler C, Knecht A, Drollette ES, Raine LB, Scudder MR, Kao SC & Hillman CH. Aerobic fitness is associated with greater hippocampal cerebral blood flow in children. *Developmental Cognitive Neuroscience*. 2016; 20:52-58.
34. Chambliss KL & Shaul PW. Estrogen modulation of endothelial nitric oxide synthase. *Endocrine Reviews*. 2006; 23:665– 686.

35. Chemick NS & Longobardo GS. Oxygen and carbon dioxide gas stores of the body. *Physiological Reviews*. 1970; 50:196-243.
36. Chia M, Armstrong N & Childs D. The assessment of children's anaerobic performance using modifications of the Wingate anaerobic test. *Pediatric Exercise Science*. 1997; 9(1):80-89.
37. Christian D, Todd C, Hill R, Rance J, Mackintosh K, Stratton G & Brophy S. Active children through incentive vouchers–evaluation (ACTIVE): A mixed-method feasibility study. *BioMed Central Public Health*. 2016; 16(1):890.
38. Chugani HT. A critical period of brain development: Studies of cerebral glucose utilization with PET. *Preventive Medicine*. 1998; 27(2):184-188.
39. Chung A, Backholer K, Wong E, Palermo C, Keating C & Peeters A. Trends in child and adolescent obesity prevalence in economically advanced countries according to socioeconomic position: A systematic review. *Obesity Reviews*. 2016; 17(3):276-295.
40. Cooke JP. The pivotal role of nitric oxide for vascular health. *The Canadian Journal of Cardiology*. 2004; 20:7B-15B.
41. Cooper DM, Kaplan MR, Baumgarten L, Weiler-Ravell D, Whipp BJ & Wasserman K. Coupling of ventilation and CO₂ production during exercise in children. *Pediatric Research*. 1987; 21(6):568-72.
42. Cooper DM, Weiler-Ravell D, Whipp BJ & Wasserman K. Aerobic parameters of exercise as a function of body size during growth in children. *Journal of Applied Physiology*. 1984; 56:628-634.
43. Costigan SA, Eather N, Plotnikoff RC, et al. Preliminary efficacy and feasibility of embedding high intensity interval training into the school day: A pilot randomized controlled

- trial. *Preventative Medicine Reports*. 2015; 2:973–9.
44. Curtelin D, Morales-Alamo D, Torres-Peralta R, Rasmussen P, Martin-Rincon M, Perez-Valera M, Siebenmann C, Pérez-Suárez I, Cherouveim E, Sheel AW, Lundby C & Calbet J AL. Cerebral blood flow, frontal lobe oxygenation and intra-arterial blood pressure during sprint exercise in normoxia and severe acute hypoxia in humans. *Journal of Cerebral Blood Flow & Metabolism*. 2017; p.0271678X17691986.
 45. Davignon J & Ganz P. Role of endothelial dysfunction in atherosclerosis. *Circulation*. 2004; 109(23):III-27.
 46. DeBellis MD, Keshavan MS, Beers SB, Hall J & Frustaci K. Sex differences in brain maturation during childhood and adolescence. *Cerebral Cortex*. 2001; 11:552–557.
 47. De Araujo ACC, Roschel H, Picanço AR, do Prado DML, Villares SMF, de Sa Pinto AL & Gualano B. Similar health benefits of endurance and high-intensity interval training in obese children. *Public Library of Online Science One*. 2012; 7(8):e42747.
 48. Demirkaya S, Uluc K, Bek S & Vural O. Normal blood flow velocities of basal cerebral arteries decrease with advancing age: A transcranial Doppler sonography study. *The Tohoku Journal of Experimental Medicine*. 2008; 214(2):145-149.
 49. Drummond JC. The lower limit of autoregulation: Time to revise our thinking? *Anesthesiology*. 1997; 86:1431–1433.
 50. Einstein EH, Song LH, Villela NL, Fasani-Feldberg GB, Jacobs JL, Kim DO, Nathawat A, Patel D, Bender RB & Peters DF. Anomalous origin of the left vertebral artery from the aortic arch. *Aorta*. 2016; 4(2): 64-67.
 51. Ellis LA, Ainslie PN, Armstrong VA, Morris LE, Simair RG, Sletten NR, Tallon CM & McManus AM. Anterior cerebral blood velocity and end-tidal CO₂ responses to exercise

- differ in children and adults. *American Journal of Physiology-Heart and Circulatory Physiology*. 2017; pp.ajpheart-00034.
52. Eston RG, Lambrick DM & Rowlands AV. The perceptual response to exercise of progressively increasing intensity in children aged 7–8 years: Validation of a pictorial curvilinear ratings of perceived exertion scale. *Psychophysiology*. 2009; 46(4):843-851.
 53. Fakhouri TH, Hughes JP, Brody DJ, Kit BK & Ogden CL. Physical activity and screen-time viewing among elementary school-aged children in the United States from 2009 to 2010. *Jama Pediatrics*. 2013; 167(3):223-229.
 54. Fenton RA, Rubio R & Berne RM. Adenosine and the acid-base state of vascular smooth muscle. *Journal of Applied Physiology*. 1981; 51(1):179–84.
 55. Fisher JP, Hartwich D, Seifert T, Olesen ND, McNulty CL, Nielsen HB, Van Lieshout JJ & Secher NH. Cerebral perfusion, oxygenation and metabolism during exercise in young and elderly individuals. *Journal of Physiology*. 2013; 591:1859–1870.
 56. Fox KR, Cooper A & McKenna J. The school and promotion of children’s health-enhancing physical activity: Perspectives from the United Kingdom. *Journal of School Health*. 2004; 23:338–58.
 57. Fjørtoft I, Kristoffersen B & Sageie J. Children in schoolyards: Tracking movement patterns and physical activity in schoolyards using global positioning system and heart rate monitoring. *Landscape and Urban Planning*. 2009; 93(3):210-217.
 58. Freedman DS, Mei Z, Srinivasan SR, Berenson GS & Dietz WH. Cardiovascular risk factors and excess adiposity among overweight children and adolescents: The Bogalusa heart study. *The Journal of Pediatrics*. 2007; 150(1):12-17.
 59. Gabrielsen TO & Greiz T. Normal size of ICA, MCA. *Acta Radiological: Diagnosis*. 1970;

10:1–10.

60. Gaesser GA & Brooks GA. Metabolic bases of excess post-exercise oxygen consumption: A review. *Medicine and Science in Sports and Exercise*. 1984; 16(1):29.
61. Giardino ND, Friedman SD & Dager SR. Anxiety, respiration, and cerebral blood flow: Implications for functional brain imaging. *Comprehensive Psychiatry*. 2007; 48(2):103-112.
62. Gibbs FA, Maxwell HP & Gibbs EL. Volume flow of blood through the brain of man at rest, during hyperventilation and while breathing high CO₂. *Federation Proceedings*. 1946; 5:33.
63. Gotoh F, Tazaki Y & Meyer JS. Transport of gases through brain and their extravascular vasomotor action. *Experimental Neurology*. 1961; 4:48–58.
64. Gratas-Delamarche A, Mercier J, Ramonatxo M, Dassonville J & Prefaut C. Ventilatory response of prepubertal boys and adults to carbon dioxide at rest and during exercise. *European Journal of Applied Physiology and Occupational Physiology*. 1993; 66(1):25-30.
65. Grubb RL Jr, Raichle ME, Eichling JO, Ter-Pogossian MM. The effects of changes in PaCO₂ on cerebral blood volume, blood flow, and vascular mean transit time. *Stroke*. 1974; 5(5):630–9.
66. Hales PW, Kawadler JM, Aylett SE, Kirkham FJ & Clark CA. Arterial spin labeling characterization of cerebral perfusion during normal maturation from late childhood into adulthood: Normal “reference range” values and their use in clinical studies. *Journal of Cerebral Blood Flow and Metabolism*. 2014; 34:776–784.
67. Hansen HS, Froberg K, Hyldebrandt N & Nielsen JR. A controlled study of eight months of physical training and reduction of blood pressure in children: The Odense schoolchild study. *British Medical Journal*. 1991; 303(6804):682-685.
68. Heyman A, Fillenbaum GG, Welsh-Bohmer KA, Gearing M, Mirra SS, Mohs RC, Peterson

- BL & Pieper CF. Cerebral infarcts in patients with autopsy-proven Alzheimer's disease CERAD, part XVIII. *Neurology*. 1998; 51(1):159-162.
69. Hoedt-Rasmussen K, Sveinsdottir E & Lassen NA. Regional cerebral blood flow in man determined by intra-arterial injection of radioactive inert gas. *Circulation Research*. 1966; 18:237–247.
70. Hofman A, Ott A, Breteler MM, Bots ML, Slooter AJ, van Harskamp F, van Duijn CN, Van Broeckhoven C & Grobbee DE. Atherosclerosis, apolipoprotein E, and prevalence of dementia and Alzheimer's disease in the Rotterdam Study. *The Lancet*. 1997; 349(9046):151-154.
71. Iadecola C & Nedergaard M. Glial regulation of the cerebral microvasculature. *Nature and Neuroscience*. 2007; 10:1369–1376.
72. Ide K, Eliasziw M & Poulin MJ. The relationship between middle cerebral artery blood velocity and end-tidal PCO₂ in the hypocapnic-hypercapnic range in humans. *Journal of Applied Physiology*. 2003; 95:129–37.
73. Ingvar DH & Lassen NA. Quantitative determination of regional cerebral blood- flow in man. *The Lancet*. 1961; 278:806–807.
74. Ingvar DH, Cronqvist S, Ekberg R, Risberg J & Hoedt-Rasmussen K. Normal values of regional cerebral blood flow in man, including flow and weight estimates of gray and white matter. A preliminary summary. *Acta Neurologica Scandinavica Supplementum*. 1965; 14:72–78.
75. Janssen I, Katzmarzyk PT, Boyce WF, Vereecken C, Mulvihill C, Roberts C, Currie C & Pickett W. Comparison of overweight and obesity prevalence in school-aged youth from 34 countries and their relationships with physical activity and dietary patterns. *Obesity Reviews*.

- 2005; 6(2):123-132.
76. Jorgensen LG, Perko M, Hanel B, Schroeder TV & Secher NH. Middle cerebral artery flow velocity and blood flow during exercise and muscle ischemia in humans. *Journal of Applied Physiology*. 1992; 72(3):1123-1132.
77. Juonala M, Jarvisalo MJ, Mäki-Torkko N, Kähönen M, Viikari JS & Raitakari OT. Risk factors identified in childhood and decreased carotid artery elasticity in adulthood: The cardiovascular risk in young finns study. *Circulation*. 2005; 112(10):1486-1493.
78. Kalaria RN. Cerebral vessels in ageing and Alzheimer's disease. *Pharmacology & Therapeutics*. 1996; 72(3):193-214.
79. Karaki H & Weiss GB. Effect of transmembrane pH gradient changes on potassium-induced relaxation in vascular smooth muscle. *Blood Vessels*. 1981; 18(1-2):36-44.
80. Kennedy C & Sokoloff L. An adaptation of the nitrous oxide method to the study of the cerebral circulation in children; Normal values for cerebral blood flow and cerebral metabolic rate in childhood. *Journal of Clinical Investigation*. 1957; 36(7):1130.
81. Kety SS & Schmidt CF. The determination of cerebral blood flow in man by the use of nitrous oxide in low concentrations. *American Journal of Physiology*. 1945; 143:53-66.
82. Kety SS & Schmidt CF. The nitrous oxide method for the quantitative determination of cerebral blood flow in man: Theory, procedure and normal values. *Journal of Clinical Investigation*. 1948; 27:476-483.
83. Koh SX & Lee JK. S100B as a marker for brain damage and blood-brain barrier disruption following exercise. *Sports Medicine*. 2014; 44:369-385.
84. Kontos HA, Wei EP, Raper AJ, Patterson JL Jr. Local mechanism of CO₂ action of cat pial arterioles. *Stroke*. 1997; 8(2):226-9.

85. Kopelman PG. Obesity as a medical problem. *Nature*. 2000; 404:635–43.
86. Kuvin JT, Rämet ME, Patel AR, Pandian NG, Mendelsohn ME & Karas RH. A novel mechanism for the beneficial vascular effects of high-density lipoprotein cholesterol: Enhanced vasorelaxation and increased endothelial nitric oxide synthase expression. *American Heart Journal*. 2002; 144(1):165-172.
87. Lassen NA. Cerebral blood flow and oxygen consumption in man. *Physiological Reviews*. 1959; 39:183–238.
88. Lassen NA & Munck O. The cerebral blood flow in man determined by the use of radioactive krypton. *Acta Physiologica Scandinavica*. 1955; 33:30–49.
89. Lautenschlager NT, Cox K & Cyarto EV. The influence of exercise on brain aging and dementia. *Biochimica et Biophysica Acta*. 2012; 474–481.
90. Leung J, Kosinski PD, Croal PL & Kassner A. Developmental trajectories of cerebrovascular reactivity in healthy children and young adults assessed with magnetic resonance imaging. *The Journal of Physiology*. 2016; 594(10):2681-2689.
91. Liang J, Matheson BE, Kaye WH & Boutelle KN. Neurocognitive correlates of obesity and obesity-related behaviors in children and adolescents. *International Journal of Obesity*. 2014; 38(4):494-506.
92. Lin TW, Wang JN & Kan CD. Cerebral hyperperfusion syndrome after surgical repair of congenital supravulvular aortic stenosis. *Ann Thoracic Surgery*. 2015; 100:e51–e54.
93. Lucas SJ, Tzeng YC, Galvin SD, Thomas KN, Ogoh S & Ainslie PN. Influence of changes in blood pressure on cerebral perfusion and oxygenation. *Hypertension*. 2010; 55:698–705.
94. Lucas SJ, Cotter JD, Brassard P & Bailey DM. High-intensity interval exercise and cerebrovascular health: Curiosity, cause, and consequence. *Journal of Cerebral Blood Flow*

- & *Metabolism*. 2015; 35(6):902-911.
95. MacDougall JD, Tuxen DSDG, Sale DG, Moroz JR & Sutton JR. Arterial blood pressure response to heavy resistance exercise. *Journal of Applied Physiology*. 1985; 58(3):785-790.
96. Mackintosh KA, Niezen G & Eslambolchilar P. Mission possible: Using ubiquitous social goal sharing technology to promote physical activity in children. *Movement, Health and Exercise*. 2016; 5:1–14.
97. Madsen PL & Secher NH. Near-infrared oximetry of the brain. *Progress in Neurobiology*. 1999; 58:541–560.
98. Madsen PL, Sperling BK, Warming T, Schmidt JF, Secher NH, Wildschjødtz G, Holm S & Lassen NA. Middle cerebral artery blood velocity and cerebral blood flow and O₂ uptake during dynamic exercise. *Journal of Applied Physiology*. 1993; 74, 245–250.
99. Maggio P, Salinet ASM, Panerai RB & Robinson TG. Does hypercapnia-induced impairment of cerebral autoregulation affect neurovascular coupling? A functional TCD study. *Journal of Applied Physiology*. 2013; 115:491–497.
100. McManus AM, Ainslie PN, Green DJ, Simair RG, Smith K & Lewis N. Impact of prolonged sitting on vascular function in young girls. *Experimental Physiology*. 2015; 100(11):1379-1387.
101. McManus AM, Cheng CH, Leung MP, Yung TC & Macfarlane DJ. Improving aerobic power in primary school boys: a comparison of continuous and interval training. *Internation Journal of Sports Medicine*. 2005; 26:781–786.
102. McManus AM, Chu EY, Yu CC & Hu Y. How children move: Activity pattern characteristics in lean and obese Chinese children. *Journal of Obesity*. 2011.

103. McMurray RG, Harrell JS, Bangdiwala SI, Bradley CB, Deng S & Levine A. A school-based intervention can reduce body fat and blood pressure in young adolescents. *Journal of Adolescent Health*. 2002; 31(2):125-132.
104. Meyer AA, Kundt G, Lenschow U, Schuff-Werner P & Kienast W. Improvement of early vascular changes and cardiovascular risk factors in obese children after a six-month exercise program. *Journal of the American College of Cardiology*. 2006; 48(9):1865-1870.
105. Mintun MA, Fox PT & Raichle ME. A highly accurate method of localizing regions of neuronal activation in the human brain with positron emission tomography. *Journal of Cerebral Blood Flow and Metabolism*. 1989; 9:96–103.
106. Moore RD, Wu CT, Pontifex MB, O’Leary KC, Scudder MR, Raine LB, Johnson CR & Hillman CH. Aerobic fitness and intra-individual variability of neurocognition in preadolescent children. *Brain and Cognition*. 2013; 82(1):43-57.
107. Moraine JJ, Lamotte M, Berre J, Niset G, Leduc A & Naeije R. Relationship of middle cerebral artery blood flow velocity to intensity during dynamic exercise in normal subjects. *European Journal of Applied Physiology and Occupational Physiology*. 1993; 67:35–38.
108. Nguyen JC, Killcross AS & Jenkins TA. Obesity and cognitive decline: Role of inflammation and vascular changes. *Neuroinflammation and Behaviour*. 2015; 100.
109. Nourry C, Deruelle F, Guinhouya C, Baquet G, Fabre C, Bart F, Berthoin S & Mucci P. High-intensity intermittent running training improves pulmonary function and alters exercise breathing pattern in children. *European Journal of Applied Physiology*. 2005; 94(4):415-423.

110. Nybo L, Moller K, Pedersen BK, Nielsen B & Secher NH. Association between fatigue and failure to preserve cerebral energy turnover during prolonged exercise. *ActaPhysScand*. 2003; 179:67– 74.
111. Ogawa M, Fukuyama H, Harada K & Kimura J. Cerebral blood flow and metabolism in multiple system atrophy of the Shy-Drager syndrome type: A PET study. *Journal of Neurological Science*. 1998; 158:173–179.
112. Ogoh S & Ainslie PN. Cerebral blood flow during exercise: mechanisms of regulation. *Journal of Applied Physiology*. 2009; 107(5):1370-1380.
113. Ogoh S, Hayashi N, Inagaki M, Ainslie PN & Miyamoto T. Interaction between the ventilatory and cerebrovascular responses to hypo- and hypercapnia at rest and during exercise. *Journal of Physiology*. 2008; 586:4327–4338.
114. Ohuchi H, Kato Y, Tasato H, Arakaki Y & Kamiya T. Ventilatory response and arterial blood gases during exercise in children. *Pediatric Research*. 1999; 45:389-396.
115. Okazawa H, Yamauchi H, Toyoda H, Sugimoto K, Fujibayashi Y & Yonekura Y. Relationship between vasodilatation and cerebral blood flow increase in impaired hemodynamics: A PET study with the acetazolamide test in cerebrovascular disease. *Journal of Nuclear Medicine*. 2003; 44(12):1875-1883.
116. Olesen J, Paulson OB, Lassen NA. Regional cerebral blood flow in man determined by the initial slope of the clearance of intra-arterially injected ¹³³Xe. *Stroke*. 1971; 2:519– 540.
117. Onis MD, Onyango AW, Borghi E, Siyam A, Nishida C & Siekmann J. Development of a WHO growth reference for school-aged children and adolescents. *Bulletin of the World Health Organization*. 2007; 85(9):660-667.

118. Panerai RB, Evans DH & Naylor AR. Influence of arterial blood pressure on cerebrovascular reactivity. *Stroke*. 1999; 30:1293–1295.
119. Paulson OB, Hasselbalch SG, Rostrup EG, Knudsen M & Pelligrino D. Cerebral blood flow response to functional activation. *Journal of Cerebral Blood Flow and Metabolism*. 2011; 30:2–14.
120. Paulson OB, Strandgaard S & Edvinson L. Cerebral autoregulation. *Cerebrovascular Brain and Metabolism Reviews*. 1990; 2:161–191.
121. Pearson HA. Diseases of the blood. In: Behrman RE, Vaughn EC (eds) Textbook of Pediatrics. WB Saunders, Philadelphia, 1983; 1204-1257.
122. Poole DC & Jones AM. Oxygen uptake kinetics. *Comprehensive Physiology*. 2012.
123. Poulin MJ, Liang PJ, Robbins PA. Dynamics of the cerebral blood flow response to step changes in end-tidal PCO₂ and PO₂ in humans. *Journal of Applied Physiology*. 1996; 81(3):1084–95.
124. Querido JS & Sheel AW. Regulation of cerebral blood flow during exercise. *Sports Medicine*. 2007; 37(9):765-782.
125. Rasmussen AR, Wohlfahrt-Veje C, de Renzy-Martin KT, Hagen CP, Tinggaard J, Mouritsen A, Mieritz MG & Main KM. Validity of self-assessment of pubertal maturation. *Pediatrics*. 2015; 135(1):86-93.
126. Ratel S & Blazeovich AJ. Are Prepubertal Children Metabolically Comparable to Well-Trained Adult Endurance Athletes?. *Sports Medicine*. 2017; 1-9.
127. Ratel S, Duche P, Hennegrave A, Van Praagh E & Bedu M. Acid-base balance during repeated cycling sprints in boys and men. *Journal of Applied Physiology*. 2002; 135(2):479-

- 485.
128. Ratel S, Lazaar N, Williams CA, Bedu M & Duche P. Age differences in human skeletal muscle fatigue during high-intensity intermittent exercise. *Acta Paediatrica*. 2003; 92(11):1248-1254.
129. Regan RE, Fisher JA & Duffin J. Factors affecting the determination of cerebrovascular reactivity. *Brain and behavior*. 2014; 4(5):775-788.
130. Salmon JO, Ball K, Crawford D, Booth M, Telford A, Hume C, Jolley D & Worsley A. Reducing sedentary behaviour and increasing physical activity among 10-year-old children: Overview and process evaluation of the 'Switch-Play' intervention. *Health Promotion International*. 2005; 20(1):7-17.
131. Sato K, Fisher JP, Seifert T, Overgaard M, Secher NH & Ogoh S. Blood flow in internal carotid and vertebral arteries during orthostatic stress. *Experimental Physiology*. 2012; 97(12):1272-1280. 2012 (A).
132. Sato K, Ogoh S, Hirasawa A, Oue A, Sadamoto T. The distribution of blood flow in the carotid and vertebral arteries during dynamic exercise in humans. *Journal of Physiology*. 2011; 589: 2847–2856.
133. Sato K, Sadamoto T, Hirasawa A, Oue A, Subudhi AW, Miyazawa T et al. Differential blood flow responses to CO₂ in human internal and external carotid and vertebral arteries. *Journal of Physiology*. 2012; 590:3277–3290. 2012 (B).
134. Satterthwaite TD, Shinohara RT, Wolf DH, Hopson RD, Elliott MA, Vandekar SN, Ruparel K, Calkins ME, Roalf DR, Gennatas ED & Jackson C. Impact of puberty on the evolution of cerebral perfusion during adolescence. *Proceedings of the National Academy of Sciences*. 2014; 111(23):8643-8648.

135. Schönig M, Walter J & Scheel P. Estimation of cerebral blood flow through color duplex sonography of the carotid and vertebral arteries in healthy adults. *Stroke*. 1994; 25(1):17-22.
136. Seals DR, Victor RG. Regulation of muscle sympathetic nerve activity during exercise in humans. *Exercise and Sport Scientist Review*. 1991; 19:313–349.
137. Secher NH, Seifert T & Van Lieshout JJ. Cerebral blood flow and metabolism during exercise: Implications for fatigue. *Journal of Applied Physiology*. 2008; 104(1):306-314.
138. Seong J, Lieber BB & Wakhloo AK. Morphological age-dependent development of the human carotid bifurcation. *Journal of Biomechanics*. 2005; 38:453–465.
139. Serrador JM, Picot PA, Rutt BK, Shoemaker JK & Bondar RL. MRI measures of middle cerebral artery diameter in conscious humans during simulated orthostasis. *Stroke*. 2000; 31(7):1672-1678.
140. Seydel HG. The diameters of the cerebral arteries of the human fetus. *Anatomical Record*. 1964; 150:79–86.
141. Skilton MR & Celermajer DS. Endothelial dysfunction and arterial abnormalities in childhood obesity. *International Journal of Obesity*. 2006; 30(7):1041-1049.
142. Skoog I, Kalaria RN & Breteler MM. Vascular factors and Alzheimer disease. *Alzheimer Disease & Associated Disorders*. 1999; 13:S106-S114.
143. Skoog I, Nilsson L, Persson G, Lernfelt B, Landahl S, Palmertz B, Andreasson LA, Oden A & Svanborg A. 15-year longitudinal study of blood pressure and dementia. *The Lancet*. 1996; 347(9009):1141-1145.
144. Skow RJ, MacKay CM, Tymko MM, Willie CK, Smith KJ, Ainslie PN & Day TA. Differential cerebrovascular CO₂ reactivity in anterior and posterior cerebral circulations.

- Respiratory Physiology & Neurobiology*. 2013; 189(1):76-86.
145. Smith KJ & Ainslie PN. Regulation of cerebral blood flow and metabolism during exercise. Accepted in *Experimental Physiology*. 2017; doi: 10.1113/EP086249.
146. Smith KJ, MacLeod D, Willie CK, Lewis NCS, Hoiland RL, Ikeda K, Tymko MM, Donnelly J, Day TA, MacLeod N, Lucas SJE & Ainslie PN. Influence of high altitude on cerebral blood flow and fuel utilization during exercise and recovery. *Journal of Physiology*. 2014; 592:5507–5527.
147. Smith KJ, Wong LE, Eves ND, Koelwyn GJ, Smirl JD, Willie CK & Ainslie PN. Regional cerebral blood flow distribution during exercise: Influence of oxygen. *Respiratory Physiology & Neurobiology*. 2012; 184(1):97-105.
148. Strong WB, Malina RM, Blimkie CJ, Daniels SR, Dishman RK, Gutin B, Hergenroeder AC, Must A, Nixon PA, Pivarnik JM & Rowland T. Evidence based physical activity for school-age youth. *The Journal of Pediatrics*. 2005; 146(6):732-737.
149. Subudhi AW, Lorenz MC, Fulco CS & Roach RC. Cerebrovascular responses to incremental exercise during hypobaric hypoxia: Effect of oxygenation on maximal performance. *American Journal of Physiology: Heart and Circulatory Physiology*. 2008; 294:H164–H171.
150. Sullivan GW, Sarembock IJ & Linden J. The role of inflammation in vascular diseases. *Journal of Leukocyte Biology*. 2000; 67(5):591-602.
151. Tandon PS, Zhou C, Lozano P & Christakis DA. Preschoolers' total daily screen time at home and by type of child care. *The Journal of Pediatrics*. 2011; 158(2):297-300.
152. Timmons BW & Bar-Or O. RPE during prolonged cycling with and without carbohydrate ingestion in boys and men. *Medicine and Science in Sports and Exercise*. 2003; 35:1901–

1907.

153. Tinken TM, Thijssen DH, Hopkins N, Black MA, Dawson EA, Minson CT, Newcomer SC, Laughlin MH, Cable NT & Green DJ. Impact of shear rate modulation on vascular function in humans. *Hypertension*. 2009; 54(2):278-85.
154. Tolfrey K & Armstrong N. Child-adult differences in whole blood lactate responses to incremental treadmill exercise. *British journal of sports medicine*. 1995; 29(3):196-199.
155. Tontisirin N, Muangman SL, Suz P, Pihoker C, Fisk D, Moore A, Lam AM & Vavilala MS. Early childhood gender differences in anterior and posterior cerebral blood flow velocity and autoregulation. *Pediatrics*. 2007; 119(3):e610-e615.
156. Townsend JR, Stout JR, Morton AB, Jajtner AR, Gonzalez AM, Wells AJ, Mangine GT, McCormack WP, Emerson NS, Robinson EH & Hoffman JR. Excess post-exercise oxygen consumption (EPOC) following multiple effort sprint and moderate aerobic exercise. *Kinesiology*. 2013; 45(1):16-21.
157. Tremblay MS, LeBlanc AG, Janssen I, Kho ME, Hicks A, Murumets K, Colley RC & Duggan M. Canadian sedentary behaviour guidelines for children and youth. *Applied Physiology, Nutrition, and Metabolism*. 2011; 36(1) 59-64. 2011 (A).
158. Tremblay MS, LeBlanc AG, Kho ME, Saunders TJ, Larouche R, Colley RC, Goldfield G & Gorber SC. Systematic review of sedentary behaviour and health indicators in school-aged children and youth. *International Journal of Behavioral Nutrition and Physical Activity*. 2011; 8(1):1. 2001 (B).
159. Tremblay MS & Willms JD. Is the Canadian childhood obesity epidemic related to physical inactivity?. *International Journal of Obesity*. 2003; 27(9):1100-1105.
160. Trost SG, Kerr LM, Ward DS & Pate RR. Physical activity and determinants of physical

- activity in obese and non-obese children. *International Journal of Obesity*. 2001; 25(6):822.
161. Tzeng YC & Ainslie PN. Blood pressure regulation IX: Cerebral autoregulation under blood pressure challenges. *European Journal of Applied Physiology*. 2014; 114:545–559.
162. Udomphorn Y, Armstead WM & Vavilala MS. Cerebral blood flow and autoregulation after pediatric traumatic brain injury. *Pediatric Neurology*. 2008; 38:225-234.
163. van Mook WN, Rennenberg RJ, Schurink GW, van Oostenbrugge RJ, Mess WH, Hofman PA & de Leeuw PW. Cerebral hyperperfusion syndrome. *The Lancet Neurology*. 2005; 4(12):877-88.
164. Van Praagh E & Doré E. Short-term muscle power during growth and maturation. *Sports Medicine*. 2002; 32(11):701-728.
165. Weston KS, Wisløff U & Coombes JS. High-intensity interval training in patients with lifestyle-induced cardiometabolic disease: A systematic review and meta-analysis. *British Journal of Sports Medicine*. 2014; 48(16):1227-1234.
166. Vavilala MS, Bowen A, Lam AM, Uffman JC, Powell J, Winn HR, & Rivara FP. Blood pressure and outcome after severe pediatric traumatic brain injury. *Journal of Trauma and Acute Care Surgery*. 2003; 55:1039-1044. 2003 (A).
167. Vavilala MS, Kincaid MS, Muangman SL, Suz P, Rozet I & Lam AM. Gender differences in cerebral blood flow velocity and autoregulation between the anterior and posterior circulations in healthy children. *Pediatric Research*. 2005; 58(3):574-578.
168. Vavilala MS, Lee LA & Lam AM. The lower limit of cerebral autoregulation in children during sevoflurane anesthesia. *Journal of Neurosurgical Anesthesiology*. 2003; 15(4):307-312. 2003 (B).
169. Verlhac S. Transcranial Doppler in children. *Pediatric Radiology*. 2011; 41(1):153-165.

170. Williams CA, Ratel S & Armstrong N. Achievement of peak during a 90-s maximal intensity cycle sprint in adolescents. *Canadian Journal of Applied Physiology*. 2005; 30(2):157-171.
171. Willie CK, Colino FL, Bailey DM, Tzeng YC, Binsted G, Jones LW, Haykowsky MJ, Bellapart J, Ogoh S, Smith KJ & Smirl JD. Utility of transcranial Doppler ultrasound for the integrative assessment of cerebrovascular function. *Journal of Neuroscience Methods*. 2011; 196(2):221-237.
172. Willie CK, Macleod DB, Shaw AD, Smith KJ, Tzeng YC, Eves ND, Ikeda K, Graham J, Lewis NC, Day TA & Ainslie PN. Regional brain blood flow in man during acute changes in arterial blood gases. *The Journal of Physiology*. 2012; 590(14):3261-75.
173. Willie CK, Tzeng YC, Fisher JA & Ainslie PN. Integrative regulation of human brain blood flow. *Journal of Physiology*. 2014; 592:841–859.
174. Wintermark M, Lepori D, Cotting J, Roulet E, van Melle G, Meuli R, Maeder P, Regli L, Verdun FR, Deonna T, Schnyder P & Gudinchet F. Brain perfusion in children: Evolution with age assessed by quantitative perfusion computed tomography. *Pediatrics*. 2004; 113:1642–1652.
175. Wintermark M, Sesay M, Barbier E, Borbely K, Dillon WP, Eastwood JD, Glenn TC, Grandin CB, Pedraza S, Soustiel J-F, Nariai T, Zaharchuk G, Caille J-M, Dousset V & Yonas H. Comparative overview of brain perfusion imaging techniques. *Stroke*. 2005; 36:e83–99.
176. Zhang R, Zuckerman JH, Giller CA, Levine BD. Transfer function analysis of dynamic cerebral autoregulation in humans. *American Journal of Physiology*. 1998; 274:H233.